ASIAN JOURNAL OF PHARMACEUTICAL AND CLINICAL RESEARCH



Vol 6, Issue 4, 2013 ISSN - 0974-2441

Research Article

IN VIVO UTILIZATION OF SEAFOOD PROCESSING WASTES FOR CULTIVATION OF THE MEDICINAL MUSHROOM (GANODERMA LUCIDUM) USING AGRO-INDUSTRIAL WASTE

S.SUBBU LAKSHMI

Department of Microbiology, Kamaraj college, Thoothukudi, Tamil Nadu, India. Email: daisyrani11@yahoo.com

Received: 19 July 2013, Revised and Accepted: 11 August 2013

ABSTRACT

Objective: To examine the utilization of seafood processing wastes for artificial cultivation of medicinal mushroom and composting in laboratory condition

Method: The selected agro-industrial wastes (e.g., coir pith, woodchips, sugarcane bagasse) were mixed with fishery waste in specific ratio (1:1). The substrates which are not mixed with fishery waste are regarded as control. All the above materials (1:1,control) were allowed to decompose about 15 days. The composted materials were placed in heat resistant transparent polyethylene bags. Each sterile bag was then aseptically inoculated with *G.lucidum*. The bags were then incubated under ambient temperature and controlled humidity.

Results: The maximum biological yield per bed was obtained with sugarcane bagasse control bed 64.78 g/bed. The lowest yield was observed in woodchips (1:1) 4.6 g/bed. Based on mass obtained of *G.lucidum* the best substrates were ordered of like woodchips>coir>sugarcane. Conclusion: The scope of this work is sea food wastes could be used to cultivate a medicinal mushroom while at the same time promoting environmental sustainability.

Keywords: Mushroom, fishery waste, solid substrates, biological yield.

INTRODUCTION

Fish waste can also be utilized in the production of organic fertilizers and composts, which have significant benefits over chemical based products. Fish waste can result in water quality problems at marinas with large numbers of fish landings or at marinas that have limited fish landing but poor flushing. The amount of fish disposed of in to a small area such as a marina can exceed that existing naturally in the water at any one time. Fish waste decomposes, which requires oxygen. In sufficient quantity, disposal of fish waste can thus be a cause of dissolved oxygen depression as well as odor problems [8, 18]. The catching and processing of fish generates a significant amount of waste. Of a total UK fish and shellfish resource of 851,984 tones, it is estimated that 43% (359,964 tones) ends up as products for human consumption and the remainder (492,020 tones) is classed as waste. The majority of waste is produced in the on-shore processing sector (35% of the resource) whereas discards and processing waste at sea produce smaller quantities (17% and 5%respectively of the resource). The fish industry developed around fishing ports when landings were plentiful and there was little concern about environmental impacts. Fish waste is rich in valuable minerals, enzymes, pigments and flavors that are required by many including food, agriculture, aquaculture industries pharmaceuticals. Possible alternatives include silage production, which has potential in livestock feeding. Basidiomycetous fungi (mushrooms) can be defined as "macro fungi" with distinctive fruiting bodies that are large enough to be seen by the naked eye and to be picked by hand. It is estimated that there are approximately 1.5 million species of mushrooms in the world of which approximately 70,000 species are described. About 10,000 of the known species belong to the macro fungi of which about 5,000 species are edible and over 1,800 species are considered to have medicinal properties[3]. The fruiting body, mycelia and spores of G. lucidum contain approximately 400 different bio active compounds which mainly include triterpenoids, polysaccharides, nucleotides, sterols, steroids, fatty acids, proteins/peptides & trace [19,12,17,16,26].A successful artificial cultivation of Ganoderma lucidum has been reported on most broad-leaf hardwood trees and commonly used species include oak, pecan, elder, choke cherry, and plum[4,6].It is normally cultivated on solid substrates or other lignocelluloses materials such as straw, sawdust and log [24,25]. Ganoderma species can be cultivated on the sawdust which

may originate from different kinds of trees[30,20].Hardwood sawdust is the basic substrate for the cultivation of most medicinal mushrooms [5] and the growth and development of them varied from one tree species to another[1]. Hardwood sawdust and wheat bran mixture may be used in production of Ganoderma lucidum [21,35]. Ganoderma lucidum has been reported to have a number of pharmacological effects including immunomodulating, atherosclerotic, anti-inflammatory, analgesic, chemopreventive, antitumor, radio protective, sleep promoting, antibacterial, antiviral (including anti -HIV), hypolipidemic, anti fibrotic, hepatoprotective, diabetic, anti-oxidative and radial scavenging, anti- aging , hypoglycemic and anti-ulcer properties.[34,4,15,13,19,12,32,33,26].Recently more studies demonstrated that G. lucidum contained anti bacterial constituents that are able to inhibit gram-positive and / or gram negative bacteria [34,25,13,19,11,26,27]. Ganoderma lucidum may also have hypoglycemic activity, anti-inflammatory effects, and cytotoxicity toward hepatoma cells[9,2]. Bioactive polysaccharides in mushrooms can often be extracted from mycelia of the species without waiting for a full fruiting body to develop [28,14]. The aim of the study is to reduce environmental pollution while producing medicinal mushroom.

