



## DEVELOPMENT AND VALIDATION OF HPTLC METHOD FOR THE DETERMINATION OF $\beta$ - BOSWELLIC ACID FROM *BOSWELLIA SERRATA* ROXB. (EXUDATE)

PAWAR R.K. <sup>1</sup>, SHARMA SHIVANI <sup>2</sup>, SINGH K.C. <sup>2</sup> AND SHARMA RAJEEV KR. <sup>3</sup>

<sup>1,3</sup> Pharmacopoeial Laboratory for Indian Medicine, Ghaziabad-201 002 (U.P.), <sup>2</sup> Department of chemistry, R.S.S. (P.G.) College, Pilkhuwa, Ghaziabad (U.P.), India Email: pawarplim@gmail.com

### ABSTRACT

A simple, rapid, selective and quantitative HPTLC method has been developed for determination of  $\beta$ -Boswellic acid in different samples of *Boswellia serrata* Roxb. exudate. The n-hexane extract of *Boswellia serrata* Roxb. (exudate) samples were applied on TLC Aluminium plate pre coated with Silica gel 60 GF<sub>254</sub> and developed using Toluene : Ethyl acetate: Formic acid (5:4.5:0.5) v/v as a mobile phase. The plate was sprayed (derivatized) with Anisaldehyde- Sulphuric Acid reagent followed by heating at 110°C for 10 minutes and detection and quantification were carried out densitometrically using an UV detector at wavelength of 530 nm. Content of marker compound in the samples were found similar.

**Keywords:**  $\beta$ -Boswellic acid, *Boswellia serrata* Roxb., Kundru exudate, Shallaki, HPTLC.

### INTRODUCTION

*Boswellia serrata* Roxb. Ex Colebr. Syn. *Boswellia serrata* var. *glabra* (Roxb) Bennett, *Boswellia glabra* Roxb. (Family- Burseraceae) is well known as Kundru or Shallaki and distributed in dry forests from Punjab to West Bengal and in Peninsular India. Common at the foot of Western Himalaya, in Rajasthan, Gujrat, Maharashtra, Madhya Pradesh, Bihar Orissa Andhra Pradesh and further south in the Peninsula. Reported to be threatened in North Eastern Region of India. It is a medium size to large sized, deciduous tree, upto 18 m in height, an evergreen and spiny tree. Leaves are alternate, imparipinnate and crowded towards the end of branches; leaflets 17-31, opposite, sessile, ovate or ovate-lanceolate, crenate and pubescent.

Flowers are small and white, in axillary racemes or panicles. Drupes the fruits are about 1.2 cm long and trigonous, splitting into 3 valves and subtended by the woody disk. Seeds are compressed and pendulous. Flowering is during March-April and fruiting in winter season. The oleo-resin exudates out during winter and gets deposited on the various parts of tree trunk. The oleo-resin is secreted in the schizogenous duct in the bark which are scattered just below the bast fibres.

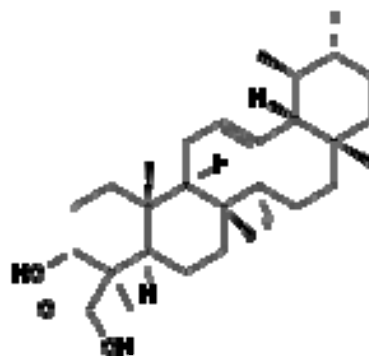
The oleo-resin exudes as colourless semi-fluid liquid which gradually becomes whitish to golden yellow and solidifies slowly with time. Sometimes, it is reddish brown, greenish yellow or dull yellow to orange in colour. Oleo-gum resin exudates and bark are used in medicine. The bark is sweet, acrid, cooling and tonic. It is good for asthma, dysentery, ulcers, haemorrhoids and skin diseases. Gum-resin exudates obtained from the plant is sweet, bitter, astringent and commercially known as Indian oilbanum or Indian frankincense or Sallai guggul.

It is used as antipyretic, expectorant, diuretic, choleric, antiseptic, antidiarrhetic, diaphoretic, stomachic and urethrorrhea, orchopathy, bronchitis, asthma, cough, skin diseases, ulcers, tumours, cystic breast, chronic laryngitis, jaundice and arthritis. It shows anti-inflammatory and anti-arthritis activity have been mainly attributed to a component in the resin containing  $\beta$ -Boswellic acid and is used for rheumatic patients [1-9].

