



## OPTIMIZATION OF PROCESS VARIABLES FOR THE PREPARATION OF CHITOSAN-ALGINATE NANOPARTICLES

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### ABSTRACT

The aim of this study is to prepare and drug loaded nanoparticles of Gemcitabine, an anticancer drug and optimization in terms of chemical properties, drug concentration, polymer concentration, cross-linking agent and stirring speed. Nanoparticles of Gemcitabine were fabricated using chitosan polymer and pregelated Sodium alginate by Iontropic pregelation method. Calcium chloride was also included in the formulation for pregelation of sodium alginate. Prepared chitosan and alginate suspension further crosslinked with Glutaraldehyde. Different formulations of nanoparticles were prepared using different concentrations of chitosan, stirring speed, time of rotation and polymer to drug ratio in the nanoparticles. The average particle size ranged between 230 nm to 627 nm. Drug entrapment ranged between 72.12%-92.89%. The result indicated that the drug loaded nanoparticles of Gemcitabine showed optimum particle size and maximum drug entrapment with drug polymer ratio 05:75, cross-linking agents 02 ml, stirring rate 800 rpm and stirring time 90 min.

**Keywords:** Nanoparticles, Chitosan, Sodium alginate, Calcium chloride, ionotropic pre gelation

### INTRODUCTION

The colloidal carriers based on biodegradable and bio-compatible polymeric systems have largely influenced the controlled and targeted drug delivery concept. Nanoparticles are sub-nanosized colloidal structures composed of synthetic or semi-synthetic polymers<sup>1</sup>.

Colloidal drug delivery systems offer a number of advantages over conventional dosage forms. Due to their small particle size, colloidal preparations lend themselves to parenteral administration and may be useful as sustained release injections for the delivery to a specific organ or target site. Targeting the drug to the desired site of action would not only improve the therapeutic efficiency but also enable a reduction of the amount of drug which must be administered to achieve a therapeutic response, thus minimizing unwanted toxic effects<sup>2</sup>.

Nanoparticles useful for sustained drug release can be also obtained by electrostatic interaction between alginate and chitosan<sup>3</sup>. Both alginate and chitosan have been widely used in drug delivery<sup>4</sup>. Chitosan is a natural cationic polysaccharide derived by deacetylation of chitin, a copolymer consisting of combined units of glucosamine and N-acetyl glucosamine<sup>5, 6</sup>. In the pharmaceutical field chitosan's advantageous biological properties have prompted its extensive study as a carrier both of drugs<sup>7, 8</sup> and of proteins<sup>9, 10</sup>. Drug loaded nanoparticles made of polyelectrolytes complexation have shown potential for use as drug delivery systems<sup>11, 12</sup>. Polyelectrolyte complexes are formed by interactions between macromolecules that carry oppositely charged ionizable groups<sup>13</sup>. A more selective drug delivery was achieved using water soluble drug-polymer conjugates<sup>14</sup>.

Gemcitabine, a nucleoside analog related to cytarabine, is one of the most effective cytotoxic agents for non small cell lung cancer (NSCLC). It is a pyrimidine antimetabolite that is anabolized sequentially to the nucleoside monophosphate, diphosphate, and triphosphate intracellularly. This drug may be a cell cycle-specific agent inhibiting DNA synthesis, and it also induces apoptosis<sup>15</sup>.

### MATERIALS AND METHODS

Chitosan (deacetylation degree 85%) low MW (50 kDa) was obtained as a gift sample from central institute of fishre technology (Trivendram, Kerala). Sodium alginate (low viscosity), Calcium chloride and Glutaraldehyde of analytical grade were purchased from Loba chemicals (Pune). Gemcitabine pharmaceutical grade (as per USP) was obtained as a gift sample from Shilpa Medicare Limited, (Raichur), Karnataka, India. Glacial acetic acid of analytical

grade was procured from Qualigens Fine Chemicals. A549 human non small cell lung cancer cell line Purchased from NCCL (Pune). All other chemicals were of analytical grade and used as received. Double distilled water was used throughout the study. Magnetic stirrer was used of Rami.

