

SYNTHESIS, CHARACTERIZATION AND MEDICAL EFFICACY OF CR(III) COMPLEXES OF SULPHONYL-UREAS, AS ORAL ANTIDIABETICS

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ABSTRACT

Synthesis, characterization and hypoglycemic activity of chromium(III) complexes with Gliclazide (GLC), Glibenclamide (GLB) and Glimeperide (GLM) oral antidiabetic allopathic drugs have been studied. The conductometric titration using monovariation method indicates that complexes are non-ionic and L₂M type. Analytical data agrees with the molecular formulae of complexes viz., (C₁₅H₂₁N₃O₃S)₂Cr·2H₂O, (C₂₃H₂₈N₃O₅SCl)₂Cr·2H₂O, (C₂₄H₃₄N₄O₅S)₂Cr·2H₂O. Structure of complexes was assigned as octahedral in which ligand molecules lie horizontally joining the central chromium atom and two water molecules attached vertically with the metal. Infra-red spectral studies confirm the coordination of sulphonyl oxygen on one side and enolic oxygen from other side with metal ion, IR, Mass and ¹H-NMR studies supports structure IV for the complex proposed on the basis of analytical data. Magnetic susceptibility suggests that the complexes are diamagnetic, Thermal studies supports the presence of ligand moieties and coordinated water. X-ray diffraction data also supports the complex formation and symmetries.

Keywords: HPMC-5CPS, Gliclazide, Glibenclamide and Glimeperide complexes. Hypoglycemic activity

INTRODUCTION

Chromium is an essential metal that appears to have beneficial role in regulation of insulin action, metabolic syndrome and cardiovascular disease. Chromium function in our bodies is critical without it, the hormone insulin would not work. Most of the people are familiar with insulin as the shot diabetes give themselves in order to control their high blood sugar. But what most of the people don't realize is that insulin is the "Master hormone of our metabolism, it is not only controls blood sugar levels and many other aspects of carbohydrate break down and storage but also directs much of the metabolism involving fat, proteins and energy (calories). Because insulin requires chromium to function properly¹. Chromium reduces insulin resistance, this essential trace element could therefore have wide ranging effects on high blood pressure and abnormal blood lipid in addition to lowering blood sugar².

Kaats *et al.*,³ suggest that supplementation with chromium picolinate can lead significant improvement in body consumption when a BCI (Body Consumption Index) is used as the outcome criterion that represent a sum of the net gain in non fat mass added to sum of the net losses of body fat. Chromium is a true potentiator of insulin and is known as glucose tolerance factor (GTF) trivalent chromium Cr(III) has been claimed to be a constituent of glucose tolerance factor. Schwartz and Mertz (1959) showed that trivalent chromium (Cr³⁺) cured the impaired glucose tolerance observed in rats on a chromium deficient diet.

Pandey *et al.*,⁴ studied the Cr(III) complexes with Di-benzyl sulphide ligand.

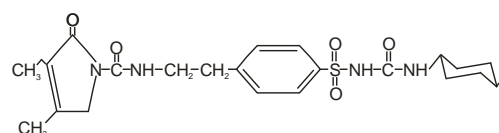
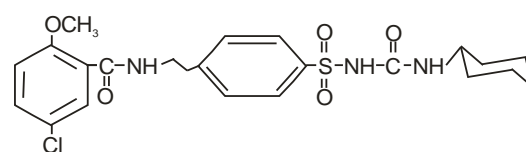
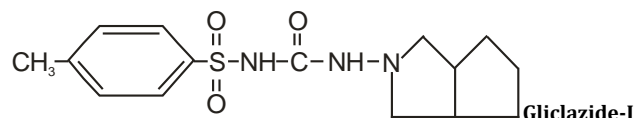
Louise *et al.*⁵, synthesized homoleptic trimethyl silylacetylde complex of Cr(III), LiF *et al.*⁶, Synthesize Cr(III) complex and compare anti-hyperglycemic activity shows enhanced antidiabetic activity.

Yang X *et al.*⁷, synthesize a new chromium complex-chromium (phenylalanine) improves insulin responsiveness and reduces whole body glucose tolerance.

Complexation of sulphonylureas with transition and inner transition metal has been studied in detail by Yoshinaga and Yamamoto⁸⁻⁹ (1966 a,b), Iqbal *et al.*¹⁰ (1984, 1985, 1986), Dury and Al-Jibori¹¹ (2012) Shahriare and Ghammany¹² (2012) Modhaviya¹³ (2012), AbdulVudood *et al.*¹⁴ (2012), Sunilkumar and Sharma¹⁵ (2012) and Jacob and Iqbal¹⁶ (2010)

A perusal of available literature shows that systemic study on complexation of chromium(III) with sulphonyl ureas is relatively scanty. The study of chemistry and chemical reaction of coordination

compounds helps in establishing structure activity relationship. It has been reported that the biological activity of metal complexes is more potent and less toxic as compared to the free ligand Singel¹⁷ (1982), Brown¹⁸ (1982), Phipps¹⁹ (1976), Williams²⁰ (1976), Lippard²¹ (1983), Meares and Wensel²² (1984). In view of the above and in continuation of our work, it is interesting to have an insight into the synthesis of chromium(III) complexes with Gliclazide, Glibenclamide and Glimeperide and to study various structural aspects of the isolated complexes. here, the synthesis and characterization of chromium(III) complexes with sulphonyl ureas have been described for following drugs.



EXPERIMENTAL

Ligand-Metal ratio

a) Pure gliclazide m.p. 180°C (Lit.179.5-180.5), 0.005M, pure Glibenclamide m.p. 172.08°C (Lit.170.5-173.5), 0.005M and Glimeperide m.p. 207.00°C (Lit. 206.5-208.00), 0.005M, with chromium chloride 0.01M prepared (AnalaR grade) were separately prepared in purified 90% ethanol, Gliclazide, Glibenclamide and Glimeperide (20 ml.) was diluted to 200 ml. each and titrated conductometrically against chromium chloride at 29±1°C. Results were plotted in the form of graph which indicates ligand metal ratio as 2:1 (L₂M)

b) Formation of 2:1 (L₂M) ratio was also confirmed by Job's method¹⁵ of continuous variation as modified by Turner and Anderson¹⁶, using Δconductance as index property. From these values the stability constant (logk) and free energy change (-ΔF), were also calculated (Irving and Rossotti²³⁻²⁵ (1953, 1954, 1955), Willard *et al.*²⁶, 2000) Tables 1 and 2, fig 1 and 2 given only for Gliclazide chromium complex.

Synthesis of complexes

The chemicals used in this synthesis were all of analytical grade. A weighed quantity of Gliclazide, Glibenclamide and Glimeperide (2mol.) was dissolved separately in minimum quantity of 90% ethanol. The chromium chloride solution was prepared by dissolving it separately in the same solvent. Metallic solution was added slowly with stirring into the solution of the ligand at room temperature maintaining the pH between 6.0 to 6.5 by adding dilute NaOH solution. On refluxing the mixture for 3h and on cooling, the complexes separated out, which were filtered off, washed well with ethanol and finally dried in vacuum and weighed.