MATERIALS AND METHODS

Tissue Culture Technique

A large healthy and fresh mushroom *G.lucidum* collected from the Chozhavanthan at Madurai district, Tamil Nadu and India. It should be cleaned with 75% alcohol. The mushroom should be split in half by hand longitudinally and some inside tissue taken from the upper part of the stripe. It should be placed centrally on the surface of the sterilized Rose Bengal agar with a sterilized needle and kept at room temperatures (28°C \pm 2) for 8 days. Within two or three days some white, delicate mycelia will be produced from the small piece of the tissue. About eight days later the mycelium will grow rapidly and cover the surface of the agar medium. Then it will be ready to transfer to spawn substrate to make spawn. This spawn is used as inoculums for cultivation of mushroom.

Spawn production

The sorghum was cleaned manually to remove inert matter, stubble and debris. The cleaned grain was soaked in 0.5% Cu SO $_4^{\rm for}$ 10 min and the soaked grain was thoroughly washed and soaked in tap water for 2 hours. Thereafter, the soaked grain was drained and the excess water removed. Spawn containers should be made out of heat resistant material: mostly glass and polypropylene (PP). About 250 g of grain medium was filled in to polypropylene bag. It was sealed by

covered by a piece of paper by tying a rubber band around the neck. Autoclaved at 121°C, 15 psi, for 30 min and the sterilized bag was allowed to cool for 24 hours. Then immediately inoculated with mycelial culture of *G.lucidum* maintained on Rose Bengal agar.

using cotton wool plugged with poly vinyl chloride pipe ring and

Fish Compost preparation

Fishery wastes were collected from the food processing unit at Thoothukudi which contains head, tail, shells, intestine, fins, dead fishes and so on. The wastes obtained were brought to the laboratory and the un compostable materials such as shells and large bones were removed and the remaining wastes were cut into small uniform size pieces. Disease-free agro-industrial wastes were collected from Thoothukudi district which were cut in to small pieces (2-3cm) and sun-dried in order to achieve proper drying. The selected agro-industrial wastes were mixed with fishery wastes in specific ratio 1:1(500:500) which are not mixed with fishery wastes are regarded as control. All the above materials (1:1,control) were heaped in a separate plastic container and allowed to decompose for about 15 days. During the decomposition, water was sprayed over the materials with turning and restacking for every day to produce homogenize composed. Finally, the Composted materials were used for mushroom cultivation.

Mushroom bed preparation

Polypropylene bags of 14×18; 14×12 size were taken and the bottom of the bag was tied with a thread to provide a flat circular bottom of the mushroom beds. Composted materials were placed in polypropylene bags were sterilized at 121°C for 2 hrs, after cooling, the spawning was done 3-5 layers. The neck of the bag was prepared by using heat resistant PVC tube and plugged with cotton, then transferred to the culture room at 25-32°C temperature, 85-95% relative humidity. Water was sprayed 3-4 times per day and proper aeration was maintained in culture house to develop the fruiting bodies. After 15-20 days, the pinheads of G.lucidum emerged from the mouth of polypropylene bags. The cap formation of Ganoderma lucidum initiated in 2-3 days after opening the mouths of bags. Fruiting bodies were harvested when the caps become completely red and the white margin disappeared [22]. Experimental design was a Completely Randomized Block with three replicates.Data on period of after completion of mycelium running, biological yield, days of first harvest, number of fruiting bodies, length, diameter, biological efficiency, moisture content of mushroom and dry yield were recorded. The biological efficiency (BE) percentage [fresh weight of harvested mushrooms/dry matter content of the substrate] x100 [23], productivity (P)percentage (Mushroom dry weight / Compost initial dry weight) x 100 and percentage of consumption (Before cultivation -After cultivation) /100 was calculated. Compared the means with one-way ANOVA. The difference was statistically significant when p < 0.05.

