



IN SEARCH OF NEW LEADS: A CLOSER LOOK AT THE THERAPEUTIC POTENTIAL OF THE CONSTITUENTS OF *MILLETTIA THONNINGII*, *MILLETTIA PACHYCARPA* AND THEIR STRUCTURAL ANALOGUES

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ABSTRACT

This review discusses the potential applications of isoflavonoids from *Millettia thonningii* and *M. pachycarpa* in developing new pharmaceutical agents based on folkloric anecdotes and evidences from pharmacological and biochemical assays and to encourage further research into their pharmacological applications. *Millettia thonningii* is a deciduous plant indigenous to tropical West Africa, *M. pachycarpa*, is a climbing shrub indigenous to South-East Asia and other species have been used in folk-medicine for the treatment of inflammatory diseases, chronic diseases and several pathogenic diseases. Scientific research has implicated several prenylated isoflavonoids as being useful antioxidants and used in the management of radical-mediated diseases such as cancer, diabetes, ischemic heart diseases, Alzheimer's and Parkinson's diseases etc. Chemical investigations into these plants have revealed that they contain several isoflavonoids which bear structural resemblance to some of the isoflavonoids already in allopathic medical usage. Plants continue to maintain their historical stead as a store house of important drug candidates and source of new "leads" for synthetic modifications to improve activity through optimization of pharmacodynamic and pharmacokinetic properties. Natural products have also been found useful in specific pharmacological probes, a potential which is grossly underestimated. The alpinumisoflavones which have been isolated from the *Millettia thonningii* have shown high toxicities to the brine shrimp while isolates from *M. pachycarpa* have also shown anti-estrogenic and anticancer properties. Information obtained from crystal structural studies of these alpinumisoflavonoids coupled with their molecular and electronic distribution properties can further our understanding of their therapeutic potential and their observed bioactivities. The alpinumisoflavones are characterized by a fused tricyclic ring system which contains nearly coplanar benzopyrone ring fragments and a puckered six membered pyran ring that adopts a half-chair conformation with inter and intra molecular O-H...O and C-H...O contacts. A phenyl ring attached to the benzopyrone moiety shows out of plane twist with various degrees of torsion depending on the substitution on the phenyl ring.

Keywords: *Millettia thonningii*, *M. pachycarpa*, Isoflavones, Folk-medicine, crystal structure, flavonoids, Alpinumisoflavones.

INTRODUCTION

Millettia thonningii (Schum-Thonn) Baker is a deciduous tree which grows in tropical climates all over the world. It belongs to the family Papilionaceae and is indigenous to tropical West Africa and is found in moist areas and occasionally arid savanna regions¹. It is among the commonest of close to 150 plants in this genus scattered all over the world. It can grow as tall as 20m with green or grey bark and pinnately shaped leaves which are arranged alternately on the branches. Flowering of the plant normally starts around September while the fruits start appearing in December. Both seed dispersal and vegetative propagation are used as means of propagation².

M. pachycarpa Benth (*M. taiwaniana*) is a deciduous climbing shrub indigenous to South-East Asian tropical forests. It belongs to the family Fabaceae and can grow to a height of about 6m. It has large clusters pea-shaped flowers that are lilac coloured. The stem is usually brown or grey with dark brown seeds. The flowering usually occurs in July and August³.

Ethnobotanical uses of *Millettia* species

Millettia thonningii has been used in traditional medical practice in alleviating several ailments. According to folkloric anecdotes, Francophone countries in West Africa use a bark infusion to treat constipation in children⁴. In Nigeria however, it is reported that the leaf extracts of this plant cure diarrheal symptoms as well as dysentery. The pulverized roots and bark decoction are reported to

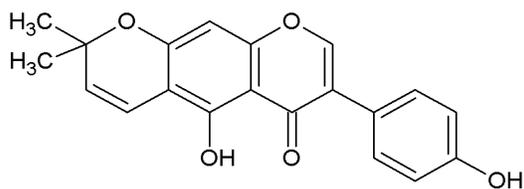
relief symptomatic episodes of dysmenorrhea and amenorrhea as well as analgesic against intestinal pains. It is also drunk as a "blood purifier" and as a de-wormer. The leaf juice is said to be lethal to the Bulinus snail, a water snail which is a vector for *Schistosoma cercariae* which causes schistosomiasis, a parasitic disease endemic in Africa, Far East and South America^{4,5}.

Millettia oblate Dunn is used to treat bladder troubles with the root decoction used as a cure for cough and stomach ache. *Millettia lasiantha* is used as an aphrodisiac either by chewing the roots or drinking aqueous decoction of it while gargling root extracts of *Millettia makodensis* Harms gives relief from toothache⁶.

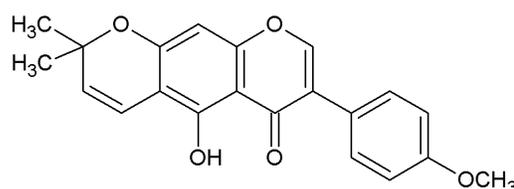
Roots extracts of *Millettia pachycarpa* Benth are used in China for fishing by pouring the root extracts in the water body which poisons and kills them. Saponins and rotenone related compounds have been implicated in this activity⁷. Elsewhere, extracts of *Millettia barberi* Dunn are used for this purpose while the dried and pulverized bark of the tree is used as a snuff against sinusitis and headaches⁸. In India, *Millettia auriculata* is used as an effective insecticide⁹ as well as applied to cattle sores to kill vermin. Seeds of *Millettia ferruginea* Baker however are used as vermifuge against roundworms¹⁰.

Isolated compounds from various parts of *Millettia thonningii*

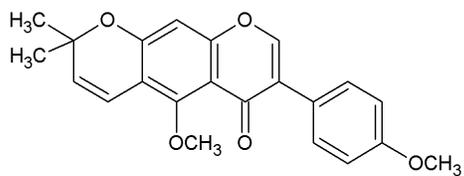
Several compounds have been isolated from several parts of the plant. The following are some examples.



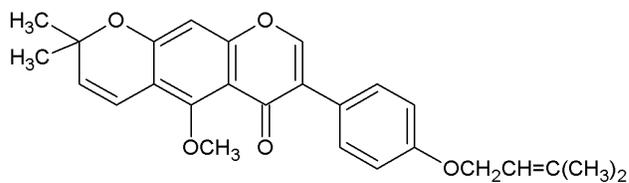
Alpinumisoflavone [1]



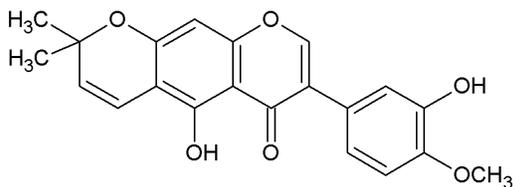
4'-O-methylalpinumisoflavone [2]



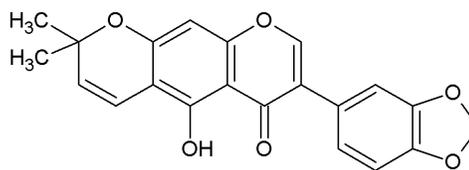
O,O-dimethylalpinumisoflavone [3]



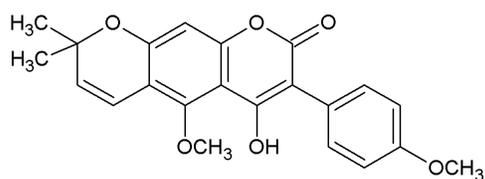
5-O-methyl-4-O-(3-methylbut-2-en-1-yl)alpinumisoflavone [4]



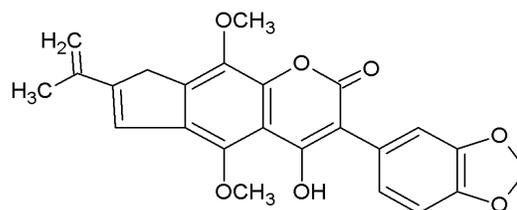
3',5-dihydroxy-4-methoxy-2'',2''-dimethylpyrano(5'',6'',7'')-isoflavone [5]



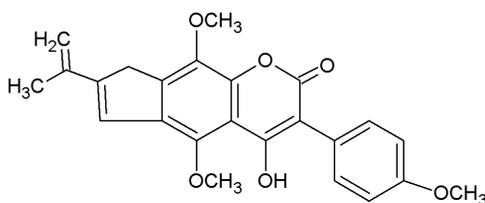
Robustone [6]



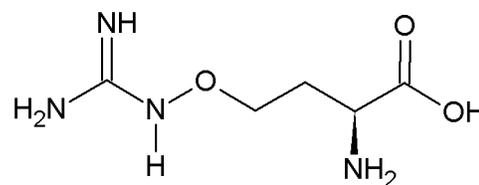
Robustic acid [7]



