

EVALUATION OF ENZYMATIC AND NON-ENZYMATIC ANTIOXIDANT POTENTIAL OF *EVOLVULUS ALSINOIDES* (L.)L.

D.GOMATHI, M.KALAISELVI, G.RAVIKUMAR AND C.UMA*

Department of Biochemistry, Karpagam University, Coimbatore – 641 021, Email: umaradhakrishnan29@gmail.com

Received:8 February 2012, Revised and Accepted: 7 May 2012

ABSTRACT

Introduction: Antioxidants are free radical scavengers which protect the human body against free radicals that causes various pathological conditions such as ischemia, anemia, asthma, arthritis, inflammation and aging process. Some of the antioxidant enzymes and non-enzymatic molecules widely distributed in the biological system which are capable of scavenging free radicals. The study was conducted to evaluate the antioxidant potential of *Evolvulus alsinoides* by using enzymatic and non enzymatic parameters. **Methods:** The enzymatic antioxidants (Superoxide dismutase, Catalase, Glutathione-s-transferase, Glutathione peroxidase, Peroxidase, Ascorbate oxidase and Polyphenoloxidase) and non-enzymatic antioxidants (Ascorbic acid, Total reduced glutathione and α -Tocopherol) activities were determined. **Result:** The results obtained in the present study showed the significant levels of enzymatic and non-enzymatic antioxidants in *Evolvulus alsinoides* (L.)L. **Conclusion:** From above study it is concluded that, the *Evolvulus alsinoides* showed promising antioxidant activity which can be used as effective protecting agents against oxidative stress and various diseases.

Keywords: *Evolvulus alsinoides*, Enzymatic antioxidants, Ascorbic acid, Total reduced glutathione, 50% ethanol

INTRODUCTION

Environmental stress adversely affects plant performance and often results in significant reductions in crop yield and quality worldwide. The exposure of plants to environmental stresses such as drought stress, heat stress, chilling stress, salt stress and plant diseases can result in the production of reactive oxygen species (ROS) that contributes to diminished plant performance^{1,2,3}. Reactive oxygen species (ROS) like peroxy radicals, hydroxyl radicals, superoxide anions, singlet oxygen (species), the ferryl or perferryl ions, hydrogen peroxide ions, and peroxy nitrates, are highly reactive molecules generated by biochemical redox reactions that occur as a part of normal cell metabolism. Oxidative defense is provided by a number of enzymes and vitamins, including retinol, tocopherol, ascorbic acid and glutathione. When free radical levels increase, individuals may become deficient in these protective antioxidants resulting in oxidative stress⁴. In order to limit oxidative damage under stress condition plants developed a series of detoxification systems that breakdown the highly toxic ROS^{5,6}.

Antioxidants are free radical scavengers which protect the human body against free radicals that causes various pathological conditions such as ischemia, anemia, asthma, arthritis, inflammation and aging process⁷. In nature, AOX are grouped as exogenous or endogenous. The endogenous group includes enzymes (and trace elements part-of) like superoxidase dismutase (Zn, Mn and Cu), glutathione peroxide (Se) and catalase, and proteins like albumin, transferrin, ceruloplasmin, metallothionein and haptoglobin. The most important exogenous AOX are dietary phytochemicals (such as polyphenols, quinones, flavonoids, catechins, coumarins, terpenoids) and the smaller molecules like ascorbic acid (Vitamin C), α -tocopherol (Vitamin-E)⁸.

Plants have developed an array of defense strategies (antioxidant system) to cope up with oxidative stress. The function of this antioxidant system is to scavenge the toxic radicals produced during oxidative stress and thus help the plants to survive through such conditions⁹. Numerous studies have shown that aromatic and medicinal plants are sources of diverse nutrient and non-nutrient molecules, many of which display antioxidant and antimicrobial properties which can protect the human body against both cellular oxidation reaction and pathogens¹⁰. These mechanisms employed ROS-scavenging enzymes, such as superoxide dismutase (SOD), catalase (CAT), and peroxidase (POD)¹¹. Therefore, the great interest has been recently focused on the natural foods, medicinal plants and phytocostituents due to their well-known abilities to scavenge free radicals (i.e. antioxidant power)^{12,13}.

