ASIAN JOURNAL OF PHARMACEUTICAL AND CLINICAL RESEARCH



Vol 6, Issue 3, 2013 ISSN - 0974-2441

Research Article

LARVICIDAL POTENTIAL OF SOME INDIAN MEDICINAL PLANT EXTRACTS AGAINST AEDES AEGYPTI (L.)

M. S. SHIVAKUMAR¹,* R. SRINIVASAN² AND D. NATARAJAN²

Department of Biotechnology, Periyar University, Salem 636-011, Tamil Nadu, India. Email: skentomology@gmail.com

Received: 1 February 2013, Revised and Accepted: 3 March 2013

ABSTRACT

The larvicidal potential of different solvent crude (hexane, chloroform, ethyl acetate, acetone and methanol) leaf extracts of four plants (*Blepharis maderaspatensis, Elaeagnus indica, Maesa indica, Phyllanthus wightianus* and *Memecylon edule*) was tested against the fourth-instar larvae of *Aedes aegypti*. Insecticidal susceptibility tests were carried out using WHO standard method and the mortality was observed after 24-h exposure. All the tested extracts showed moderate to good larvicidal activities. However, the maximum larval mortality was detected in acetone extract of *E. indica* (LC_{50} 90.89, LC_{90} 217.21 and LC_{99} 441.88 ppm) followed by *M. indica* acetone extract (LC_{50} 173.21, LC_{90} 289.86 and LC_{99} 441.04 ppm). These results revealed that larvicidal properties of the selected plants and encourages further effort to investigate the bioactive compounds in those extracts that might possess good larvicidal properties when it will be isolated in pure form.

Keywords: Blepharis maderaspatensis, Elaeagnus indica, Maesa indica, Phyllanthus wightianus, Memecylon edule, dengue vector mosquito.

INTRODUCTION

Mosquitoes are vector for various disease including malaria, yellow fever, filariasis Japanese encephalitis and chikungunya. Among these mosquito borne diseases dengue fever dengue hemaorrhagic fever, yellow fever and chikungunya are endemic in Southeast Asia and Africa[1]. It is transmitted by *Aedes aegypti* (Linn.). One of the methods available for controlling the mosquitoes is use of synthetic insecticides. Mosquitoes develop genetic resistance to synthetic insecticides[2] and even to biopesticides such as *Bacillus sphaericus*[3]. Also synthetic insecticides adversely affect the environment by contaminating air, water, and soil. There is a urgent need to find alternatives to the synthetic insecticides which is more potent and low-cost.

Plants are rich source of alternative agents for control of mosquitoes, because they possess bioactive chemicals, which act against limited number of species including specific target-insects and are eco-friendly[4]. Traditionally plant based products have been used in human communities for many centuries for managing insects. Several secondary metabolites present in plants serve as a defense mechanism against insect attacks. These bioactive chemical may act as insecticides, antifeedants, moulting hormones, oviposition deterrents, repellents, juvenile hormone mimics, growth inhibitors, antimoulting hormones as well as attractants. Plant based pesticides are less toxic, delay the development of resistance because of its new structure and easily biodegradable[5].

Several plant extracts and isolated compounds from different plant families have been evaluated for their promising larvicidal activities[6]. About 2000 species of terrestrial plants have been reported for their insecticidal properties [7]. Search for eco-safe, low cost and a highly potential insecticide for the control of mosquitoes needs the preliminary screening of plants to evaluate their insecticidal activities.

Plant based products does not have any hazardous effect on ecosystem. Recent research has proved that effectiveness of plant derived compounds, such as saponine[8], steroids[9][10], isoflavonoids[11], essential oils[12], alkaloids and tannins[13] has potential mosquito larvicides. Plant secondary metabolites and their synthetic derivatives provide alternative source in the control of mosquitos[14].

The present investigation was carried out to validate the larvicidal potential of different solvent extracts of four (Blepharis maderaspatensis (L.) B. Heyne ex Roth., Elaeagnus indica Servett., Maesa indica (Roxb.) DC, Phyllanthus wightianus Müll.Arg. and Memecylon edule Roxb.) medicinal plants against fourth instar Ae. aegypti larvae. All the plants were selected based on their ethnobotanical literatures and least explored. This is first hand report on larvicidal activity of all the selected plants against Ae. aegypti larvae.

MATERIALS AND METHODS

Plant material

Healthy leaves of *B. maderaspatensis* (Acanthaceae), *E. indica* (Elaeagnaceae), *M. indica* (Myrsinaceae) *P. wightianus* (Phyllanthaceae) and *M. edule* (Melastomataceae) were collected from various regions of Eastern Ghats of Tamil Nadu, India. The plants were identified with the references of standard books and herbariums from the Natural Drug Research Laboratory (NDRL), Department of Biotechnology, Periyar University, Salem, India. The plant materials were cleaned, air-dried at room temperature for two weeks and coarsely powdered.