In addition to use for arthritis this gummy resin is also mentioned in traditional Ayurvedic texts as a remedy for diarrhea, dysentery, ringworm, boils, fevers (antipyretic) skin and blood diseases, cardiovascular diseases, mouth sores, vaginal discharges, hair loss, jaundice, hemorrhoids, syphilitic diseases, irregular menses and to stimulate the liver. Modern medicine and pharmacology point to *Boswellia serrata*'s use as an anti arthritic, anti-inflammatory, anti-hyperlipidemic (control blood lipids), antiatherosclerotic (anti-coronary plaque), analgesic (pain-reliever) and hepatoprotective (protects the liver). [10-14]

The gum resin contains a mixture of triterpene acids known as Boswellic acid ( $\alpha, \beta, \gamma$  boswellic acid) acetyl-  $\beta$  boswellic acid, 11-keto-  $\beta$ -boswellic acid, acetyl-11-keto-  $\beta$ -boswellic acid and their derivatives [10]. Volatile oil contains  $\alpha$ -thujene,  $\alpha$ -phellandrene,  $\beta$ -phellandrene,  $\alpha$ -terpineol, d-limonene, myrcene,  $\alpha$ -terpene, *p*-cymene [11]; a diterpene alcohol serratalol and four tetracyclic triterpene acids 3-  $\alpha$ - acetoxytirucall -8,24-dien-21-oic acid, 3-ketotirucall-8, 24-dien-21-oic acid, 3-  $\alpha$ - hydroxytirucall -8,24-dien-oic acid, 3-  $\beta$ - hydroxytirucall -8,24-dien-21-oic acid. [1-14, 18].

It is also found to contain arabinose, rhamnose, glucose, galactose, fructose, idose, galacturonic acid and  $\beta$  sitosterol isolated from gum [12-17]. Essential oil from gum gave phenol-o-cresol, m-cresol, p-cresol, thymol, and carvacrol and carboxylic acid-  $\alpha$ -campholenic acid, 2,2,4-trimethylcyclopent-3-en-1-yl acetic acid and campholytic acid [2].



Structure of  $\beta$ -boswellic acid

The research has implicated [16] a beneficial role for the resin in the treatment of osteoarthritis, soft tissue rheumatism, low back pain, gout and rheumatoid arthritis. A creeping crippling disease causing great physical suffering, it is possible to alleviate physical pain, increase movement (mobility) and prevent further tissue injury through proper treatment. Treatment with *Boswellia serrata*, on the other hand Boswellic acid [17] "significantly reduced the infiltration of leucocytes into the knee joint" in turn significantly reducing inflammation causing immune white blood-cell response.

So that Boswellic acid is an active constituent and used as a marker. Literature survey reveals that the TLC and HPTLC methods are reported but no method as yet is reported for the determination of  $\beta$ -Boswellic acid in *Boswellia serrata* Roxb. exudates.

With increasing demand for herbal products in medicines and cosmetics there is an urgent need for standardization. So the aim of

the work is to develop a simple, rapid, selective and cost effective HPTLC method for the determination of  $\beta$ -Boswellic acid in *Boswellia serrata* exudate.

## MATERIAL AND METHOD

### Plant material

The Kundru exudate was procured from the Local Market, Ghaziabad. It was identified and authenticated by the Botanists of Pharmacopoeial Laboratory for Indian Medicine, Ghaziabad. One genuine sample also taken from the Museum of Pharmacopoeial Laboratory for Indian Medicine, Ghaziabad.

### Equipment

A Cammag (Switzerland) HPTLC system equipped with a sample applicator Linomat V, Twin trough glass Chamber (20x10 cm<sup>2</sup>) with SS lid, TLC Scanner III, Reprostar III and Wincats an integrated Software 4.02 (Switzerland), Rotavapour.

### Chemical & reagents

Analytical grade; Alcohol, Toluene, ethyl acetate, Formic acid, Chloroform, Methanol, Anisaldehyde, Sulphuric acid and n-Hexane were used; obtained from S.D. Fine Chem. Ltd. (Mumbai, India). TLC Aluminium pre coated plate with Silica gel 60 GF<sub>254</sub> (20x10 cm<sup>2</sup>; 0.2 mm thick) used were obtained from E. Merck Ltd. (Mumbai, India). Reference standard-  $\beta$ -Boswellic acid procured from Natural Remedies Pvt. Ltd., Bangalore, India.

