

SCREENING OF FIFTEEN INDIAN AYURVEDIC PLANTS FOR ALPHA-GLUCOSIDASE INHIBITORY ACTIVITY AND ENZYME KINETICS

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ABSTRACT

Alpha-glucosidase inhibitory activity of fifteen Indian medicinal plants has been evaluated by *in vitro* enzyme assay. Methanol extracts of *Cyperus rotundus* (tubers), *Plumbago zeylanica* (root), *Symplocos racemosa* (bark), and *Terminalia arjuna* (bark) had displayed 100% inhibition with the IC₅₀ value of 3.98 µg/ml, 3.46 µg/ml, 8.16 µg/ml and 0.69 µg/ml, respectively. Bark extract of *Terminalia arjuna* is highly effective against alpha-glucosidase activity even at nanogram concentration. Plant parts of *Piper retrofractum* (stem), *Zingiber officinale* (rhizome), *Acorus calamus* (rhizome), *Picrorhiza kurroa* (rhizome), *Marsdenia tenacissima* (stem), *Clerodendron serratum* (root), and *Rubia cordifolia* (root) are not effective and they require high concentration to exhibit inhibition. Potential plants that show maximum inhibition at low concentration (<10 µg/ml) were subjected to kinetic analysis to determine the mode of inhibition of the enzyme. *Cyperus rotundus*, *Symplocos racemosa* and *Terminalia arjuna* exhibited uncompetitive inhibition and *Plumbago zeylanica* had displayed mixed inhibition to alpha-glucosidase enzyme activity. From the enzyme assay, we infer that *Cyperus rotundus*, *Plumbago zeylanica*, *Symplocos racemosa* and *Terminalia arjuna* contain potential alpha-glucosidase inhibitors that can be exploited for its use in the treatment of diabetes.

Keywords: Alpha-glucosidase inhibition, Kinetics, Medicinal plants, Diabetes

INTRODUCTION

The world prevalence of diabetes among adults (aged 20–79 years) was 6.4%, affecting 285 million adults (2010), and will increase to 7.7% and 439 million adults by 2030¹. In India, the number of adults with diabetes was 50.8 million (2010) and will be expected to reach 87 million (2030), with a mean annual increment of 1.8 million¹. Type 2 diabetes mellitus (T2DM) is a heterogeneous metabolic disorder, characterized by defects in insulin secretion and insulin sensitivity². Absolute or relative deficiency of insulin results in hyperglycemia in diabetic individuals³. Delayed insulin secretion immediately after meal leads to sudden surge in blood glucose level known as 'hyperglycemic spikes'⁴. This 2-hour post-prandial plasma glucose will range from 140 to 199 mg/dL (impaired glucose tolerance) and then rise to greater than 200 mg/dL in the case of diabetics. Post-prandial phase is associated with macrovascular and microvascular diabetic complications and it is the major independent risk factor for cardiovascular disease (CVD)⁵. In the evolution of diabetic interventions, there is an effective preventive strategy to treat this risky post-prandial phase: Unabsorbed carbohydrates (oligosaccharides and disaccharides) are bound to the alpha-glucosidase enzymes located in the brush border of the enterocytes of the jejunum in small intestine and on cleavage to monosaccharides, they are immediately absorbed in the upper jejunum causing hyperglycemia⁶. On administering alpha-glucosidase inhibitors with carbohydrates, they compete with the binding of oligosaccharides and prevent their cleavage to monosaccharides, thereby slowing the digestion process and rise in post-prandial blood glucose level⁷. Acarbose, voglibose and miglitol are commercial alpha-glucosidase inhibitors that are considered as first-line treatment for diabetic individuals with post-prandial hyperglycemia. On giving them along with other oral hypoglycaemic agents like metformin, sulfonylurea improves glycaemic control (reduced HbA1c). However, these alpha-glucosidase inhibitors have prominent gastrointestinal side effects like flatulence, diarrhoea, and abdominal discomfort⁸. This warrants the search for alternative natural herbal medicines that have fewer side effects than the available inhibitors for the treatment of diabetes. The current study aims to evaluate fifteen Indian Ayurvedic medicinal plants for alpha-glucosidase inhibitory activity *in vitro*. Plants used are *Aconitum heterophyllum* Wall, *Acorus calamus* Linn, *Clerodendron serratum* Linn, *Cyperus rotundus* Linn, *Marsdenia tenacissima* (Roxb.) Moon, *Mesua ferrea* Linn, *Nigella sativa* Linn, *Picrorhiza kurroa* Royle ex benth, *Piper*

retrofractum Vahl, *Plumbago zeylanica* Linn, *Rubia cordifolia* Linn, *Saussurea lappa* C.B.C.L, *Symplocos racemosa* Roxb, *Terminalia arjuna* (Roxb.) Wight & Arn, *Zingiber officinale* Rosc. From this study we report that *Cyperus rotundus*, *Plumbago zeylanica*, *Symplocos racemosa* and *Terminalia arjuna* have the most potent alpha-glucosidase inhibiting components since they show maximum inhibition even at nanogram and microgram quantities. Kinetic analysis done on these plants showed that *Plumbago zeylanica* exhibited a mixed (non-competitive- uncompetitive) type of inhibition, whereas the other three plants inhibited the enzyme in an uncompetitive manner. This study shows that these plants contain potential alpha-glucosidase inhibitors which can be exploited further for the isolation of active components used in the treatment of diabetes.

MATERIALS AND METHODS

Materials

The plant materials were purchased from Amman Ayurvedic shop, Vellore, India and ground to yield a powdered form for solvent extraction. *p*-Nitrophenyl- α -D-glucopyranoside (PNPG), Yeast alpha-glucosidase (EC 3.2.1.20), sodium phosphate salts and sodium carbonate were purchased from Sisco (SRL), India. The 96-well microplate reader was purchased from Bio Tek USA Inc. and Voglibose tablets (2 mg) were purchased from Bayer Pharma.

Plant background

Grounded plant parts (250 g) were extracted using methanol (250 ml) in Soxhlet apparatus for 8 hours. Then, the extract was evaporated to dryness and the final dry crude extract was stored in dark at -20°C until used for the experiments. The standard drug, Voglibose was dissolved in distilled water, centrifuged at 6000 rpm and the supernatant was taken and appropriately diluted.

Aconitum heterophyllum Wall, Ranunculaceae, Ativishaa (Sanskrit)

The plant is commonly found in alpine and sub-alpine region of the Himalayas at altitudes between 1,800-4,500 m. The roots of the plant have medicinal properties. Various butyrylcholinesterase and acetylcholinesterase inhibitors were extracted from the roots of the plant⁹. Nortriterpenoid alkaloids were extracted from the roots of these plants having antibacterial activity¹⁰. Ethanolic extracts of this plant stimulates phagocytic function while inhibiting the humoral component of the body's immune system, thus acting as an immunomodulator¹¹.

Acorus calamus Linn, Araceae, Vacha (Sanskrit)

Pectic polysaccharide obtained from this plant activates macrophages and stimulate Th1 response¹². Ethanolic extract of the leaves promoted wound-healing activity in rats¹³. An inhibitory effect of the aqueous extract on water-bloom forming species of algae has been indicated¹⁴. Ethyl acetate extracts showed insulin releasing and alpha-glucosidase inhibitory activity¹⁵ and they published IC₅₀ value of 0.41 µg/ml.

