ABSTRACT

The objective of the present investigation was to search for an effective natural release retardant that can be used as an alternative source for synthetic excipients in tablet formulation. Tamarind seed polysaccharide (TSP) was isolated and the yield was found to be 76.66%. The microbiological studies confirmed that TSP does not support microbial growth. Sustained release tablets of Metformin hydrochloride were prepared using the isolated TSP. The release profile of the prepared tablets were compared with the formulations prepared with HPMC K 4M and a marketed product. The drug content of all the formulations ranged from 99.74 to 101 %. Formulation F7 showed release up to 11 h which was similar to the marketed product. The results indicated that a decrease in release rate of the drug was observed by increasing the polymer concentration. The release data when fitted into various kinetic equations revealed that the drug release was by matrix and Peppas model release with R² values above 0.9. Stability studies of formulation F7 was carried out and was found to be stable with no change in physical appearance and minimal decrease in drug content. Thus, a stable, safe natural release retardant could be isolated from tamarind seed polysaccharide, which can be effectively used in tablet formulation.

Keywords: Tamarind seed polysaccharide; Metformin hydrochloride; HPMC K 4 M; Sustained release, Drug delivery.

INTRODUCTION

Administration of drug by oral route is one of the important methods to impart systemic effects. Over the years research is going on for identification and utilisation biocompatible polymeric material from natural sources in development of oral sustained release dosage forms. Plant polysaccharide, has shown to be useful for the development of drug delivery systems for specific drug delivery. Among hydrophilic polymers, polysaccharides which are relatively complex carbohydrates are the choice material due to their non toxicity. They provide good mechanical properties for applications as fibres, films, adhesives, rheology modifiers, hydrogels, emulsifiers and drug delivery agents. Such polysaccharides are also called texture modifiers wherein they provide appropriate texture to the products. Being natural they have certain drawbacks like purity, source and microbial contamination. If these factors can be identified and controlled, polysaccharides can be good substitute for synthetic polymer. Tamarind seed polysaccharide (TSP) which is a glucosaminoglycan derivative extracted from the kernel of seeds of Tamarindus indica Linn. Family Leguminoseae. It is a polymer that consists of cellulose type spine that carries xylose and galactoxylate substituents.

The polysaccharide constitutes about 65% of the seed component and is neutral, hydrophilic polymer. Tamarind seed polysaccharide possesses properties like high viscosity, broad pH tolerance, nanocarcinogenicity, mucoadhesive nature, and biocompatibility. It is used as stabilizer, thickener, gelling agent and binder in food and pharmaceutical industries. It is a branched polysaccharide with a main chain of β-[d-(1,4)-linked glucopyranosyl units, and that a side chain consisting of a single d-xylopyranosyl unit attached to every second, third, and fourth d-glucopyranosyl unit through a β-[d-(1,6) linkage. One d galactopyranosyl unit attaches to one of the xylopyranosyl units through a β-[d-(1, 2) linkage. The molecular weight of tamarind seed polysaccharide (TSP) is within the ranges of 2.5×10³ to 6.5×10⁵. Metformin hydrochloride is 1, 1-dimethyl biguanide hydrochloride, which is white, crystalline; hygroscopic powder and used as an oral hypoglycaemic agent specifically for type-2 diabetes mellitus. It is given in divided doses of 0.5–3g daily. Metformin controls blood glucose levels in persons with type 2 diabetes by reducing glucose production in the liver and increasing sensitivity to existing insulin. Metformin is slowly and incompletely absorbed from the gastrointestinal tract; the absolute bioavailability of a single 500 mg dose is 50 to 60% and reduced when taken with food. The plasma elimination half-life is about 2 to 6 h after oral doses. Its pH is 6.68 and pKa is 12.4. The aim of present study is to design sustained release matrix tablet of Metformin hydrochloride using tamarind seed polysaccharide as release retardant.

MATERIALS AND METHODS

Materials

Tamarind kernel powder was purchased from Sri Balaji industries, Bangalore. Metformin hydrochloride was purchased from Yarrow Chem Products, Mumbai. Dicalcium phosphate, magnesium stearate, and purified talc were purchased from S.D. Fine Chemicals, Mumbai, India. Absolute ethanol was purchased from Central Drug House, Mumbai. All other chemicals used were of analytical reagent grade.