The elemental analysis of the isolated complexes were carried out using the reported method Jeffery *et al.*²⁷, (1989), Mohammed²⁸ (1989), Scott²⁹ (1960) while chromium was estimated as chromium oxide.

The IR spectrum of the ligands as well as of the complexes were recorded on Perkin Elmer Spectrum RX1 model FTIR (CDRI Lucknow) India, ¹H-NMR spectra of the ligand and isolated complexes were recorded on a Bruker AM-200 Spectrometer (CDRI Lucknow) and d₆-DMSO was used as a solvent (Fig.-3 and Fig.-4)

For the thermal studies, thermogravimetric analysis (TGA/DTA) technique was adopted at IIT Bombay (Mumbai) India. X-ray diffractometer model Rigaku D-max/B, with 12KW rotating Anode X-ray generator was used for scanning the ligand, metal salt and respective complexes, radiation used was Cu, Kα (1W=1.5060Å). The samples were scanned in the range 10.0084 to 79.9804°(2θ) powder data were indexed using computer software (FPSUIT V 2.0)

From stoichiometry and analytical data, the composition of the complex comes out to be (C₁₅H₂₁N₃O₃S)₂Cr·2H₂O, (C₂₃H₂₈O₅N₃SCI)₂Cr·2H₂O and (C₂₄H₃₄N₄O₅S)₂Cr·2H₂O for GLC, GLB and GLM respectively which favours 2:1 (L₂:M) ratio. The tentative common structure IV assigned to the complexes on the basis of analytical data is further supported by Thermogravimetric study and XRD data. Cullity³⁰(1978), Bragg and Bragg³¹ (1993), Guinier³² (1952), Henry *et al.*,³³ (1951)

RESULTS AND DISCUSSION

Thermal Studies

The thermal studies using TGA method shows that the complex obtained is a well knit one with water of hydration. It has a sharp melting peak indicating that the entire process of decomposition took place in a temperature range (120-510°C) It is also devoid of water of hydration and hence the structure losses its crystallinity and water of coordination and gets melted simultaneously. Chromium is a highly electropositive and hence the complex decompose at higher temperature³⁴⁻³⁶ of (783.°k) Other kinetic parameters like energy of activation, free energy (E_a), entropy change (ΔS), apparent entropy (S*), order of reaction (n) and frequency factor (Z) etc calculated, using Sharp-Wentworth and Freeman-Carroll methods³⁷⁻³⁸ (Table-10). The magnetic studies indicate that the complex is diamagnetic in nature.

Infra-red spectral studies

The IR spectra of ligand and isolated complexes were recorded the range 4000-400 cm⁻¹. The assignments of the infrared spectral bands are based on literature. (Table 5 Fig 3)

The strong band in its region of 3355cm⁻¹ (GLC), 3340cm⁻¹ (GLB) and 3350 cm⁻¹(GLM) indicates the presence of coordinated water which was further confirmed by thermal studies.

The proposed structure for the isolated complexes also supported by IR absorption with reference to pioneer workers, Weissberger³⁹ (1956), Cotton⁴⁰ (1960), Nakamoto⁴¹ (1963), Rao⁴² (1963), Bellamy⁴³ (1964). IR bands obtained at 1143 ± 20 cm⁻¹, which is the characteristic of combination frequencies resulting from C=O stretching vibration and M-O stretch. Absorption bands due to C=N stretching Dyer *et al.*⁴⁴, (1966) vibrations are also found in the region 2522 ± 5cm⁻¹. The complexes also displays a band at 1142 cm⁻¹. Which is considered to be associated with νSO₂-N band (Table 5) Iqbal *et al.*,

¹H-NMR Studies

The ¹H-NMR spectral data are given in (table 6). It was observed that the singlet due to the amide (NH) proton around (δ8.74) in the spectrum of the ligand disappeared in the complex shows the formation of M-O band. This also confirms the deprotonation of amide NH group through enolisation (the appearance of > C=N stretching band observed in IR spectra)

Other feature of NMR spectrum were the aromatic proton resonances located and the presence of unresolved multiplet suggestive of excessive deshielding of aromatic protons Slichter⁴⁵ (1963), Akitt⁴⁶ (1973), Siewers⁴⁷ (1973), Dixit and Singh⁴⁸, (2001), Pandey *et al.*⁴⁹, (2003) Iqbal *et al.*,

Electronic Spectral Studies

Three spin allowed transitions at ν₁ ⁴A_{2g} → ⁴T_{2g}, ν₂ ⁴A_{2g} → ⁴T_{2g}(F) and ν₃ ⁴A_{2g} → ⁴T_{2g}(P) (Cotton *et al.*, 1999) are observed suggesting for octahedral Cr(III) complex. The diffused reflectance spectrum of the Cr(III) chelate shows three absorption bands between 18000 to 28000 cm⁻¹ which are in agreement with those in the literature for octahedral Cr(III) complexes Cotton *et al.*⁵⁰, (1999).

X-Ray Diffraction Studies

X-ray diffraction studies also confirms the complexes and formation of a new bonds the number of peaks in Gliclazide, Glibenclamide and Glimeperide are 13,11 and 9 respectively (Table 7) and that of chromium chloride is 8 thus indicating that complexes formed are well kit one moreover the X-ray pattern of neither Gliclazide, Glibenclamide and Glimeperide nor chromium chloride are seen in diffractogram of the complexes. all the reflections present are new ones and the patterns are fairly strong. On comparing the pattern obtained with available literature, it is came evident that its pattern is not in good agreement with available information and thus confirms the formation of totally new complexes (Fig-9)

The X-ray diffraction pattern of Cr(III) complexes has been determined for 2θ range from 5.0084° to 79.97884° from the cell data and crystal lattice parameters one can conclude that Cr(III) complexes with GLC, GLB and GLM are having monoclinic crystal system (Table 7, only one diffractogram and cell data is enclosed of Gliclazide-Cr(III) complex) keeping in view all these observations and results the following structure of Gliclazide, Glibenclamide and Glimeperide, chromium complexes can be proposed for the isolated complexes.

The general structure (IV) of the complexes is further supported by the values of ¹H-NMR as well as IR frequencies (table 5 and 6.). Moreover the enolization of N¹ nitrogen is not possible because its hydrogen is simultaneously attracted from the group SO₂ from one side -C=O on the other side the participation of N² hydrogen in enolization is supported from the fact that -R group is electron releasing while on other side carbonyl oxygen is proton attracting.

