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**Research Article** 

# ANTIFERTILITY POTENTIAL OF SOME MEDICINAL PLANTS IN MALES: AN OVERVIEW

# SURESH C. JOSHI<sup>A,\*</sup>, AKSHA SHARMA<sup>A</sup> AND MRIDULA CHATURVEDI<sup>B</sup>

<sup>a</sup>Reproductive Toxicology Unit, Center for advanced studies, Department of Zoology, University of Rajasthan, Jaipur 302055 India, <sup>b</sup>Vedic Women's College, Jaipur, India. Email: s\_c\_joshi2003@rediffmail.com

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### ABSTRACT

The development of an effective, reversible and safe male contraceptive has been the focus of research around the world for more than 30 years. This review concentrates on those recent advances in science and technology that offer possible inroads for shifting the paradigm for male-based contraception. A large number of scientists are searching for a relatively cheap, widely available, easily accepted and effective contraceptive of plant origin that is equally non-invasive, non-hormonal in action, non-toxic and relatively long acting. Medicinal plants are important elements of indigenous medical system in India as well as in other countries. In these days, the use of traditional medicines has received considerable interest and a large number of plants have been screened for their antifertility activity. Thus, the present review includes a brief account of research reports on plants with antifertility potential.

Keywords: Contraceptive, Male-based, Non-hormonal, Antifertility Potential, Effective

## INTRODUCTION

Fertility control is an issue of global and national public health concern. There is a global need to support individuals in familyplanning due to the increasing growth rate of the world's population with its negative impact on environment, economic growth and poverty reduction in underdeveloped countries<sup>1</sup>. About 90% of the world's contraceptive users are women. Though considerable progress has been made in the development of highly effective, acceptable and reversible methods of contraception in females, progress and possibilities on males are still slow and limited<sup>2,3</sup>. Aware of this responsibility, health organizations and pharmaceutical companies continue to financially support or actively pursue research towards new contraceptive approaches<sup>4</sup>. Current methods of contraception result in an unacceptable rate of unintended pregnancies and many side effects also<sup>5,6</sup>. A large number of chemical agents have been known but all tend to lead to total spermatogenic arrest and, ultimately, to irreversible sterility7. As concerns regarding side effects of existing male contraceptive methods prevent universal acceptance<sup>8, 9</sup>, the development of additional male methods of fertility control can provide tremendous social and public health benefits. There are relatively few realistic approaches currently being pursued which include (a) the suppression of sperm production, (b) disruption of sperm maturation and/or function, and (c) interruption of sperm transport<sup>10,1</sup>. and inhibitors Contraceptive vaccines, of spermatogenesis and sperm motility, provide a potential for nonhormonal male contraceptives<sup>11</sup>. It has, therefore, become necessary to use biologically active botanical substances or fertilityregulating agents of plant origin which are ecofriendly in approach and interfere with the natural patterns of reproduction<sup>12</sup>. Male antifertility drugs can induce contraception by interfering with spermatogenesis progression. Their action mechanism is correlated with the apoptosis of spermatogenic cells<sup>13</sup>.

In our country as well as in the world, there are several medicinal plants associated with antifertility properties<sup>14-16</sup>. Although very few contraceptives have been developed from plant extracts, their potentiality has not been determined accurately, and their mode of action has been beyond our knowledge until now because there are many problems in assessing plant extract including batch to batch variation and a lack of definite active portion of the extract used for the development of herbal contraceptives<sup>17</sup>.