Clerodendron serratum Linn, Verbenaceae, Bharangi (Sanskrit)

Ethanolic extract of the roots of *Clerodendron* has hepatoprotective activity against carbon tetrachloride induced toxicity in rats¹⁵. Alcoholic extract of the roots of this plant has anti-inflammatory activity¹⁶. It was also shown to have antinociceptive and antipyretic activity in animal model.

Cyperus rotundus Linn, Cyperaceae, Musta (Sanskrit)

The rhizome of this plant has been reported to play a major role in the protection of neurodegenerative disorders due to its antioxidant and free radical scavenging activity¹⁷. Ethanolic extract of this plant has analgesic properties¹⁸. Hydro-alcoholic extract of this plant displaying antiviral effect against Herpes Simplex¹⁹. Hydro-ethanolic extract of *Cyperus* reduces blood glucose level significantly in alloxan induced diabetic rats²⁰. Ethanol extract of aerial parts of this plant showed marked protection against convulsions induced by chemo convulsive agents in mice²¹.

Marsdenia tenacissima (Roxb.) Moon, Asclepiadaceae, Madhuras (Sanskrit)

A significant antitumor effect of this herb has been reported in experimental and clinical applications²². Novel pregnane glycosides were isolated from roots of this plant²³. Tenacigenin derived from this plant had shown to reverse multidrug resistance in cancerous cells²⁴.

Mesua ferrea Linn, Clusiaceae, Nagakesara (Sanskrit)

Estrogenic and progestational activity of this plant on mice and humans has been reported²⁵. Recently it has been shown that Calophyllolide isolated from *Mesua* is effective in reducing the increased capillary permeability (induced in mice by Histamine, 5-HT and bradykinin). Main use of stamen has been described to control bleeding in menorrhagia and piles. Xanthonenes, a number of 4-phenylcoumarin derivatives, friedelin and triterpenes have been isolated from the plant. Xanthonenes are isolated from the heartwood; coumarin derivatives from the seeds; canophyllal, canophyllol and canophyllic acid from the leaves.

Nigella sativa Linn, Ranunculaceae, Kalongi (Sanskrit)

Volatile oil extracted by hydro-distillation contains thymoquinone (3.8 %) which is involved in anti-inflammatory activities *in vivo* and *in vitro*²⁶ and the therapeutic potential of thymoquinone in pancreatic cancer has been proved²⁷. It also showed its potential adjuvant effects to improve immunotherapy in the treatment of allergic Rhinitis²⁸. Fixed oil and water extract of this plant (0.1% v/v) has shown to considerably reduce formation of sickle cells due to its calcium antagonistic and antioxidant activities²⁹. Oral administration of the ethanolic extract of *N. sativa* seeds to diabetic rats reduced hyperglycemia³⁰. Methanolic extract of *Nigella* significantly inhibits glucose utilisation in the intestine of rats³¹. Petroleum ether extract of the seeds of this plant has shown to exert insulin sensitising action. *N. sativa* treatment in streptozotocin-induced diabetic rats reduced lipid peroxidation and serum nitric oxide levels to a significant level by increasing antioxidant enzyme activity³². Oil extracted from the plant blocks nitric oxide overproduction and ceases morphine-induced tolerance and dependence in mice³³. Methanolic extracts of this plant possess potent CNS depressant and analgesic activity³⁴.

Picrorhiza kurroa Royle ex benth, Scrofulariaceae, Katuka (Sanskrit)

A glycoside, picroliv isolated from this plant is hepatoprotective. It has cholorectic and anti-cholestatic effects in rats and guinea pigs³⁵.

It also has anti-viral and immuno-modulatory effect. Picroliv restored cadmium induced abnormalities in the liver of male rats³⁶. It has anti-inflammatory, anti-carcinogenic³⁷ and antioxidant effects³⁸. Methanolic extract of the plant has healing potential in indomethacin induced stomach ulcers in mice³⁹. Tannins derived from the plant inhibit cyclooxygenase and lipid peroxidation⁴⁰.

Piper retrofractum Vahl, Piperaceae, Chavya (Sanskrit)

Aqueous extracts of the plant have potent larvicidal activity⁴¹. Ethanolic extract of the plant has larvicidal and insecticidal properties⁴². Fruits of this plant have potent anti-bacterial and anti-fungal properties⁴³. Phenolic amides obtained from the plant have potent antioxidant properties⁴⁴.

Plumbago zeylanica Linn, Plumbaginaceae, Chitraka (Sanskrit)

Plumbagin isolated from this plant possess significant anticancer activity⁴⁵ and anti-inflammatory effects⁴⁶. Hexane and chloroform extract of this plant had significant larvicidal activity⁴⁷. Acetone and ethanolic extracts of the leaves of this plant had reversible concentration dependent oestrogenic and anti-oestrogenic activity⁴⁸. Methanolic extract of the root of this plant showed promising antioxidant activity⁴⁹.

Rubia cordifolia Linn, Rubiaceae, Manjistha (Sanskrit)

Anti-oxidative constituents were isolated from ethyl acetate extract of this plant⁵⁰. Alcoholic extract of this plant increased brain gamma-amino-n-butyric acid levels and decreased brain dopamine and plasma corticosterone levels⁵¹. The extract also inhibited acidity and ulcer formation, decreased blood sugar level in alloxan treated animals⁵¹.

Saussurea lappa C.B.C.L, Asteraceae, Kustha (Sanskrit)

Anti-hepatotoxic activity of the aqueous-methanol root extract has been reported in mice⁵². Ethanol extract of this plant inhibited *Streptococcus mutans* in a dose dependent manner⁵³. The plant contains cholinergic and a calcium antagonist ingredient which is helpful for use in constipation and spasms⁵⁴. Antifungal constituents were isolated from the roots of the plant⁵⁵.

Symplocos racemosa Roxb. Symplocaceae, Lodhra (Sanskrit)

Glycosides isolated from this plant are reported to inhibit thymidine phosphorylase whose overexpression is linked to angiogenesis⁵⁶. New phenolic glycosides of salirepin series in the n-butanol fraction of the bark of *S. racemosa* has been isolated⁵⁷. Ethyl substituted glycoside that inhibits lipoxygenase⁵⁸ and phenolic glycoside that inhibits human nucleotide pyrophosphatase phosphodiesterase has been identified⁵⁹.

Terminalia arjuna Roxb. Wight & Arn. Combretaceae, Arjuna (Sanskrit)

This plant is widely known to prove comprehensive relief to the people suffering from cardio-vascular diseases, especially hyperlipidemia and ischemic heart disease. Some important findings related to the above mentioned activity has been studied⁶⁰. Anti-inflammatory, immunomodulatory and antinociceptive activity of the bark in mice and rats has been studied⁶¹. *T. arjuna* bark extract attenuated catecholamine-induced myocardial fibrosis and oxidative stress⁶². It has antimicrobial activity against multi drug resistant (MDR) strains of fungi and bacteria of clinical origin⁶³. Oleanane-type triterpene glycosides have been isolated and they suppress the release of nitric oxide and superoxide from macrophages and also inhibit aggregation of platelets⁶⁴. The effect of *T. arjuna* extract on adriamycin-induced micronuclei formation in cultured human peripheral blood lymphocytes has been studied⁶⁵. *T. arjuna* has been found to be useful in diabetes associated with ischemic heart disease⁶⁶.