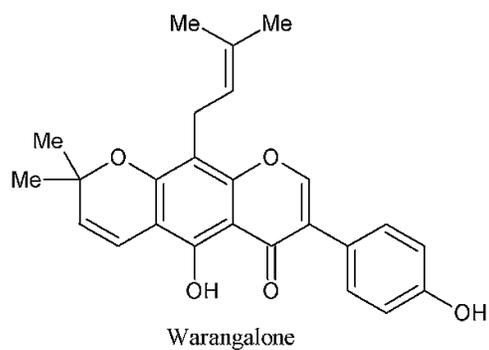
Thonningine-A [8]



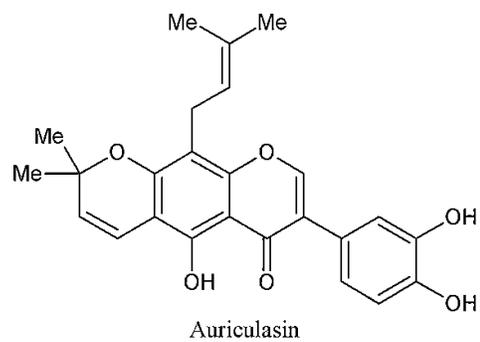
Thonningine - B [9]



Canavanine [10]

Fig. 1: Some isolated compounds from *Milletia thonningii*.

Warangalone



Auriculasin

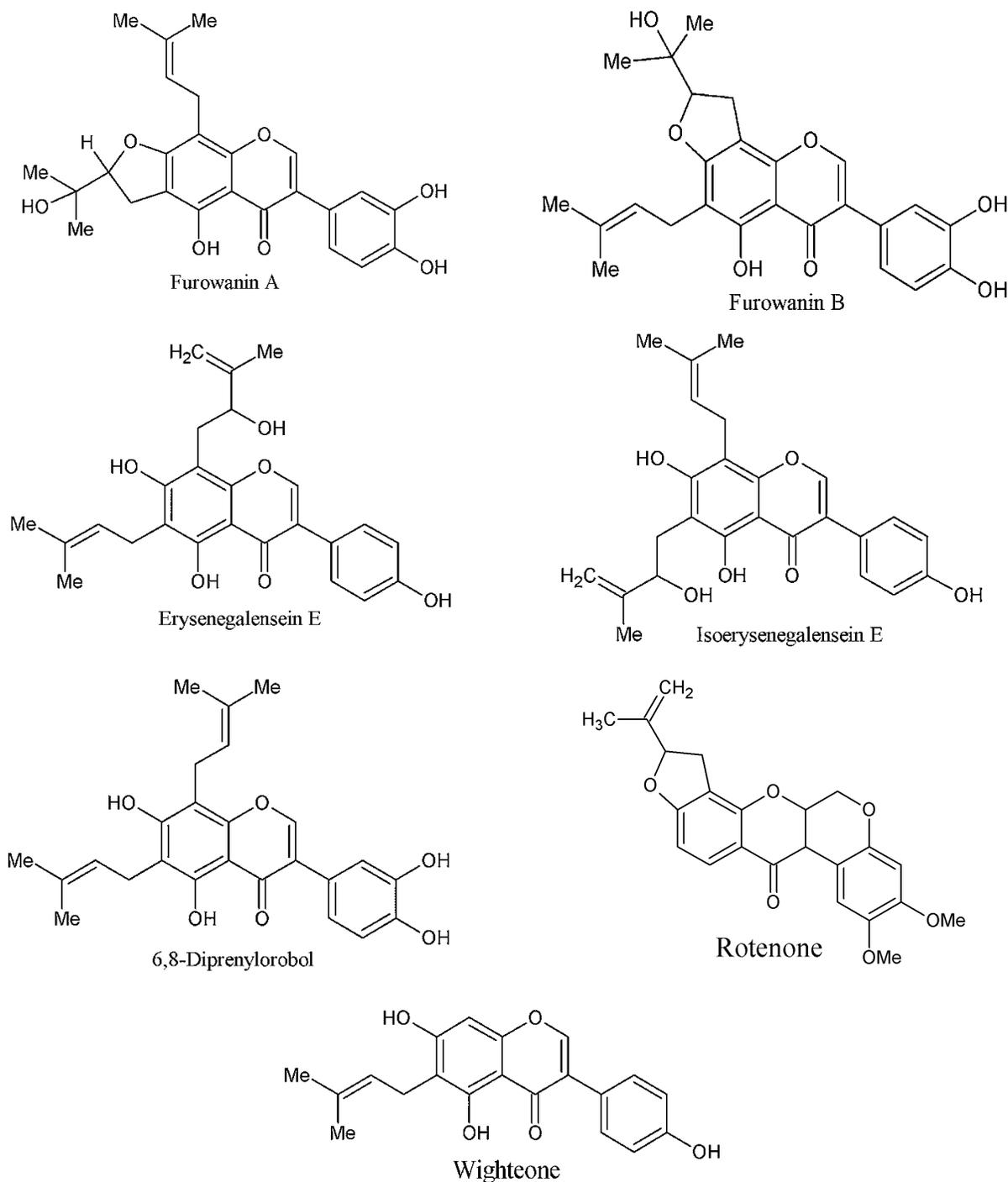


Fig. 2: Some isolated compounds from *Milletia pachycarpa*.

Some compounds isolated from *M. pachycarpa*.

The above are some of the compounds isolated from *M. pachycarpa* also known as *M. taiwaniana*. Compounds with similar structural backbone have been isolated from other species of *Milletia*.

Biosynthesis of Flavonoids

Alpinumisoflavones share ancestry with a general class of compounds called flavonoids which are a group of over 8000 polyphenolic compounds that occur exclusively but with very wide distribution in plant species. They are characterized by a phenylbenzopyrone structure (C6-C3-C6), and their categorization is

based on the level of saturation and opening of the central pyran ring, into: flavones, flavanols, isoflavones, flavonols, flavanones, and flavanonols. They are produced biogenetically from two main primary synthetic pathways- the acetic acid pathway and the shikimate pathway which produces phenylalanine and tyrosine¹¹. The flavonoids are formed in plants and participate in the light-dependent phase of photosynthesis during which they catalyze electron transport¹². Phenylalanine and tyrosine are converted to cinnamic acid and parahydroxycinnamic acid, respectively, by the action of phenylalanine and tyrosine ammonia lyases¹³. The rest of the biosynthetic pathway is summarized below.

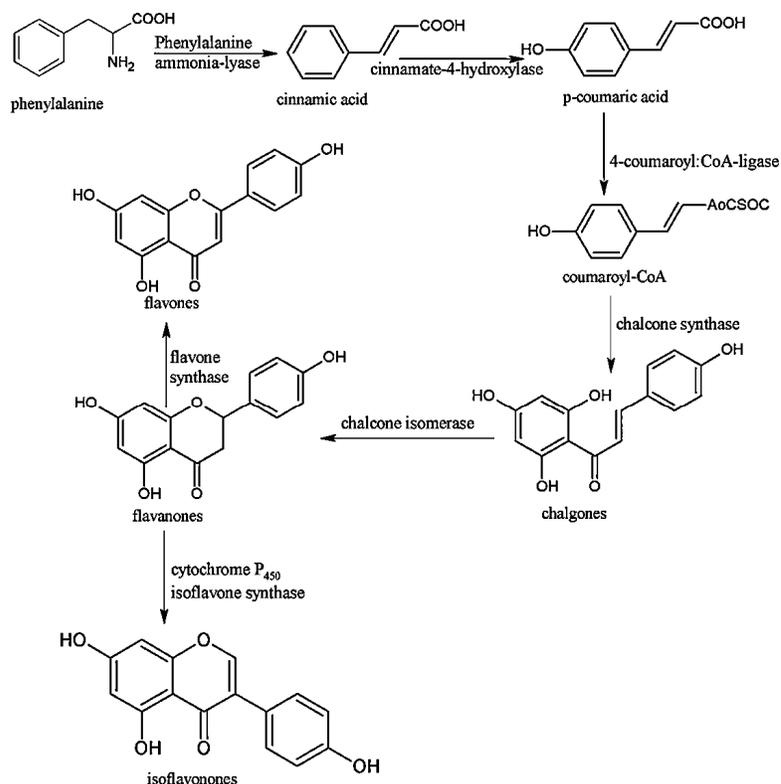


Fig. 3: Biosynthetic route to flavonoids

Pharmacological profile of constituents of *Millettia thonningii* and *M. pachycarpa*

Over the years, scientific research continues to accrue evidences in support of the folkloric use of plants for curative purposes. Phytochemical research has led to isolation of several different compounds from plants, most of the time with entirely different bioactivities. These numerous compounds with myriad of bioactivities may somehow give credence to the use of one decoction of a plant in curing several different ailments in our traditional African herbal practices. Unfortunately however, many people who do not trust the therapeutic potential of herbal medicines question the use of one herb in curing a 'million' diseases without the benefit of a scientific evidence of the bioactive components to support or contradict these claims.