### Sample & standard preparation

#### Sample preparation

1g of coarsely powdered drug samples were extracted with 10 ml n-Hexane for 24 hours by cold extraction method. The extracts were filtered by Whatmann filter paper and make up to 10 ml in a volumetric flask.

#### Standard preparation

5mg of standard  $\beta$ -Boswellic acid dissolved in 3ml of n-Hexane and made up to 5ml in standard volumetric flask.

### H.P.T.L.C (High Performance Thin Layer Chromatography)

TLC Aluminium pre coated plate with Silica gel60 GF<sub>254</sub> (20x10 cm<sup>2</sup>; 0.2 mm thick) was used with Toluene : Ethyl acetate: Formic acid (5:4.5:0.5) V/V as mobile phase. N-hexane extract of samples and  $\beta$ -Boswellic acid standard solution applied on plate by using Linomat V applicator. Cammag Twin Trough Glass Chamber (20x10 cm<sup>2</sup>) with SS lid was used for development of TLC plate.

The Twin Trough Glass Chamber was saturated with mobile phase for 30 minutes. TLC plate was developed to 8 cm distance above the position of the sample application. The plate was removed from the chamber and air dried at room temperature.

This plate was sprayed (derivatized) with Anisaldehyde- Sulphuric Acid reagent followed by heating at 110°C for 10 minutes and HPTLC finger print profile was snapped by Cammag Reprostar III, before derivatization under UV 254 nm, 366 nm and after derivatization (Fig.1).

The derivatized plate was scanned immediately using Camag TLC Scanner III at wavelength 530nm. Wincats an integrated Software 4.02 was used for the detection as well as for the evaluation of data.

### Method Validation and Recovery study

To study the accuracy and precision of the proposed method, recovery experiment was carried out. To a fixed amount of n-hexane

extract of samples, the standard solution of  $\beta$ -Boswellic acid was added (ratio 9:1 v/v) and total amount of standard  $\beta$ -Boswellic acid were determined. Percent recovery was calculated from the amount of  $\beta$ -Boswellic acid found via graph (Table No. 3).

### Linearity of detector response, assay and recovery

In order to establish linearity, standard solution of  $\beta$ -Boswellic acid (1mg/ml) applied on TLC Aluminium pre coated plate with Silica gel60 GF<sub>254</sub> (20X10 cm<sup>2</sup>; 0.2 mm thick), 10 $\mu$ l, 5 $\mu$ l, 1 $\mu$ l on Track No. S1, S2 & S3 respectively and for assay, 9 $\mu$ l of n-hexane extract of both samples applied on Track No. T1 & T2 and for recovery study, the n-hexane extract of both samples were spiked with standard  $\beta$ -Boswellic acid solution (ratio 9:1v/v) and applied 10 $\mu$ l on Track No. T3& T4 on the same plate.

TLC plates were developed to 8 cm distance above the position of the sample application and removed from the chamber and air dried at room temperature.

This HPTLC finger print profile was snapped by Cammag Reprostar III, before derivatization under UV Light 254 nm, 366 nm and after derivatization (Fig.1). The plate was derivatized with Anisaldehyde-Sulphuric Acid reagent followed by heating at 110°C for 10 minutes and scanned immediately using Camag TLC Scanner III at wavelength 530nm.

Wincats an integrated Software 4.02 was used for the detection as well as for the evaluation of data. It was observed that  $\beta$ -Boswellic acid appeared at R<sub>f</sub> 0.84 (dark violet colour). The peaks, graph and spectra obtained were given in Fig.2 and 3 and R<sub>f</sub> values, colour of bands (Table No.1), quantity of  $\beta$ -Boswellic acid, linearity, standard deviation & regression coefficient found via graph (Table No. 2) and calculated quantity of  $\beta$ -Boswellic acid & % recovery were given in Table No. 3.