Table 1 and 2: Gliclazide with Chromium Chloride (Jobs Methods)

Ratio	Gliclazide - 0.005M Solvent - 90% Ethanol			Cond. x 10 ⁻⁴ Mhos (C ₁ +C ₂ -C ₃)	Corrected Δ Cond. x 10 ⁻⁴ Mhos	Ratio	CrCl ₃ -0.005M Temp - 27±1°C			Conductance x10 ⁻⁴ Mhos (C ₁ +C ₂ -C ₃)	Corrected Δ Cond. x 10 ⁻⁴ Mhos
	Gliclazide - 0.002M Solvent - 90% Ethanol						CrCl ₃ -0.002M Temp - 27±1°C				
	S:L C ₁	M:S C ₂	M:L C ₃				S:L C ₁	M:S C ₂	M:L C ₃		
00/12	0.18	2.45	2.50	0.13	0.00	00/12	0.13	1.80	1.87	0.06	0.00
01/11	0.40	2.28	2.44	0.24	0.12	01/11	0.28	1.72	1.83	0.17	0.13
02/10	0.67	2.24	2.42	0.49	0.39	02/10	0.49	1.61	1.79	0.31	0.27
03/09	0.86	2.13	2.38	0.61	0.51	03/09	0.68	1.50	1.73	0.45	0.41
04/08	1.11	2.03	2.32	0.82	0.74	04/08	0.86	1.43	1.71	0.58	0.54
05/07	1.25	1.79	2.30	0.74	0.65	05/07	0.45	1.25	1.18	0.52	0.48
06/06	1.41	1.56	2.32	0.65	0.56	06/06	0.98	1.09	1.62	0.45	0.41
07/05	1.55	1.30	2.25	0.60	0.51	07/05	1.03	0.96	1.59	0.40	0.36
08/04	1.73	1.00	2.25	0.48	0.40	08/04	1.09	0.76	1.56	0.29	0.27
09/03	1.90	0.70	2.22	0.38	0.36	09/03	1.16	0.64	1.53	0.27	0.24
10/02	2.05	0.45	2.20	0.30	0.22	10/02	1.26	0.42	1.49	0.19	0.16
11/01	2.19	0.21	2.19	0.21	0.14	11/01	1.37	0.23	1.47	0.13	0.10
12/00	2.27	0.04	2.20	0.11	0.00	12/00	1.47	0.08	1.50	0.05	0.00

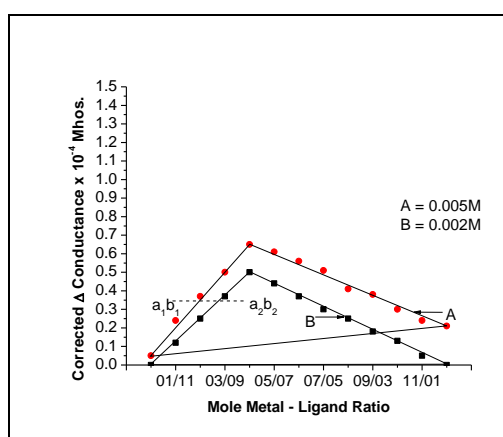
Modified Jobs Method:Gliclazide With Chromium Chloride (CrCl₃)

Fig. 1

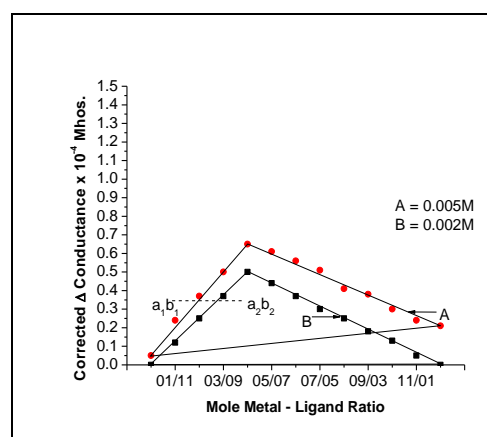


Fig-2

Table 3: Physico-chemical and analytical data of GLC-Cr, GLB-Cr and GLM-Cr complexes

S.N.	Complexes	Colour	Yield (%)	M.P. (°C)	Molar conductance Ω ⁻¹ cm ⁻¹ mol ⁻¹
1	(C ₁₅ H ₂₁ N ₃ O ₃ S) ₂ Cr·2H ₂ O	Green	50.61	180	29.50
2	(C ₂₃ H ₂₈ O ₅ N ₃ SCI) ₂ Cr·2H ₂ O	Green	69.75	181	27.40
3	(C ₂₄ H ₃₄ N ₄ O ₅ S) ₂ Cr·2H ₂ O	Green	54.61	180	29.50

Table 4: Elemental analysis of GLC-Cr, GLB-Cr and GLM-Cr complexes

S.N.	Molecular formulae of complexes	Molecular weight (gm/mole)	% Analysis found (calculated)							
			C	H	N	O	S	Cl	Metal	H ₂ O
1	(C ₁₅ H ₂₁ N ₃ O ₃ S) ₂ Cr·2H ₂ O	732.824	48.72 (49.12)	4.25 (5.45)	11.23 (11.46)	13.12 (13.10)	8.15 (8.73)	-	7.84 (7.09)	4.74 (4.91)
2	(C ₂₃ H ₂₈ O ₅ N ₃ SCI) ₂ Cr·2H ₂ O	1040.004	53.24 (53.07)	5.24 (5.19)	8.18 (8.07)	15.11 (15.38)	6.28 (6.15)	6.11 (6.82)	5.18 (4.99)	3.94 (3.46)
3	(C ₂₄ H ₃₄ N ₄ O ₅ S) ₂ Cr·2H ₂ O	1069.23	53.61 (53.87)	6.27 (6.17)	10.41 (10.47)	17.90 (17.95)	5.90 (5.98)	-	4.26 (4.86)	3.84 (3.36)

Table 5: Specific IR assignment of sulphonyl ureas-chromium complexes.

IR frequencies (cm ⁻¹)	Assignments		
	GLC-Cr	GLB-Cr	GLM-Cr
670±5	680±2	665±10	Metal oxygen bond
909	980	908±10	Aromatic ring vibration
998	1020±20	990±20	S=O frequencies (LJB/359) Bellamy 1964
1018	1055	1060	C-O of Chelate ring
1142±5	1120	1128	SO ₂ -N frequency
1442	1435±10	1437	Six membered enolic ring structure modified in complex

1643±5	1655±10	1640±20	C-O stretching frequency (KN/184) Nakamoto 1963
2522	2550±5	2520±10	C=N stretching frequency
3355	3340±10	3350	Coordinated water
710±5	705±10	708±5	Ar-S linkage (LJB/355) Bellamy 1964
813±10	820±10	810±10	1-4 disubstituted benzene ring frequency