Determination of moisture content

The moisture content of the mushroom was estimated by drying in an oven at 80°C for three consecutive days. It was cooled in a desiccator and weighed. The moisture content was calculated by the following formula.

Moisture content (%) = Fresh weight – Dry weight

Fresh weight – Bry weight

RESULT AND DISCUSSION

Development of fruiting body

The reproductive growth of *Ganoderma lucidum* varied from one substrate to another. The first pin head formation within 16 days, was recorded on woodchips control. The highest time required for pin head formation (47.66 days) was recorded in sugar cane waste 1:1. The lowest period of fruiting body formation (64.00 days) was observed in woodchips control .The highest period of fruiting body formation (102.33 days) was observed in sugar cane waste 1:1. [10] demonstrated that Polypore mushroom was found to be rich in protein, fiber and cellulose. Polypore mushroom is found to grow on rotting hard wood logs. So, attempt was made to cultivate the species in laboratory condition. [31] also showed that *G.lucidum* grown on a wide variety of dead or dying trees, commercial production in the wood.

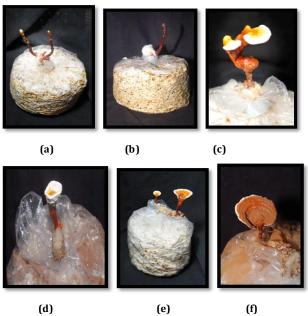


Fig.1:Growth of Ganoderma lucidum in (a)Woodchips control(b) Woodchips 1:1 (c) Coir pith control (d) Coir pith 1:1 (e) sugarcane control (f) sugarcane 1:1

Size of fruiting body

Length and diameter of fruiting bodies produced on agro-industrial wastes and fishery wastes ranged from 5.80 to 12.00 cm and 1.16 to 9.40 cm respectively (Fig 2). The highest length of fruiting body (12.00 cm) was recorded on coir pith control. While highest diameter of fruiting body (9.40 cm) was found on coir pith control. The least length of fruiting body in sugarcane waste (1:1) (5.80 cm) and diameter of fruiting body (1.16 cm) were recorded in woodchips (1:1) respectively.[31] reported that *G.lucidum* is traditionally cultivated in solid cultures. Solid cultures are used to obtain basidiocarps for tonic or tea. *G.lucidum* mycelium or spawn have normally been produced in solid cultures using substrates such as grain, sawdust or wood. The advantage of solid stage fermentation (SSF) over other technique is that a concentrated product can be obtained from a cheap substrate such as an agricultural residue with little pre-treatment or enrichment.

X 100

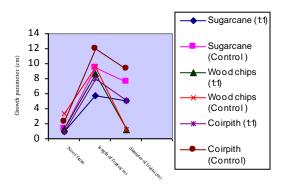
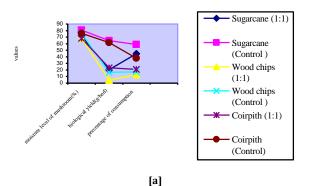


Fig.2: Effect of agro-industrial wastes on the development and size of fruiting bodies of *G.lucidum*

Biological yield, dry yield and biological efficiency

Significant difference was observed in biological yield, dry yield and biological efficiency of Ganoderma mushroom on agro-industrial wastes (Fig 3a and Fig 3b). The highest biological yield (64.78g/bed) was obtained from sugarcane waste control (Fig 1e) followed by coir pith control (61.73g/bed) (Fig 1c), coir pith 1:1 (23.15 g/bed) (Fig 1d), sugarcane waste 1:1 (20.33g/bed) (Fig 1f) and woodchips control (16.7g/bed) (Fig 1a).woodchips 1:1 gave poorest yield (4.6g/bed) (Fig 1b).Dry yield was observed in coir pith control(14.73g/bed) followed by Sugarcane (12.25g/bed), Coirpith1:1(7.45g/bed), Sugarcane 1:1 (5.70g/bed), Woodchips Control (4.0g/bed) and Wood chips 1:1 (1.49g/bed). The biological efficiency was highest (12.95%) sugarcane waste control in which was followed by coir pith control (12.34%), coir pith 1:1(4.63%) ,sugarcane waste 1:1 (4.066%)and woodchips control (3.34%) while lowest biological efficiency (0.92%) was observed in woodchips 1:1.[29] also suggested that various substrates for G.lucidum cultivation have been investigated as a supplement for substrate mixer. A successful artificial cultivation has been reported on solid substrates, utilizing saw dust and agricultural waste as the main components. The suitability of rice bran, rice husks, coconut fiber, peanut hulls, corn, sorghum and sugarcane bagasse as supplements for the substrate mixture and rice bran, ground corn and ground sorghum were found good supplements compared to some other agricultural residues such as rice husk, coconut fiber, peanut hull and sugar cane bagasse. Fig 3 shows effect of agroindustrial waste on percentage of food consumption, moisture level and productivity of Ganoderma lucidum.