Isolation of TSP

To 30 g of tamarind kernel powder, 200 ml of cold distilled water was added to prepare slurry. This slurry was added to 800 ml of boiling distilled water. The solution was boiled for 20 mins in a water bath with continuous stirring. The resulting clear thin solution was kept overnight. The solution was centrifuged at 5000 rpm for 20 mins to separate the foreign matter. The supernatant liquid was separated and poured into twice the volume of absolute ethanol by continuous stirring. The precipitate was collected using muslin cloth and was washed with absolute ethanol. The product was then dried at room temperature for 24 h.

Evaluation of isolated TSP powder

Phytochemical examination

Preliminary tests were performed to confirm the presence of polysaccharide by ruthenium red test and Molisch’s test.

Micromeritic properties of TSP

Isolated TSP powder was evaluated for bulk density, tap density, bulkiness, angle of repose, Hausner’s ratio and Carr’s index.

Specific gravity of 1% solution

A clean and dry empty specific gravity bottle was weighed then the bottle was completely filled with distilled water and weighed. After cleaning and drying, the bottle was completely filled with 1% TSP solution and weighed.
Solubility studies
The solubility of TSP was checked using different solvents. 50 mg of TSP powder was dissolved in 5 ml of solvents like water, ethanol, acetone, methanol and ether.

Determination of melting point
Melting point was determined by a capillary tube in melting point apparatus.

Loss on drying
LOD was carried out as per the method mentioned in Indian Pharmacopoeia[12].

Total tablet weight
Dicalcium phosphate, TSP, HPMC K4M, Metformin hydrochloride, TSP was subjected to IR spectral studies using FTIR spectrophotometer (FTIR 8400 S, Shimadzu, Japan) at the scanning range of 4000-400 cm⁻¹.

pH of 1% solution
The pH of the 1% TSP solution was measured using a digital pH meter.

Moisture content
The moisture content of the powder was carried out by thermogravimetric method using IR moisture balance.

Viscosity of 1% solution
The viscosity of 1% solution was determined using Brookfield viscometer using spindle no.2 (Brookfield, LVDVIT) at 5,10,20,50 and 100 rpm.

Determination of swelling index
Swelling index was determined in water, 0.1N HCl and phosphate buffer pH 6.8. 1 g of TSP powder was introduced into a 25 ml glass stopped measuring cylinder; 25 ml of solvent was added. The mixture was shaken thoroughly at every 10 min interval for 1 h and allowed to stand for 3 h at room temperature. The volume occupied by the material was measured[13].

Particle size analysis
The particle size of TSP powder was determined using optical microscope (Olympus Lite image). A total of 100 particles were counted and the average particle size in micrometers was determined.

Determination of ash value
Ash values like total ash, acid insoluble ash and water soluble ash was determined following the method reported in literature[14].

Microbial content determination
Total microbial content was determined by Streak plate (surface plating) method using agar plates.

Compatibility studies
FTIR studies: The pure drug, Metformin Hydrochloride and the physical mixture of pure drug with TSP powder, HPMC K4M in the ratio 1:1 were subjected to FTIR spectrophotometer (FTIR 8400 S, Shimadzu, Japan) at the scanning range of 4000-400 cm⁻¹.

DSC studies: Metformin hydrochloride was mixed with the polymers and the DSC analysis of each sample was carried out under the temperature range of 0-300ºC, heating rate 10ºC/min, nitrogen atmosphere (20ml/min) and alumina as reference standard. DSC was performed for the samples metformin hydrochloride, TSP, HPMC K4M, pure drug with TSP (1:1) and pure drug with HPMC K4M (1:1). The apparatus was indium cyclohexane calibrated[15].