Zingiber officinale Rosc. Zingiberaceae, Sunthi (Sanskrit)

Fractions obtained from the rhizome of this plant show antimicrobial and antioxidant effects⁶⁷. A formulation of the plant was effective in controlling alcohol hangover symptoms⁶⁸. Dried fermented ginger improved intestinal function⁶⁹. Ethanolic extract of

this plant had remarkable inhibitory activity against multi-drug resistant bacterial and fungal strains⁷⁰. The plant has larvicidal constituents⁷¹ and anti-invasive property of the constituents of the plant in hepatocarcinoma cells has been studied⁷². Ginger promoted glucose transport in muscle cell line⁷³. The protective action of ginger oil on gastric ulcer induced by aspirin in rats has been investigated⁷⁴. Ethanolic extract of ginger has protective effect against renal damage induced by alcohol⁷⁵. Aqueous extract of ginger rhizome produced significant increase in insulin levels and decrease in fasting glucose levels in diabetic rats⁷⁶. Treatment with the extract also caused reduction in serum cholesterol levels, blood pressure and serum triglycerides in diabetic rats.

Inhibition assay for alpha-glucosidase activity

Alpha-glucosidase inhibition was analysed using kinetic end-point assay described by Pistia-Brueggeman with few modifications. Alpha-glucosidase inhibitory activity was performed following the modified method of Pistia Brueggeman and Hollingsworth⁷⁷. In a 96-well plate reader, a reaction mixture containing 50 µl of phosphate buffer (50 mM; pH 6.8), 10 µl of alpha-glucosidase (1 U/ml) and 20 µl of plant extract of varying concentrations was pre-incubated for 5 min at 37°C, and then 20 µl of 1 mM PNPG was added to the mixture as a substrate. After further incubation at 37°C for 30 min, the reaction was stopped by adding 50 µl of sodium carbonate (0.1M). All the enzyme, inhibitor and substrate solutions were made using the same buffer. Voglibose was used as a positive control and water as negative control. The yellow colour produced (due to *p*-nitrophenol formation) was quantitated by colorimetric analysis and reading the absorbance at 405 nm. Each experiment was performed in triplicates, along with appropriate blanks.

The % inhibition has been obtained using the formula:

$$\% \text{ inhibition} = \frac{\{\text{Absorbance (control)} - \text{Absorbance (sample)}\}}{\text{Absorbance (control)}}$$

IC₅₀ value is defined as the concentration of extract inhibiting 50% of alpha-glucosidase activity under the stated assay conditions. In case of significant inhibition, IC₅₀ values were determined by linear regression by fitting to a sigmoidal dose-response equation with variable slope. All values are represented as Mean ± Standard Deviation.

Kinetics of alpha-glucosidase inhibition

The mode of inhibition of plant extracts against alpha-glucosidase activity was measured with increasing concentrations of PNPG (0.125, 0.25, 0.5 and 1 mM) as substrate in the absence or presence of the plant extracts at two different concentrations for each plant extract. The enzyme reaction was performed by incubating the

mixture at 37°C for 30 min and optimal doses of the plant extracts were determined based on the results from inhibitory activity assay as described earlier. Mode of inhibition of plant extracts was determined by Lineweaver-Burk plot analysis of the data calculated following Michaelis-Menten kinetics^{78,79}. Experimental inhibitor constant (K_i) values were determined by secondary plots. The theoretical value of K_i

$$K_i = V_{max} \cdot I / (V_m - V_{max})$$

RESULTS AND DISCUSSION

Extracts obtained from several Ayurvedic plants have been reported for their anti-diabetic activity. But these plants have not been evaluated for the mechanism of action to control blood glucose level. In the present study, fifteen Indian Ayurvedic medicinal plants were evaluated for their alpha-glucosidase inhibitory potential. Plants used in this study along with the parts used and inhibition values are listed in Table 1. Before our study, except *Acorus*, none of the other plants have been studied for their alpha-glucosidase inhibitory effect *in vitro*. On screening 15 plant extracts, 4 extracts exhibited 100 % inhibition, while 10 extracts showed 10-100% AGI activity (10-100%). Only one extract (*Acorus calamus*) showed less than 10% inhibition. Voglibose, an available drug for alpha-glucosidase inhibition showed IC₅₀ value of 21.98±2.91 µg/ml.

Extracts with 100% inhibitory effect on the alpha-glucosidase inhibition at 100 µg/ml

The extracts of *Terminalia arjuna*, *Plumbago zeylanica*, *Cyperus rotundus*, *Symplocos racemosa* had displayed 100% inhibition at the concentration of 100 µg/ml (Table 1 and Fig. 1). The bark extract of *Terminalia arjuna* was the most potential inhibitor of the enzyme based on the IC₅₀ value (0.69 µg/ml). On comparison to the control drug Voglibose with the IC₅₀ value of 21.98±2.91 µg/ml, all these four plants displayed maximum inhibition at lower concentrations.

Extracts with 40-100% inhibitory effect on the alpha-glucosidase inhibition

The extracts of *Mesua ferrea* and *Aconitum heterophyllum* had shown moderate potency of 45.9% and 40.3% at 100 µg/ml, respectively.

Extracts with 10-40% inhibitory effect on the alpha-glucosidase inhibition

The extracts of the following plants showed inhibition against alpha-glucosidase at 100 µg/ml. *Nigella sativa* (28.7%), *Rubia cordifolia* (29.7%), *Clerodendron serratum* (32.3%), *Marsdenia tenacissima* (22.1%), *Picrorhiza kurroa* (26.2%), *Saussurea lappa* (19.9%), *Zingiber officinale* (16.9%), and *Piper retrofractum* (15%).

Table 1: Inhibitory effect of plant extracts on alpha-glucosidase (Positive control: Voglibose (21.98±2.91 µg/ml))

| Botanical name | Family | Parts used | Percentage inhibition at 100 µg/ml | IC ₅₀ (µg/ml) |
|-------------------------------|-----------------|------------|------------------------------------|--------------------------|
| <i>Aconitum heterophyllum</i> | Ranunculaceae | Rhizome | 40.3 | 166.5±13.63 |
| <i>Acorus calamus</i> | Araceae | Rhizome | 7.5 | 401.06±15.96 |
| <i>Clerodendron serratum</i> | Verbenaceae | Root | 32.3 | 264.66±8.92 |
| <i>Cyperus rotundus</i> ** | Cyperaceae | Tubers | 100 | 3.98±0.55 |
| <i>Marsdenia tenacissima</i> | Asclepiadaceae | Stem | 22.1 | 298.43±4.79 |
| <i>Mesua ferrea</i> | Clusiaceae | Dried buds | 45.87 | 128.75±7.9 |
| <i>Nigella sativa</i> | Ranunculaceae | Seeds | 28.7 | 172.81±12.6 |
| <i>Picrorhiza kurroa</i> | Scrofulariaceae | Rhizome | 26.2 | 313.53±10.84 |
| <i>Piper retrofractum</i> | Piperaceae | Stem | 15.0 | 670.24±32.97 |
| <i>Plumbago zeylanica</i> ** | Plumbaginaceae | Root | 100 | 3.46±0.53 |
| <i>Rubia cordifolia</i> | Rubiaceae | Root | 29.7 | 253.42±4.88 |
| <i>Saussurea lappa</i> | Asteraceae | Rhizome | 19.9 | 401.76±5.17 |
| <i>Symplocos racemosa</i> ** | Symplocaceae | Bark | 100 | 8.16±0.28 |
| <i>Terminalia arjuna</i> ** | Combretaceae | Bark | 100 | 0.69±0.08 |
| <i>Zingiber officinale</i> | Zingiberaceae | Rhizome | 16.9 | 650.33±7.1 |