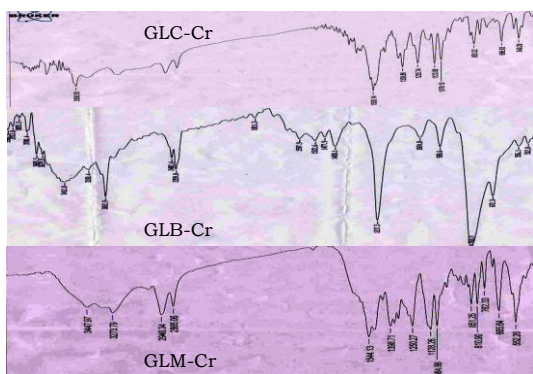


Fig. 3: Infra-red spectra of GLC-Cr, GLB-Cr and GLM-Cr complexes

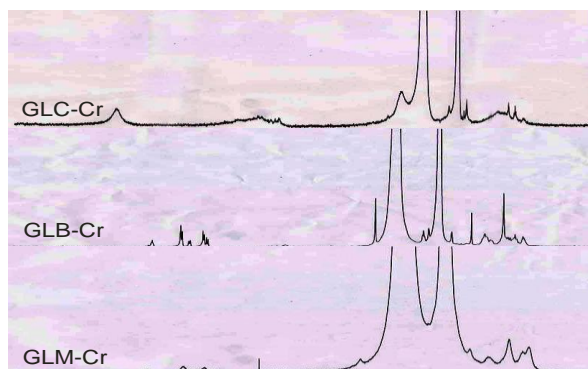


Fig-4: ¹H-NMR spectra of GLC-Cr, GLB-Cr and GLM-Cr Complexes

Table 6: ¹H-NMR Assignment of GLC-Cr, GLB-Cr and GLM-Cr complexes

COMPLEXES	Assignment
GLC-Cr	δ0.944-1.855 (t,H,CH ₃), δ2.264-2.826 (d,H,O-H), δ3.396-4.461 (s,2H,NH), δ7.129 (d,4H,Ar-H), δ7.679 (t,6H,Ar-H), δ7.993 (s,6H,Ar-H) and δ11.343-11.519 (t,H,Cr-OH ₂).
GLB-Cr	δ1.407-1.986 (d,2H,CH ₂), δ2.034-2.852 (d,2H,O-CH ₃), δ2.935-3.866 (d,3H,NH), δ7.129-7.230 (d,3H,Ar-H), δ7.485-7.669 (d,4H,Ar-H) and δ8.239-8.255 (t,H,Ar-H).
GLM-Cr	δ1.11-1.44 (CH ₂), δ1.71-1.96 (CH ₂), δ3.00-3.14 (NH), δ3.77 (-O-CH), δ6.42 (aromatic), δ3.77 (NH-CO-Cr), δ6.40 (aromatic), δ6.83-6.89 (aromatic), δ7.30-7.51 (aromatic), δ7.87-7.90 (aromatic), δ7.97-7.94 (aromatic), δ8.11-8.15 (aromatic) and δ10.14 (H,Cr-OH ₂)

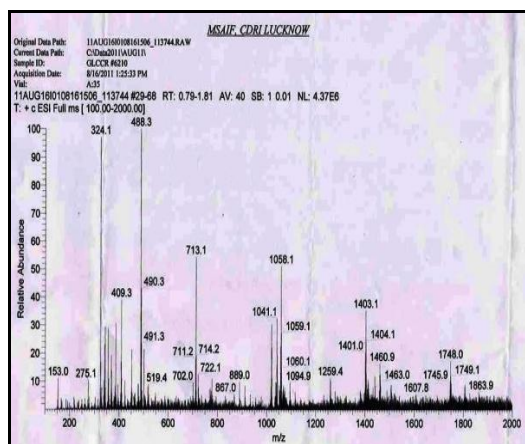


Fig. 5: Mass Spectrum of GLC-Cr Complex

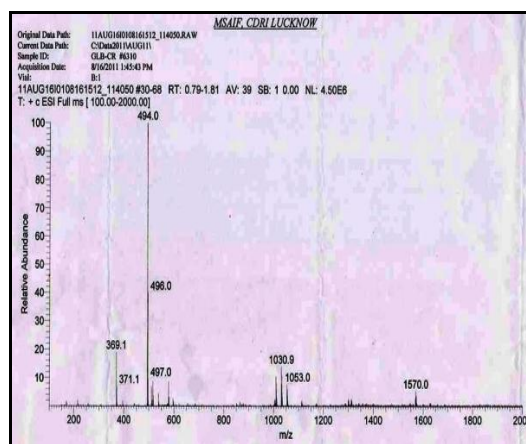


Fig. 6: Mass Spectrum of GLB-Cr Complex

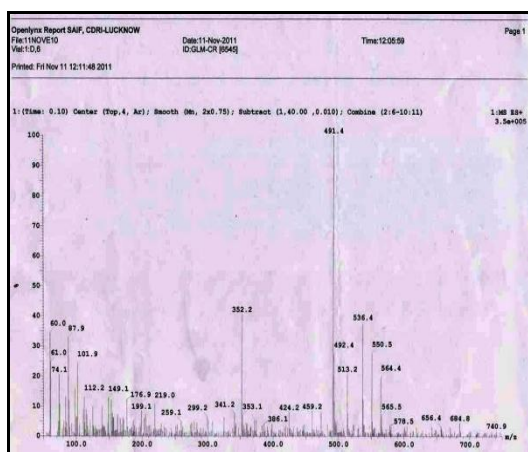


Fig. 7: Mass Spectrum of GLM-Cr Complex



Fig. 8: TGA Curve of GLC-Cr Complex

Table 7(A): Cell data of crystal parameter of GLC-Cr Complex

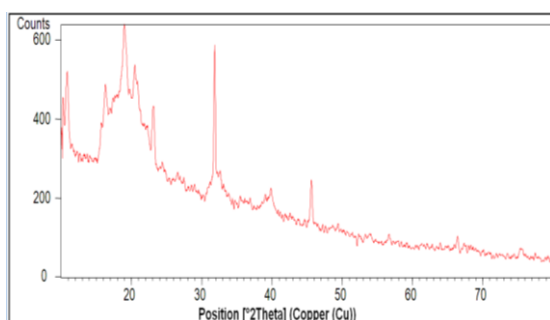


Fig.9: X-ray diffractogram of Gliclazide-Cr complex

$a(\text{\AA}) = 21.6990$ Volume($abc\sin\beta$) $\text{\AA}^3 = 13880.931$
 $b(\text{\AA}) = 23.1881$ D_{cal} = 4.02100 g/cm³
 $c(\text{\AA}) = 27.5891$ D_{obs} = 4.03241 g/cm³
 Standard deviation = 0.024%
 Crystal system = Monoclinic
 $\alpha = 90^\circ, \beta = 89.4^\circ, \gamma = 90^\circ$
 Porosity(%) = 2.837
 Density = 0.05329g/cm³
 Particle size = 15.794microns

Table 7 (B): Cell data of crystal parameter of GLC-Cr complex

2θ	I/I ₀	D _(Obs)	D _(Cal)	h	k	l
10.5540	69.89	8.38237	8.46731	1	0	3
16.3437	60.23	5.42369	5.42470	4	0	0
19.0109	99.40	4.66834	4.64949	2	3	4
19.8736	82.38	4.46760	4.46387	3	4	1
20.4293	53.81	4.34731	4.34770	1	4	4
22.0439	100.00	4.03241	4.02100	5	2	1
31.8513	52.99	2.80964	2.80664	7	1	4
39.3397	29.09	2.29037	2.28905	1	4	11
45.6199	37.40	1.98697	1.98754	9	3	7