Formulation of matrix tablets
The tablets were prepared by wet granulation technique using varying concentrations of TSP and HPMC K4M as a release retardant. To the TSP powder, dicalcium phosphate and Metformin hydrochloride (previously passed through sieve no. 80) were added and granulated using 5% PVP solution (previously dissolved in alcohol) as granulating agent. The wet mass was passed through sieve no. 12 and the granules obtained were dried at 45ºC for 30 mins. The dried granules were subjected to dry screening by passing through mesh no. 16 superimposed on mesh no. 24 and then the granules were lubricated with the mixture of talc and magnesium stearate. The granules were compressed into tablets using 12 mm capsule shaped punch in rotary tablet press (Rimek RSB-4 mini press, Cadmach). A similar procedure was adopted to prepare Metformin hydrochloride tablets using HPMC K4M as release retardant in two different ratio. Total 9 formulations were prepared using TSP and HPMC K4M. The composition of tablet formulations is shown in Table 1.

Table 1: Formulation of Metformin hydrochloride tablets

<table>
<thead>
<tr>
<th>Ingredients</th>
<th>Formulation code (Quantity in mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>F1</td>
</tr>
<tr>
<td>Metformin Hydrochloride</td>
<td>500</td>
</tr>
<tr>
<td>TSP</td>
<td>50</td>
</tr>
<tr>
<td>HPMC K4M</td>
<td>--</td>
</tr>
<tr>
<td>Dicalcium phosphate</td>
<td>343</td>
</tr>
<tr>
<td>PVP solution (5%)</td>
<td>qs</td>
</tr>
<tr>
<td>Talc</td>
<td>5</td>
</tr>
<tr>
<td>Magnesium stearate</td>
<td>2</td>
</tr>
<tr>
<td>Total tablet weight</td>
<td>900</td>
</tr>
</tbody>
</table>

Evaluation of tablets

Pre compression studies of granules
Various parameters like bulk density, tap density, angle of repose, Hausner’s ratio, Carr’s index, and bulkiness were determined.

Post compression studies of the prepared formulations
Test for weight variation, hardness, friability and thickness were determined for compressed tablets.

Drug content determination
Ten tablets of each formulation were powdered. Powder equivalent to 500 mg of Metformin hydrochloride was weighed and transferred to 100 ml volumetric flask, initially about 50 ml of phosphate buffer 6.8 was added and the flask was shaken thoroughly and the volume was made up to 100 ml with the buffer solution. The resulting solution was filtered. From this 5 ml was taken and diluted to 100 ml. From this 2ml was taken and diluted to 100 ml. From the resulting solution drug content was estimated at 234 nm using UV spectrophotometer taking phosphate buffer as blank.

In-vitro drug release
Drug release studies were carried out using USP XXIII dissolution test apparatus, rotating paddle method (Electro Lab, Mumbai, India). The study was conducted at 37±0.5ºC and 50 rpm. The dissolution medium used was 900 ml of phosphate buffer pH 6.8 and study was carried up to 12 h. 2 ml of sample was withdrawn at different time intervals and replaced with fresh medium in order to maintain sink condition. The withdrawn samples were diluted suitably and drug content was estimated spectrophotometrically at 234 nm, using phosphate buffer as blank.
In-vitro release mechanism

In vitro drug release mechanism was determined by using PCP DISSO V3 software.

Similarity factor (f2) analysis

The primary purpose of f2 is to compare the closeness of two products under consideration. Generally similarity factor in the range of 50-100 is acceptable. The similarity factor between the 2 formulations F7 and F10 was determined using the data obtained from the drug release studies. The data were analyzed by the formula,

\[ f_2 = 50 \log \left( \left\{ 1 + \frac{1}{N} \sum_{i=1}^{N} (R_i - T_i)^2 \right\}^{1/2} \times 100 \right) \]

Where, \( R_i \) and \( T_i \) are the cumulative percentage dissolved at each of the selected N time points of the reference and test product respectively.

Stability studies

Accelerated stability studies were carried out according to ICH guidelines. During the study period the formulations were packed in HDP containers and stored at 40 ± 2°C and 75 ± 5% RH for a period of 6 months in a stability chamber. The samples were tested at the interval of 2 months and evaluated for the drug content and dissolution studies and the shelf life of the prepared tablet formulations were predicted.