Preparation of extracts

Powdered plant materials were extracted successively by using different solvents of increasing polarity (hexane, chloroform, ethyl acetate, acetone and methanol) in soxhlet apparatus for 18 h and the extractives were filtered through Whatman filter paper No. 4 then the extracts were concentrated at 40° C in vacuum and stored at 4° C for this investigations.

Test insects

Ae. aegypti, larvae was obtained from National Centre for Disease Control (NCDC) Coonoor, Tamil Nadu and maintained at Department of Biotechnology, Periyar University Salem. Larvae were fed a diet of Brewer's yeast and powdered dog biscuits in the ratio of 3:1, kept at $27 \pm 2^{\circ}$ C and 75% - 85% relative humidity (RH), with a photoperiod of 14:10 LD for the larval growth. Late third instars to early fourth instars larva were used for larval bioassay which obtained from the stock culture maintained at Department of Biotechnology, Periyar University, Salem.

Larvicidal bioassay

The larvicidal activity of crude extracts of the selected plants was assessed by the protocol of WHO[15] with some modifications and as per the method of Rahuman $et\ a[16].$ For the bioassay in a container 25 fourth instar larvae were kept in 249 ml of distilled water with 1 ml of extracts (400 ppm) in DMSO. Tween-80 was used as an emulsifier at concentration of 0.02% (v/v). The chamber containing the control larvae received 1 ml of DMSO served as negative control. After 24 hours exposures the dead larvae were counted and corrected by using Abbott's[17] formula and the percentage mortality was recorded from the average of six replicates.

Dose-response bioassay

Based on the preliminary screening results, in which above 90% mortality of larvae occurs alone, were subjected to dose-response larvicidal bioassay. The desired mortality percentage was observed in acetone and ethyl acetate extracts of *E. indica*, ethyl acetate extract of *B. maderaspatensis* and acetone extract of *M. indica* at 400 ppm concentration were subjected to dose dependent bioassay. Different concentrations (50-400 ppm) of the above mentioned crude extracts were tested for larvicidal activity described by

WHO[15]. The average mortality percentages of six replicates were recorded and corrected by using Abbott's formula.

Statistical analysis

Data were analyzed using one-way ANOVA. Significant differences between treatments were determined using Tukey's multiple range tests (P \leq 0.05). LC₅₀, LC₉₀ and LC₉₉ values were calculated using probit analysis¹⁸.

RESULTS AND DISCUSSION

The results of larvicidal efficacy of different solvent extracts of the selected plants were shown in Table 1. All the plant extracts showed good to moderate effect on fourth instar larvae of Ae. aegypti after 24 h of exposure at 400 ppm (0.04%) concentration. The highest mortality (100%) was observed in acetone extracts of E. indica and M. indica. Significant (p>0.05) activity was detected in ethyl acetate extracts of E. indica (97%) and E. indica E. indica (90%) followed by E. indica chloroform extract (85%). Most of the extracts of E. indica E.

 $Table \ 1: Larvicidal \ activity \ of \ different \ solvent \ leaf \ extracts \ of \ selected \ plants \ against \ 4^{th} \ instar \ larvae \ of \ \textit{Ae. aegypti} \ at \ 400 \ ppm \ (0.04\%)$

Plant names	% Mortality*							
	Methanol	Acetone	Ethyl acetate	Chloroform	Hexane			
B. maderaspatensis	8.0±1.0 a	48.0±6.0	90.6±0.5 a	26.6±2.3	10.6±1.1 a			
E. indica	22.6±2.0 b	100±0.0 b	97.3±0.5 ba	21.3±0.5	34.6±1.1			
M. indica	24.0±2.0 cb	100±0.0 cb	14.6±1.5 c	85.3±1.5 c	6.6±3.0 ca			
M. edule	5.3 ± 1.5^{da}	4.00 ± 0.0	10.6±0.5 cd	1.3±0.5	17.3±0.5			
P. wightianus	42.6±1.1	73.3±1.5	78.6±2.0	82.6±1.1 ec	70.6±2.0			

Control—Nil mortality, Total no of larvae =25, *-Mean value of six replicates ± SD. Significant at p>0.05 level.