### Preparation of drug-loaded alginate nanoparticles

Alginate/chitosan particles were prepared in a two-step procedure based on the ionotropic pre-gelation of polyanion with calcium chloride followed by polycationic crosslinking through an adapted protocol initially described<sup>13</sup>, but modified according to ideal pre-gelation stoichiometric ratio and time of drug association by Peniche-Covas et al 1995. 7.5 ml of 18 mM calcium chloride solution was added drop wise for 60 min under gentle stirring (800 rpm) into a beaker containing 117.5 ml of a 0.063% alginate solution to provide an alginate pre-gel. Then, 25 ml of different concentration (0.05-0.09%) chitosan solution was added drop wise into the pre-gel over 90 min. The pH of alginate and chitosan solutions was initially set to 4.9 and 4.6, respectively. A colloidal dispersion at pH 4.7 formed upon polycationic chitosan addition, visible as the Tyndall effect. Nanoparticles were stirred for 30 min to improve curing and subsequently collected by centrifugation (20,000g/45 min) at 4°C. For Gemcitabine-loaded nanoparticles, 5 mg of insulin was mixed with the alginate solution before calcium chloride addition.

Drug-loaded nanoparticles were recovered by centrifugation at 19,000 rpm for 30-45 min and washed thrice with distilled water to obtain the final pellet.

Glutaraldehyde (GLA) cross-linking nanoparticles were prepared as follows: a known mass of 0.25% (w/w) GLA solution was dropped in CS-ALG suspension or drug loaded CS-ALG suspension under magnetic stirring. This mixture was further stirred for three hours under room temperature (G-CS-ALG) [13].

### Optimization

#### Optimization of formulation variables

Various formulation variables were optimized to prepare nanoparticle viz. polymer concentration and cross-linking agent concentrations. The effect of these variables on the particle size, shape, size distribution entrapment efficiency was studied.

#### Optimization of process variables

Process various variables that could affect the preparation and properties of final preparations were optimized i.e. stirring speed

(500, 600, 700, 800, 1000 rpm.) and stirring time (hrs.). Effect of these variables on particle size, shape, size distribution and entrapment efficiency was studied.

#### Estimation of entrapped drug in nanoparticle

The 50 mg of nanoparticles was dispersed in 50 ml of PBS pH 7.4 for 24 hrs. and homogenate was centrifuged at 300rpm for 5 min., and the supernatant was assayed for 5-FU, spectrophotometrically. The percentage drug entrapped was calculated and reported in table (1, 2, 3, 4 and 5).

#### RESULTS AND DISCUSSION

Using Ionotropic pregelation method, average particle size and entrapment with high drug contents could be produced (Fig. 1,2,3,4 & 5).

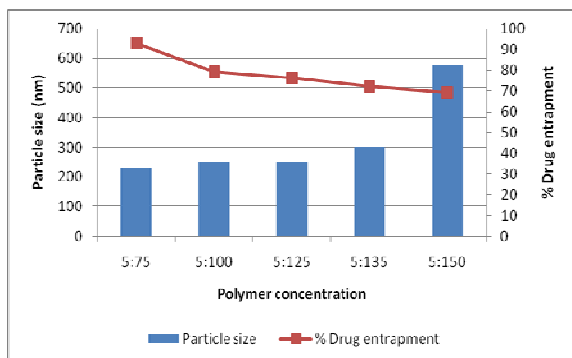
In the study of drug: polymer (Sodium alginate) ratio, Sodium alginate concentration was effected the particle size from 230-575 and entrapment of drug and 92.89%-69.00%.

The drug concentration also effect the partical size and drug entrapment with respect the concentration of chitosan from 229-480 and 69.97%-91.98% respectively. The same parameters were affected with using the different concentration of cross-linking agents 232-589 and 83.65%-90.88%.

Table no. 4. Indicates that the effect of stirring rate on the particle size and drug entrapment. An increase in stirring speed, the particle size of nanoparticles was reduced from 627-236 and entrapment of drug content was increases from 78.00%-90.00%. Stirring time (at 800 rpm) also effected the nanoparticle size and drug entrapment from 557-236 and 78.00%-90.90%.