RESULTS AND DISCUSSION

The percentage yield of TSP was found to be 76.66%. The identification of the isolated polysaccharide was confirmed by Ruthenium red test as the polysaccharide stained pink in colour. Phytochemical tests showed presence of carbohydrates which was confirmed by Molisch’s test and Barfoed’s test.

The isolated TSP powder was soluble in hot water forming a viscous colloidal solution, but sparingly soluble in cold water and insoluble in ethanol, methanol, acetone and ether. Specific gravity of 1% TSP solution in distilled water was found to be 1.1254 and pH was found to be 6.84. Melting point was found to be 180°C, loss on drying was 20% and moisture content was found to be 2.4%. The viscosity of 1% solution at varying rpm 5,10,20,50 and 100 was found to be 6.75, 2.72, 1.41, 0.98, and 0.81 cps respectively. As the rpm increases the viscosity was found to be decreasing thereby exhibiting shear thinning nature of the TSP solution. The isolated TSP powder swelled to 33.33% in water, 28.57% in 0.1N hydrochloric acid and 7% in phosphate buffer pH6.8. The average particle size of the isolated TSP powder was found to be 6.63±0.24 µm.

The values of total ash, water soluble ash and acid insoluble ash was 2.14, 1.63 and 0.66%. The values of bulk density, tap density, Hausner’s ratio and bulkness of TSP powder were 0.740±0.05 g/cc, 0.833±0.06 g/cc, 1.166±0.01 and 1.400±0.02 respectively. The values of angle of repose and compressibility index of TSP powder were found to be 15.83±0.01° and 14.33±0.04% indicating excellent flow properties. The total microbial count determined by streak plate method showed negligible amount of microbes at the end of 7th day of the study. The microbial count was 46 CFU/g at the end of 7th day (Fig. 1). The isolated TSP powder when observed under microscope showed asymmetric particles with rough surface.

FT-IR spectrum of metformin hydrochloride showed peak at 1064.63 cm⁻¹ corresponding to NH stretching in a secondary amine. The peak at 3166.90 cm⁻¹ indicated OH stretch corresponding to carboxylic group. At 1573.81 cm⁻¹ indicated C-C stretch (in ring), 937.34 cm⁻¹ and 925.29 cm⁻¹ indicated O-H bend confirming the drug as Metformin hydrochloride. The principal absorption peaks of tamarind seed polysaccharide were found at 1045.35 cm⁻¹ and 1063.93 cm⁻¹ indicating C-O-C and ether group, respectively. The absorption peaks of C-O-C and ether group made the polysaccharide more resistant to degradation upon storage.

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and 18.98% indicating satisfactory compressibility index of the granules and formulations F4, F5, F6, F8, F9 have the values ranged between 8.31 to 8.84% indicating excellent compressible property. All the formulations showed thickness in the range of 5.0 to 7.0 mm and friability less than 1% which was within the prescribed limits. The average weight deviation percentage of 20 tablets from each formulation was less than ± 5% and all the tablets were well within the Pharmacopoeial limit. All the formulations showed good hardness values ranging from 3.26 to 4.8 kg/cm². The prepared tablets showed high degree of drug content uniformity ranging from 99.7 to 101%, indicating uniform mixing of all the ingredients during granulation.

Fig. 2: IR Spectra of (a) Metformin hydrochloride (b) Isolated TSP powder (c) HPMC K 4M (d) Physical mixture of Metformin hydrochloride and TSP (e) Physical mixture of Metformin hydrochloride and HPMC K 4M
Fig. 3: DSC thermogram of (a) Metformin hydrochloride (b) Isolated TSP powder (c) HPMC K 4M (d) Physical mixture of Metformin Hydrochloride with TSP powder (e) Physical mixture of Metformin hydrochloride with HPMC K 4M