** Potential plants showing 100% inhibition.

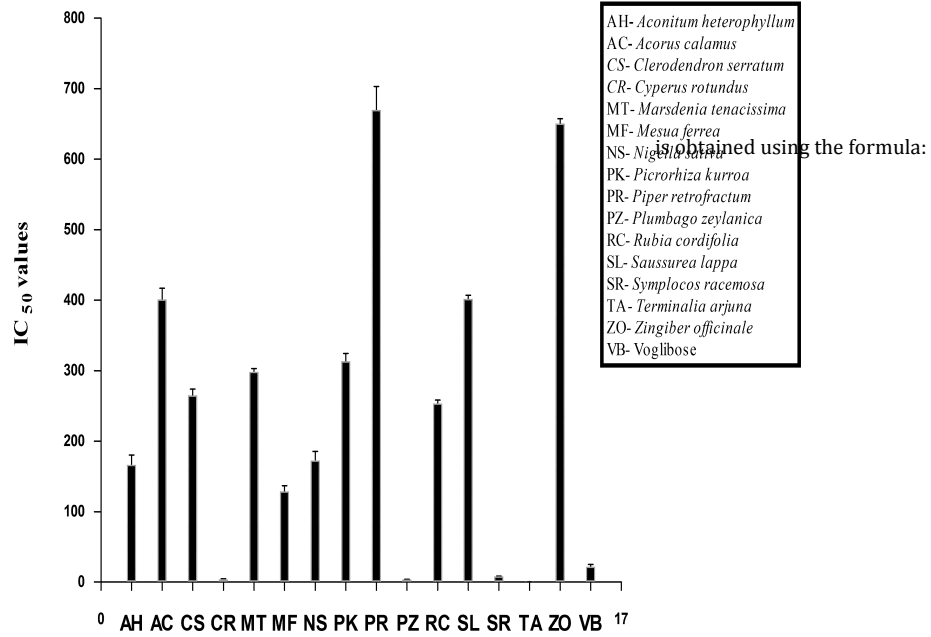


Fig. 1: Graphical representation of the IC₅₀ values for the plant extracts and standard (Voglibose)

Following these results, kinetic study has been performed on the four extracts (*C. rotundus*, *P. zeylanica*, *S. racemosa* and *T. arjuna*) showing 100% inhibition against alpha-glucosidase activity. Lineweaver-Burk (LB) plot has been used to determine the mode of inhibition (Fig. 2A-2D) and experimental inhibitor constant (K_i) values were determined by secondary plots (Fig. 3A-3D). The tuber extract of *C. rotundus* contains uncompetitive inhibitor(s) of alpha-glucosidase (Fig. 2A and 3A) and there is a reduction of V_{max} from 1.54mM.min⁻¹ to 0.67mM.min⁻¹ and K_m from 2.68 mM to 1.16 mM. The root extract of *P.zeylanica* has mixed inhibitor(s) of alpha-

glucosidase (Fig. 2B and 3B) and there is a reduction in V_{max} from 1.82 mM.min⁻¹ to 1.28 mM.min⁻¹ and K_m from 3.37 mM to 2.49 mM. The bark extract of *S. racemosa* (Fig. 2C and 3C) contains uncompetitive inhibitor(s) of alpha-glucosidase and there is a reduction of V_{max} from 0.41 mM.min⁻¹ to 0.23 mM.min⁻¹ and the K_m from 0.73 mM to 0.40 mM. The bark extract of *T. arjuna* contains uncompetitive inhibitor(s) of alpha-glucosidase (Fig. 2D and 3D) and the V_{max} reduced from 1.19 mM.min⁻¹ to 0.48 mM.min⁻¹ and the K_m from 2.14 mM to 0.87 mM. The extracts of *S. racemosa* and *T. arjuna* requires low substrate concentration to reach half the maximal velocity.

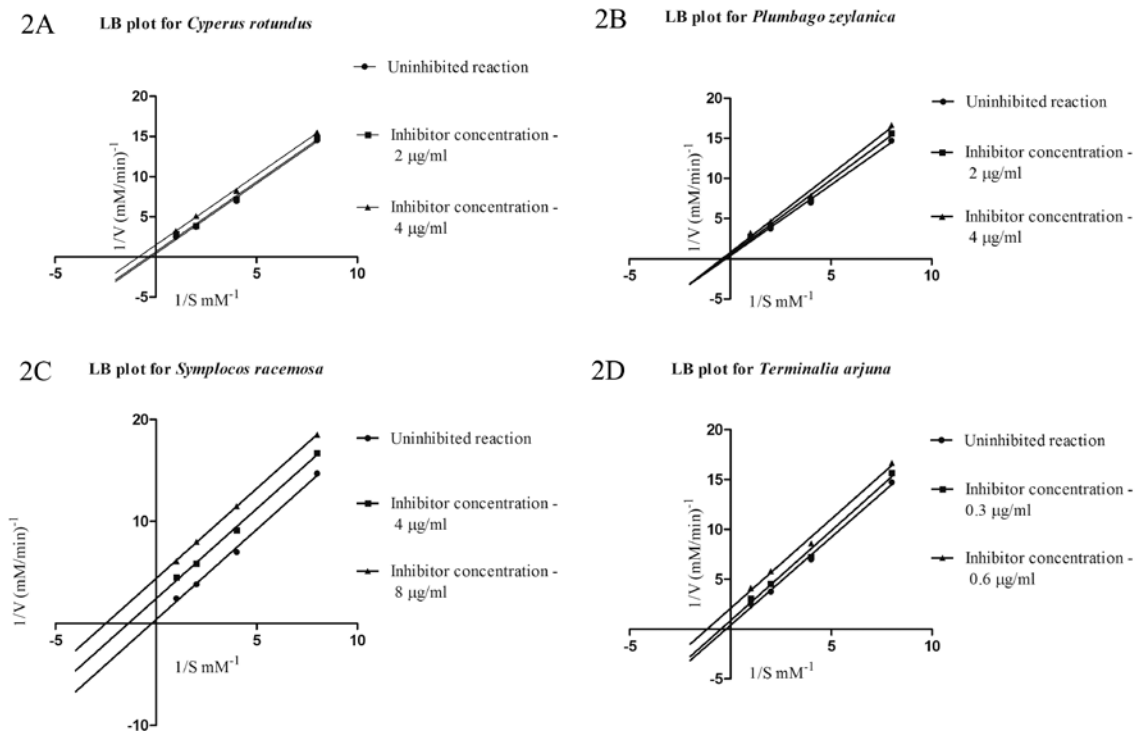


Fig. 2A-2D: Lineweaver-Burk plot for kinetic analysis of alpha-glucosidase inhibition by plant extracts

Yeast alpha-glucosidase was treated with various concentration of PNP-G (0.125-1 mM) in the absence or presence of the extract (*Cyperus*, *Plumbago*, *Symplocos*) at the concentration range 2 µg/ml

and 8 µg/ml. In the case of *Terminalia*, the extract at a concentration of 0.3 µg/ml and 0.6 µg/ml has been used. The enzyme reaction was performed by incubating the mixture at 37°C for 30 min.