Evolvulus alsinoides (L.)L. is a perennial herb belonging to the family *Convolvulaceae* with a small woody and branched root stock.¹⁴ It is

an important medicinal plant that grows in the open and grassy places almost throughout the India and subtropical countries of the world. Plant extracts have been used in traditional medicine for treatment of bronchitis, asthma and brain disorders like insanity, epilepsy, nervous disability, and scrofula. This extract also has exhibited antioxidant and immunomodulatory activities¹⁵. It has a known nootropic and anti-inflammatory activity^{16,17}. The aim of the present work was to study the antioxidant properties of whole plant of *Evolvulus alsinoides* (L.)L.

MATERIALS AND METHODS

Sample collection and extraction

The whole plant of *Evolvulus alsinoides* (L.)L. used for the investigation was obtained from Coimbatore District, Tamilnadu, India. The plant was authenticated by Dr.P.Satyanarayana, Botanical Survey of India, TNAU Campus, Coimbatore. The voucher number is BSI/SRC/5/23/2011-12/Tech.-514. Fresh plant material was washed under running tap water. The samples were prepared by grinding one gram each of whole plant material in 5 ml of 50% ethanol, separately, in a pre-chilled mortar and pestle and the extracts were centrifuged at 10,000 g at 4°C for 10 minutes. The supernatants thus obtained were used within four hours for various enzymatic and non-enzymatic antioxidant assays.

Estimation of enzymatic and non-enzymatic antioxidants in *Evolvulus alsinoides*

The enzymatic antioxidant activities of superoxide dismutase, catalase, glutathione peroxidase, glutathione s transferase, peroxidase, ascorbate oxidase and polyphenol oxidase were determined spectrophotometrically by using the methods of Das *et al.*, 2000, Sinha, 1972 Rotruck *et al.*, 1973, Habig *et al.*, 1974, Addy and Goodman, 1972, Vines and Oberbacher, 1965¹⁸⁻²³. The non-enzymatic antioxidants like vitamin C, α -tocopherol were studied by the method of Sadasivam and Manickam, 1996²⁴ and the total reduced glutathione were estimated by Boyne and Ellman, 1972 method²⁵.

RESULT AND DISCUSSION

Free radicals can be scavenged through chemoprevention by utilizing natural antioxidant compounds present in foods and medicinal plants. Almost all organisms are well protected against free radical damage by enzymes such as superoxide dismutase and catalase, or compounds such as ascorbic acid, tocopherols and glutathione. Some medicinal plants have been shown to have both chemopreventive and/or therapeutic effects on human diseases²⁶.

Figure 1 shows the activity of SOD and catalase of *Evolvulus alsinoides*. The levels were found to be 49.8 ± 0.13 units/mg protein and 180.3 ± 1.36 μ mole of H_2O_2 consumed/min/mg proteins. SOD is one of the antioxidant enzymes that play a key role in cellular defense against ROS²⁷. Similarly, CAT is also one of the principal

antioxidant enzymes; it eliminates H_2O_2 by transforming it into H_2O and O_2 . The stimulation of SOD activity along with CAT seemed to play a protective role against membrane damage as Cu is particularly toxic to membranes²⁸.