Table 2: Data for pre compression studies of the prepared granules

<table>
<thead>
<tr>
<th>Parameters</th>
<th>F1</th>
<th>F2</th>
<th>F3</th>
<th>F4</th>
<th>F5</th>
<th>F6</th>
<th>F7</th>
<th>F8</th>
<th>F9</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bulk density(^a) (g/cc)</td>
<td>0.44±0.04</td>
<td>0.39±0.05</td>
<td>0.397±0.04</td>
<td>0.402±0.04</td>
<td>0.495±0.01</td>
<td>0.508±0.04</td>
<td>0.431±0.06</td>
<td>0.353±0.02</td>
<td>0.285±0.02</td>
</tr>
<tr>
<td>Tap density(^a) (g/cc)</td>
<td>0.65±0.05</td>
<td>0.53±0.04</td>
<td>0.472±0.06</td>
<td>0.441±0.05</td>
<td>0.572±0.01</td>
<td>0.641±0.01</td>
<td>0.532±0.01</td>
<td>0.385±0.06</td>
<td>0.311±0.04</td>
</tr>
<tr>
<td>Angle of repose(^a) (θ)</td>
<td>10.31±0.70</td>
<td>13.76±0.28</td>
<td>31.0±1.25</td>
<td>30.75±0.55</td>
<td>38.65±1.48</td>
<td>38.84±1.56</td>
<td>38.41±1.85</td>
<td>34.60±3.83</td>
<td>38.65±0.53</td>
</tr>
<tr>
<td>Rate of flow(^a) (sec)</td>
<td>2.31±0.15</td>
<td>2.64±1.46</td>
<td>2.81±0.49</td>
<td>3.05±0.44</td>
<td>12.47±0.67</td>
<td>12.51±0.52</td>
<td>14.88±0.46</td>
<td>6.63±0.30</td>
<td>12.47±0.25</td>
</tr>
<tr>
<td>Carr’s index(^a) (%)</td>
<td>32.30±1.29</td>
<td>21.53±1.06</td>
<td>15.88±1.04</td>
<td>8.94±1.32</td>
<td>8.36±2.59</td>
<td>8.56±2.42</td>
<td>18.98±2.30</td>
<td>8.31±1.81</td>
<td>8.36±1.81</td>
</tr>
<tr>
<td>Hausner’s ratio(^a)</td>
<td>1.47±0.02</td>
<td>1.35±0.02</td>
<td>1.188±0.01</td>
<td>1.097±0.03</td>
<td>1.091±0.04</td>
<td>1.094±0.02</td>
<td>1.234±0.02</td>
<td>1.090±0.02</td>
<td>1.091±0.01</td>
</tr>
<tr>
<td>Bulkiness(^a)</td>
<td>2.27±0.05</td>
<td>2.56±0.03</td>
<td>2.518±0.02</td>
<td>2.487±0.02</td>
<td>3.508±0.05</td>
<td>1.968±0.04</td>
<td>2.320±0.03</td>
<td>2.832±0.04</td>
<td>2.832±0.04</td>
</tr>
</tbody>
</table>

\(^a\) Mean ± SD, n = 3.
The results of dissolution study indicated that the formulations F1, F2, F3, F4, F5, F6, F7, F8, and F9 showed 99.8, 100.63, 100.1, 108.35, 104.64, 100.36, 101.80, 101.80, 99.54 and 99.28% release respectively. Parallelly a marketed formulation was subjected to dissolution studies to compare the release profile of the prepared formulations. The drug released between 3 to 12 h from the tablet formulations (Fig.4). The formulation F1 showed 99.38% at the end of 3rd h and formulation F7 showed 101.80% release at the end of 11th h. It was observed that as the polymer concentration increased the drug release was retarded. This may be due to increase in polymer concentration or increase path length diffusion. The release of formulation F7 was found similar to the release of marketed formulation F10.