Table 8: Mass Spectra Data of the Gliclazide complexes

Sl. No.	Compound Formula Weight (g/mol)	MS/EI	Assignment
1.	(C ₁₅ H ₂₁ N ₃ O ₃ S) ₂ Cr·2H ₂ O 732.824	722.1 323.412(cal).324.1(fnd)	[M+2H] ²⁺ , [M-2H ₂ O+H] ⁺ , [L+H] ⁺
2.	(C ₂₃ H ₂₈ O ₅ N ₃ SCl) ₂ Cr·2H ₂ O 1040.004	1031 494(cal).496(fnd)	[M-2H] ²⁺ , [M-2H ₂ O+H] ⁺ , [L+H] ⁺
3.	(C ₂₄ H ₃₄ N ₄ O ₅ S) ₂ Cr·2H ₂ O 1069.23	1071.32 490(cal).491.4(fnd)	[M+2H] ²⁺ , [M-2H ₂ O+H] ⁺ , [L+H] ⁺

Table 9: Thermogravimetric Data and Decomposition Temperature Range of Gliclazide-Complexes

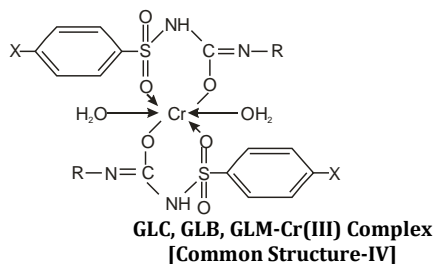
GLC-Complexes	Loss of crystalline water molecular		Decomposition step, temperature range (°C), mass loss (%)						Mass of residue left over (%)
			First Step		Second Step		Third Step		
			Temp. range (°C)	Mass loss (%)	Temp. range (°C)	Mass loss (%)	Temp. range (°C)	Mass loss (%)	
(GLC) ₂ Cr·2H ₂ O	40-120	4.74(F) 4.91(C)	120-200	15.23(F)	200-300	31.94(F)	300-510	42.28.55(F)	10.54(F)
				23.18(C)	47.74(C)	100.0(C)			
(GLB) ₂ Cr·2H ₂ O	40-130	3.94(F) 5.86(C)	130-310	9.14(F)	310-350	14.47(F)	350-510	17.82(F)	3.55(F)
				23.18(C)	47.74(C)	100.0(C)			
(GLC) ₂ Cr·2H ₂ O	40-130	5.24(F) 2.89(C)	130-250	9.84(F)	250-350	61.33(F)	350-510	69.71(F)	0.96(F)
				36.81(C)	69.29(C)	100.0(C)			

F=Found, C=Calculated

Table 10: Thermogravimetric data of Metal complexes of Gliclazide drug with corresponding to heating rate of 10°C/min.

Complexes	Decomposition Temp. (°C)	%Wt. loss	Ea(Kj/mole)		ΔS* (Kj/mole)	ΔF (Kj/mole)	Z	S*	n
			F.C.	S.W.					
(C ₁₅ H ₂₁ N ₃ O ₃ S) ₂ Cr·2H ₂ O	40-120	4.74	33.67	32.66	-33.98	-10.26227	322.8	-48.5380	1.01
	120-200	15.23	55.14	54.38	-82.07	-34.66047	268.3		
	200-300	31.94	109.37	108.38	-116.8	-72.65703	252.3		
	300-510	42.28	111.63	112.36	-118.7	-89.65035	189.2		
(C ₂₃ H ₂₈ O ₅ ClN ₃ S) ₂ Cr·2H ₂ O	40-130	3.94	29.77	28.68	-24.59	-7.421	284.3	-43.2123	0.94
	130-310	9.14	67.76	66.14	-67.76	-29.94992	263.0		
	310-350	14.47	114.2	113.92	-116.50	-72.4653	252.0		
	350-510	17.82	118.3	116.06	-121.20	-89.5425	203.2		
(C ₂₄ H ₃₄ N ₄ O ₅ S) ₂ Cr·2H ₂ O	40-125	3.28	52.66	52.16	-43.48	-13.12178	269.2	-44.2381	0.99
	125-270	19.91	85.84	85.13	-82.05	-39.54421	269.2		
	270-410	26.67	138.23	135.14	-101.2	-43.09237	257.6		
	410-510	33.11	144.33	148.25	-121.5	-54.23253	246.5		

FC=Freeman and Carroll, SW = Sharp and Wentworth



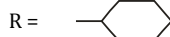
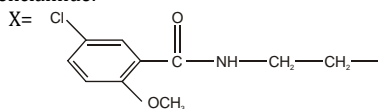
Where,

Gliclazide :

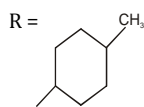
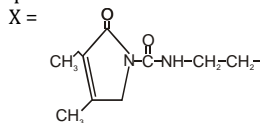
X = CH₃



Glibenclamide:



Glimeperide:



Mass Spectral Studies

The mass spectra of Cr(III) complexes⁵¹⁻⁵⁴ of the ligands Gliclazide, Glibenclamide and Glimeperide peaks attributable to the related molecular ions m/z 722.1[M-2H]²⁺, m/z 1031[M-2H]²⁺, m/z 1042[M-H]⁺ respectively.

The observed free ligand Gliclazide, Glibenclamide and Glimeperide peaks for Cr(III), m/z 324.1 [L+H]⁺, m/z 496 [L+H]⁺, m/z 491.4 [L+H]⁺. The mass spectra of the complexes were given in Fig. 5, 6 and 7. The values in the table which have high abundance were reported in table number 8.

Hypoglycemic Studies

Pharmacology is mainly concerned with the responses of living organisms to chemical stimuli. One may further divide the subject from a medical view point, into pharmacodynamics and pharmacotherapy, the former is concerned with the response of living organism to chemical stimuli in the absence of disease, while the later with the response the organism to such stimuli in a pathogenic state. This is the phase of pharmacology (i.e. pharmacotherapy) which is of special interest to the physician.

Pharmacotherapy includes the treatment of the sick with drugs and therefore is of prime importance in practice of medicine. It is fundamental to the health-economy of the people. A compound or a complex which is to be recommended as a drug of utility, must be capable of easy absorption and excretion. It is also essential that neither the absence itself nor the metabolic products thereof should exercise toxicity or any adverse side effect to the patient⁵⁵⁻⁵⁷.

1. Animal Study- Where necessary such tests should be carried out on animals as rats, rabbits and dogs. When a substance has given satisfactory results for the aforesaid animals then only it may be tried on monkeys and men.

2. Dosage forms- HPMC-5CPS enteric coated granules/pellets in capsule shell.

Composition of enteric coated granules

- | | |
|------------------------|-------------|
| 1. Drug | - 5 mg/dose |
| 2. Lactose | - 100 mg. |
| 3. Starch/sugar sphere | - 100 mg. |
| 4. HPMC- 5CPS | - 10 mg. |
| 5. Water | - q.s. |

Preparation of Enteric Coated Grannules

Make a blend of drug and lactose paste the blend through 100# sieve **HPMC-5CPS** is dissolved in water (2% solution) keep the starch/sugar spheres in conventional coating pan and using HPMC-5CPS solution layering of blend is carried out by conventional method. Pellets prepared, dried in tray dryer at 50°C. Dried pellets are filled in capsule shells.⁵⁸⁻⁶⁰

Hypoglycemic Study on animals(Folin Wu method)⁶⁸

Pharmacological studies were carried out on male albino rats weighing 150 to 200 g. Animals were divided in three groups A, B and C each group containing eleven animals, so selected that the total weight of animals in each group remained the same.