Concentration of SOD and Catalase in *Evolvulus alsinoides*

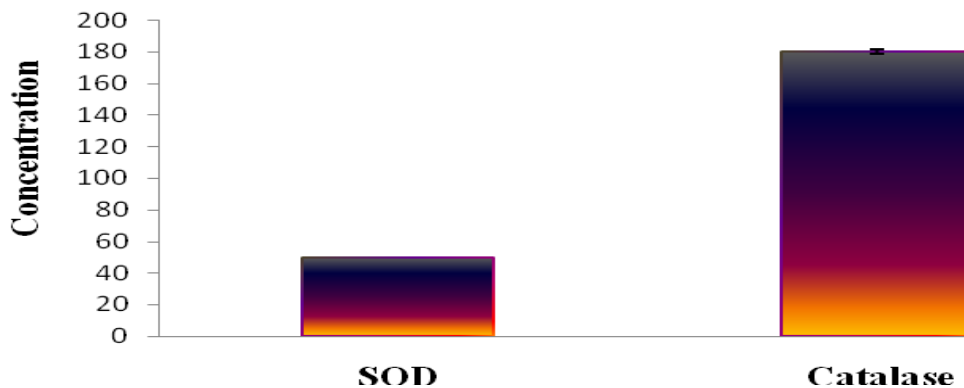


Figure 1

Values are expressed as mean \pm SD (n=3)

Units: SOD - Units/mg protein ; Catalase - μ mole of H_2O_2 consumed/min/mg protein

Glutathione peroxidase is the general name of an enzyme family with peroxidase activity whose main biological role is to protect the organisms from oxidative damage. The enzyme plays an important role in peroxide detoxification. Glutathione peroxidases utilize the reducing equivalents of glutathione to reduce hydrogen peroxide and it may be the main mechanism for protection against the deleterious effects of hydroperoxides. Glutathione reductase also known as GSR or GR is an enzyme that reduces glutathione disulfide (GSSG) to the sulfhydryl form GSH which is an important cellular antioxidant. Glutathione reductase plays an important role in protecting hemoglobin, red cell enzymes, and biological cell membranes against oxidative damage by increasing the level of

reduced glutathione (GSSGR) in the process of aerobic glycolysis¹⁰.

Glutathione peroxidase, Glutathione S transferase and Peroxidase levels in fresh plant were depicted in figure 2 which shows the high level of GPx, peroxidase activity (781.4 ± 1.46 μ g of glutathione oxidized/min/mg protein and 467 ± 0.90 μ moles/g tissue) and 353.1 ± 1.03 μ moles of CDNB - GSH conjugate formed/min/mg protein. Peroxidases are referring to heme containing enzymes which are able to oxidise organic and inorganic compounds using hydrogen peroxide as co-substrate. The non-specificity of peroxidase makes the enzyme suitable to a broad range of electron donor substrates²⁹.

GPx, GST and Peroxidase activity of *Evolvulus alsinoides*

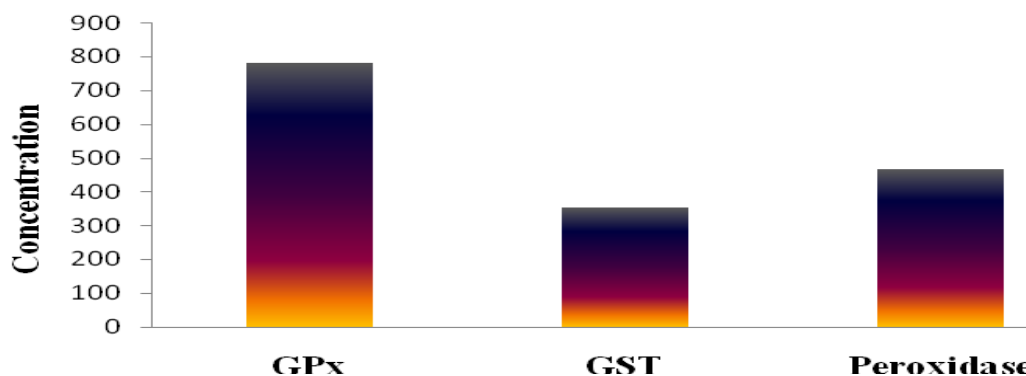


Figure 2

Values are expressed as mean \pm SD (n=3)