While the patronage of herbal medicines still remains very low among the elite, plants continue to maintain the historical stead as important source of new drug candidates for the management of several diseases and also as source of new 'leads' for synthetic modifications to optimize pharmacokinetic and pharmacodynamic properties. They may be useful for specific pharmacological and biochemical probes¹⁴. According to the WHO, 80% of the world population continues to rely mainly on traditional medicines for their health care¹⁵.

Millettia thonningii and *M. pachycarpa* have been used in folk-medicine for the treatment of pains, pathogenic diseases and radical-mediated diseases for ages. In trying to ascertain the efficacy or otherwise of the constituents of these plants in justification of their folkloric use, several bioactivity tests have been conducted on some extracts of various parts of the plants. In addition several bioactivity tests have been carried out on extracts of some other plants which have constituents that are also found in *Millettia thonningii* and *M. pachycarpa*. The following chronicles some of the major biochemical assays carried out on some of the constituents of

these plants.

Anti-estrogenic and anticancer activity

Hypoxia is a condition characterized by lack of oxygen in the tissues of the body of organisms and is concomitant with cancers and tumors when oxygen demand of the rapidly dividing cells are not met. In dealing with this situation, the body's feedback mechanisms cause hypoxic tumor cells to activate the transcription of genes that function to promote anaerobic metabolism as well as those that initiate tumor angiogenesis¹⁶⁻¹⁸. A common feature of this situation is that some regions of tumor cells are constantly hypoxic which causes them to become more aggressive and resistant to treatment¹⁹ in their bid to adapt to the low oxygen and nutrients supply.

First described by Semenza and Wang²⁰ as transcriptional activator responsible for the hypoxic induction of erythropoietin and for that matter a key regulator of oxygen homeostasis, hypoxia-inducible factor-1 (HIF-1) a heterodimer of the basic helix-loop-helix PER-ARNT-Sim proteins has a basic structure made up of constitutively expressed HIF-1 β subunit and a HIF-1 α (HIF-1 α and HIF-1 β /ARNT) subunits²¹ inhibition of which offers a major biochemical target for the discovery of hypoxia-selective anticancer drugs. HIF-1 has been found to regulate the expression of genes involved in processes such as immortalization, genetic instability, dedifferentiation and stem cell maintenance, tumor angiogenesis, metabolic reprogramming, survival and resistance to apoptosis, migration/invasion and metastasis, and treatment resistance. These it does by promoting hypoxic adaptation and survival by increasing oxygen delivery and decreasing oxygen consumption, expressing growth factors for autocrine signaling, suppressing cell death, and promoting metastasis^{22, 23}. Research based on animal models suggest that, proliferating cells express vascular endothelial growth factor (VEGF) gene, which stimulates angiogenesis to provide the additional perfusion that is required to maintain oxygenation of an increased number of cells hence HIF-1 inhibition reduced tumor vascularity

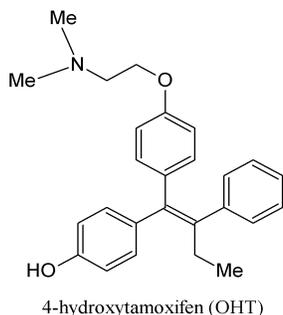
and retarded tumor growth hence HIF-1 α protein expression is indicative of the advanced state of the disease, metastasis, treatment resistance, and poor prognosis in cancer patients^{22, 24-28}. A combination of chemotherapy or radiation with HIF-1 has been found to produce enhanced treatment outcomes in preclinical studies^{29, 30}.

Liu Yang *et al*³¹ using natural product chemistry based approach in the search for HIF-1 inhibitors with the conviction that small molecule HIF-1 inhibitors represent potential drug leads that will suppress tumor growth and enhance chemotherapy/radiation by inhibiting hypoxia-induced gene expression report after studying the lipid extract of *Lonchocarpus glabrescens* strong inhibition of HIF-1 by the constituents of this plant. Alpinumisoflavone and 4'-O-methylalpinumisoflavone which are copiously abundant in seeds and the root-bark of *Milletia thonningii* are the two compounds implicated in this bioactivity. In human breast tumor T47D cells, Alpinumisoflavone and 4'-O-methylalpinumisoflavone inhibited hypoxia-induced HIF-1 activation with IC₅₀ values of 5 and 0.6 μ M, respectively.

At the concentrations that inhibited HIF-1 activation, 4'-O-methylalpinumisoflavone inhibited hypoxic induction of HIF-1 target genes (*CDKN1A*, *GLUT-1*, and *VEGF*), tumor angiogenesis *in vitro*, cell migration, and chemotaxis. 4'-O-methylalpinumisoflavone inhibits HIF-1 activation by blocking the induction of nuclear HIF-1 α protein, the oxygen-regulated subunit that controls HIF-1 activity. Mechanistic studies indicate that, unlike rotenone and other mitochondrial inhibitors, 4'-O-methylalpinumisoflavone represents the first small molecule that inhibits HIF-1 activation by simultaneously suppressing mitochondrial respiration and disrupting protein translation *in vitro*. This unique mechanism distinguishes 4'-O-methylalpinumisoflavone from other small molecule HIF-1 inhibitors that are simple mitochondrial inhibitors or flavanoid-based protein kinase inhibitors.

When isoflavonoids isolated from the leaves of *Milletia pachycarpa* (*M. taiwaniana*) were tested for the inhibitory potential to the growth of human leukemia HL-60 cells, furowanin-A, Warangalone, Isoerysenalensein-E, and euchenone b₁₀ showed the most significant cytotoxicity. Furowanin-A, Warangalone, and Isoerysenalensein-E, were found to induce apoptosis in HL-60 cells through activation of the caspase-9/caspase-3 pathway, which is triggered by mitochondrial dysfunction³².

Isoerysenalensein E and 6, 8- diprenylorobol were found to be the most potent antagonists of the estrogen receptor (ER) with their anti-estrogenic activity comparable to that of 4-hydroxytamoxifen, a metabolite of tamoxifen and a stronger ER antagonist than tamoxifen. Warangalone and auriculasin were found to be weak ER inhibitors, and alpinumisoflavone and erysenalensein E were non-ER inhibitors³³. These finding has been corroborated by Ito *et al*³⁴.



Anti-infective properties

When 10g of powdered seeds of *Milletia pachycarpa* (*M. taiwaniana*) were extracted with 100ml of water, acetone and ethanol and applied as fine spray against the adult housefly, *Musca domestica*, the fully grown larvae of the cabbage worm *Pieris rapae* and the silk worm *Bombyx mori*, the ethanol extract was found to be the most toxic against the cabbage worm, while the acetone extract was most effective against housefly. The aqueous extract was found

to be toxic to the housefly, cabbage worm and *Pareva* larvae. Rotenone and rotenoid group of compounds have been established as the active constituents in these extracts³⁵.

Mukerja and Tripathi³⁶, reported that the ether extract of the seeds of *Milletia pachycarpa* was found to be one-third as toxic as dichlorodiphenyltrichloromethane (DDT) when used as contact poison for houseflies. At a concentration of 0.5%, the extract was 80-90% lethal to silkworm larvae while at 1% it was 100% lethal to the eggs of silkworm.

Bishnupada Roy *et al*³⁷, report that, crude ethanol, methanol, and acetone fractions of *M. pachycarpa* when assayed against *Railletina echinobothrida*, the intestinal cestode parasite of domestic fowl, in an effort to verify the putative anthelmintic efficacy and cestocidal potential of this plant realized that *in vitro* exposure of the worm to the extracts at a concentration of 25 mg/mL in phosphate buffered saline (at 37°C \pm 1°C) caused distortions and disruption of mitochondria, nucleus, nucleolus, nuclear membrane, basal lamina, and tegumental vacuolization in the distal cytoplasm leading to scar formation in the surface.

Perrett *et al*³⁸, investigating the potential anthelmintic properties of *Milletia thonningii*, found that a chloroform extract of the seeds when topically applied to mouse skin prior to exposure to *Schistosoma mansoni* cercariae, showed molluscicidal and cercaricidal activity. Subsequent re-infection was inhibited for as long as the extracts of *Milletia thonningii* were present on the surface of the animal skin perhaps justifying the use of the plant as an anthelmintic and a purgative according to folklore^{4,5}. The compounds implicated for this activity are thought to be the isoflavonoid alpinumisoflavone and its derivatives. In furtherance of the work done above, *in vitro* bioactivity study by Laddiard *et al*³⁹, and later corroborated by Maillard *et al*⁴⁰, shows that the extracts of the seeds of this plant are lethal to *Schistosoma mansoni* miracidia, cercariae, and adult worms. Robustic acid and other coumarin compounds present in the seeds and alpinumisoflavones are said to be responsible for this observed bioactivity.

