Academic Sciences

International Journal of Pharmacy and Pharmaceutical Sciences

ISSN- 0975-1491

Vol 3, Issue 4, 2011

Research Article

DEVELOPMENT OF FORMULATION CONTAINING EXTRACTS OF ARISTOLOCHIA BRACTEATA AND CASSIA TORA – EVALUATION OF ANTIMICROBIAL ACTIVITY

JAYASUTHA J^{1*}, MONIC JOSEPHINE NITHILA S², VEERABATHINI RAJINIKANTH²

¹SRM College of Pharmacy, SRM University, Kattankulathur, Kancheepuram, Tamilnadu, India, ²Brown's College of Pharmacy, Khammam, Andhra Pradesh, India Email: jayasutha15@gmail.com

Received: 1 Aug 2011, Revised and Accepted: 9 Sep 2011

ABSTRACT

The aqueous and ethanolic extract of *Aristolochia bracteata* and *Cassia tora* was screened for antimicrobial activity having the concentration of 2 mg, 4 mg and 6 mg by disc diffusion method, using nutrient agar medium against Gram positive bacteria such as Staphylococcus aureus, Staphylococcus albus, Bacillus subtilis, Streptococcus faecalis and Gram negative bacteria such as Escherichia coli, Klebshiella aerogenes, Proteus vulgaris and Pseudomonas aeruginosa. Then the extract also tested against fungi Candida albicans and Aspergillus niger. *Ciprofloxacin* and *Clotrimazole* are used as a reference standard for bacteria and fungi respectively. Among the two extracts of *Aristolochia bracteata* and *Cassia tora*, ethanol extract was found to have the significant activity against bacteria. Nil effect at 2 & 4mg levels and very minimum effect at 6mg level produced by the aqueous extracts of both the plants. Antifungal activity assessment also indicates that the tested fungal strains are more susceptible to aqueous extract of both the plants.

Keywords: Aristolochia bracteata, Cassia tora, Antibacterial activity, Antifungal activity

INTRODUCTION

Aristolochia bracteata is used in traditional medicine as a gastric stimulant and in the treatment of cancer, lung inflammation, dysentery and snake bites¹. This plant belongs to the family Aristolochiaceae. It has insecticidal properties. Its roots and leaves are bitter and antihelmintic, and are medicinally important. Almost every part of the plant has medicinal usage. Identifying bioactive compounds and establishing their health effects are active areas of scientific enquiry². *Cassia tora* (Leguminosae) is a wild crop and grows in most parts of India as a weed. According to ayurveda, the leaves and seeds are useful in leprosy, ringworm, flatulence, colic, dyspepsia, constipation, cough, bronchitis, cardiac disorders⁴. The present study aims to determine the antimicrobial activity of *Aristolochia bracteata* and *Cassia tora*.

MATERIALS AND METHODS

Collection of the plant materials

The plants *Aristolochia bracteata* and *Cassia tora* is widely found throughout India. For our work the plants were collected in and around Thanjavur, Tamilnadu, India. The leaves were collected and they are dried in room temperature. Then they are crushed in to small pieces and these dried small pieces are finely powdered.

Extraction

The extraction is done by maceration process and the powder material was extracted with water and ethanol. About one hundred grams of the powdered leaf material was mixed with four hundred ml (1:4) of the above solvents and subjected to extraction by cold maceration method. The set up were kept at room temperature for seven days for complete extraction during the period of maceration, the content was shaken occasionally to mix and enhance extraction. After seven days the extract were filtered through muslin cloth. The extract was collected in suitable container and it was subjected to concentration by evaporation.

Preliminary qualitative phytochemical examination of aqueous and ethanolic extracts of *Aristolochia bracteata* and *Cassia tora*

Both the extracts were evaluated for preliminary phytochemical screening for identification of its active constituents like alkaloids, glycosides, phytosterol, fixed oils and fats, saponins, tannins and phenolic compounds. Proteins and free amino acids, gums, mucilages, flavonoids, lignin.

Antimicrobial Screening

The aqueous and ethanolic extracts of Aristolochia bracteata and Cassia tora was screened for antimicrobial activity by disc diffusion method, using nutrient agar medium against Gram positive bacteria such as Staphylococcus aureus, Staphylococcus albus, Bacillus subtilis, Streptococcus faecalis and Gram negative bacteria such as Escherichia coli, Klebshiella aerogenes, Proteus vulgaris and Pseudomonas aeruginosa. Then the extract also tested against fungi Candida albicans and Aspergillus niger. Ciprofloxacin and Clotrimazole are used as a reference standard for bacteria and fungi respectively. The plates were divided in to 5 sections according to the number of standards and sample solution to be used with the help of the glass marker. Sterile cotton swab was dipped in to the inoculums; excess fluid was removed by rotating the swab with firm pressure. The standard disc and the sterile discs loaded with 2, 4 & 6 mg of the extracts were placed into one section of the plate. Then the plates were incubated at 37°C for 18 - 24 hours till perfect growth was observed. The inhibitory zone was measured with the help of an antibiotic zone reader. The corresponding solvents used for extraction acts as control.