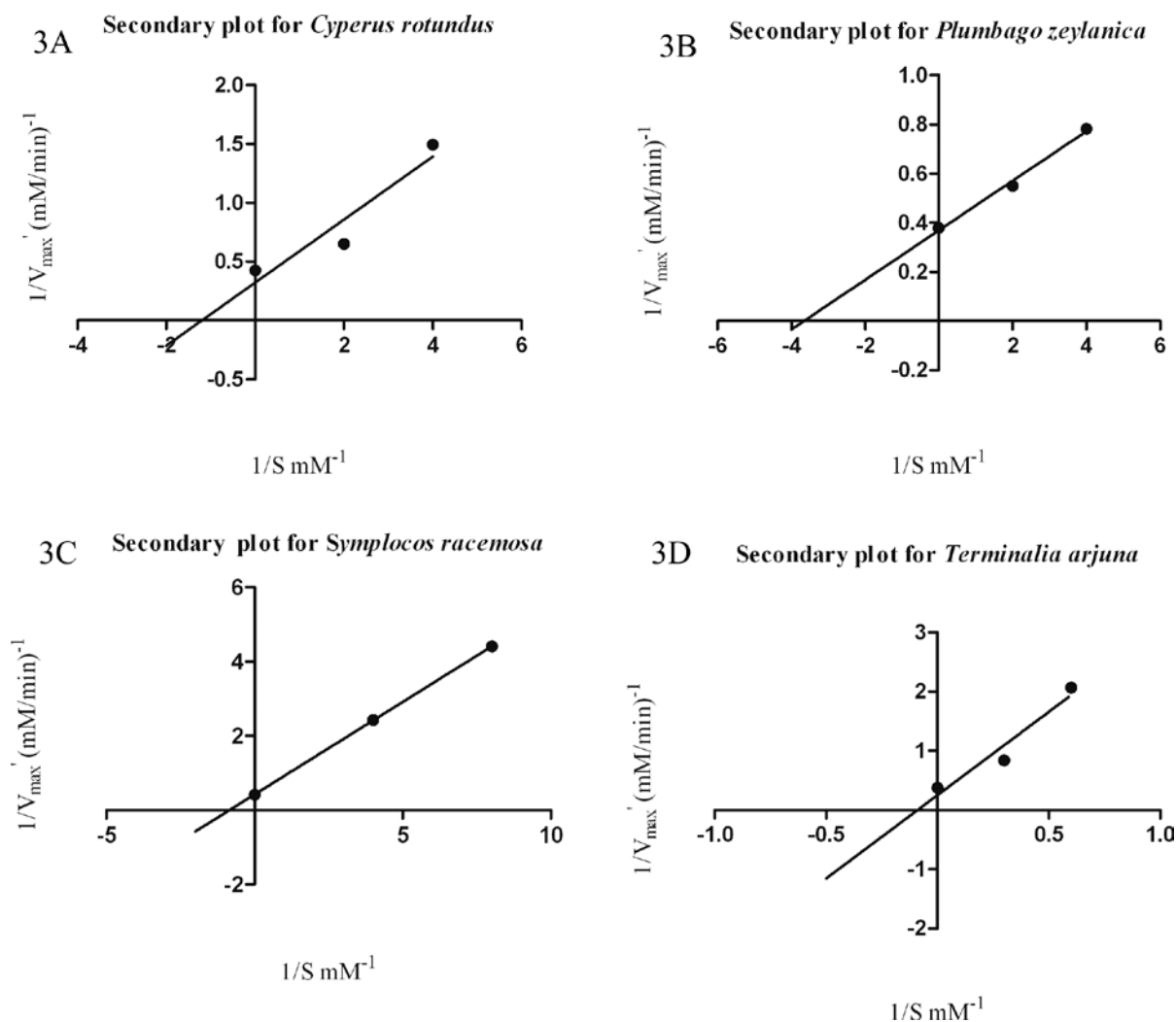


Fig. 3A-3D: Secondary plots for the kinetic analysis of alpha-glucosidase by plant extracts

Table 2: Kinetic analysis of alpha-glucosidase inhibition by the plant extracts

| Extract concentration (µg/ml) | <i>C. rotundus</i> | | <i>P. zeylanica</i> | | <i>S. racemosa</i> | | <i>T. arjuna</i> | |
|----------------------------------|--------------------|-----------------------------------|---------------------|-----------------------------------|--------------------|-----------------------------------|------------------|-----------------------------------|
| | K_m (mM) | V_{max} (mM.min ⁻¹) | K_m (mM) | V_{max} (mM.min ⁻¹) | K_m (mM) | V_{max} (mM.min ⁻¹) | K_m (mM) | V_{max} (mM.min ⁻¹) |
| 0 | 4.099 | 2.353 | 4.654 | 2.639 | 4.709 | 2.666 | 4.654 | 2.639 |
| 2 | 2.676 | 1.536 | 3.373 | 1.821 | 0.726 | 0.412 | 2.145 | 1.189 |
| 4 | 1.165 | 0.669 | 2.488 | 1.279 | 0.400 | 0.227 | 0.866 | 0.484 |

Inhibitor concentration (µg/ml) was plotted on the X-axis and $1/V$ (mM/min)⁻¹ values obtained from the Lineweaver-Burk plot was plotted on the Y axis. The point on the X-axis where the line intersects the axis is the experimental inhibitor constant (K_i).

Table 2 shows the kinetics of alpha-glucosidase inhibition by the plant extracts. In all the four plants, on increasing the substrate concentration of the extract (from 2 µg/ml to 4 µg/ml), there is a decrease in the values for maximum velocity (V_{max}) and the substrate concentration required to reach half the maximum velocity (K_m).

In *C. rotundus*, K_m value of PNP-glycoside for alpha-glucosidase was 2.49 mM and the experimental K_i value was 1.21 µg/ml. The extract of *P. zeylanica* had shown the K_m value of 1.17mM and the experimental K_i value was 3.68 µg/ml. In *S. racemosa*, K_m value of PNP-glycoside for alpha-glucosidase was 0.40 mM and the experimental K_i value was 0.77 µg/ml. This K_i value is close to that of the calculated K_i value (0.74 µg/ml).

The extract of *T. arjuna* had shown the K_m value of 0.87 mM and the experimental K_i value of 0.09 µg/ml. The commercial alpha-glucosidase inhibitor, Voglibose is a competitive inhibitor of the enzyme and it is required at higher concentration to reduce the post-prandial glucose level. The uncompetitive inhibitors bind to

the enzyme-substrate complex, lowering the K_m and the maximum enzyme activity (V_{max}).

CONCLUSION

Diabetes mellitus is a progressive metabolic disorder affecting majority of the population across the world. There are various measures to manage and treat this aggravating disease. The main effect of diabetes is increase in glycemic level and therefore to reach normoglycemic level, along with insulin and other oral hypoglycemic agents like sulfonylureas, biguanides⁸⁰, Thiazolidinediones (TZD), alpha-glucosidase inhibitors (AGI) and incretin mimetics (GLP-1, GIP, DPP-4 inhibitors)⁸¹ are being used. Alpha-glucosidase inhibitors delay the action of alpha-glucosidases to break complex carbohydrates in to simple sugars, thereby lowering the absorption of glucose. These inhibitors play a vital role in reducing the post-prandial hyperglycemia. As a consequence of their pharmacological action, alpha-glucosidase inhibitors also cause a concomitant decrease in post-prandial plasma insulin and gastric inhibitory polypeptide (GIP) and a rise in late post-prandial plasma glucagon-like peptide 1 (GLP-1) levels⁸². In individuals with normal or impaired glucose tolerance with hyperinsulinemia, alpha-glucosidase inhibitors decrease hyperinsulinemia and improve insulin sensitivity⁸³. Post-prandial hyperglycemia contributes to raise in glycated haemoglobin (HbA1c) levels, which as an indicator of total glycemic load, is tightly correlated with the incidence of micro and macroangiopathy in Type 2 diabetes. It can induce or deteriorate fasting hyperglycemia and be associated with coagulation activation and/or lipid metabolism abnormalities⁸⁴. Epidemiological data from the United Kingdom Prospective Diabetes Study (UKPDS) also showed that there is a 14–16% decrease in macrovascular complications for every 1% absolute reduction in glycated haemoglobin⁸⁵. Alpha-glucosidase inhibitors like acarbose, miglitol and voglibose are used in conjunction with other anti-diabetic drugs. But these inhibitors have some side effects like flatulence and diarrhea. This indicates that newer AGI's with lesser side-effects needs to be discovered.