Units: GPx - μ g of glutathione oxidized/min/mg protein ; GST - μ moles of CDNB - GSH conjugate formed/min/mg protein
Peroxidase - μ moles/g tissue

The polyphenol oxidase (PPO) comprising of catechol oxidase and laccase is an enzyme that catalyzes the aerobic oxidation of variety of phenolic substrates in the plant material¹⁰ and ascorbate oxidase is an enzyme which catalyzes the one-electron oxidation of ascorbate with the concomitant four-electron reduction of dioxygen to water^{30,31}. Our results for ascorbate oxidase, glucose-6-phosphate-dehydrogenase and polyphenol oxidase in fresh plant sample were

represented in figure 3 which shows the high level of ascorbate oxidase and glucose-6-phosphate-dehydrogenase when compared to polyphenol oxidase. According to the results of Murata *et al.*, 2008³², ascorbate oxidase is a member of the multicopper oxidase family and can be obtained from higher plants such as green zucchini squash and cucumber and fungal species.

Concentration of Polyphenol oxidase, Ascorbate oxidase and Glucose 6 phosphate dehydrogenase in *Evolvulus alsinoides*

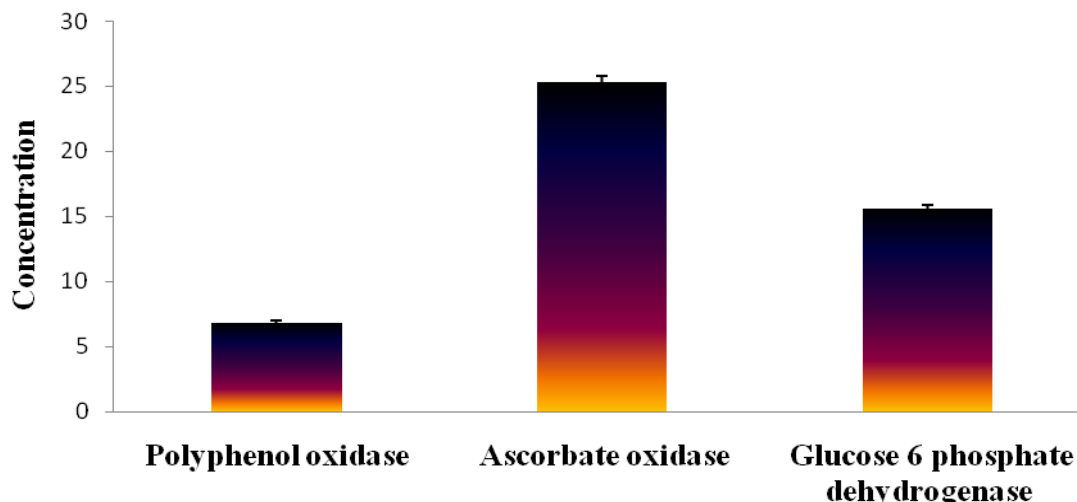


Figure 3

Values are expressed as mean ± SD (n=3)

Units: Polyphenol Oxidase - $\mu\text{moles/g tissue}$; Ascorbate oxidase - Unit/g tissue ; Glucose 6 phosphate dehydrogenase Units/ mg protein

Reactive oxygen species (ROS) get special attention lately due to many factors such as drought, cold, heat, herbicides and heavy metals. All of these factors lead to increasing number and accumulation of ROS in plant cells³³. Scientific research shows that ROS are harmful to the cell because they can raise the oxidative level through loss of cellular structure and function³⁴. ROS detoxification agents in cells include antioxidative enzymes such as ascorbate oxidase, peroxidase, catalase and ascorbate peroxidase. Johnson *et al.*, 2003³⁵ said that ROS detoxification agents also includes non

enzymatic antioxidants such as flavones, anthocyanins, carotenoids and ascorbic acid. The levels of total reduced glutathione and vitamin C of *Evolvulus alsinoides* were found to be $57.3 \pm 0.51 \mu\text{g/mg protein}$ and $365.18 \pm 0.94 \mu\text{g/mg protein}$ respectively (Figure 4). Ascorbic acid has been proposed to have roles on regulation of photosynthesis³⁶. Ascorbic acid is readily oxidised to monodehydro ascorbic acid as part of its antioxidant function³⁷. Ascorbic acid prevents or reducing oxidative damage was reported in ketoconazole treated drought stressed *C. roseus*³⁸.