Animals of all the three groups were kept in experimental conditions and were fed on a fixed particular diet (i.e. milk and bread). When animals were acclimatized to the laboratory conditions then fasting blood sugar was estimated colorimetrically (as abridged in Table 1) for four days using Folin and Wu method¹⁷ to ascertain an average blood sugar level. On fourth day doses of Gliclazide, Glibenclamide and Glimeperide were given separately to the animals of group A, B and C respectively (Table 2, 3 and 4).

In case of group (A) animals, after the oral administration of Gliclazide (5.0 mg/kg) fall in blood sugar was noted with increasing duration of time i.e. after ½, 1½, 3½ and 5½ hours.

The peak time of the effect of Gliclazide has been found to be 1 to 2 hours and duration of action to be 4 to 6 hours.

Thereafter animals of group A, were further maintained for normal blood glucose level by way of feeding them on same experimental diet for three more days (i.e. without giving drug or complex). On the 8th day when it was confirmed that all the eleven animals of this groups have returned to their normal blood sugar level, then only the animals were orally administered a dose of Gliclazide-Cr(III) complex (5.0 mg/kg) and fasting blood glucose was recorded again with increasing duration of time (table 2)

Taking fresh groups of animals and using the same procedure, hypoglycemic activity of Glibenclamide and Glimeperide (oral antidiabetics) were also studied and compared with their complexes. In case of Glibenclamide fasting blood sugar was recorded after 2, 4, 6, 8 and 10 hrs. The peak time of the effect of these drugs was proved to be about 4 to 6 hrs. and duration of action to be upto 10 to 12 hrs. After the administration of drugs.

On the days of administration of drugs or its complex the diet was given to the animals after final observation i.e. at 2.0 p.m. to group A animals and 4.0 p.m. to the animals of group B and C (Table 2-4).

Table 11: Colorimetric estimation of blood glucose of male albino rats.

Test Sample	Glucose Standard-I	Glucose Standard-II	Glucose Standard-III
0.05 ml. blood + 3.9 ml. copper reagent + 0.05 ml. sodium tungstate (to coagulate protein) and centrifuge the solution.	0.01% standard glucose solution (I)	0.0025% standard glucose solution (II)	
2.0 ml. supernatant liquid of the sample + 2 ml. Harding's B-solution (NaHCO ₃ + potassium oxalate + sodium tartrate)	2 ml. glucose solution + 2 ml. Harding's B - solution	2 ml. glucose + 2 ml. Harding's B-solution	2 ml. Cu reagent + 2 ml. Harding's B-solution
Sample (a)	Sample (b)	Sample (c)	Sample (d)
$\frac{\text{test reading} \times 80}{\text{Standard (i)reading}} =$ glucose/100 ml. of blood in mg.	II	$\frac{\text{test reading} \times 200}{\text{Standard (ii)reading}} =$ glucose/100 ml. of blood in mg.	

Table 12(a):- Hypoglycemic Activity of Gliclazide Alone

Fasting sugar estimated in mg/100 ml. of blood of albino rats (Group A)(11 Animals)									Least blood sugar values	Fall in blood sugar	% fall in blood sugar	Average % fall in blood sugar
I day	II day	III day	IV day	Average Values	IV day after oral administration of Gliclazide 5 mg/kg							
7.30 am.					8 am	9 am	11 am	1 pm				
99	101	103	98	100.25	71	70	69	72	69	31.25	31.17	33.88
102	99	98	96	98.75	73	72	69	68	68	30.75	31.14	
99	100	101	103	100.75	70	69	72	71	69	31.75	31.51	
99	98	101	102	100	69	65	68	71	65	35	35.00	
96	98	102	103	99.75	62	68	65	75	62	37.75	37.84	
100	102	103	99	101	68	64	72	78	64	37	36.63	
100	102	103	99	101	68	64	72	78	64	37	36.63	

Table - 12(b):- HYPOGLYCEMIC ACTIVITY OF GLICLAZIDE AND ITS CHROMIUM COMPLEX

Fasting sugar estimated in mg/100 ml. of blood of albino rats (Group A)									Least blood sugar values	Fall in blood sugar	% fall in blood sugar	Average % fall in blood sugar
V day	VI day	VII day	VIII day	Average Values	VIII day after oral administration of Gliclazide complex							
7.30 am.					8 am	9 am	11 am	1 pm				
98	102	101	99	100	72	73	68	75	68	32	32.00	33.31
103	101	99	98	100.25	71	69	71	74	69	31.25	31.17	
99	102	98	101	100	71	68	71	78	68	32	32.00	
98	101	100	102	100.25	68	66	65	74	65	35.25	35.16	
98	99	102	101	100	63	68	69	73	63	37	37.00	
100	103	101	99	100.75	69	68	71	73	68	32.75	32.51	
100	103	101	99	100.75	69	68	71	73	68	32.75	32.51	

On IV day drug was administered orally.

On VII day complex was administered orally at 7.35 am.

N.B. All readings are represented in whole number.

Table 13(a): Hypoglycemic Activity Of Glibenclamide Alone

Fasting sugar estimated in mg/100 ml. of blood of albino rats (Group B)									Least blood sugar values	Fall in blood sugar	% fall in blood sugar	Average % fall in blood sugar	
I day	II day	III day	IV day	Average Values	IV day after oral administration of Glibenclamide 5 mg/kg								
6.00 am.					8 am	10 am	12 noon	2 pm	4 pm				
99	103	104	98	101	70	69	68	73	74	68	33	32.67	33.72
102	100	99	97	99.5	70	68	72	74	72	68	31.5	31.66	
101	100	102	103	101.5	70	69	70	72	73	69	32.5	32.02	
100	102	101	101	101	69	68	64	71	75	64	37	36.63	
98	100	99	102	99.75	63	64	69	70	72	63	36.75	36.84	
100	102	103	98	100.75	69	68	72	72	74	68	32.75	32.51	
100	102	103	98	100.75	69	68	72	72	74	68	32.75	32.51	

Table 13(b): Hypoglycemic Activity of Glibenclamide and its Chromium Complex

Fasting sugar estimated in mg/100 ml. of blood of albino rats (Group B)									Least blood sugar values	Fall in blood sugar	% fall in blood sugar	Average % fall in blood sugar	
V day	VI day	VII day	VIII day	Average Values	VIII day after oral administration of Glibenclamide complex								
6.00 am.					8 am	10 am	12 noon	2 pm	4 pm				
100	101	103	99	100.75	71	61	74	79	80	61	39.75	39.45	33.61
103	101	100	98	100.5	70	69	70	76	78	69	31.5	31.34	
101	100	103	99	100.75	70	68	69	70	74	68	32.75	32.51	
101	103	102	102	102	68	69	72	74	76	68	34	33.33	
99	101	102	100	100.5	67	69	68	70	74	67	33.5	33.33	
101	103	101	99	101	69	69	73	74	76	69	32	31.68	
101	103	101	99	101	69	69	73	74	76	69	32	31.68	

On IV day drug was administered orally at 6.10 am.