In investigating the possible mode of this anti-schistosomal bioactivity using rat liver as the test organ, Laddiard *et al*^{39,41}, found that the seed extracts of *Milletia thonningii* inhibited the site I mitochondrial electron transport system's NADH dehydrogenase at concentrations of 30-159mg/l. While these bioactivities have been unambiguously established, it is not clear which of the many isoflavones are responsible for these bioactivities and how the substitution pattern and their resulting molecular structures influence these bioactivities.

Monoamineoxidase enzyme inhibition

The surge in Parkinson's and Alzheimer's diseases which are aging-related neurodegenerative diseases has increased immensely the interest in the search for compounds which have this therapeutic potential. It has been observed that selegiline, rasagiline and lazabemide, which are inhibitors of Monoamine oxidase- B implicated in these neurodegenerative diseases have also been observed to have protective effects on neuronal tissues increasing the interest in compounds which have this therapeutic potential⁴². In assessing the therapeutic potential of the fruits of *Cudrania tricuspidata*, it was found that they contain prenylated flavones which were found to be potent Monoamine oxidase-A&B (MAO-A/B) inhibitors in concentration dependent manner and could be possible therapeutic candidates for the Parkinson's and Alzheimer's diseases. However further pharmacological investigations and *in vivo* physiological functions remain unelucidated. Some of the compounds found to be responsible for this (MAO-A/B) inhibitory effects included 4'-O-methylalpinumisoflavone and alpinumisoflavone⁴³, which are copiously abundant in *Milletia thonningii*. The slight differences in IC₅₀ values 23.9, and 25.8 μ M, respectively found for 4'-O-methylalpinumisoflavone and alpinumisoflavone makes it imperative for other compounds in this plant to be investigated.

Allelopathic uses

In search of potential herbicides, phytotoxic effects of the extracts of

the aerial part of *Pueraria phaseoloides* were evaluated using Petri dish bioassay on the potential inhibition of the seed germination of the weed species of *Mimosa pudica*, *Senna obtusifolia*, *Senna occidentalis* and *Urena lobata*. Arruda et al⁴⁴, report that, two of the compounds isolated from this plant and used for the bioassay are 4'-O-methylalpinumisoflavone and alpinumisoflavone. The inhibition on germination on the various weeds was found to be more selective. At 3ppm, the inhibition of 4'-O-methylalpinumisoflavone on *S. obtusifolia* was found to be the strongest at 77% while alpinumisoflavone inhibited *U. obata* at 60%, highest observed in the experiment.

Pharmacological profile of structural analogues of the constituents of *Milletia thonningii* and *Milletia pachycarpa*

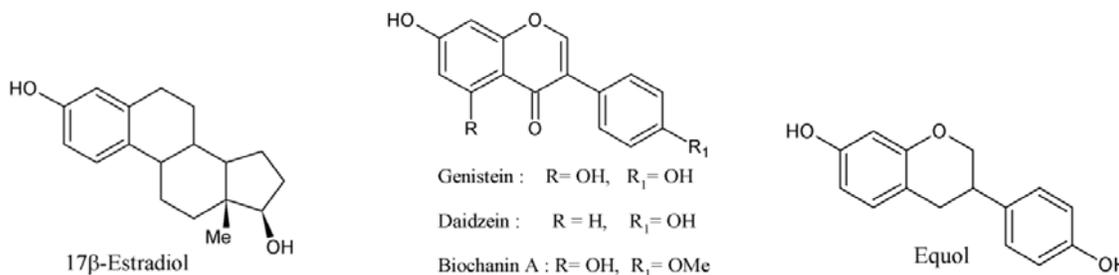
The constituents of *Milletia thonningii* share a lot of structural semblance with some very well-known flavonoids such as the catechins, epicatechins, genistein, daidzein, quercetin, Licopyranocoumarin, Glycycomarin, Glycherrisoflavone, kaempferol, Glabrene, silybin and many others with well documented pharmacological profile. The structural diversity and its ubiquitous distribution in plant species give some credence to the assertion that flavonoids may have existed in nature for over one billion years affording interactions with evolving organisms over the eons and undergoing structural transformations sometimes without losing its precursor. If the theory of evolution is anything to go by, then flavonoids must possess some important functions in nature which have been critical to the plants to have been retained in vascular plants throughout evolution⁴⁵. Indeed this long existence of

flavonoids also means a long association with various animal species that have come through the evolutionary ladder and perhaps may be a justification for their array of biochemical and pharmacological activities in living systems.

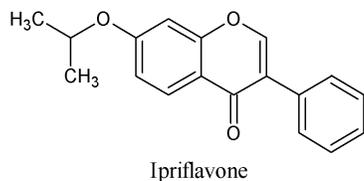
Flavonoid compounds undoubtedly have critical effects in the physiological and biochemical systems of plants imputed with functions such as antioxidants, enzyme inhibitors, precursors of toxic substances, pigments and light screens⁴⁶. Furthermore they play active roles in photosensitization and energy transfer, the actions of plant growth hormones and growth regulators, the control of respiration, photosynthesis, morphogenesis, and sex determination, as well as defense against infection⁴⁷.

Estrogenic and antitumor bioactivity

A phenolic ring and a distance of 11.5Å between two hydroxyl groups in genistein are some structural features it shares with the most potent of the endogenous estrogen steroids, 17β- estradiol. These seemingly trivial features enable genistein to exert both estrogenic and anti-estrogenic activity by binding competitively to estrogen receptors and sex hormone binding proteins. Equol which is a non-steroidal estrogen produced as a metabolic product of isoflavonoids such as daidzein by gastrointestinal bacterial flora has proven to have beneficial effects in incidences of prostate cancer and some physiological changes accompanying menopause and together with genistein, are able to dislodge bound estrogen and testosterone from human sex steroid binding proteins resulting in their delayed clearance and hence availing the hormones to target cells⁴⁸⁻⁵⁰.



Neonatal administration of genistein has been found to confer protection against chemically-induced mammary tumors in rats by inducing increased latency, reduced tumor incidence and multiplicity, and more rapid maturation of undifferentiated end buds to differentiated lobules⁵¹. Biochanin A (4'-methoxygenistein) copiously abundant in chickpea on the other hand, is an active cancer chemopreventant in animal models. The mechanism of inhibition of these isoflavones in breast cancers, cell growths and the development of chemically induced cancers in the stomach, bladder, lung, prostate, and blood involves stimulation of a signal transduction pathway leading to apoptosis⁵². Genistein blocks Epidermal Growth Factor (EGF) mediated tyrosine phosphorylation *in vivo* in human epidermal carcinoma cells but appears neither to induce phosphorylation of EGF receptors nor other tyrosine kinase substrates. It is suggested hence that the inhibition of cell growth is through modulation of transforming growth factor (TGF) β1 signaling pathways.



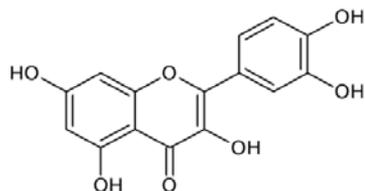
The development of ipriflavone as an oral treatment for acute leukemias and osteoporosis by increasing bone calcium retention, inhibition of bone breakdown, promotion of the activity of bone-building cells and reduction in the pain of osteoporotic fractures was based on the observation that genistein when targeted specifically to

leukemia cell lines via a linkage to a monoclonal antibody, was found to be a strong inhibitor of the growth of the leukemia cells by selectively inhibiting CD-19-associated tyrosine kinase activities, resulting in death of human B-cell precursor leukemia cells⁵³. However, in several cell systems in which genistein inhibits growth, the therapeutic window (index) has been found to be very wide and only exerts toxicity at concentrations greatly in excess of those at which it first exerts its biological and pharmacological effects making it a potentially important molecule for dietary cancer chemoprevention.

Protein kinases mediate phosphorylation of amino acid residues in proteins resulting in subsequent changes in the conformational identity of the protein and are found to be responsible for controlling several cellular functions. Protein tyrosine kinases (PTK) found in many different types of cells are implicated in focal adhesions which are perfunctory links to the extracellular matrix within which biochemical signaling of proteins at sites of integrin binding and clustering takes place invariably regulating cell transformation and growth, expression of genes, cell-cell adhesion interactions, cell motility, and shape⁵⁴⁻⁵⁶. Genistein was not only found to selectively inhibit PTK and at higher concentrations protein histidine kinase (PHK), but also has the propensity to inhibit DNA topoisomerases I and II which introduce transient breaks in linear DNA sequences and participate in several genetically related processes, including replication, transcription, recombination, integration, and transposition⁵⁷. They are also found to possess antioxidant and cell cycle inhibitory activity⁵⁸.