Concentration of TRG, Vitamin C and Vitamin E in *Evolvulus alsinoides*

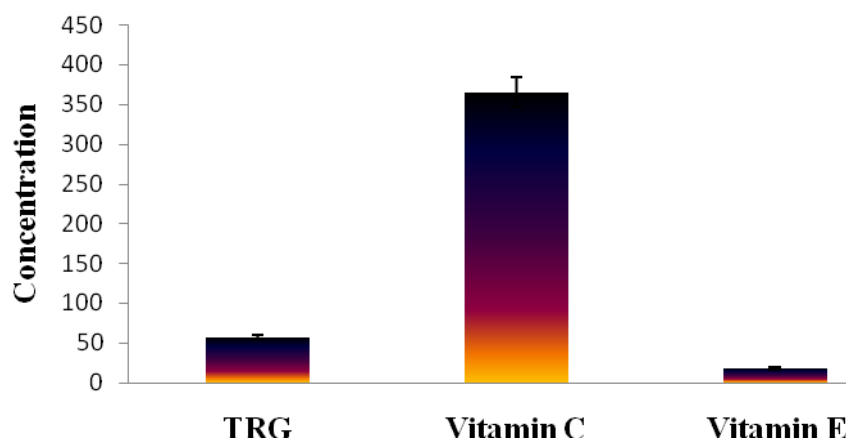


Figure 4

Values are expressed as mean ± SD (n=3)

Units: Total reduced glutathione - $\mu\text{g/mg protein}$; Vitamin C - $\mu\text{g/mg protein}$; Vitamin E - $\mu\text{g/mg protein}$

Vitamin E (α -tocopherol) is probably the most important lipid-soluble antioxidant protecting membranes, lipids and lipoproteins³⁹. Vitamin E is one of the few nutrients for which supplementation with higher than recommended levels have been shown to enhance immune response and resistance to diseases⁴⁰. Many studies have suggested that high intake of Vitamin E may slow down the development and progression of atherosclerosis. Some clinical trials

also reported beneficial effects of Vitamin E supplementation in the secondary prevention of cardiovascular events^{41,42}.

CONCLUSION

The results of the above study clearly indicated that the presence of significant level of enzymatic and non-enzymatic antioxidant activity in *Evolvulus alsinoides* that could protect against oxidant and free

radical injuries. Thus, the effective source of *Evolvulus alsinoides* (L.)L. could be employed in all medicinal preparations to combat diseases associated with oxidative stress including cancer, diabetes and related disorders.