**Table 3: Data for post compression studies of the prepared formulations**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>F1</th>
<th>F2</th>
<th>F3</th>
<th>F4</th>
<th>F5</th>
<th>F6</th>
<th>F7</th>
<th>F8</th>
<th>F9</th>
</tr>
</thead>
<tbody>
<tr>
<td>Weight variation (mg)</td>
<td>90±0.02</td>
<td>90±0.02</td>
<td>90±0.01</td>
<td>90±0.03</td>
<td>90±0.02</td>
<td>90±0.02</td>
<td>90±0.02</td>
<td>90±0.02</td>
<td>91±0.02</td>
</tr>
<tr>
<td>Hardness (kg/cm²)</td>
<td>4.5±0.24</td>
<td>3.6±0.41</td>
<td>4.2±0.72</td>
<td>4.4±0.41</td>
<td>4.6±0.40</td>
<td>4.2±0.36</td>
<td>3.46±0.30</td>
<td>4.8±0.20</td>
<td>4.4±0.41</td>
</tr>
<tr>
<td>Friability (%)</td>
<td>0.86±0.06</td>
<td>0.72±0.04</td>
<td>0.93±0.04</td>
<td>0.92±0.02</td>
<td>0.94±0.04</td>
<td>0.84±0.02</td>
<td>0.80±0.03</td>
<td>0.13±0.02</td>
<td>0.79±0.05</td>
</tr>
<tr>
<td>Thickness (mm)</td>
<td>6.88±0.04</td>
<td>7.90±0.12</td>
<td>7.85±0.06</td>
<td>7.96±0.04</td>
<td>5.66±0.28</td>
<td>6.96±0.88</td>
<td>5.89±0.16</td>
<td>7.24±0.02</td>
<td>5.94±0.06</td>
</tr>
<tr>
<td>Drug content (%)</td>
<td>99.74±0.25</td>
<td>100±0.22</td>
<td>99.92±0.30</td>
<td>100±0.24</td>
<td>100±0.50</td>
<td>99.8±0.45</td>
<td>99.94±0.26</td>
<td>100±0.36</td>
<td>100±0.41</td>
</tr>
</tbody>
</table>

*Mean ± SD, n = 3.

The drug release kinetics of formulations F1, F3, F6, F8, and F9 was predicted to be Matrix model with R² values 0.9539, 0.9935, 0.9764, 0.9894 and 0.9883 respectively, indicating the release of drug from matrix as a square root of time dependent process based on Fickian diffusion. Formulations F2, F4, F5, and F7 showed Peppas model with R² values 0.9909, 0.9934, 0.9957, and 0.9909 respectively. By incorporating the first 60% of release data, mechanism of release was indicated according to Korsmeyer where ‘n’ is the release exponent, indicative of mechanism of drug release. Fickian diffusional release and case-II relaxational release are the limits of this phenomenon. Fickian diffusional release occurs by the usual molecular diffusion of the drug due to a chemical potential gradient. Case-II relaxational release is the drug transport mechanism associated with stress and state-transition in hydrophilic glassy polymers which swell in water or biological fluids. The release exponent values of F1, F2 and F6 were 0.1354, 0.1570 and 0.3299 which was beyond the limits of Korsmeyer model so-called power law. The power law can only give limited insight into the exact release mechanism of the drug. The release exponent values of F3, F5, F7, F8, and F9 were 0.5343, 0.6715, 0.6142, 0.4794 and 0.5347, indicating non-Fickian diffusion/anomalous drug release. Formulation F7 showed Peppas model with R² values 0.9909 and release exponent values of 0.6142 found to have similar release profile as compared with marketed product. The similarity factor (f₂) value between F7 and F10 (Marketed product) formulations was found to be 50.65. This study confirmed that the release of Metformin hydrochloride from the prepared tablets (F7) was similar to that of the marketed tablet (F10). Based on this formulation F7 containing 350 mg of TSP powder was optimized and subjected for stability studies. The stability studies indicated that the tablets do not show any physical changes during the study period and the drug content was found above 97% for the formulation F7 at the end of 6 months. Formulation F7 showed a maximum of 99.36% drug content and drug release of 92.18% after 11 hours at the end of 6 months. This indicated that formulation F7 was stable at accelerated storage condition and there was no appreciable change in the physical appearance of the prepared tablets.

**CONCLUSION**

Tamarind seed polysaccharide being a natural polysaccharide do not possess any toxicity or side effects and was found to be suitable and effective for retarding the drug release of the selected drug Metformin Hydrochloride. Hence, it can be administered to patients safely, thereby improving the patient compliance by releasing the drug in a sustained manner.

**ACKNOWLEDGEMENT**

The authors are thankful to Gokula Education Foundation for providing the necessary facilities to carry out the research work. The authors thank IISc, Bangalore for carrying out DSC studies.
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