Apart from its inhibitory effects on PTK and Protein Kinase C (PKC) activity, quercetin has been found capable of inhibiting nuclear kinase II-catalyzed phosphorylation of isolated nuclear proteins in

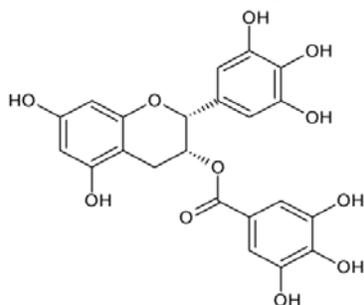
HeLa cells using Guanidine Tri Phosphate (GTP) as phosphate donor⁵⁹.



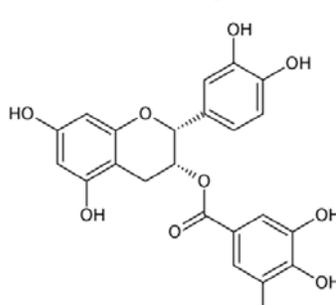
Quercetin

Anti-retroviral activity

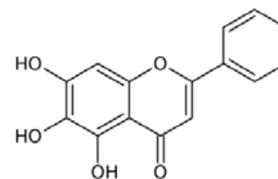
Viruses continue to be a continuous source of worry for mankind being responsible for deadly diseases such as H1N1, Ebola, AIDS, some cancers and many other diseases. Retroviruses are Ribonucleic acid (RNA) viruses that copy their genomes into Deoxyribonucleic acid (DNA) during their replication. Reverse transcriptase (RT) enzymes transcribe the infecting RNA chains of these viruses into complementary DNA molecules that integrate into the host cell genome and this could lead to a permanent genetic alteration. Thus RTs have the potential to copy any RNA into DNA, even in the absence of specific transfer RNA (tRNA) primers such as Human T-



(-)- Epigallocatechin gallate



(-)- Epicatechin gallate



Baicalein

Based on the observation that baicalein isolated from a Chinese traditional drug which is a flavone, inhibited HIV-reverse transcriptase (HIV-RT), a survey of the flavonoid family was carried out in which was found that (-)-epicatechin gallate (ECg) and (-)-epigallocatechin gallate (EGCg) which are major components of Japanese green tea strongly inhibited HIV-RT with IC₅₀ values between 10-20ng/ml concentration^{62,63}.

Myricetin, morin, quercetin, and fisetin are some of the flavonoids suspected to be potential nonpeptidic inhibitors of the HIV protease enzyme with IC₅₀ values in the range of 10 to 50 μM as reported by Brinkworth *et al*⁶⁴ while quercetin has been found to also strongly inhibit integrase activity⁶⁵.

Antioxidant activity

The evolutionary choice to use oxygen for respiration and oxygen containing compounds for diverse biological activities is not one without consequences. The ubiquitous presence of oxygen in the body of animals comes with it the production of other more "reactive oxygen species" (ROS) such as superoxide (O₂⁻), hydroxyl radical (·OH), peroxy radical (ROO·), alkoxy radical (RO·), hydrogen peroxide (H₂O₂), singlet oxygen (¹O₂), triplet oxygen (³O₂), hypochlorous acid (HOCl) and many others. The production of ROS in the body is essential for proper functioning but in excess, they pose several dangers to the health of individuals. When activated upon xenobiotic entry into the body of organisms, phagocytic cells such as monocytes, neutrophils, eosinophils, and macrophages generate ·O₂· as a result of increased oxygen consumption (respiratory burst), catalyzed by a membrane-bound NADPH oxidase system and are responsible for their bacteriocidal and tumoricidal functions^{66,67}.

lymphotropic viruses 1 & 2, Murine leukemia virus, Human immunodeficiency virus (HIV) 1 & 2 and Mouse mammary tumor virus. Encoded by the retrovirus RNA and packaged inside each viral capsid during the production of new virus particles, reverse transcriptase is described as an unusual DNA polymerase that utilizes either RNA or DNA as a template. Upon entry into a cell, the enzyme RT which accompanies the single-stranded RNA of the retrovirus makes a DNA copy of the RNA strand to form a DNA-RNA hybrid helix, which is then further transformed by the same enzyme to a DNA double helix. A virus-encoded integrase that catalyzes the insertion of the viral DNA into virtually any site on a host-cell chromosome recognizes the two ends of the linear viral DNA after which the integrated viral DNA is transcribed by host-cell RNA polymerase to produce numerous identical types. Enzymatic activities present in retrovirus are; RNA-dependent DNA polymerase (reverse transcriptase; RT), DNA-dependent DNA polymerase, Ribonuclease H (RNase H), Integrase and Protease⁶⁰.

In their search for potential natural products that could inhibit key enzymes in the synthesis of DNA, Spedding *et al*⁶¹, reported that quercetin was one of the three most active naturally occurring flavonoids that inhibited three reverse transcriptases (RT) viz: avian myeloblastosis RT, Rous-associated virus-2 RT, and Moloney murine leukemia virus (MMLV) RT when poly(rA)oligo(dT) 12-18 or rabbit globin messenger RNA (mRNA) were used as templates.

While ROS generated by phagocytes play an important physiological function, they can also cause cellular damage to bio-systems which is a major contributor to degenerative diseases such as cancer, atherosclerosis, stroke, myocardial infarction, trauma, arthritis, ischemia/reoxygenation injury, and aging⁶⁸, peroxidation of membrane lipids, oxidative damage to nucleic acids and carbohydrates, and the oxidation of sulfhydryl and other susceptible groups in proteins^{69,70}.

In an attempt to ensure healthy life devoid of the dangers posed by ROS, defense to the body is provided by antioxidant systems which are involved in radical mop up by scavenging and quenching radicals hence detoxifying the body of ROS in the cell through enzymatic and non-enzymatic systems. The best known antioxidant molecules are vitamins A, E, and β-carotene, ascorbic acid, urate, ubiquinol, retinoids and carotenoids⁷¹⁻⁷³.

Many *in vitro* studies conducted on an array of compounds with the hope of finding potent radical scavengers found flavonoids as the most promising candidates, which contribute to the inhibition of lipid peroxidation and oxidation of low density lipoproteins (LDL) implicated in the pathogenesis of coronary heart diseases by decreasing the susceptibility of LDL to oxidation⁷⁴⁻⁷⁶. The protective effects of flavonoids in biological systems are ascribed to their capacity to transfer free radical electrons, chelate metal catalysts⁷⁷, activate antioxidant enzymes⁷⁸, reduce α-tocopherol radicals⁷⁹ and inhibit oxidases⁸⁰.

Consumption of food products rich in antioxidants has been amply demonstrated to confer a lot of health benefits. For example diets rich in rice, maize and beans have been shown to exhibit anticarcinogenic activity by inhibiting protease enzymes as well as interfering with the formation of ROS⁸¹. The agents present in these

diets to which are attributed these functions are flavonoids. Constituent of various types of teas which are basically flavonoids (catechins and theaflavins) have also been found to be anticarcinogenic and chemopreventive agents⁸², radioprotective agents, antibacterial, antifungal and antiviral⁸³⁻⁸⁶.

Crystal structure of Alpinumisoflavones and their bioactivity

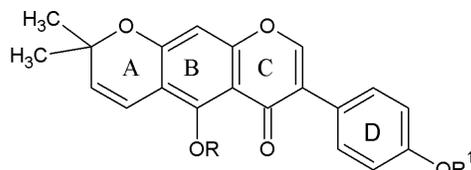
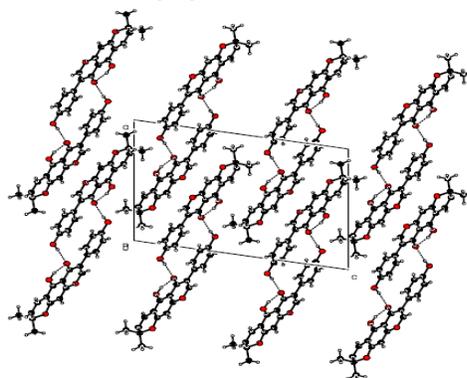


Fig. 4: General structure of the Alpinumisoflavones

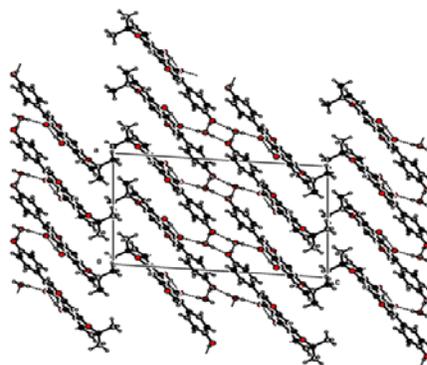
The discovery of the electronic nature of drug-receptor interactions has opened the frontiers for crystal and bioactive engineering in a bid to improve predictability of bioactive compounds based on crystal structural studies. The electron-conformational method of pharmacophore identification and bioactivity prediction has seen great improvement in the usage of an atomic index of orbital and charge controlled interaction to better represent the ligand (substrate) in its interaction with the bioreceptor and the multi-conformational problem is considered in view of ligand-receptor binding states, resulting in essential simplification of the expression of bioactivity⁸⁷. Systematic study hence of some of the compounds isolated from this plant, specifically the alpinumisoflavones (1-4)

and their semi-synthetic analogues, have focused on crystal structure and activity⁸⁸⁻⁹¹, with the hope that the crystal structure, molecular and electronic properties can deepen our understanding of their observed bioactivities.