Therefore, we have screened potential anti-diabetic plants for alpha-glucosidase inhibition. From the study we have identified *Cyperus rotundus*, *Plumbago zeylanica*, *Symplocos racemosa* and *Terminalia arjuna* for potential alpha-glucosidase inhibitory activity compared to the standard (voglibose). Particularly, *Terminalia arjuna* is effective even in nanogram quantities. Therefore, our plan is to investigate its efficiency *in vivo* using animal models. Kinetic studies performed on these effective plants exhibit uncompetitive inhibition for *Cyperus rotundus*, *Symplocos racemosa* and *Terminalia arjuna*. This indicates that the substrate binding could cause a conformational change to take place in the enzyme such that it reveals an inhibitor binding site. *Plumbago zeylanica* exhibited non-competitive-uncompetitive inhibition. This indicates that the inhibitor constant for free enzyme is lesser than the inhibitor constant for enzyme-substrate complex. Also, the binding of the inhibitor to the free enzyme or the enzyme-substrate complex is mutually independent.

Competing interests

The authors have no conflicting interests.

Authors' contribution

AB carried out the *in vitro* enzyme assay, kinetic analysis, prepared figures, and drafted the manuscript. MSS has contributed in standardizing the assay and figure preparation. KT conceived of the study, designed, involved in preparing the manuscript and interpretation. All authors read and approved the final manuscript.

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REFERENCES

- Shaw JE, Sicree RA, Zimmet PZ. Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Res Clin Pract.* 2010; 87: 4-14.

- Nyenwe EA, Jerkins TW, Umpierrez GE, Kitabchi AE. Management of type 2 diabetes: evolving strategies for the treatment of patients with type 2 diabetes. *Metabolism.* 2011; 60: 1-23.
- Gerich JE. The genetic basis of type 2 diabetes mellitus: impaired insulin secretion versus impaired insulin sensitivity. *Endocr Rev.* 1998; 19: 491-503.
- Ceriello A. Postprandial hyperglycemia and diabetes complications: is it time to treat? *Diabetes.* 2005; 54: 1-7.
- Bonora E, Muggeo M. Postprandial blood glucose as a risk factor for cardiovascular disease in Type II diabetes: the epidemiological evidence. *Diabetologia.* 2001; 44: 2107-14.
- Puls W, Krause HP, Müller L, Schutt H, Sitt R, Thomas G. Inhibitors of the rate of carbohydrate and lipid absorption by the intestine. *Int J Obes.* 1984; 8 Suppl 1: 181-90.
- Fonseca V. Clinical significance of targeting post-prandial and fasting hyperglycemia in managing type 2 diabetes mellitus. *Curr Med Res Opin.* 2003; 19: 635-41.
- Hollander P. Safety profile of acarbose, an alpha-glucosidase inhibitor. *Drugs.* 1992; 44 Suppl 3: 47-53.
- Nisar M, Ahmad M, Wadood N, Lodhi MA, Shaheen F, Choudhary MI. New diterpenoid alkaloids from *Aconitum heterophyllum* Wall: Selective butyrylcholinesterase inhibitors. *J Enzyme Inhib Med Chem.* 2009; 24: 47-51.
- Ahmad M, Ahmad W, Ahmad M, Zeeshan M, Obaidullah, Shaheen F. Norditerpenoid alkaloids from the roots of *Aconitum heterophyllum* Wall with antibacterial activity. *J Enzyme Inhib Med Chem* 2008; 23: 1018-1022.
- Atal CK, Sharma ML, Kaul A, Khajuria A. Immunomodulating agents of plant origin. I: Preliminary screening. *J Ethnopharmacol* 1986; 18: 133-141.
- Belska NV, Guriev AM, Danilets MG, Trophimova ES, Uchasova EG, Ligatcheva AA et al. Water-soluble polysaccharide obtained from *Acorus calamus* L. classically activates macrophages and stimulates Th1 response. *Int Immunopharmacol.* 2010; 10: 933-42.
- Jain N, Jain R, Jain A, Jain DK, Chandel HS. Evaluation of wound-healing activity of *Acorus calamus* Linn. *Nat Prod Res* 2010; 24: 534-41.
- Hu GJ, Zhang WH, Shang YZ, He L. Inhibitory effects of dry *Acorus calamus* extracts on the growth of two water bloom-forming algal species. *Ying Yong Sheng Tai Xue Bao* 2009; 20: 2277-82.
- Vidya SM, Krishna V, Manjunatha BK, Mankani KL, Ahmed M, Singh SD. Evaluation of hepatoprotective activity of *Clerodendrum serratum* L. *Indian J Exp Biol* 2007; 45: 538-542.
- Narayanan N, Thirugnanasambantham P, Viswanathan S, Vijayasekaran V, Sukumar E. Antinociceptive, anti-inflammatory and antipyretic effects of ethanol extract of *Clerodendron serratum* roots in experimental animals. *J Ethnopharmacol* 1999; 65: 237-241.
- Lee CH, Hwang DS, Kim HG, Oh H, Park H, Cho JH et al. Protective effect of *Cyperus rotundus* rhizoma against 6-hydroxydopamine-induced neuronal damage. *J Med Food* 2010; 13: 564-571.
- Pal D, Dutta S, Sarkar A. Evaluation of CNS activities of ethanol extract of roots and rhizomes of *Cyperus rotundus* in mice. *Acta Pol Pharm* 2009; 66: 535-541.
- Soltan MM, Zaki AK. Antiviral screening of forty-two Egyptian medicinal plants. *J Ethnopharmacol* 2009; 126: 102-107.
- Raut NA, Gaikwad NJ. Antidiabetic activity of hydro-ethanolic extract of *Cyperus rotundus* in alloxan induced diabetes in rats. *Fitoterapia* 2006; 77: 585-588.
- Dilip KL. Determination of brain biogenic amines in *Cynodon dactylon* Pers. and *Cyperus rotundus* L. treated mice. *International Journal of Pharmacy and Pharmaceutical Sciences* 2009; 1: 190-197.
- Qian J, Hua H, Qin S. Progress on antitumor effects of *Marsdenia tenacissima*. *Zhongguo Zhong Yao Za Zhi* 2009; 34: 11-13.
- Xia ZH, Xing WX, Mao SL, Lao AN, Uzawa J, Yoshida S et al. Pregnane glycosides from the stems of *Marsdenia tenacissima*. *J Asian Nat Prod Res* 2004; 6: 79-85.
- Hu YJ, Shen XL, Lu HL, Zhang YH, Huang XA, Fu LC et al. Tenacigenin B derivatives reverse P-glycoprotein-mediated