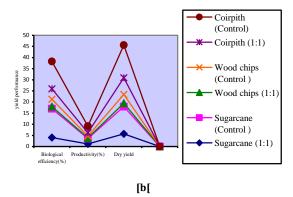


Fig.3: Effect of agro-industrial wastes on yield of *Ganoderma* lucidum (a)moisture level of mushroom, biological yield and percentage of consumption (b) biological efficiency, productivity and dry yield.

CONCLUSION

This research is to looking at the possible use of fish waste and agroindustrial wastes for mushroom cultivation, which would provide medicinal food and encourage the biological conversion processes of agro-industrial wastes. Composting fish waste is a relatively new, practical and an environmental sound alternative to disposing of fish waste. It is economical, fairly odorless and a biologically beneficial practice for seafood operations. The study has shown that composting is safe, Practical and a cost effective solution for the seafood industry.

REFERENCE

- S.M. Ayodele, E.O. Akpaja and F. Anyiador. Evaluation of yield of *Lentinus squarrosulus* (Mont.) Singer on selected economic tree species. *Pak. J. Biol. Sci.* 10(23) 4283-4286 (2007).
- X.F. Bao, C.Q. Liu, J. Fang and X.Y. Li. Structural and immune logical studies of a major polysaccharide from spores of Ganoderma lucidum (Fr.) Karst. Carbohydr Res 332:67-74 (2001).
- 3. S.T. Chang. Ganoderma The leader in production and technology of mushroom nutriceuticals. *In Proc 6th Int symp Recent Adv. Ganoderma lucidum* Res (Ed. B.K. Kim, I.H. Kim and Y.s. Kim) pp 43-52. The pharmaceutical Society of Korea, Seoul, Korea (1995).
- 4. S.T. Chang and J.A. Buswell. *Ganoderma lucidum*(Curt.:Fr.)P.Karst.(Aphyllophoromyceti-deae)預 mushrooming medicinal mushroom. *Int.J.Med.Mushrooms*,1 (2),139 · 46 Mushrooms 2004,6 (1) (1999).
- A.W. Chen. Cultivation of the medicinal mushroom Ganoderma lucidum (Curt.:Fr.) P. Karst. (Reishi) in North America. Int. J. Med. Mushroom 1: 263-282 (1999).
- K.L. Chen and D.M. Chao. Ling Zhi (Ganoderma species). In: Chinese Medicinal Mycology. (Ed.) K. T. Hsu, United Press of Beijing Medical University and Chinese United Medical University. Beijing, China pp. 496-517 (1997).
- A.W. Chen. A practical guide for synthetic-log cultivation of medicinal mushroom *Grifola frondosa* (Dich. Fr) S.Fr.Gray (Maitake). *Int. J. Med. Mush* 1:153-168 (1999).
- DNREC. State of Delaware Marina Guidebook. Delaware Department of Natural Resources and Environmental Control, Dover, DE (1990).
- S.K. Eo, Y.S. Kim, C.K. Lee and S.S. Han. Antiviral activities of various water and methanol soluble substances isolated from *Ganoderma lucidum J.Ethnopharmacol* 68,129 · 36 (1999b).
- Fauzia Hafiz, Fauzia Begum, Abu Md Imtiaz Hossain, Sahana Parveen, Majeda Begum, Zaibun Nessa, Motahar Hossain, John Liton Munshi. Ganoderma resinaceum Bangladesh Journal of Scientific and Industrial Research Vol 42, No 2 (2007).