The toxicity of dose–response larvicidal bioassay was given in Table 2. According to preliminary screening results, four extracts were subjected to dose–response larvicidal bioassay which has above 90% larval mortality. Among them significant mortality rate was

observed in acetone extract of E. indica with LC₅₀ LC₉₀ and LC₉₉ values of 90, 217 and 441 ppm respectively followed by acetone extract of M. indica with LC₅₀ LC₉₀ and LC₉₉ values of 173, 289 and 441 ppm respectively. The larvicidal activity of the different selected plant extracts were found to be dose depended. E. indica ethyl acetate extract shows considered mortality with LC₅₀ LC₉₀ and LC₉₉ values of 151, 456 and 1121 ppm respectively.

Table 2: Dose-response larvicidal bioassay of different solvent leaf extracts against 4th instar larvae of A. aegypti

Plant names	Extracts	Conc. (ppm)	% Mortality*	LC ₅₀ ± SE (ppm) (LCL-UCL)	LC ₉₀ ± SE (ppm) (LCL-UCL)	LC ₉₉ ± SE (ppm) (LCL-UCL)	χ ^{2 (df=4)}
B. maderaspatensis	Ethyl acetate	100	18.6 ± 0.5	197.6 ± 0.2 (181.6-213.8)	438.0 ± 0.3 (381.6-531.3)	838.3 ± 0.8 (664.3-1174.3)	4.9
		150	30.6 ± 0.5				
		200	42.6 ± 2.0				
		250	62.6 ± 1.5				
		300	77.3 ± 1.5				
		400	90.6 ± 2.5				
E. indica	Acetone	50	24.0 ± 1.0	90.8 ± 0.1 (80.1-101.1)	217.2 ± 0.2 (191.5-254.8)	441.8 ± 0.6 (358.7-587.5)	5.2
		100	50.6 ± 0.5				
		150	70.6 ± 1.1				
		200	89.3 ± 3.7				
		300	97.3 ± 1.1				
		400	100.0 ± 0.0				
E. indica	Ethyl acetate	50	18.6 ± 1.5	151.2 ± 0.4 (93.3-224.2)	456.1 ± 0.5 (284.6-2005.6)	1121.7 ± 1.2 (527.0-16044.2)	20.0
		100	26.6 ± 1.1				
		150	41.3 ± 3.5				
		200	52.0 ± 3.6				
		300	80.0 ± 4.0	,			
		400	97.3 ± 0.5				
M. indica	Acetone	100	16.0 ± 1.0	173.2 ± 0.7 (135.9-206.9)	289.8 ± 0.9 (237.2-448.8)	441.0 ± 2.2 (325.6-968.0)	17.8
		150	30.6 ± 1.5				
		200	52.0 ± 2.6				
		250	78.6 ± 3.5				
		300	98.6 ± 0.5				
		400	100.0 ± 0.0				

Control- Nil mortality, Significant at p<0.001 level, *-Mean value of six replicates \pm SD, LC= Lethal Concentration, LCL=Lower Confidence

Limit, UCL=Upper Confidence Limit, SE=Standard Error, $\chi 2$ =chi-square and df=degree of freedom.

Nowadays, the control of mosquitoes at larval stage is focused with plant extracts. The advantage of targeting mosquito at the larval stage is they cannot escape from their breeding sites until the adult emergences and also to reduce the overall pesticide use to control of

adults by aerial application of adulticidal chemicals. Bioactive crude extracts or isolated phyto-constituents from the plant could be used as alternative to the currently used synthetic insecticides. The biological activity of plant extracts might be due to various compounds, including phenolics, terpenoids, and alkaloids present in plants[19].

Our preliminary screening for larvicidal properties of different solvent leaf extracts of four plants among 20 extracts, 4 extracts gave high larvicidal potency with low lethal concentrations (LC₅₀ <197 ppm) against 4th instar larvae of Ae. aegypti. Likewise, Cavalcanti et al[12] reported that the larvicidal activity of essential oils of Brazilian plants against Ae. aegypti and observed the LC₅₀ to range from 60 to 533 ppm. Similarly, Rahuman and Venkatesan²⁰ screened the petroleum ether extracts of Citrullus colocynthis; methanol extracts of Cannabis sativus, Cannabis indica and Momordica charantia; and acetone extract of Trichosanthes anguina against the larvae of Ae. aegypti the LC₅₀ values are 74.57, 309.46, 492.73, 199.14, and 554.20 ppm respectively which supports the present results were comparably good.