On VII day complex was administered orally at 6.10 am.

N.B. All readings are represented in whole number.

Table 14(a):- Hypoglycemic activity of Glimeperide alone.

Fasting sugar estimated in mg/100 ml. of blood of albino rats (Group C)										Least blood sugar values	Fall in blood sugar	% fall in blood sugar	Average % fall in blood sugar
I day	II day	III day	IV day	Average Values	IV day after oral administration of Glimeperide 5 mg/kg								
7.00 am.					8 am	10 am	12 noon	2 pm	4 pm				
102	101	98	99	100	61	65	67	70	72	61	39	39.00	
97	98	100	101	99	71	68	69	72	74	68	31	31.31	
99	96	99	100	98.5	63	67	61	70	71	61	37.5	38.07	37.42
99	98	100	101	99.5	69	64	59	64	72	59	40.5	40.70	
98	100	103	102	100.75	64	65	67	61	73	61	39.75	39.45	
101	102	99	98	100	68	64	69	72	78	64	36	36.00	

Table 14(b):- Hypoglycemic activity of Glimeperide and its chromium complex

Fasting sugar estimated in mg/100 ml. of blood of albino rats (Group C)						Least blood sugar values	Fall in blood sugar	% fall in blood sugar	Average % fall in blood sugar				
V day	VI day	VII day	VIII day	Average Values	VIII day after oral administration of Glimeperide complex								
7.00 am.					8 am					10 am	12 noon	2 pm	4 pm
94	92	96	92	93.5	60	52	48	71	72	48	45.5	48.66	
91	90	91	90	90.5	67	50	49	44	74	44	46.5	51.38	
94	93	94	93	93.5	68	52	49	72	44	44	49.5	52.94	53.41
96	90	95	93	93.5	67	50	49	44	71	44	49.5	52.94	
96	100	99	98	98.25	69	53	47	41	73	41	57.25	58.27	
100	99	96	98	98.25	68	52	48	43	71	43	55.25	56.23	

On IV day drug was administered orally at 7.05 am.

On VII day complex was administered orally at 7.05 am.

N.B. All readings are represented in whole number.

As the reported metal complexes of Gliclazide, Glibenclamide and Glimeperide drugs are able to dissociate at stomach pH, therefore its dosages, to be given to subject animals should be such that it should not be dissociate in stomach i.e. at pH 1.2 for this complexes prepared enteric coated to make at the drug bioavailable as it is, i.e. at duodenum and small intestinal pH (5.5 to 6.8 pH)

Drug is coated with a polymer *HPMC-5CPS* (Hydroxy propylmethyl cellulose) which does not permit drug to dissolve in stomach (i.e. pH 1.2) and such polymer dissolves rapidly at dedenal pH (5 above 5.5) thus drug releases at 5.5 pH and is available for absorption. At this pH complex at stable, non-dissociatable and absorbable. Therefore dosage forms for animals study is prepared as enteric coated, polymerized in this dosage forms are not soluble at pH 12. This drug delivery system is adopted for further study⁶⁰⁻⁶⁷

The hypoglycemic effects of Gliclazide, Glibenclamide and Glimeperide the well known sulphonyl ureas, were investigated on the blood sugar levels of male albino rats by using Folin Wu method.⁶⁸ . Analysis of data presented in table 2,3 and 4 would show that all these drugs caused a marked decrease in blood sugar level to the extent of 33.88%, 33.11% and 33.72% while their complexes reduced the blood sugar level to 33.61%, 37.42% and 53.41% respectively.

This blood sugar lowering effect of sulphonyl ureas seems to be related to the stimulation of insulin secretion on the other hand, many studies have strongly indicated the presence of long term or extra pancreatic action of sulphonyl ureas⁶⁹. The hypoglycemic activity of sulphonyl ureas may also be attributed to the stimulation of glycolysis and to the inhibition of glycogenesis in the liver by itself or by enhancing insulin action.

Further, on comparing the hypoglycemic effect of complexes of these sulphonyl ureas in relation to time, it becomes evident from tables 2, 3 and 4 that on the whole the maximum fall in blood sugar was observed after 1½, 6.0 and 5.0 hrs. with the administration of Gliclazide, Glibenclamide and Glimeperide complexes respectively. On comparing the hypoglycemic effect of these complexes with their parent drugs, it was revealed that in the three groups Gliclazide-Cr(III), Glibenclamide-Cr(III) and Glimeperide-Cr(III) treated albino rat had lowest blood sugar level being 49.72 mg/100 ml., 45.45 mg/100 ml. and 53.46 mg/100 ml. respectively on an average. These facts clearly indicate a better hypoglycemic activity of complexes as compared to their parent drugs which is in agreement with the earlier findings of Iqbal and co-worker⁷¹. This improved hypoglycemic activity may be related to smaller particle size of metal complexes than drugs as on

complexation particle size is reduced which may promote the ratio of absorption of complexes in gastro-intestinal tract.

Results of the present work are also in conformity with the hypoglycemic effect of copper-phenformin complex over parent drug phenformin as mentioned by Piccini *et al.*,⁷⁰.

These interesting observations on metal-complexes of oral sulphonylureas used as anti-diabetic agents for lowering blood sugar concentration may likely substantiate the use of these complexes after extensive clinical studies.

REFERENCES

1. Barry Mennen. : Dietry Chromium an overview, executive information Bureau Inc.
2. Reaven G.M., : Role of Insulin Resistance Human disease,Diabetes 37,1595 Dec.(1998)
3. Kaats G.R.,Blum K.,Fisher J.A.,and Adelman J.A. : Current Therapeutic Res. 57(10);747-765,Oct(1996)
4. Pandey R.N.,Rajnish Kumar Singh and Kalpana: Asian J.of Chem.23(6),2739-2741(2011)
5. Louise A.,Berben and Jeffery.R.,Lang :Inorg.Chem.J. 44,8459-8468(2005)
6. Li F,Wu Y,Zou Y,ZhaoT.,Xhang M.,Feng W.,,Yang L: Food Chem. Toxicol 50(5),1623-1631 Epub.Feb-18(2012)
7. Yang X,Palanichamy K.,Onto A.C.,Rao M.N.,Fang C.X.,J,and Sreejayan N. : FEBS Lett. Feb, 28,579(6),1458-1464(2012)
8. Yashinaga I.,Yammamoto Y. : E ndocrinologie (Gen.), 50,3(1966a)
9. Yashinaga I.,Yammamoto Y. :J.Osaka 1,3(1966b)
10. Iqbal, S.A., George J.,and Zaafarany. : Jou of Saudi Chem Soc. 14; 345-350 (2010)
11. Dury and Al-Jibori. : Orient J. Chem. 28(2), 781-786 (2012)
12. Shahriare Ghammamy: Orient J. Chem. 28(2), (2012)
13. Modhavadiya V.A. : Orient J. Chem. 28(2), 921-925 (2012).
14. Abdul Vudood., Manish Kumar and Saxena P.N. : Orient J. Chem. 28(2), 1019-1023 (1012)
15. Sunil Kumar and Sharma T.R. : Orient J. Chem. 28(2), 963-967 (2012).
16. Iqbal.S.A., George J. and Zaafarany, I. : Journal of Saudi Chemical Society,14,345-350(2010)
17. Singel,H.(Ed.),Dekker: Metal Ions in Biological SystemsVol-14(1982)
18. Brown D.A., : Metal Ions in Biological SystemsVol-14,125,(1982)
19. Phipps,D.A. : Metals and Metabolism, Oxford University Press p.63(1976)