- Y. Gao, S.H. Zhou, M. Huang and A. Xu. Anti-bacterial and antiviral value of the genus *Ganoderma* P.Karst.species (Aphyllophoro-mycetideae): a review. *Int. J. Med.Mushrooms* 5 (3), 235 46 (2003).
- 12. Y.H. Gao and S.F. Zhou. The immune modulating effects of *Ganoderma lucidum. Int J Med Mush* 4: 1-11(2002).
- C.H. Hobbs. Medicinal Mushrooms: An Exploration of Tradition, Healing, and Culture, 2nd Ed.; Botanica Press, Inc.: Santa Cruz, CA, USA (1995).
- 14. N. Hatvani. Antimicrobial effect of the culture fluid of *Lentinus edodes* mycelium grown in submerged liquid culture. *Int. J. Antimicrob. Agents* 17:71–74 (2001).
- S.C. Jong and J.M. Birmingham. Medicinal benefits of the mushroom *Ganoderma*. Adv. Appl. Microbiol 37:101-134 (1992).
- H.W. Kim, B.K. Kim. Biomedical triterpenoids of Ganoderma lucidum(Curt.:Fr.)P.Karst. (Aphyllophoromycetideae). Int. J. Med. Mushrooms 1 (2), 121 • 38 (1999).
- T. Mizuno. Reishi, Ganoderma lucidum and Ganoderma tsugae: bioactive substances and medicinal effects. Food Rev. Int. 11 (1), 151 · 66 (1995).
- W.G. McDougal, R.S. Mustain, L.S. Slotta and J.M. Milbrat. Marina Flushing and Sedimentation. In *Proceedings of a Specialty Conference on Innovations in Port Engineering and Development in the 1990's*, American Society of Civil Engineers pp. 323-332 (1986).
- D.J. McKenna, K. Jones, K. Hughes. Reishi Botanical Medicines. The Desk reference for Major Herbal Supplements ,2nd Ed.;The Haworth Herbal Press:New York,London, Oxford 825 • 55 (2002).
- P. Olei. Mushroom Cultivation. Backhuys Publishers, Leyden, The Netherlands p. 429(2003).
- A. Peksen and G. Yakupoglu. Tea waste as a supplement for the cultivation of *Ganoderma lucidum*. World J. Microbiol. Biotechnol 25: 611-618 (2009).
- D.J. Royse. Specialty Mushrooms. ASHS Press, Arlington, VA pp. 464-475 (1996).
- D.J. Royse. Effect of spawn run time and substrate nutrition on yield and size of the shiitake mushroom. *Mycologia* 77:756-762 (1985).
- H. Riu, G. Roig and J. Sancho. Production of carpophores of Lentinus edodes and Ganoderma lucidum grown on cork residues. Microbiologia. SEM. 13: 97-103 (1997).

- P. Stamets. Growing of Gourmet and Medicinal Mushrooms.
 3rd Edn, Ten Speed Press, Berkeley, California, USA. p. 574 (2000).
- 26. J. Smith, N. Rowan and R. Sullivan.Medicinal Mushrooms.Their Therapeutic Properties and Current Medical Usage with Special Emphasis on Cancer Treatment;Special Report Commis- sioned by Cancer Research UK;The University of Strathclyde in Glasgow 256 (2002).
- I. Suay ,F. Arenal , F.Asenio, A. Basiilio, M.Cabello and M.T. Diez. Screening of basid-iomycetes for antimicrobial activities. *Antonie van Leeuwenhoek*. 78: 129-139 (2000).
- C.H. Song, Y.J. Jeon, B.K. Yang, K.S. Ra and H.I. Kim. Anticomplementary activity of endo-polymers produced from submerged mycelial culture of higher fungi with particular reference to *Lentinus edodes*. *Biotechnol. Lett.* 20:741–744 (1998).
- S. Triratana, S. Thaithatgoon and M. Gawgla. Cultivation of *Ganoderma lucidum* in sawdust bags. ISMS Vol. 13, Part 2, Article 20 (Article abstract) (1991).
- 30. S.P. Wasser. Reishi or Ling Zhi (Ganoderma lucidum). Encyclopedia of Dietary Supplements. pp. 603-622 (2005).
- R. Wagner, D.A. Mitchell, G.L. Sassaki, M.A. L.A. Amazonas and M. Berovic. Current techniques for the cultivation of Ganoderma lucidum for the production of biomass, ganoderic acid and polysaccharides. Food Technol. Biotechnol. 41(4):371-382 (2003).
- 32. S.P. Wasser and A.L. Weis. Medicinal properties of substances occurring in higher Basidiomycetes mushrooms: current perspectives (review).*Int.J. Med.Mushrooms* 1 (1),31 2 (1999).
- 33. S.P. Wasser. Medicinal mushrooms as a source of antitumor and immune modulating polysaccharides. *Appl. Microbiol. Biotechnol* 60, 258 74 (2002).
- 34. P.S. Wasser, L.A. Weis and C. Chem. Medicinal mushroom *Ganoderma lucidum-reishi* mushroom. *In: E. Nero (ed)*. San Antonio-Haifa-Kiev (1997).
- F.C. Yang, C. Hsieh and H.M. Chen .Use of stillage grain from a rice-spirit distillery in the solid state fermentation of Ganoderma lucidum. Proc. Biochemistry 39(1): 21-26 (2003).