## RESULTS AND DISCUSSION

Of the various mobile phases tried, the mobile phase containing Toluene : Ethyl acetate: Formic acid (5:4.5:0.5) v/v and the active principle  $\beta$ -Boswellic acid resolved as a dark violet colour band at R<sub>f</sub> 0.84 very efficiently from the other components in n-hexane extract of *Boswellia serrata* Roxb. (exudates) (Fig.1).

Sharp peaks of  $\beta$ -Boswellic acid (Standard and samples) were obtained when the plate was scanned at wavelength 530nm (Fig.2). Quantity of  $\beta$ -Boswellic acid found in samples were obtained automatically (Table No. 2) via graph (Fig.3) and %  $\beta$ -Boswellic acid found in samples and % recovery were calculated (Table No.3).

Quantity of  $\beta$ -Boswellic acid found in Local Market Sample, Ghaziabad (U.P.) is 1.5678mg in 1g drug sample (0.15678% w/w) and quantity of  $\beta$ -Boswellic acid found in Museum Sample of PLIM, Ghaziabad is 1.7100 mg in 1g drug sample (0.17100%w/w). The % recovery of  $\beta$ -Boswellic acid in Local Market Sample, Ghaziabad (U.P.) is 98.10% w/w and 97.05%w/w in Museum Sample of PLIM, Ghaziabad (U.P.). The mean % recovery was 97.58%.

The accuracy and reproducibility of the method was established by means of recovery experiment. The mean recovery was close to 100% which indicates the accuracy of the method.

The robustness of the method was studied, during method development, by determining the effect of small variation, of mobile phase composition ( $\pm 2\%$ ), chamber saturation period, development distance, derivatization time, and scanning time (10% variation of each). No significant change of R<sub>f</sub> or response to  $\beta$ -Boswellic acid was observed, indicating the robustness of the method.

Table 3

Sr. No. ↓	Sample from →	Local market sample, ghaziabad	Museum sample of plim, ghaziabad
1.	Quantity of $\beta$ -Boswellic acid in 1g	1.5678mg	1.7100 mg
2.	% $\beta$ -Boswellic acid	0.15678% w/w	0.1710% w/w
3.	% Recovery	98.10% w/w	97.05% w/w

Table 1

Sr. No.	Detection/visualization	Kundru exudate (Track No. T1, T2, T3 and T4)		Standard- $\beta$ -Boswellic acid (Track No. S1, S2 and S3)	
		R <sub>f</sub> values	Colour of band	R <sub>f</sub> values	Colour of band
1.	Under UV 254 nm	0.52	dark grey	-	No significant band
		0.56	dark grey		
		0.69	dark grey		
		0.76	dark grey		
		0.89	dark grey		
2.	Under UV 366 nm	0.52	blue	-	No significant band
		0.69	blue		
		0.89	bright sky blue		
3.	After derivatization	0.13	light violet	0.62	dark violet
		0.20	light violet		
		0.27	light violet		
		0.34	dark violet		
		0.40	violet		
		0.52	violet		
		0.62	dark violet		
		0.69	dark violet		
		0.76	dark brown		
		0.81	dark brown		
0.89	dark brown				

Table 2

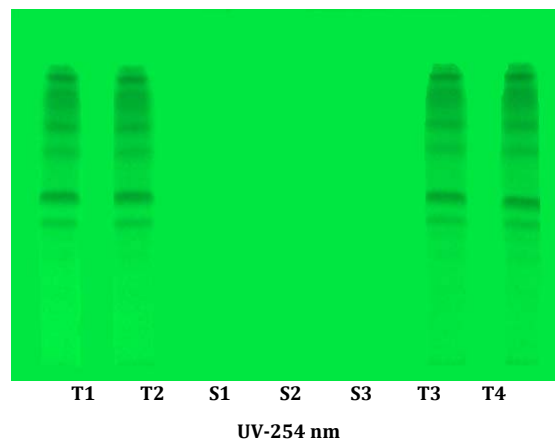
Sr. No.	Track No.	Volume applied on plate	Quantity applied on plate	Quantity of $\beta$ -Boswellic acid via graph	Linearity & Regression Coefficient and Standard deviation via graph
1.	T1	9 $\mu$ l	900 $\mu$ g	1.424 $\mu$ g	
2.	T2	9 $\mu$ l	900 $\mu$ g	1.562 $\mu$ g	$Y = 4717.263 + 54.644 * X + -0.027 * X^2$
3.	S1	10 $\mu$ l	10 $\mu$ g	10.000 $\mu$ g	$r = 0.99999 \quad \text{sdv} = 0.00\%$
4.	S2	5 $\mu$ l	5 $\mu$ g	5.000 $\mu$ g	
5.	S3	1 $\mu$ l	1 $\mu$ g	1.000 $\mu$ g	
6.	T3	(9+1) $\mu$ l	900 $\mu$ g+1 $\mu$ g	2.397 $\mu$ g -1 $\mu$ g = 1.397 $\mu$ g	
7.	T4	(9+1) $\mu$ l	900 $\mu$ g+1 $\mu$ g	2.516 $\mu$ g -1 $\mu$ g = 1.516 $\mu$ g	