- multidrug resistance in HepG2/Dox cells. *J Nat Prod* 2008; 71: 1049-1051.
25. Meherji PK, Shetye TA, Munshi SR, Vaidya RA, Antarkar DS, Koppikar S et al. Screening of *Mesua ferrea* (Nagkesar) for estrogenic & progestational activity in human & experimental models. *Indian J Exp Biol* 1978; 16: 932-933.
 26. Geng D, Zhang S, Lan J. Analysis on chemical components of volatile oil and determination of thymoquinone from seed of *Nigella glandulifera*. *Zhongguo Zhong Yao Za Zhi* 2009; 34: 2887-2890.
 27. Banerjee S, Azmi AS, Padhye S, Singh MW, Baruah JB, Philip PA et al. Structure-activity studies on therapeutic potential of Thymoquinone analogs in pancreatic cancer. *Pharm Res* 2010; 27: 1146-1158.
 28. İşik H, Cevikbaş A, Gürer US, Kiran B, Uresin Y, Rayaman P et al. Potential Adjuvant Effects of *Nigella sativa* Seeds to Improve Specific Immunotherapy in Allergic Rhinitis Patients. *Med Princ Pract* 2010; 19: 206-211.
 29. Ibraheem NK, Ahmed JH, Hassan MK. The effect of fixed oil and water extracts of *Nigella sativa* on sickle cells: an in vitro study. *Singapore Med J* 2010; 51: 230-234.
 30. Kaleem M, Kirmani D, Asif M, Ahmed Q, Bano B. Biochemical effects of *Nigella sativa* L seeds in diabetic rats. *Indian J Exp Biol* 2006; 44: 745-748.
 31. Arayne MS, Sultana N, Mirza AZ, Zuberi MH, Siddiqui FA. In vitro hypoglycemic activity of methanolic extract of some indigenous plants. *Pak J Pharm Sci* 2007; 20: 268-273.
 32. Kanter M, Coskun O, Korkmaz A, Oter S. Effects of *Nigella sativa* on oxidative stress and beta-cell damage in streptozotocin-induced diabetic rats. *Anat Rec A Discov Mol Cell Evol Biol* 2004; 279: 685-691.
 33. Abdel-Zaher AO, Abdel-Rahman MS, Elwasei FM. Blockade of Nitric Oxide Overproduction and Oxidative Stress by *Nigella sativa* Oil Attenuates Morphine-Induced Tolerance and Dependence in Mice. *Neurochem Res*. 2010 Oct; 35: 1557-65.
 34. Al-Naggar TB, Gómez-Serranillos MP, Carretero ME, Villar AM. Neuropharmacological activity of *Nigella sativa* L. extracts. *Journal of Ethnopharmacology* 2003; 88: 63-68.
 35. Verma PC, Basu V, Gupta V, Saxena G, Rahman LU. Pharmacology and chemistry of a potent hepatoprotective compound Picroliv isolated from the roots and rhizomes of *Picrorhiza kurroa* royle ex benth. (kutki). *Curr Pharm Biotechnol* 2009; 10: 641-649.
 36. Yadav N, Khandelwal S. Therapeutic efficacy of Picroliv in chronic cadmium toxicity. *Food Chem Toxicol* 2009; 47: 871-879.
 37. Anand P, Kunnumakkara AB, Harikumar KB, Ahn KS, Badmaev V, Aggarwal BB. Modification of cysteine residue in p65 subunit of nuclear factor-kappaB (NF-kappaB) by picroliv suppresses NF-kappaB-regulated gene products and potentiates apoptosis. *Cancer Res* 2008; 68: 8861-8870.
 38. Dhuley JN: Effect of picroliv administration on hepatic microsomal mixed function oxidases and glutathione-conjugating enzyme system in cholestatic rats. *Hindustan Antibiot Bull* 2005; 47-48: 13-19.
 39. Banerjee D, Maity B, Nag SK, Bandyopadhyay SK, Chattopadhyay S. Healing potential of *Picrorhiza kurroa* (Scrophulariaceae) rhizomes against indomethacin-induced gastric ulceration: a mechanistic exploration. *BMC Complement Altern Med* 2008; 8: 3.
 40. Zhang Y, DeWitt DL, Murugesan S, Nair MG. Novel lipid-peroxidation- and cyclooxygenase-inhibitory tannins from *Picrorhiza kurroa* seeds. *Chem Biodivers* 2004; 1: 426-441.
 41. Chansang U, Zahiri NS, Bansiddhi J, Boonruad T, Thongsrirak P, Mingmuang J et al. Mosquito larvicidal activity of aqueous extracts of long pepper (*Piper retrofractum* vahl) from Thailand. *J vector Ecol* 2005; 30: 195-200.
 42. Komalamisra N, Trongtokit Y, Palakul K, Prummongkol S, Samung Y, Apiwathnasorn C et al. Insecticide susceptibility of mosquitoes invading tsunami-affected areas of Thailand. *Southeast Asian J Trop Med Public Health* 2006; 37 Suppl 3: 118-122.
 43. Limyati DA, Juniar BL. Jamu Gendong, a kind of traditional medicine in Indonesia: the microbial contamination of its raw materials and endproduct. *J Ethnopharmacol* 1998; 63: 201-208.
 44. Nakatani N, Inatani R, Ohta H, Nishioka A. Chemical constituents of peppers (*Piper* spp.) and application to food preservation: naturally occurring antioxidative compounds. *Environ Health Perspect* 1986; 67: 135-142.
 45. Chen CA, Chang HH, Kao CY, Tsai TH, Chen YJ. Plumbagin, isolated from *Plumbago zeylanica*, induces cell death through apoptosis in human pancreatic cancer cells. *Pancreatology* 2009; 9: 797-809.
 46. Checker R, Sharma D, Sandur SK, Khanam S, Poduval TB. Anti-inflammatory effects of plumbagin are mediated by inhibition of NF-kappaB activation in lymphocytes. *Int Immunopharmacol* 2009; 9: 949-958.
 47. Maniafu BM, Wilber L, Ndiege IO, Wanjala CC, Akenga TA. Larvicidal activity of extracts from three *Plumbago* spp against *Anopheles gambiae*. *Mem Inst Oswaldo Cruz* 2009; 104: 813-817.
 48. Edwin S, Joshi SB, Jain DC. Antifertility activity of leaves of *Plumbago zeylanica* Linn. in female albino rats. *Eur J Contracept Reprod Health Care* 2009; 14: 233-239.
 49. Maryam Z, Farrukh A, Iqbal A. The in vitro antioxidant activity and total phenolic content of four Indian medicinal plants. *International Journal of Pharmacy and Pharmaceutical Sciences* 2009; 1: 88-95.
 50. Lu Y, Hu R, Dai Z, Pan Y. Preparative separation of antioxidative constituents from *Rubia cordifolia* by column-switching counter-current chromatography. *J Sep Sci*. 2010; 33: 2200-5.
 51. Patil RA, Jagdale SC, Kasture SB. Antihyperglycemic, antistress and nootropic activity of roots of *Rubia cordifolia* Linn. *Indian J Exp Biol* 2006; 44: 987-992.
 52. Yaesh S, Jamal Q, Shah AJ, Gilani AH. Antihepatotoxic activity of *Saussurea lappa* extract on D-galactosamine and lipopolysaccharide-induced hepatitis in mice. *Phytother Res* 2009; 24 Suppl 2: 229-232.
 53. Yu HH, Lee JS, Lee KH, Kim KY, You YO. *Saussurea lappa* inhibits the growth, acid production, adhesion, and water-insoluble glucan synthesis of *Streptococcus mutans*. *J Ethnopharmacol* 2007; 111: 413-417.
 54. Gilani AH, Shah AJ, Yaesh S. Presence of cholinergic and calcium antagonist constituents in *Saussurea lappa* explains its use in constipation and spasm. *Phytother Res* 2007; 21: 541-544.
 55. Rao KS, Babu GV, Ramnareddy YV. Acylated flavone glycosides from the roots of *Saussurea lappa* and their antifungal activity. *Molecules* 2007; 12: 328-344.
 56. Miszczak-Zaborska E, Smolarek M, Bartkowiak J. Influence of the thymidine phosphorylase (platelet-derived endothelial cell growth factor) on tumor angiogenesis. Catalytic activity of enzyme inhibitors. *Postepy Biochem* 2010; 56: 61-66.
 57. Ahmad VU, Rashid MA, Abbasi MA, Rasool N, Zubair M. New salirepin derivatives from *Symplocos racemosa*. *J Asian Nat Prod Res* 2007; 9: 209-215.
 58. Abbasi MA, Ahmad VU, Zubair M, Nawaz SA, Lodhi MA, Farooq U et al. Lipoxygenase inhibiting ethyl substituted glycoside from *Symplocos racemosa*. *Nat Prod Res* 2005; 19: 509-515.
 59. Choudhary MI, Fatima N, Abbasi MA, Jalil S, Ahmad VU, Atta-ur-Rahman. Phenolic glycosides, a new class of human recombinant nucleotide pyrophosphatase phosphodiesterase-1 inhibitors. *Bioorg Med Chem* 2004; 12: 5793-5798.
 60. Mahmood ZA, Sualeh M, Mahmood SB, Karim MA. Herbal treatment for cardiovascular disease the evidence based therapy. *Pak J Pharm Sci* 2010; 23: 119-124.
 61. Halder S, Bharal N, Mediratta PK, Kaur I, Sharma KK. Anti-inflammatory, immunomodulatory and antinociceptive activity of *Terminalia arjuna* Roxb bark powder in mice and rats. *Indian J Exp Biol* 2009; 47: 577-583.
 62. Kumar S, Enjamoori R, Jaiswal A, Ray R, Seth S, Maulik SK. Catecholamine-induced myocardial fibrosis and oxidative stress is attenuated by *Terminalia arjuna* (Roxb.). *J Pharm Pharmacol* 2009; 61: 1529-1536.
 63. Khan R, Islam B, Akram M, Shakil S, Ahmad A, Ali SM et al. Antimicrobial activity of five herbal extracts against multi drug