REFERENCES

- Grill D, Tauze M, Dekok L. Significance of glutathione to plant adaptation to the environment. Kluwer Academic Publishers; Boston: MA. 2001.
- Rahimizadeh M, Habibi D, Madani H, Mohammadi GN, Mehraban A, Sabet AM. The effect of micronutrients on antioxidant enzymes metabolism in Sunflower (*Helianthus annuus* L.) under Drought stress. HELIA 2007;30 Suppl 47: 167-174
- Cicek N, Cakirlar H. Changes in some antioxidant enzyme activities in six soybean cultivars in response to long-term salinity at two different temperatures. Gen. App. Plant Physiology 2008;34 (3 Suppl 4): 267-280
- Sayed MSB, Khanum S, Tul-Kubra K, Ullah, MA, Chowdhury JA, Azad MAK et al. Antioxidant status and oxidative DNA damage of Bangladeshi dockyard laborers. Journal of Pharmacy Research. 2011;4 Suppl 10: 3271-3274
- Liu X, Huang B. Heat stress injury in relation to membrane lipid peroxidation in creeping bentgrass. Crop Sci 2000;40: 503-510.
- Almeselmani M, Deshmukh PS, Sairam RK, Kushwaha SR, Singh TP. Protective role of antioxidant enzymes under high temperature stress. Plant Sci 2006;171: 382-388.
- Gacche RN, Potlawar SG, Shegokar HD, Jadhav A.D. Evaluation of enzymatic and non enzymatic antioxidant potential of *Vitis vinifera* L. Asian J. Exp. Biol. Sci. Spl 2010;45-49
- Mon MM, Maw SS, Oo ZK. Quantitative Determination of Free Radical Scavenging Activity and Anti-tumor Activity of Some Myanmar Herbal Plants. World Academy of Science Engineering and Technology 2011;75: 524-530.
- Mandal S, Yadav S, Yadav S, Nema, RK. Antioxidants: A review. Journal of Chemical and Pharmaceutical Research 2009;1 Suppl 1: 102-104.
- Manjusha GV, Rajathi K, Alphonse JKM, Meera KS. Antioxidant Potential and Antimicrobial activity of *Andrographis paniculata* and *Tinospora Cordifolia* against pathogenic organisms. Journal of Pharmacy Research 2011;4 Suppl 2: 452-455
- Xu HW, Lu Y, Tong SY, Song FB. Lipid peroxidation, antioxidant enzyme activity and osmotic adjustment changes in husk leaves of maize in black soils region of Northeast China. African Journal of Agricultural Research 2011;6 Suppl 13: 3098-3102
- Galvez M, Martin-Cordero C, Houghton PJ, Ayuso MJ. Antioxidant activity of methanol extracts obtained from *Plantago* species. J. Agric. Food Chem 2005;53: 1927-1933.
- Nickavar B, Kamalinejad M, Izadpanah H. *In vitro* free radical scavenging activity of five *salvia* species Pak. J. Pharm. Sci. 2007;20 Suppl 4: 291-294
- Austin DF. *Evolvulus alsinoides* (*Convolvulaceae*). An American herb in the old world. Journal of Ethnopharmacology 2008;117 Suppl 2: 185-198.
- Gupta AD, Pundeer V, Bande G, Dhar S, Ranganath IR, Kumari GS. Evaluation of antioxidant activity of four folk antidiabetic medicinal plants of India. Pharmacologyonline 2009;1: 200-208.
- Singh A. Review of Ethnomedicinal uses and pharmacology of *Evolvulus alsinoides* Linn. Ethnobotanical leaflets 2008;12: 734-740.
- Omogbai BA, Eze FA. Phytochemical screening and susceptibility of bacteria pathogens to extracts of *Evolvulus alsinoides*. Science world journal 2011;6: 5-8.
- Das K, Samanta L, Chainy GBN. A modified spectrophotometric assay of superoxide dismutase using nitrite formation by superoxide radicals. Ind J Biochem Biophys 2000;37: 201-204.
- Sinha AK. Colorimetric assay of catalase. Anal Biochem 1972;47: 389-394.
- Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: Biochemical role as a component of glutathione peroxidase. Science 1973;179: 588-590.
- Habig WH, Pabst MJ, Jakoby WB. Glutathione S-Transferases: The First Enzymatic Step In Mercapturic Acid Formation. J Biol Chem 1974;249: 7130.