Not much diversity has been found among the crystal structures of the alpinumisoflavones studied so far. The tricyclic rings A/B/C are fused. The benzopyrone moieties consist of rings B and C of the compounds and are nearly coplanar. They show an out-of-plane twist with the phenyl ring D. However, the slight differences in substitution patterns have not only resulted in remarkable differences in bioactivities⁸⁹, but also produced some remarkably different crystal packing with various kinds of inter and intra-molecular hydrogen bonds some of which have been utilized in the preparation of solvates in a bid to improving its aqueous solubility properties to make bioassays *in vitro* even easier⁹¹. Electrostatic potential maps derived from non-spherical electron density using the Invariom method based on the approximation of locality of molecular electron density, compared to brine shrimp lethality tests conducted on some of the isolates, O,O-dimethylalpinumisoflavone, alpinumisoflavone and 5-O-Methyl-4'-O-(3-methyl-but-2-en-1-yl) alpinumisoflavone⁸⁹, indicates that the presence of high electron density around the O atoms of ring B and D are important for activity. Hence increasing the electron density in these areas by introducing more electron donating substituents R and R¹ without altering the conformations in these regions significantly may lead to higher activity *in vitro*. The following structures show some of diverse crystal structure packing found in the molecules so far studied.



Part of the crystal packing of alpinumisoflavone viewed on the *a-c* plane. Hydrogen bonds are shown as broken lines⁹⁰.



Part of the crystal packing of (alpinumisoflavone + H₂O) viewed along *a-c* plain showing the formation of molecular chains of fused rings. Hydrogen bonds are shown in broken lines⁹¹.

Concluding remarks

The influence of lipid peroxidation in biological cells as being involved in the pathogenesis of many age-related diseases has long been known. Free radicals or Reactive Oxygen Species mainly produced by reactions such as abnormal oxidation and breakdown of fats and hydrogen peroxide have been implicated in the

detoxification of invading organisms and chemical substances. Stray free radicals however, can also initiate lipid peroxidation in healthy cells, damage proteins and nucleic acids and other biomolecules through uncontrolled cleavages resulting in mutations that lead to chronic ailments. It has been observed that there is some positive link between accumulated free radical damage and some age-related diseases such as coronary heart diseases, diabetes, cancer etc and

neurodegenerative diseases such as Alzheimer's and Parkinson's^{92,93}. Dietary flavonoids have been found to be able to repair a range of oxidative radical damage sustained by DNA with high intake of it having been associated with reduced risks to degenerative diseases⁹⁴.

Uncontrolled neo-vascularization leads to imbalances in the capillaries in the body and can promote angiogenesis dependent diseases such as growth of solid tumors, arthritis and inflammations making its inhibition important in the control of these diseases⁹⁵. Plant extracts found to be rich in flavonoids have been found to possess high antioxidant and anti-angiogenic activities⁹⁶. Perhaps the need to pay more attention to HIF-1 inhibitors has become more pertinent now not only because there is no approved drug that specifically targets hypoxic tumor cells but also as reported by Liu Yang *et al*³¹ of the unique mechanism of the 4'-O-methylalpinumisoflavone by simultaneously suppressing mitochondrial respiration and disrupting protein translation *in vitro*. The high inhibitory activity of 4'-O-methylalpinumisoflavone for HIF-1 was attributed by the authors to its high lipophilicity which increases its bioavailability. If this assertion is anything to go by, then the more lipophilic structural analogues of these compounds may perhaps be better candidates for this bioactivity. Constituents of *Millettia thonningii* and *M. pachycarpa* may be useful in this stead.

The discovery of monoamine oxidase inhibition potential of some constituents of this plant as already alluded to, make these plants potentially useful in the search for new drug candidates and "Lead" compounds for the synthesis of drugs to tackle the numerous radical mediated diseases. It must also be noted that anti-inflammatory effect as well as anti-microbial properties of these plants cannot be discounted having shown activity towards some pathological organisms such as worms. Utilization of the crystal structure information to model various receptor binding and docking experiments would further bring to light the therapeutic potentials of the constituents of these plants.

REFERENCES

- Hutchinson J, Dalziel JM. Flora of West Tropical Africa 1958 (Revised by Keay R.W.J. (1963). 2nd Edition, vol. 1.p. 527. Crown Agents, London.
- Troyte Stephen. A Revision of the Genus *Millettia*, Wight et Arn. J. Linn Soc. Bot. 1912; 41 (280): 123- 139.
- Miquel FAW. *Millettia pachycarpa* Benth. Pl. jungh. 1852; 250.
- Irvine FR. Wood Plants of Ghana. 1st Ed. Oxford University Press. 1961.
- Abbiw D. Useful Plants of Ghana. London: Intermediate Technology Development Group and the Royal Botanical Gardens, Kew. 1990.
- Kokwaro JO. Medicinal Plants of East Africa. Kampala-Nairobi-Dar-es-Salaam: East African Literature Bureau. 1976.
- Chopra RW, Badwar RL, Ghosh S. Poisonous Plants in India. 1949; 1: 391-393. Calcutta: Government of India Press.
- Crombie L. Chemistry of the natural rotenoids. Fortschr Chem Org Nat. 1963; 21: 275-325.
- Tattersfield F, Martin JT, Howes FN. Some Fish- Poison Plants and Their Insecticidal Properties. Bulletin of Miscellaneous Information. 1940; 5: 169-180.
- Watt JM, Breyer-Brandwijk MG. The Medicinal and Poisonous Plant of Southern and Eastern Africa. 2ed, Edinburgh and London: E & S Livingstone limited. 1962; 631.
- Bravo L. Polyphenols: Chemistry, dietary sources, metabolism, and nutritional significance. Nutrition Reviews. 1998; 56(11): 317-333.
- Das DK. Naturally occurring flavonoids: Structure, chemistry, and high performance liquid chromatography methods for separation and characterization. Methods Enzymol. 1994; 234: 410-420.
- Wagner H, Farkas L. Synthesis of flavonoids, 1975, in The Flavonoids. Part I (Harborne J. B, Mabry T. J and Mabry H eds) pp 127-213, Academic Press, NewYork.
- Balandrin MF, Kinghorn DA, Fansworth NR. Plant derived Natural Products in Drug Discovery and Development. American Chemical Society Symposium Series 534. American Chemical Society, Washington DC. 1993; 2.
- Gurib-Fakim A. Medicinal plants: traditions of yesterday and drugs of tomorrow. Molecular Aspects of Medicine. 2006; 27: 1-93.
- Harris AL. Hypoxia - a key regulatory factor in tumour growth. Nat Rev Cancer. 2002; 2(1): 38-47.
- Le QT, Denko NC, Giaccia AJ. Hypoxic gene expression and metastasis. Cancer Metastasis Rev. 2004; 23: 293-310.
- Tatum JL, Kelloff GJ, Gillies RJ, Arbeit JM, Brown JM, Chao KS, Chapman JD, Eckelman WC, Fyles AW, Giaccia AJ, Hill RP, Koch CJ, Krishna MC, Krohn KA, Lewis JS, Mason RP, Melillo G, Padhani AR, Powis G, Rajendran JG, Reba R, Robinson SP, Semenza GL, Swartz HM, Vaupel P, Yang D, Croft B, Hoffman J, Liu G, Stone H, Sullivan D: Hypoxia: importance in tumor biology, noninvasive measurement by imaging, and value of its measurement in the management of cancer therapy. Int J Radiat Biol. 2006; 82 (10):699-757.
- Graeber TG, Osmanian C, Jacks T, Housman DE, Koch CJ, Lowe SW, Giaccia AJ. Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumours. Nature. 1996; 379(6560): 88-91.
- Semenza GL, Wang GL. A nuclear factor induced by hypoxia via *de novo* protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. Mol. Cell. Biol. 1992; 12(12): 5447-5454.
- Wang GL, Semenza GL. Purification and Characterization of Hypoxia-inducible Factor 1. J. Biol. Chem. 1995; 270 (3): 1230-1237.
- Semenza GL. Targeting HIF-1 for cancer therapy. Nat. Rev. Cancer. (2003); 3: 721-732.
- Semenza GL. Evaluation of HIF-1 inhibitors as anticancer agents. Drug Discov. Today. 2007; 12: 853-859.
- Zhong H, de Marzo AM, Laughner E, Lim M, Hilton DA, Zagzag D, Buechler P, Isaacs WB, Semenza GL, Simons JW. Over expression of hypoxia-inducible factor 1alpha in common human cancers and their metastases. Cancer Res. (1999); 59 (22): 5830-5835.
- Birner P, Schindl M, Obermair A, Plank C, Breitenacker G, Oberhuber G. Over expression of hypoxia-inducible factor 1α is a marker for an unfavorable prognosis in early-stage invasive cervical cancer. Cancer Res. 2000; 60: 4693-4696.
- Bos R, Zhong H, Hanrahan CF, Mommers EC, Semenza GL, Pinedo HM, Abeloff MD, Simons JW, van Diest PJ, van der Wall E. Levels of Hypoxia-Inducible Factor-1α During Breast Carcinogenesis. J. Natl. Cancer Inst. 2001; 93: 309-314.
- Ryan HE, Poloni M, McNulty W, Elson D, Gassmann M, Arbeit JM, Johnson RS. Hypoxia-inducible Factor-1{alpha} Is a Positive Factor in Solid Tumor Growth. Cancer Res. 2000; 60: 4010-4015.
- Kung AL, Zabludoff SD, France DS, Freedman SJ, Tanner EA, Vieira A, Cornell-Kennon S, Lee J, Wang B, Wang J, Memmert K, Naegeli HU, Petersen F, Eck MJ, Bair KW, Wood AW, Livingston DM. Small molecule blockade of transcriptional coactivation of the hypoxia-inducible factor pathway. Cancer Cell. 2004; 6: 33-43.
- Unruh A, Ressel A, Mohamed HG, Johnson RS, Nadrowitz R, Richter E, Katschinski DM, Wenger RH. The hypoxia-inducible factor-1 alpha is a negative factor for tumor therapy. Oncogene. 2003; 22(21): 3213-3220.
- Moeller BJ, Dreher MR, Rabbani ZN, Schroeder T, Cao Y, Li CY, Dewhirst MW. Pleiotropic effects of HIF-1 blockade on tumor radiosensitivity. Cancer Cell. 2005; 8: 99-110.
- Liu Yang, Veena CK, Morgan JB, Mohammed KA, Jekabsons MB, Nagle DG, Zhou Yu-Dong. Methylalpinumisoflavone Inhibits Hypoxia-inducible Factor-1 (HIF-1) Activation by Simultaneously Targeting Multiple Pathways. The Journal of Biological Chemistry. 2009; 284 (9): 5859-5868.
- Ito C, Murata T, Itoigawa M, Nakao K, Kumagai M, Kaneda N, Furukawa H. Induction of Apoptosis by Isoflavonoids from the Leaves of *Millettia taiwaniana* in Human Leukemia HL-60 Cells. Planta Med. 2006; 72(5): 424-429.
- Okamoto Y, Suzuki A, Ueda K, Ito C, Itoigawa M, Furukawa H, Nishihara T, Kojima N. Anti-Estrogenic Activity of Prenylated