T1- n-Hexane extract of Local Market sample, Ghaziabad, T2- n-Hexane extract of Museum Sample of PLIM, Ghaziabad

S1-  $\beta$ -Boswellic acid standard solution (1mg/ml), S2-  $\beta$ -Boswellic acid standard solution (1mg/ml)

S3-  $\beta$ -Boswellic acid standard solution (1mg/ml), T3- n-Hexane extract (spiked with std. solution) of Local Market Sample, Ghaziabad

T4- n-Hexane extract (spiked with std. solution) of Museum Sample of PLIM, Ghaziabad



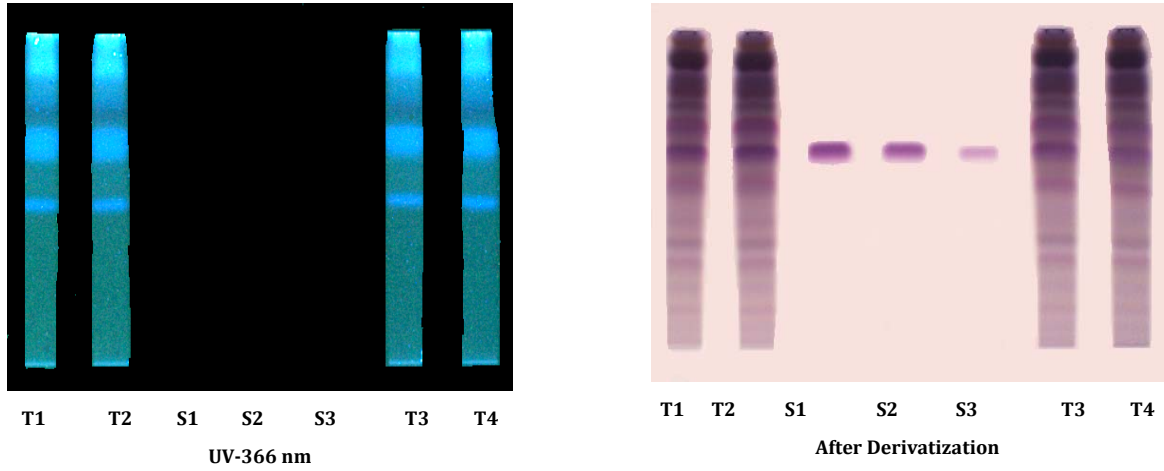


Fig. 1: T.L.C. Finger print of *Boswellia serrata* Roxb. (Exudate)