- resistant (MDR) strains of bacteria and fungus of clinical origin. *Molecules* 2009; 14: 586-597.
64. Alam MS, Kaur G, Ali A, Hamid H, Ali M, Athar M. Two new bioactive oleanane triterpene glycosides from *Terminalia arjuna*. *Nat Prod Res* 2008; 22: 1279-1288.
 65. Reddy TK, Seshadri P, Reddy KK, Jagetia GC, Reddy CD. Effect of *Terminalia arjuna* extract on adriamycin-induced DNA damage. *Phytother Res* 2008; 22: 1188-1194.
 66. Dwivedi S, Aggarwal A. Indigenous drugs in ischemic heart disease in patients with diabetes. *J Altern Complement Med*. 2009; 15: 1215-1221.
 67. Wang HM, Chen CY, Chen HA, Huang WC, Lin WR, Chen TC et al. *Zingiber officinale* (ginger) compounds have tetracycline-resistance modifying effects against clinical extensively drug-resistant *Acinetobacter baumannii*. *Phytother Res*. 2010; 24: 1825-30.
 68. Takahashi M, Li W, Koike K, Sadamoto K. Clinical effectiveness of KSS formula, a traditional folk remedy for alcohol hangover symptoms. *J Nat Med*. 2010; 64:487-91.
 69. Incharoen T, Yamauchi K, Thongwittaya N. Intestinal villus histological alterations in broilers fed dietary dried fermented ginger. *J Anim Physiol Anim Nutr (Berl)*. 2010; 94: 130-7.
 70. Khan R, Zakir M, Afaq SH, Latif A, Khan AU. Activity of solvent extracts of *Prosopis spicigera*, *Zingiber officinale* and *Trachyspermum ammi* against multidrug resistant bacterial and fungal strains. *J Infect Dev Ctries* 2010; 4: 292-300.
 71. Lin RJ, Chen CY, Lee JD, Lu CM, Chung LY, Yen CM. Larvicidal Constituents of *Zingiber officinale* (Ginger) against *Anisakis simplex*. *Planta Med*. 2010 Nov; 76: 1852-8.
 72. Weng CJ, Wu CF, Huang HW, Ho CT, Yen GC. Anti-invasion effects of 6-shogaol and 6-gingerol, two active components in ginger, on human hepatocarcinoma cells. *Mol Nutr Food Res*. 2010; 54: 1618-27.
 73. Noipha K, Ratanachaiyavong S, Ninla-Aesong P. Enhancement of glucose transport by selected plant foods in muscle cell line L6. *Diabetes Res Clin Pract*. 2010; 89: 22-6.
 74. Khushtar M, Kumar V, Javed K, Bhandari U. Protective Effect of Ginger oil on Aspirin and Pylorus Ligation-Induced Gastric Ulcer model in Rats. *Indian J Pharm Sci* 2009; 71: 554-558.
 75. Shanmugam KR, Ramakrishna CH, Mallikarjuna K, Reddy KS. Protective effect of ginger against alcohol-induced renal damage and antioxidant enzymes in male albino rats. *Indian J Exp Biol* 2010; 48: 143-149.
 76. Akhani SP, Vishwakarma SL, Goyal RK. Anti-diabetic activity of *Zingiber officinale* in streptozotocin-induced type I diabetic rats. *J Pharm Pharmacol* 2004; 56: 101-105.
 77. Pistia-Brueggeman G, Hollingsworth RI. The use of the o-nitrophenyl group as a protecting/activating group for 2-acetamido-2-deoxyglucose. *Carbohydrate Research*. 2003; 338: 455-458.
 78. Shim YJ, Doo HK, Ahn SY, Kim YS, Seong JK, Park IS et al. Inhibitory effect of aqueous extract from the gall of *Rhus chinensis* on alpha-glucosidase activity and postprandial blood glucose. *J Ethnopharmacol* 2003; 85: 283-287.
 79. Kim YM, Jeong YK, Wang MH, Lee WY, Rhee HI. Inhibitory effect of pine extract on alpha-glucosidase activity and postprandial hyperglycemia. *Nutrition* 2005; 21: 756-761.
 80. Schwab S, Diem P. Oral hypoglycaemic agents in 2009. *Ther Umsch*. 2009; 66: 677-84.
 81. Garber AJ. Incretin-based therapies in the management of type 2 diabetes: rationale and reality in a managed care setting. *Am J Manag Care*. 2010; 16 suppl 7: 187-94.
 82. Hiki M, Shimada K, Kiyonagi T, Fukao K, Hirose K, Ohsaka H et al. Single administration of alpha-glucosidase inhibitors on endothelial function and incretin secretion in diabetic patients with coronary artery disease - Juntendo University trial: effects of miglitol on endothelial vascular reactivity in type 2 diabetic patients with coronary heart disease (J-MACH) . *Circ J*. 2010; 74: 1471-8.
 83. Lebovitz HE. Adjunct therapy for type 1 diabetes mellitus. *Nat Rev Endocrinol*. 2010; 6: 326-34.
 84. Verges B. The impact of prandial glucose regulation in practice. *Diabetes Nutr Metab*. 2002 ; 15 Suppl 6: 28-32.
 85. Stratton IM, Alder AI, Neil HA, Matthews DR, Manley SE, Cull CA et al. Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. *BMJ* 2000; 321: 405-412.