The findings of present results, among the 25 tested extracts only 4 extracts has potential larvicidal activity which are comparable to earlier reports of Sakthivadivel and Daniel[21] that screened larvicidal activity of petroleum ether extracts of sixty three plants against *Cu. quinquefasciatus, An. stephensi* and *Ae. aegypti* larvae of which six found to be potential larvicides. Similarly, Pavela[22] reported the larvicidal activity methanolic extracts of thirty one Euro-Asiatic plants against *Cu. quinquefasciatus*. Likewise, Nazar *et al*[23] investigated 100 coastal plant extracts including *B. maderaspatensis* against the *Cu. quinquefasciatus* larvae of which seventeen plants were possessed larvicidal properties and also the whole plant extract of *B. maderaspatensis* showed no activity but, the present investigation revealed that larvicidal properties of *B. maderaspatensis* against *Ae. aegypti*.

The findings of present study are quite comparable with previous reports of Vinayaka et~al[24] who have reported the larvicidal activities of different solvent leaf extracts of Elaeagnus~kologa in which methanol, ethyl acetate and acetone extracts showed 100% in 15 and 20 mg/ml concentrations against Ae.~aegypti. Suwanneepromsiri et~al[25] investigated fourteen plant extracts, among those, only eight plants were showed 100% mortality against Ae.~aegypti larvae at a concentration of $100~\mu g/ml$ with LC_{90} values range between 13.9- $56.2~\mu g/ml$ to $100~\mu g/ml$ that supports present results.

The present result was supported by earlier reports of Singh et al[26] that the larvicidal activity of $Ocimum\ canum$ oil tested against $Ae.\ aegypti$ and $Cu.\ quinquefasciatus$ (LC50 301 ppm) and $An.\ stephensi$ (234 ppm). Similarly, Ansari $et\ al[27]$ was observed the larvicidal activity of $Pinus\ longifolia$ oil against $An.\ stephensi$ (LC50 112.6 ppm), $Ae.\ aegypti$ (82.1 ppm) and $Cu.\ quinquefasciatus$ (85.7 ppm). The results of our study is compared with the findings of Sumroiphon $et\ al^{28}$ who have reported that the effect of water extract of citrus seed extract showed LC50 values of 135, 319 and 127, 411 ppm against the larvae of $Ae.\ aegypti$ and $Cu.\ quinquefasciatus$ respectively.

CONCLUSION

All the tested plants possessed different range of larvicidal property which may be used as a traditional mosquito control agent. On the basis of the present investigation results we conclude that acetone, ethyl acetate extract of *E. indica*, acetone extract *M. indica* and ethyl acetate extract of *B. maderaspatensis* contains potent larvicidal bioactive principles which may be needed further purifications to have its synthetic analogues which will be carry out in future.

ACKNOWLEDGEMENTS

The authors are gratefully thank to the University Grants Commission (UGC), New Delhi (Ref. No. 37-296 / 2009 (SR)) for

financial support and Department of Biotechnology, Periyar University, Salem for providing necessary facilities.