20. Williams, D.R.: An Introduction to Bioinorganic Chemistry C.C.Thomas, p.32
21. Lippard: Platinum, Gold and other metals as chemotherapeutic Agents, American chemical Society (1983)
22. Meares, C.F., Wensel, T.G. : Metal chelates as probes of biological system. Acc.Chem.Res. 17, 202-209 (1984)
23. Irving H, Rossotti H.S. : J.Chem.Soc. 3397 (1953)
24. Irving H, Rossotti H.S. : J.Chem.Soc. 2904 (1954)
25. Irving H, Rossotti H.S. : J.Chem.Soc. 1176 (1955)
26. Willard, H., Merritt, L.L. and Dean J.A. : Instrumental methods of Analytical, 5th Ed Pearson Education publishers, Singapore, (1975)
27. Jeffery, G.H., Bassett, J., Mendham, Denney R.C. : Vogels Text book of Quantitative chemical Analysis 5th Ed. Pearson Education Publisher, Singapore, p-473.
28. Mohammed, Ashra : Talenta, 15, 559-562 (1989)
29. Scott, A. : Standard methods of chemical Analysis. Von Nostrand, p-634 (1960)
30. Cullity B.D. : Elements of X-ray diffraction 2nd Ed. Addison Wesley, publishing company, Inc. (1978)
31. Bragg, W.L. and Bragg, W.H. : The crystalline state, Chemical Heritage foundation. A general survey, vol.1 London (1952)
32. Guinier, Andrey: X-ray Crystallographic Technology I. Liger and Watts London (1993)
33. Henry N.F.M., Lipson H. and Wooster W.A. : The Interpretation of X-ray diffraction photographs, Mc Millon London (1951)
34. Madhusudhyan P.M., Krishan, K and Ninan K.N. : Thermochem. chem. Acta 97; 189 (1986)
35. Singh S. Kumar, V. Sharma S.K., and Kaumar A. : Orient. J. Chem. 26(1) 93-101 (2010).
36. Janabi, A.S., Saumadaiy, G.A., and Jhear-Allah, B.A. : Orient J. Chem. 27(4), 1465-73 (2011)
37. Tamani, B., Yangesh H. and Koohmarch G.A. : Iran. Polym. J. 14(5), 785-792 (2005)
38. Yangesh, H. and Ataei, S.M. : Iran. Polym. J. 14(5), 449-455 (2005)
39. Weisberger A. : Chemical application of spectroscopy, Interscience publishers, NY. (1956)
40. Cotton F.A. : Modern co-ordination chem.. Interscience pub. Ed. (1960)
41. Nakamoto, K: Infra-red spectra of Inorganic and co-ordination compounds, John Wiley and son's NY. (1963)
42. Rao, C.N.R. : Chemical Applications of Infra-red spectroscopy, Academic press NY. (1963)
43. Bellamy L.J. : The Infra-red spectra of complex molecules. Matheun and co. Ltd. London (1964)
44. Dyer J.R. : "Application of Absorption spectroscopy of organic compounds" Prentice Hall of India Pvt. Ltd (1966)
45. Slichter C.P. : Principles of magnetic Resonance, Harper and Row, (1963)
46. Akitt J.W. : NMR and chemistry; An Introduction to Nuclear magnetic resonance Spectroscopy Chapman and Hall, (1973)
47. Siewers R.W. : NMR Shift Reagents Academic New York, (1973)
48. Dikshit, D.D. and Singh, S.B. : Orient J. Chem. 26(1), 171-74 (2010).
49. Pandey, R.N., Nag, A.K., Pande, Prasashti and Singh S.K. : Orient J. Chem. 26(1), 109-12 (2010).
50. Cotton F.A., Wilkinson G., Murillo C.A., Bochmann M: Advanced Inorganic chemistry, 6th Ed. Wiley, New York, (1999)
51. Furniss B.S., Hannaford A.J., Smith P.W.G. and Jatchell A.R. : Vogels Text book of practical organic chemistry 5th Ed., 361-833 (1998)
52. Mendham J., Denney R.C., Barnes J.D., and Thomas M.J.K: Vogels text book of quantitative chemical analysis, 6th Ed. 745-790 (2007)
53. Basavaiah K. and Prameela H, C. Sultan M. : II Farmaco 57, 443-449 (2002)
54. Satyanand Tyagi, Saching Kumar, Amit Kumar, and Mohit Singla: II Farmaco " Acyclovir Aug. 24, Nationmaster. Com (2004) IJPWR 1(2), (MAR-JUN) (2010)
55. A. Shcroft F.W., Gribble F.M. : Journal of Diabetes and its complications; 14, 192-196 (2000), British Journal of Pharmacology, 133, 193-199 (2001)
56. Song D.K., Asharafa F.W. : Pharm. Acta. Helv. 60, 110-111 (1985)
57. Peppas N.A. : Biopharmaceutics and Pharmacokinetics, A treatise 1st New Delhi Vallabh prakash, p-58 (1995)
58. Brahmankar D.M. and Jaiswal S.B. : S.J. Pharm. Sci. 2(2), 53-58 (2009)
59. Muhammed Shahidul Islam, Moniruzzaman, Rukhuzzaman: Biochem. 1(43), 463 (1951)
60. Sanger P, and Tuppi F. : J. Org. Chem, 23, 927 (1958)
61. Marshal F.J. Sigal M.V. Idem: 43rd Congress of Medicine and Neurology Montpellier 415 (1942)
62. Frank H., and Fuchs J. : Ein neues Antidiabetisches Prinzip, Dentsch, Med. Wschr
63. Maske H. : Deut. Med. Wochschr, 81, 823 (1956)
64. Mc. Lamore W.M., Fanelli G.M. and Pan S.Y. : Ann. NY, Acad. of sci. 74, 443 (1959)
65. Varshosaz J., Tabbakhian M. and Zahrooni M. : Journal of Microcapsulation 24(3), 253-262 (2007)
66. Folin O. and Wu H. : J. Biol. Chem. 41, 367 (1920)
67. Anturlikar S.D., Chauhan B.L. and Mitra S.K. Indian J. of Physiology and Pharmacology 39, 95-100 (1995)
68. Otutu, J.O., Osabohien, E., and Efurhievwe, E.M. Orient J. Chem. 27(4), 1389-96 (2011).
69. Qureshi R., and Iqbal S.A: Indian J. Applied and pure Biol. 2(2), 65-67 (1987)
70. Piccinni F., Murazzi E., Uberti and Lucatelli I. : Pharmaco (pavia) Ed. sci. 15, 521 (1960).