RESULTS AND DISCUSSIONS

The plant materials of *Aristolochia bracteata* and *Cassia tora* were selected for the study to evaluate the formulation containing the extracts of these plants for their antimicrobial activity. Aqueous and ethanolic extracts of these plants were made by cold maceration method and the obtained extracts concentrated by evaporation and preliminary phytochemical screening were made for the obtained extracts by standard procedure.

Table 1 reveals the presence of various phytoconstituents. Both aqueous and ethanolic extracts of *Aristolochia bracteata* showed the presence of alkaloids, saponins, phytosterol, fixed oils & fats, proteins and amino acids. Presence of gums and mucilage observed in aqueous and flavonoids in ethanol extract respectively according to their solubility.

Table – 2, reveals the phytoconstituents of *Cassia tora* in aqueous and ethanol extract. Both the extracts showed the presence of alkaloids, carbohydrates, glycosides, phytosterols, fixed oils & fats and flavonoids. The presence of gums & mucilage observed in aqueous extract.

The aqueous and ethanolic extracts of both plant materials were screened for their antimicrobial potential by standard disc diffusion technique against various clinical pathogens. Also the effect was assessed at three different levels of 2, 4, & 6mg of the extracts. The inhibitory effect produced by the extracts was compared with effect produced by the standard drug *Ciprofloxacin* 5µg and *Clotrimazole* 10µg was used as reference standard.

Jayasutha et al.

Table 1: Phytochemical investigation of Aristolochia bracteata extracts

S. No	Constituents	Aqueous	Ethanol	Method
1	Alkaloids	+	+	Mayers reagent, Hagers and Wagners reagent
2	Carbohydrates	_	_	Molisch's test
3	Glycosides	_	_	Borntrager's test
4	Phytosterols	+	+	Libermann burchard
5	Saponins	+	+	Agitated with distilled water \rightarrow foam
6	Fixed oils & fats	+	+	KOH + Phenolphthalein
7	Tannin & phenols	_	_	Fecl3, Lead acetate
8	Protein & Amino acids	+	+	Millons reagent
9	Gums & mucilages	+	_	Alcohol
10	Flavonoids	_	+	Con. H ₂ SO ₄
11	Lignin	_	_	Phloroglucinol HCl

Table 2: Phytochemical investigation of Cassia tora extracts

S. No	Constituents	Aqueous	Ethanol	Method
1	Alkaloids	+	+	Drangondroffs reagent
2	Carbohydrates	+	+	Molisch's test
3	Glycosides	+	+	Borntrager's test
4	Phytosterols	+	+	Libermann burchard
5	Saponins	_	_	Distilled water
6	Fixed oils & fats	+	+	KOH + Phenolphthalein
7	Tannin & phenols	_	_	Fecl ₃ , Lead acetate
8	Protein & Amino acids	_	_	Millons reagent
9	Gums & mucilages	+	_	Alcohol
10	Flavonoids	+	+	Con H ₂ SO ₄
11	Lignin	_	_	Phloroglucinol HCl

Table 3: Antimicrobial activity of extracts of Aristolochia bracteata

S.	Test Organism	Diameter of zone of inhibition (mm)							
No		Standard	Aqueous Extracts			Ethanolic Extracts			
		Drug	S1 (2mg)	S ₂ (4mg)	S₃ (6mg)	S1 (2mg)	S ₂ (4mg)	S₃ (6mg)	
1	Staphylococcus aureus	29	-	-	7	10	12	13	
2	Staphylococcus albus	36	-	-	7	9	11	12	
3	Streptococcus faecalis	37	-	-	-	9	11	12	
4	Bacillus subtilis	25	-	-	8	8	11	13	
5	Pseudomonas aerogenosa	32	-	-	6	7	8	12	
6	Escherichia coli	27	-	-	7	11	11	21	
7	Klebshiella aerogenes	22	-	-	6	12	12	15	
8	Proteus vulgaris	28	-	-	-	13	14	16	
9	Candida albicans	10	-	-	-	10	11	13	
10	Aspergillus niger	11	-	-	-	13	14	14	

Ciprofloxacin 5µg/disc for bacteria, Clotrimazole 10µg/disc for fungi, Solvent: Ethanol