**Table 1: Effect of drug: polymer ratio on particle size and size distribution of nanoparticle.**

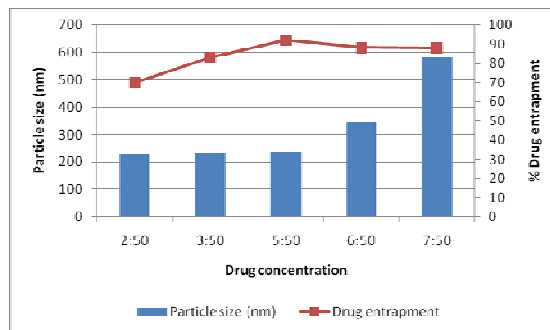
S.No.	Drug: polymer (sodium alginate) Ratio (mg)	Average particle size (nm)	Drug entrapment (%)
1	05:75	230	92.89
2	05:100	249	79.22
3	05:125	247	76.34
4	05:135	301	72.12
5	05:150	575	69.00



**Fig. 1: Effect of drug: polymer ratio on particle size and size distribution of nanoparticle.**

**Table 2: Effect of drug: polymer (chitosan) ratio on the particle size and size distribution of nanoparticle.**

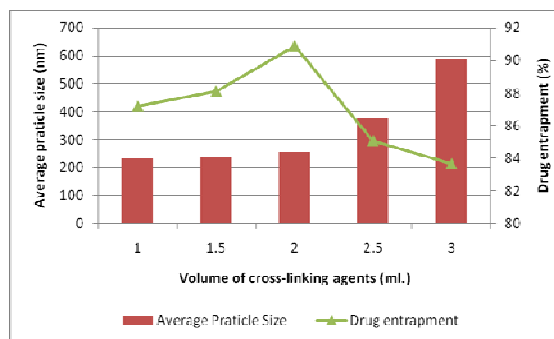
S.No.	Drug:Polymer Ratio (mg)	Average partical size (nm)	Drug Entrapment (%)
1	02:50	229	69.97
2	03:50	233	83.00
3	05:50	237	91.98
4	06:50	345	88.12
5	07:50	580	88.00



**Fig. 2: Effect of drug: polymer (Chitosan) ratio on the particle size and size distribution of nanoparticle.**

**Table 3: Effect of cross-linking agent concentration on particle size and size distribution of nanoparticle**

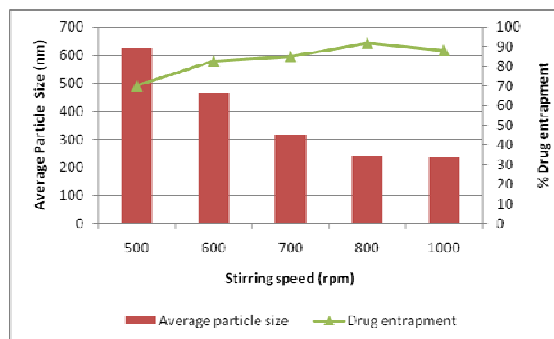
S.No.	Volume of cross-linking agents (ml.)	Average particle size (nm)	Drug entrapment (%)
1	01	232	87.20
2	1.5	239	88.11
3	02	257	90.88
4	2.5	376	85.08
5	03	589	83.65



**Fig. 3: Effect of cross-linking agent concentration on particle size and size distribution of nanoparticle.**

**Table 4: Effect of stirring rate on the particle size and size distribution of nanoparticle**

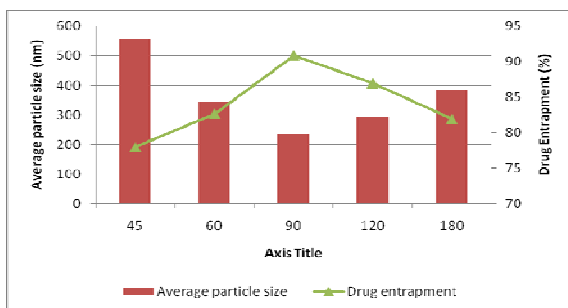
S.No.	Stirring speed (rpm)	Average particle size (nm)	Drug entrapment (%)
1	500	627	70.16
2	600	465	82.54
3	700	317	85.00
4	800	241	91.65
5	1000	236	87.90