- Addy SK, Goodman RN. Polyphenol oxidase and peroxidase in apple leaves inoculated with a virulent or an avirulent strain for *Erwinia amylovora*. Ind Phytopath 1972;25: 575-579.
- Vines HM, Oberbacher MF. Response of oxidation and phosphorylation in citrus mitochondria to arsenate. Nature 1965;206: 319-320.
- Sadasivam S, Manickam A. Biochemical method. New Age International (P) Limited; New Delhi: 2nd ed. 1996. p. 108-110.
- Boyne AF, Ellman GL. A methodology for analysis of tissue sulfhydryl components. Anal Biochem 1972;46: 639-53.
- Shajiselvin CD, Muthu, AK. *In vivo* antioxidant and lipid peroxidation effect of various extracts from whole plant of *Bauhinia purpurea* (Linn) in rat fed with high fat diet. Journal of Pharmacy Research 2011;4 Suppl 10: 3280-3281
- Bowler C, Montagu MV, Inze D. Superoxide dismutase and stress tolerance. Annu. Rev. Plant Physiol. Plant Mol. Biol 1992;43: 83- 116.
- Ahmed A, Hasnain A, Akhtar S, Hussain A, Yasin AUG, Wahid A et al. Antioxidant enzymes as bio-markers for copper tolerance in safflower (*Carthamus tinctorius* L.). African Journal of Biotechnology. 2010;9 Suppl 33: 5441-5444,
- Halliwell B. Antioxidants and human disease - A general introduction. Nutr. Reviews 1994;55: 522-544.
- Solomon EI, Sundaram UM, Machonkin TE. Multicopper oxidases and oxygenases. Chem. Rev 1996;96: 2563-2605.
- Hakiman M, Maziah M. Non enzymatic and enzymatic antioxidant activities in aqueous extract of different *Ficus deltoidea* accessions. Journal of Medicinal Plants Research. 2009;3 Suppl 3: 120-131
- Murata K, Nakamura N, Ohno H. Elucidation of the factors affecting the oxidative activity of *Acremonium* sp. HI-25 ascorbate oxidase by an electrochemical approach. Biochem. Biophys. Res. Commun 2008;367: 457-461.
- Noctor G, Foyer CH. Ascorbate and glutathione: Keeping active oxygen under control. Ann. Rev. Plant Physiol. Plant Mol. Biol 1998;49: 249-279.
- Lee DH, Young SK, Lee CB. The inductive responses of the antioxidant enzymes by salt stress in the rice (*Oryza sativa* L.). J. Plant Physiol 2001;158: 737-745.
- Johnson SM, Doherty SJ, Croy RRD. Biphasic superoxide generation in potato tubers: a self amplifying responses to stress. Plant Physiol 2003;13: 1440-1449.
- Hong-Bo S, Chu L, Jaleel CA, Zhao CX. Water-deficit stress-induced anatomical changes in higher plants. Comptes Rendus Biologies 2008;331:215-225.
- Gomathinayagam M, Jaleel CA, Lakshmanan GMA, Panneerselvam R. Changes in carbohydrate metabolism by triazole growth regulators in cassava (*Manihot esculenta* Crantz); effects on tuber production and quality. Comptes Rendus Biologies 2007;330: 644-655.
- Jaleel. Non-Enzymatic Antioxidant Changes in *Withania somnifera* with varying drought stress levels. American-

Acknowledgement

We, the authors are thankful to our Chancellor, Advisor, Vice Chancellor and Registrar of Karpagam University for providing facilities and encouragement.

- Eurasian Journal of Scientific Research 2009;4 Suppl 2: 64-67.
39. VanBakl MME, Pritzen G, Wermuth B, Weisman UN. Antioxidant and thyroid hormone status in selenium-deficient phenyl ketonuric and hyperphenyl alaninemic patients. *Am. J. Clin. Nutr* 2000;976-981.
 40. Bendich A. Vitamin C safety in humans, In: L. Packer and Fuchs, J. (eds.) *Vitamin C in health and disease*, New York: Marcel Dekker Inc; 1997. p. 367-379
 41. Meydani M. Vitamin E and prevention of heart disease in high risk patients. *Nutr. Rev.* 2000;58: 278- 281.
 42. Selvi S, Devi UP, Suja S, Murugan S, Chinnaswamy P. Comparison of Non-Enzymic Antioxidant Status of Fresh and Dried Form of *Pleurotus florida* and *Calocybe indica*. *Pakistan Journal of Nutrition* 2007;6 Suppl 5: 468-471