REFERENCES

- Maillard M, Marston A, Hostettmann K, Search for molluscicidal and larvicidal agents from plants in Baladrin M: Human Medicinal Agents from Plants. American Chemical Society. Washington DC, 1993; 534: 256-273.
- Wattal BL, Joshi GC, Das M, Role of agriculture insecticides in precipitating vector resistance. *J Communicable Diseases*, 1981; 13: 71-73.
- 3. Tabashnik B. E, Evolution of resistance to *Bacillus thuringiensis*. *Annual Review of Entomol*, 1994; 39: 47-79.
- Sukumar K, Perich MJ, Boobar LR, Botanical derivatives in mosquito control: A review. J Amer Mosquito Control Association, 1991; 7: 210-237.
- Ignacimuthu S, The root of botanicals in combating mosquitoes. Abstracts: Proceedings of symposium on recent trends in combating mosquitoes, Loyola College, Chennai, India, 19, (2000).
- Markouk M, Bekkouche K, Larhsini M, Bousaid M, Lazrek HB, Jana M, Evaluation of some Moroccan medicinal plant extracts for larvicidal activity. *J Ethnopharmacol*, 2000; 73: 93-297.
- 7. Feinstein L, *Insecticides from plants*. In: Insects: The year book of agriculture, USA, Washington, 1952; 222-229.
- 8. Wiseman Z, Chapagain BP, Larvicidal effects of aqueous extracts of *Balanites aegyptiaca* (desert date) against the larvae *of Culex pipiens* mosquitoes. *Afr J Biotechnol*, 2005; 4 (11): 1351- 1354.
- Chowdhury N, Ghosh A, Chandra G, Mosquito larvicidal activities of Solanum villosum berry extract against the dengue vector Stegomyia aegypti. BMC Complement Altern Med, 2008; 8: 10.
- Ghosh A, Chowdhury N, Chandra G, Laboratory evaluation of a phytosteroid compound of mature leaves of day jasmine (Solanaceae: Solanales) against larvae of *Culex quinquefasciatus* (Diptera: Culicidae) and non-target organisms. *Parasitol Res*, 2008; 103: 221-277.
- 11. Joseph CC, Ndoile MM, Malima RC, Nkunya MH, Larvicidal and mosquitocidal extracts, a coumarin, isoflavonoids and pterocarpans from *Neorautanenia mitis. Trans R Soc Trop Med Hyg*, 2004; 98 (8): 451-455.
- 12. Cavalcanti ESB, Morais SM, Lima MAA, Santana EWP, Larvicidal activity of essential oils from Brazilian plants against *Aedes aegypti* L. *Mem Inst Oswaldo Cruz*, 2004; 99: 541-544.
- Khanna VG, Kannabiran K, Larvicidal effect of Hemidesmus indicus, Gymnema sylvestre, and Eclipta prostrata against Culex qinquifaciatus mosquito larvae. Afr J Biotechnol, 2007; 3: 307-311.
- 14. Yang YC, Le EH, Lee HS, Lee DK, Ahn YJ, Repellency of aromatic medicinal plant extracts to *Aedes aegypti*. *J Am Mosq Control Assoc*, 2004; 20 (2): 146-149.
- World Health Organization. Guidelines for laboratory and field testing of mosquito larvicides. WHO/CDS/WHOPES/GCDPP/ 2005.13. Geneva: WHO, 2005; 9.
- 16. Rahuman AA, Gopalakrishnan G, Ghouse BS, Arumugam S, Himalayan B, Effect of *Feronia limonia* on mosquito larvae. *Fitoterapia*, 2000; 71: 553-555.
- 17. Abbott WS, A method of computing the effectiveness of an insecticide: *J Econ Entomol*, 1925; 18: 265-266.
- Finney DJ, Probit analysis, 3rd edn, New York: Cambridge University Press, 1971; 68–72.
- Rafael L, Teresinha N, Moritz JC, Maria IG, Eduardo MD, Tania SF, Evaluation of Antimicrobial and Antiplatelet Aggregation Effect of Solidago chilensis eyen. Int J Green Pharm, 2009; 3: 35-39.
- Rahuman AA, Venkatesan P, Larvicidal efficacy of five cucurbitaceous plant leaf extracts against mosquito species. Parasitol Res, 2008; 103 (1): 133–139.
- 21. Sakthivadivel M, Daniel T, Evaluation of certain insecticidal plants for the control of vector mosquitoes viz, *Culex quinquefasciatus*, *Anopheles stephensi* and *Aedes aegypti*. *Applied Entomol Zool*, 2008; 43 (1): 57-63.
- 22. Pavela R, Larvicidal activites of some Euro-Asiatic plants against *Culex quinquefasciatus* Say (Diptera: Culicidae), *J Biopesticides*, 2008; 1: 81-85.

- 23. Nazer S, Ravikumar S, Williams PG, Syed Ali M, Suganthi P, Investigated a hundred coastal plant extracts against the *Culex quinquefasciatus* larvae of which seventeen coastal plants were posses larvicidal potential, *Indian J Sci Technol*, 2009; 2 (3): 24-27.
- 24. Vinayaka KS, Prashith Kekuda TR, Nethravathi HR, Thippeswamy NB, Sudharshan SJ, Free radical scavenging and insecticidal activity of *Elaeagnus kologa* Schldl. *Drug Invention Today*, 2009; 1 (1): 74-77.
- 25. Suwanneepromsiri, Amaranaksathit, Maleeya, Kruatrachue, Usavadee Thavara, Evaluations of larvicidal activity of medicinal plant extracts to *Aedes aegypti* (Diptera: Culicidae) and other effects on a non-target fish. *Insect Sci*, 2006; 13: 179-188.
- 26. Singh NP, Kumari V, Chauhan D, Mosquito Larvicidal proberties of the leaf extract of a Herbaceous Plant, *Ocimum canum* (Family: labiatae). *J Commun Dis*, 2003; 35 (1): 43-45.
- 27. Ansari MA, Mittal PK, Razdan RK, Sreehari U, Larvicidal and mosquito repellent activities of pine (*Pinus longifolia*, Family: Pinaceae) oil. *J Vector Borne Dis*, 2005; 42: 95–99.
- 28. Sumroiphon S, Yuwaree C, Arunlertaree C, Komalamisra N, Rongsriyam Y, Bioactivity of citrus seed for mosquito-borne diseases larval control, *Southeast Asian J Trop Med Public Health*, 2006; 37 (3):123–127.