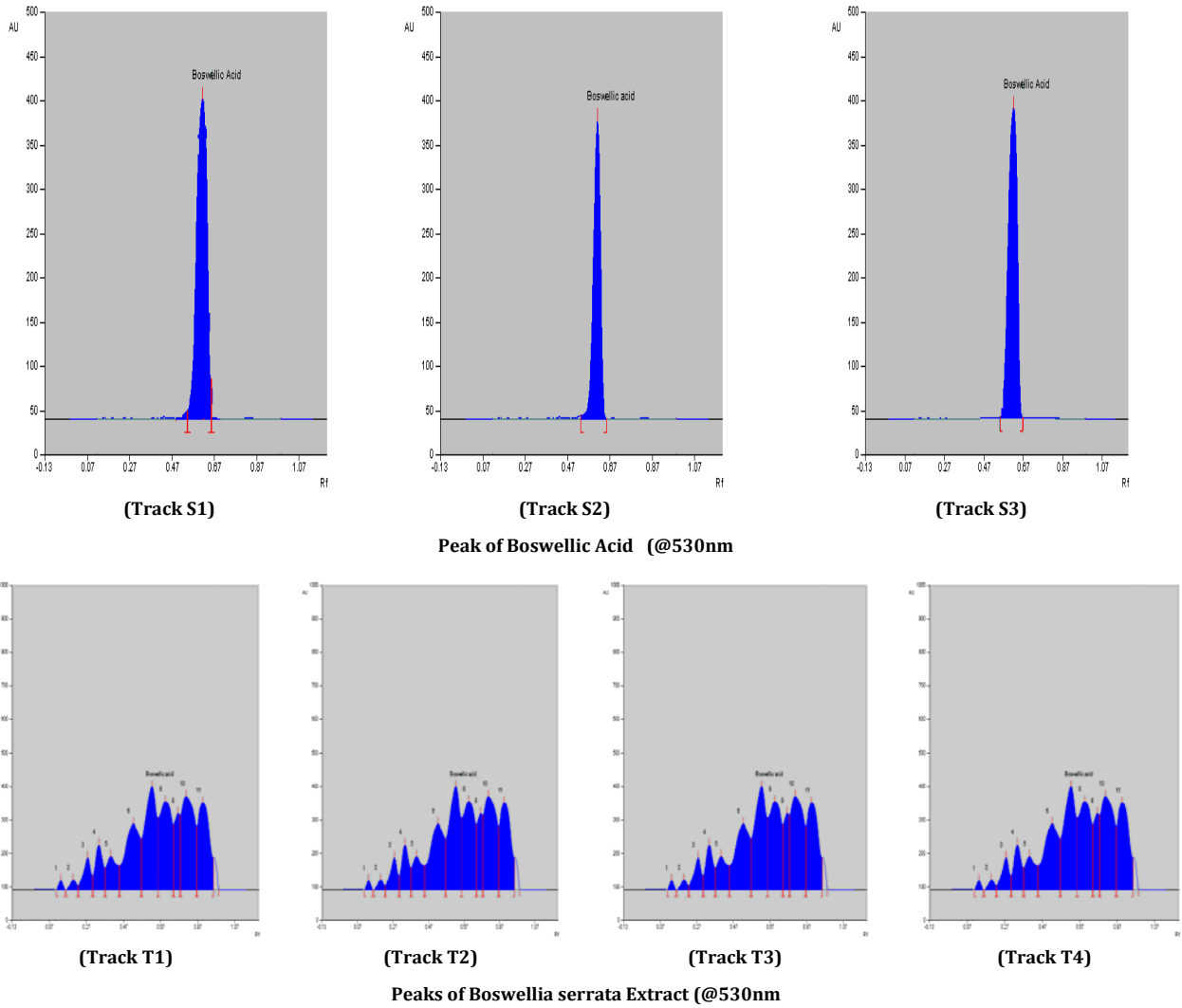
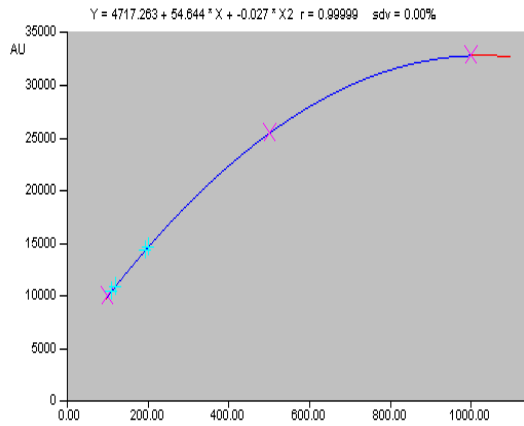
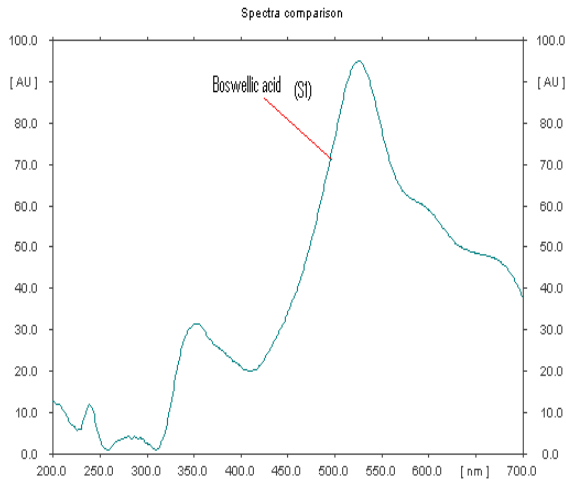