- Isoflavones from *Milletia pachycarpa*: Implications for Pharmacophores and Unique Mechanisms. Journal of Health Science. 2006; 52(2): 186-191.
34. Ito C, Itoigawa M, Kumagaya M, Okamoto Y, Ueda K, Nishihara T, Kojima N, Furukawa H. Isoflavonoids with anti-estrogenic activity from *Milletia pachycarpa*. J. Nat. Prod. 2006; 69: 138-141.
 35. Shin-Foon C, Sping L, Yee Som C. Insecticidal Action of *Milletia pachycarpa* Benth. J. Econ. Entomol. 1942; 35(1): 80-82(3).
 36. Mukherjee TD, Tripathy RL. Studies on indigenous insecticidal plants. Journal of Scientific and Industrial Research. 1956; 15C: 106-111.
 37. Bishnupada R, Shyamashree D, Veena T. Ultrastructural observations on tegumental surface of *Raillietina echinobothrida* and its alterations caused by root-peel extract of *Milletia pachycarpa*. Microscopy Research Technique. 2008; 71(11): 810-815.
 38. Perrett S, Whitfield PJ, Sanderson L, Bartlett A. The plant molluscicide *Milletia thonningii* (Leguminosae) as a topical antischistosomal agent. J Ethnopharmacol. 1995; 47: 49-54.
 39. Lyddiard, JR, Whitfield PJ, Bartlett A. Antischistosomal bioactivity of isoflavonoids from *Milletia thonningii* (Leguminosae). J. Parasitol. 2002; 88:163-170.
 40. Maillard M, Marston A, Hostettmann K. Search for Molluscicidal and Larvicidal Agents from Plants. American Chemical Society Symposium Series 534. American Chemical Society, Washington DC. 1993.
 41. Lyddiard JRA, Whitfield PJ. Inhibition of Site I mitochondrial electron transport by an extract of the seeds of *Milletia thonningii*: a potential mechanism for the plant's molluscicidal and schistosome larvicidal activity. Journal of Helminthology. 2001; 75, 259-265.
 42. Riederer P, Danielczyk W, Grunblatt E. Monoamine Oxidase-B Inhibition in Alzheimer's Disease. Neurotoxicology. 2004; 25: 271-277.
 43. Han XH, Hong SS, Hwang JS, Jeong SH, Hwang JH, Lee MH, Lee MK, Lee D, Ro JS, Hwang BY. Monoamine Oxidase Inhibitory Constituents from the Fruits of *Cudrania tricuspidata*. Arch. Pharm. Res. 2005; 28: 1324.
 44. Arruda MSP, De Araujo MQ, Lobo LT, De Souza FAP, et al. Potential allelochemicals isolated from *Pueraria phaseoloides*. Allelopathy Journal. 2005; 15 (2): 211-220.
 45. Swain T. Evolution of flavonoid compounds, 1975, in The Flavonoids (Harborne J. B, Mabry T. J and Mabry H eds) pp 109-1129, Chapman and Hall, Ltd., London.
 46. Harborne JB, Mabry TJ, Mabry H. The Flavonoids. Academic Press, New York. 1975.
 47. Smith DA, Banks SW. Formation and biological properties of isoflavonoid phytoalexins, 1986, in The Effects of Plant Flavonoids on Mammalian Cells: Implications for Inflammation, Heart Disease, and Cancer (Middleton, E Jr., Kandaswami C, and Theoharides T.C. Pharmacol Rev. 2000; 52:673-751.
 48. Barnes S, Kim H, Darley Usmar V, Patel R, Xu J, Boersma B, Luo M. Beyond ER alpha and ER beta: estrogen receptor binding is only part of the isoflavone story. J. Nutr. 2000; 130(3): 656S-657S.
 49. Akaza H, Miyana N, Takashima N, et al. Comparisons of percent equol producers between prostate cancer patients and controls: case-controlled studies of isoflavones in Japanese, Korean and American residents. Jpn. J. Clin. Oncol. 2004; 34(2): 86-9.
 50. Frankensfeld CL, McTiernan A, Aiello EJ, et al. Mammographic density in relation to daizein-metabolizing phenotypes in overweight, postmenopausal women. Cancer Epidemiol. Biomarkers Prev. 2004; 13(7): 1156-62.
 51. Fritz WA, Coward L, Wang J, Lamartiniere CA. Dietary genistein: perinatal mammary cancer prevention, bioavailability and toxicity testing in the rat. Carcinogenesis. 1998; 19: 2151-2158.
 52. Yanagihara K, Ito A, Toge T, Numoto M. Antiproliferative effects of isoflavones on human cancer cell lines established from the gastrointestinal tract. Cancer Res. 1993; 53: 5815-5821.
 53. Uckun FM, Evans WE, Forsyth CJ, Waddick, KG, Ahlgren LT, Chelstrom LM, Burkhardt A, Bolen J, Myers DE. Biotherapy of B-cell precursor leukemia by targeting genistein to CD19-associated tyrosine kinases. Science. 1995; 267: 886-891.
 54. Huang C-K. Protein kinases in neutrophils: A review. Membr Biochem. 1989; 8: 61-79.
 55. Taniguchi T, Miyazaki T, Minami Y, Kawahara A, Fujii H, Nakagawa Y, Hatakeyama M, Liu ZJ. IL-2 signaling involves recruitment and activation of multiple protein tyrosine kinases by the IL-2 receptor. Ann NY Acad Sci. 1995; 766: 235-244.
 56. Qian D, Weiss A. T cell antigen receptor signal transduction. Curr Opin Cell Biol. 1997; 9: 205-212.
 57. Okura A, Arakawa H, Oka H, Yoshinari T, Monden Y. Effect of genistein on topoisomerase activity and on the growth of [Val 12] Ha-ras-transformed NIH 3T3 cells. Biochem Biophys Res Commun. 1988; 157: 183-189.
 58. Akiyama T, Ishida J, Nakagawa S, Ogawara H, Watanabe S, Itoh N, Shibuya M, Fukami Y. Genistein, a specific inhibitor of tyrosine-specific protein kinases. J. Biol. Chem. 1987; 262: 5592-5595.
 59. Friedman DL, Kleiman NJ, Campbell FE. Nuclear protein phosphorylation in isolated nuclei from HeLa cells. Evidence that ³²P incorporation from [³²P] GTP is catalyzed by nuclear kinase II. Biochim Biophys Acta. 1985; 847:165-176.
 60. Carter John B, Saunders Venetia A. Virology: Principles and Applications. John Wiley & Sons Ltd, England. 2007.
 61. Spedding G, Ratty A, Middleton E. Inhibition of reverse transcriptases by flavonoids. Antiviral Res. 1989; 12: 99-110.
 62. Ono K, Nakane H, Fukushima M, Chermann J-C, Barre-Sinoussi F. Differential inhibition of the activities of reverse transcriptase and various cellular DNA polymerases by a traditional kampo drugs Sho-Saiko-To. Biomed. Pharmacother. 1990; 44: 13-16.
 63. Ono K, Nakane H, Fukushima M, Chermann J-C, Barre-Sinoussi F. Inhibition of reverse transcriptase activity by a flavonoid compound, 5-6-7-trihydroxyflavone. Biochem. Biophys. Res. Commun. 1989; 160: 982-987.
 64. Brinkworth RI, Stoermer MJ, Fairlie DP. Flavones are inhibitors of HIV-1 proteinase. Biochem Biophys Res Commun. 1992; 2: 631-637.
 65. Fesen MR, Kohn KW, Leteurtre F, Pommier Y. Inhibitors of human immunodeficiency virus integrase. Proc Natl Acad Sci USA. 1993; 90: 2399-2403.
 66. Curnutte JT, Babior BM. Chronic granulomatous disease. Adv Hum Genet. 1987; 16:229-297.
 67. Babior BM, Woodman RC. Chronic granulomatous disease. Semin Hematol. 1990; 27: 247-259.
 68. Davies KJA. Oxidative Damage and Repair. Pergamon Press, New York. 1991.
 69. Sies H. Oxidative Stress: From basic research to clinical application. Am J Med. 1991; 91: 31S-38S.
 70. Halliwell B, Gutteridge JMC, Cross CE. Free radicals, antioxidants, and human disease: Where are we now? J Lab Clin Med. 1992; 119: 598-620.
 71. Sies H, Krinsky NI. The present status of antioxidant vitamins and beta-carotene. Am J Clin Nutr. 1995; 62: 1229S-1300S.
 72. Krinsky NI. Overview of lycopene, carotenoids and disease prevention. Proc Soc Exp Biol Med. 1998; 218(2): 95-97.
 73. Krinsky NI. The antioxidant and biological properties of carotenoids. Ann NY Acad Sci. 1998; 854: 443-447.
 74. Castelluccio C, Paganga G, Melikian N, Bolwell GP, Pridham J, Sampson J, Rice-Evans C. Antioxidant potential of intermediates in phenylpropanoid metabolism in higher plants. FEBS Lett. 1995; 368: 188-192.
 75. Salah N, Miller NJ, Paganga G, Tijburg L, Bolwell GP, Rice-Evans C. Polyphenolic flavonols as scavengers of aqueous phase radicals and as chain-breaking antioxidants. Archives of Biochemistry and Biophysics. 1995; 322(2): 339-346.
 76. Witztum JL, Steinberg D. Role of oxidized low density lipoprotein in atherogenesis. J Clin Invest. 1991; 88: 1785-1792.
 77. Ferrali M, Signorini C, Caciotti B, Sugherini L, Ciccoli L, Giachetti D, Comproti M. Protection against oxidative damage of erythrocyte membrane by the flavonoid quercetin and its relation to iron chelating activity. FEBS Lett. 1997; 416: 123-129.