S. No	Test Organism	Diameter of zone of inhibition							
		Standard Aqueous Extracts			Ethanolic Extracts				
		Drug	S1 (2mg)	S2 (4mg)	S₃ (6mg)	S1 (2mg)	S2 (4mg)	S₃ (6mg)	
1	Staphylococcus aureus	31	-	-	8	11	11	18	
2	Staphylococcus albus	36	-	-	7	10	13	14	
3	Streptococcus faecalis	38	-	-	6	7	10	12	
4	Bacillus subtilis	25	-	-	7	7	11	12	
5	Pseudomonas aerogenosa	32	-	-	8	11	12	13	
6	Escherichia coli	26	-	-	7	12	12	13	
7	Klebshiella aerogenes	22	-	-	7	12	12	14	
8	Proteus vulgaris	28	-	-	6	11	12	12	
9	Candida albicans	10	-	-	-	8	9	10	
10	Aspergillus niger	12	-	-	-	13	14	18	

Ciprofloxacin 5 µg/disc for bacteria, Clotrimazole 10 µg/disc for fungi, Solvent: Ethanol

Table 3 & 4 showed the antimicrobial effect produced by the plant extracts against various pathogenic bacteria and fungi. This reveals that nil effect at 2 & 4mg levels and very minimum effect at 6mg level produced by the aqueous extracts of both the plants. In both ethanolic plant extracts showed increased inhibitory effect with respect to increase in level of drug concentration used against the test organisms.

The effect produced by ethanolic extract of *Aristolochia bracteata* (Table 3) was being maximum with gram negative bacteria [Escherichia coli (21mm), Kliebshiella aerogenes (15mm) and Proteus vulgaris (16mm)] than the other organisms under test. The effect produced against the fungi was being maximum as comparable with the effect of standard drug.

The ethanolic extract of *Cassia tora* (Table 4) showed higher inhibition against Staphylococcus aureus (18mm) and effect against other gram positive and gram negative bacteria under study is being moderate. The inhibition produced against fungi was found to be maximum as compared with the effect of standard drug.

CONCLUSION

We conclude that ethanolic extracts of *Aristolochia bracteata* and *Cassia tora* have remarkable antimicrobial activity. Aqueous extracts did not show significant antimicrobial activity against the microorganism under study

REFERENCES

- 1. Negi PS, Anandharamakrishnan C, Jayaprakasha GK. Antibacterial activity of *Aristolochia bracteata* roots extracts. Journal of Medicinal Food 2003; 6(4): 401-403.
- Etherton PM, Lefever M, Beecher GR, Gross MD, Keen LL, and Eiborton TD. Bioactive compounds in nutrition and health research methodologies for establishing the biological function- the antioxidant and anti-inflammatory effects of flavonoids against Atherosclerosis. Annual Review of Nutrition 2004; 24: 511-538.
- 3. Ahmad I, Mehmood Z, Mohammad F. Screening of some Indian medicinal plants for their antimicrobial properties. Journal of Ethanopharmacology 1998; 62: 183-193.
- 4. Chan MJ, Peria LM. Plant natural products with leishmanicidal activity. Natural Product Report 2001; 18: 674-688.

- 5. Desta B. Ethiopian traditional herbal drugs. Part II: Antimicrobial activity of 63 medicinal plants. Journal of Ethanopharmacology 1993; 39: 129-139.
- Karaman I, Sahin F, Gulluce M, Ogutchu H, Sengul M, Adiguzel A. Antimicrobial activity of aqueous and methanol extracts of *Juniperus oxycedrus* L. Journal of Ethanopharmacology 2003; 28(37): 1-5
- Pinner RS, Teustch L, Simenson L, Klug J, Clarke MG, Berkelman R. Trends in infectious diseases mortality in the United States. The Journal of the American Medical Association 1996; 275: 189-193.
- 8. Ambersing Rajput, Suboth Chandra Pal, Bhagavan Patil. Phytochemical screening, antibacterial activity and physicochemical evaluation of leaves of *Butea Monosperma*. 2011; 3(suppl 3): 189-91.
- 9. Devjani Chakraborty, Barkha Shah. Antimicrobial, antioxidative and anti-hemolytic activity of *Piper betel* leaf extracts. 2011; 3(suppl 3): 192-99.
- Schlumbam A, Mauch F, Vogeli U, Bollar T. Plant Chitinases are potent inhibitors of fungal growth. Nature 1986; 324: 365-367.
- 11. Seth SD, Sharma S, Bhawana S. Antibacterial activity of *Ocimum* sanctum *L*.fixed coil. Indian Journal of Experimental Biology 2004; 43(9): 835-837.
- Silva O, Duarte A, Pimental M, Viegas S, Barroso H, Machado J, Pires I, Cabrita J, Gomes E. Antimicrobial activity of *Terminalia macro ptera* root. Journal of Ethanopharmacology 1997; 57: 203-207.
- 13. Sushil Chandra Sati, Nitin Sati, O.P. Sati. Chemical investigation and screening of antimicrobial activity of stem bark of *Quercus leucotrichophora*. International Journal of Pharmacy and Pharmaceutical Sciences. 2011; 3(suppl 3): 89-91.