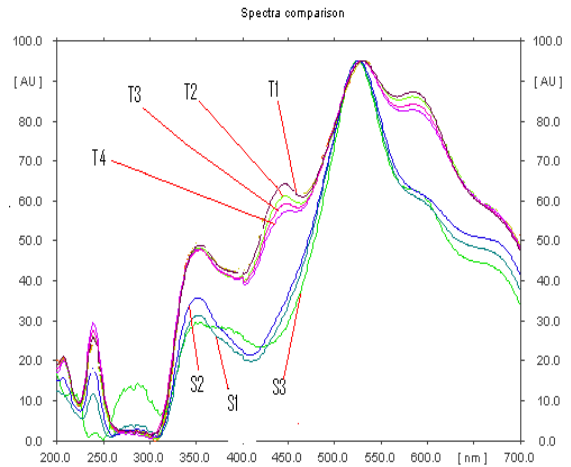
Fig. 2: Peaks of *Boswellia serrata* Roxb. (Exudate) in all Tracks



Conc. (µg) Graph Conc. vs AU

Spectra of  $\beta$ -Boswellic acid @ 530nm**CONCLUSION**

The proposed HPTLC method is simple, rapid, accurate, reproducible, selective and economic and can be used for routine quality control analysis of *Boswellia serrata* Roxb. (exudate) powder and quantitative determination of  $\beta$ -Boswellic acid in exudate powder.

Spectra of  $\beta$ -Boswellic acid in all tracks @ 530nm**Fig. 3: Graph and Spectra of *Boswellia serrata* Roxb. (Exudate)****REFERENCES**

1. Dhiman A. K., Ayurvedic Drug Plants, Daya Publishing House, Delhi, 2006, p-326-327.
2. Husain A. & et. al., Dictionary of Indian Medicinal Plant, CIMAP, Lucknow, 1992, p- 82.
3. The Wealth of India, Raw Materials, Revised Edition, Vol I, NISCAIR, CSIR, New Delhi, 1988, p-148.
4. Khare C.P., Encyclopedia of India, Rational Western Therapy, Ayurvedic and other Traditional Usage, 2004.
5. Prajapati/Purohit and et. al., A Handbook of Medicinal Plants, A Complete Source Book, Agrobio, India, 2004, p-96.
6. Sharma R.K., Govil J.N., Singh V.K., Recent progress in Medicinal plants, Vol.7, -Ethanomedicine and Pharmacognosy II, p-404.
7. Thakur R.S, Puri H.S., Akhtar Husain, Major Medicinal Plant of India, CIMAP, Lucknow, 1989, p- 124.
8. Indian Pharmacopoeia, 2007, p-2045.
9. Chatterjee Asima, Pakrashi S. C., The Treatise on Indian Medicinal Plants, Vol.-3, p- 63.
10. Elizabeth M. Williamson Major Herbs of Ayurveda, 2002, p-79.
11. Quality Standards of Indian Medicinal Plants Vol. 2, p- 19.
12. Rastogi Ram P., B.N. Mehrotra, Compendium of Indian Medicinal Plants, CDRI, PID, New Delhi, 1993, Vol. 2, p- 105.
13. Rastogi Ram P., B.N. Mehrotra, Compendium of Indian Medicinal Plants, CDRI, PID, New Delhi, Vol. 3, p-101.
14. Rastogi Ram P., B.N. Mehrotra, Compendium of Indian Medicinal Plants, CDRI, PID, New Delhi Vol. 4, p-115.
15. Pharmacognosy Reviews, Vol.1, Jan-May, 2007, p-137.
16. Sharma M.L. & et.al., Indian Journal of Pharmacology, 11 (6): 647- 652, 1989.
17. Kulkarni R.R., et al., Indian Journal of Pharmacology, 24 (1) :98-101, 1992.