78. Elliott AJ, Scheiber SA, Thomas C, Pardini RS. Inhibition of glutathione reductase by flavonoids. A structure-activity study. *Biochem Pharmacol.* 1992; 44: 1603-1608.
79. Hirano R, Sasamoto W, Matsumoto A, Itakura H, Igarashi O, Kondo K. Antioxidant ability of various flavonoids against DPPH radicals and LDL oxidation. *J Nutr Sci Vitaminol (Tokyo).* 2001; 47: 357-362.
80. Cos P, Ying L, Calomme M, Hu JP, Cimanga K, Van Poel B, Pieters L, Vlietinck AJ, Vanden Berghe D. Structure-activity relationship and classification of flavonoids as inhibitors of xanthine oxidase and superoxide scavengers. *J Nat Prod.* 1998; 61: 71-76.
81. Frenkel K, Chrzan K, Ryan C, Wiesner R, Troll W. Chymotrypsin specific protease inhibitors decrease H₂O₂ formation by activated human polymorphonuclear leukocytes. *Carcinogenesis.* 1987; 8: 1207-1212.
82. Hara Y, Matsuzaki T, Suzuki T. Angiotensin I converting enzyme activity of tea components. *Nippon Nogeikagaku Kaishi.* 1987; 61, 803-808 (in Japanese).
83. Fukai K, Ishigami T, Hara Y. Antibacterial activity of tea polyphenols against phytopathogenic bacteria. *Agric.Biol.Chem.* 1991; 55: 1895-1897.
84. Toda M, Okubo S, Hiyoshi R, Shimamura T. The bacterial activities of tea and coffee. *Lett. in Appl. Microbiol.* 1989; 8: 123-125.
85. Toda M, Okubo S, Ikigai H, Suzuki T, Shimamura T. The protective activity of the tea against infection by *Vibrio cholera*. *J. Appl. Bacteriol.* 1991; 70: 109-112.
86. Nakayama M, Toda M, Okubo S, Shimamura T. Inhibition of influenza virus infection by tea. *Lett. Appl. Microbiol.* 1990; 11: 38-40.
87. Bersuker BI, Bahçeci S, Boggs JE. Improved Electron-Conformational Method of Pharmacophore Identification and Bioactivity Prediction. Application to Angiotensin Converting Enzyme Inhibitors. *J. Chem. Inf. Comput. Sci.* 2000; 40 (6): 1363-1376.
88. Kingsford-Adaboh R, Osei-Fosu P, Asomaning WA, Weber M, Luger P. The Crystal Structures of O, O-Dimethylalpinumisoflavone and 5-O-Methyl-4'-O-(3-methylbut-2-en-1-yl)alpinumisoflavone. *Cryst. Res. Technol.* 2001; 36:107-115.
89. Kingsford-Adaboh R, Dittrich B, Hubschle CB, Gbewonyo WSK, Okamoto H, Kimura M, Ishida H. Invariom structure refinement, electrostatic potential and toxicity of 4-O-methylalpinumisoflavone, O,O-dimethylalpinumisoflavone and 5-O-methyl-4-O-(3-methylbut-2-en-1-yl)alpinumisoflavones. *Acta Crystallogr.* 2006; B62:843-849.
90. Harrison JJEK, Tabuchi Y, Ishida H, Kingsford-Adaboh R. Alpinumisoflavone. *Acta Cryst.* 2008; E64:o713.
91. Harrison JJEK, Tabuchi Y, Ishida H, Kingsford-Adaboh R. Crystal Structure of three solvated alpinumisoflavones. *Structural Chemistry.* 2008; 20(2): 203-211.
92. Behl C. The impact of antioxidants on chronic diseases in ageing and old age. *Int. J. Vitam. Nutr. Res.* 1999; 69 (3): 146-149.
93. Scott G. Antioxidants-the modern elixir? *Chemistry in Britain.* 1995; 879-882.
94. Anderson RF, Amarsinghe C, Fisher LJ, Mak WB, Packer JE. Reduction in free-radical-induced DNA strand breaks and base damage through fast chemical repair by flavonoids. *Free Radical Research.* 2000; 33 (1): 91-103.
95. Kasbauer CW, Paper DH, Franz G. Sulfated (β1-4) galato-oligosaccharide and their effects on angiogenesis. *Carbohydrate research.* 2001; 330: 427-430.
96. Nia R, Paper DH, Essien EE, Iyadi KC, Bassey AIL, Antai AB, Franz G. Evaluation of the Anti-oxidant and Anti-angiogenic effects of *Sphenocentrum jollyanum pierre*. *African Journal of Biomedical Research.* 2004; 7(3): 129-132.