**Fig. 4: Effect of stirring rate on the particle size and size distribution of nanoparticle.**

**Table 5: Effect of stirring time at 800 rpm on the particle size and size distribution of nanoparticle**

S.No.	Stirring time (min.)	Average Particle Size (nm)	Drug Entrapment (%)
1	45	557	78.00
2	60	345	82.67
3	<b>90</b>	<b>236</b>	<b>90.90</b>
4	120	295	87.00
5	180	385	81.94



**Fig. 5: Effect of stirring time at 800 rpm on the particle size and size distribution of nanoparticle.**

#### CONCLUSION

In conclusion, Ionotropic pregelation method can produce chitosan-alginate nanoparticles with optimum particle size and maximum entrapment of drug contents. The physical properties/parameters of nanoparticles can be varied by changing a number of process variables.

#### REFERENCES

1. Vivek Kumar Gupta, P.K.Karar, S.Ramesh, S.P.Mishra, Alok Gupta. Nanoparticle Formulation for Hydrophilic & Hydrophobic Drugs. *Int. J. Res. Pharm. Sci.*2010; Vol-1, Issue-2:163-169.
2. Kreuter J. Nanoparticle-based drug delivery systems. *Journal of controlled release.*1991; 16:169-176.

3. S.J. De, D. Robinson, Polymer relationships during preparation of chitosan-alginate and poly-L-lysine-alginate nanospheres. *J. Control. Release.*2003; 89 (1):101-112.
4. O. Skaugrud, A. Hagen, B. Borgersen, M. Dornish. Biomedical and pharmaceutical applications of alginate and chitosan. *Biotechnol. Genet. Eng. Rev.*1999; 16:23-40.
5. O.S. Lee, B.J. Ha, S.N. Park, Y.S. Lee. Studies on the pH-dependent swelling properties and morphologies of chitosan/calcium-alginate complexed beads. *Macromol. Chem. Phys.*1997; 198:2971-2976.
6. M.N.V. Ravi Kumar, A review of chitin and chitosan applications. *React. Funct. Polym.* 2001; 46:1-27.
7. M.A. Bayomi, S.A. Al-Suwayeh, A.M. El-Helw, A.F. Mesnad. Preparation of casein-chitosan microspheres containing diltiazem hydrochloride by an aqueous coacervation technique. *Pharm. Acta Helv.*1998; 73:187-192.
8. F.L. Mi, H.W. Sung, S.S. Shyu, Release of indomethacin from a novel chitosan microsphere prepared by naturally occurring crosslinker: examination of crosslinking and polycation/anionic drug interaction. *J. Appl. Polym. Sci.* 2001; 81:1700-1711.
9. P. Calvo, C. Remunan-Lopez, J.L. Vila-jato, M.J. Alonso. Novel hydrophilic chitosan-polyethylene oxide nano-particles as protein carriers. *J. Appl. Polym. Sci.*1997; 63:125-132.
10. B. Sarmiento, D.C. Ferreira, L. Jorgensen, M. van de Weer. Probing insulin's secondary structure after entrapment into alginate/chitosan nanoparticles. *European Journal of Pharmaceutics and Biopharmaceutics.*2007; 65:10-17.
11. D. Lochmanna, E. Jaukb, A. Zimmer. Drug delivery of oligonucleotides by peptides. *Eur. J. Pharm. Biopharm.*2004; 58:237-251.
12. W.P. Cheng, A.I. Gray, L. Tetley, T.L.B. Hang, A.G. Schatzlein, I.F. Uchegbu. Polyelectrolyte nanoparticles with high drug loading enhance the oral uptake of hydrophobic compounds. *Biomacromolecules.*2006; 7:1509-1520.
13. M. Rajaonarivony, C. Vauthier, G. Couarraze, F. Puisieux, P. Couvreur. Development of a new drug carrier made from alginate. *J. Pharm. Sci.*1993; 82:912-917.
14. C. Peniche-Covas, W. Argu'elles-Monal. Sorption and desorption of water vapour by membranes of the polyelectrolyte complex of chitosan and carboxymethyl cellulose, *Polym. Int.*1995; 38:45-52.
15. Goodman & Gilman's. The pharmacological basis of therapeutics.2006; 11:1346-1347.