Several plant products inhibit male and female fertility and may be developed into contraceptives. Even though, many indigenous plants have been shown to prevent the birth, only few plants have so far been investigated for antifertility activity<sup>18</sup>. The World Health Organization (WHO) has set up a Task Force on Plant Research for fertility regulation with an objective to find new orally active non-

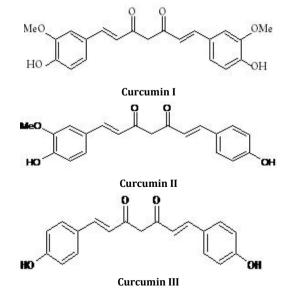
steroidal contraceptive compounds<sup>19</sup>. Various medicinal plant extracts have been tested for their antifertility activity both in male and female. Some of these plants had spermicidal effects; other caused reduction in the sperm counts and altered the mobility of the sperms. Some of them caused testicular changes and altered hormone levels<sup>20</sup>. It is necessary to use biologically active botanical substances or fertility-regulating agents of plant origin which are ecofriendly. The natural plant substances possessing mild inherent estrogenic and antiestrogenic properties offer themselves as an effective nonconventional source of contraception with less deleterious side effects<sup>21</sup>.

Plants showing antifertility potential in males are listed in table 1 and some of them are discussed below.

#### Curcurma longa

*Curcuma longa* Linn., commonly known as Turmeric, Indian saffron or Haldi belongs to family Zingiberaceae, is a perennial herb cultivated throughout India and is widely used as an antibiotic in folk medicines and as spices. Its tubers, rhizomes and oil have great importance. *C. longa* also possesses antimutagenic and anticarcinogenic properties<sup>22</sup>.

Phenolic diketone, curcumin (diferuloylmethane) (3–4%) is responsible for the yellow colour, and comprises curcumin I (94%), curcumin II (6%) and curcumin III (0.3%)  $^{23}$ .



Curcumin found to inhibit 5a-reductase, which converts testosterone to 5a-dihydrotestosterone, thereby inhibiting the growth of flank organs in hamster. Curcumin also inhibited human sperm motility and has the potential for the development of a novel intravaginal contraceptive<sup>24,25</sup>.

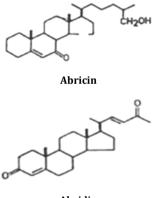
Rats fed with *Curcuma longa* aqueous and 70 % alcoholic extract for 60 days (500 mg.kg<sup>-1</sup>.day<sup>-1</sup>) showed a reduction in sperm motility and density. *C. longa* may have affected the androgen synthesis either by inhibiting the Leydig cell function or the hypothalamus pituitary axis and as a result, spermatogenesis is arrested<sup>26</sup>.

Male mice of the Parkes (P) strain were orally administered aqueous rhizome extract of *C. longa* (600 mg/kg body weight per day for 56 and 84 days) showed adverse effect of on various male reproductive organs and fertility. The treatment had adverse effects on motility, viability, morphology and number of spermatozoa in the cauda epididymidis, serum level of testosterone and on fertility. By 56 days of treatment withdrawal, however, the above parameters recovered to control levels. The results show that *C. longa* treatment causes reversible suppression of spermatogenesis and fertility, thereby suggesting the potential of this plant in the regulation of male fertility<sup>27</sup>.

### Abrus precatorius

The plant *Abrus precatorius* Linn, popularly known as Rosary pea belong to the family leguminosae (Fabaceae), is found throughout India in hedges and bushes in exposed areas<sup>28</sup>. Usually seeds are used against leucoderma, wounds, alopecia, asthma, tubercular glands, leprosy, fever, ulcer and tumor<sup>29</sup>.

Precatorine, trigonelline, choline and abrine are present in the seeds. Abricin and abridin, two steroids were also reported in the seeds; the latter exhibited anti-fertility property<sup>30</sup>.



#### Abridin

The contraceptive and toxicologic effects were observed with administration of methanolic extract (70%) of the seeds of *A. precatorius* (L.) (Fabaceae) (20 and 40 mg/kg b.wt./day) for 45 days. Treatment caused a significant decrease in caudal sperm motility, count and viability. There was a complete suppression of fertility at 40 mg/kg dose level. The decrease in weights of testis and cauda epididymis of mice at 40mg/kg level could be attributed to a loss of spermatogenic elements in testis and absence of sperms in cauda epididymis<sup>31</sup>.

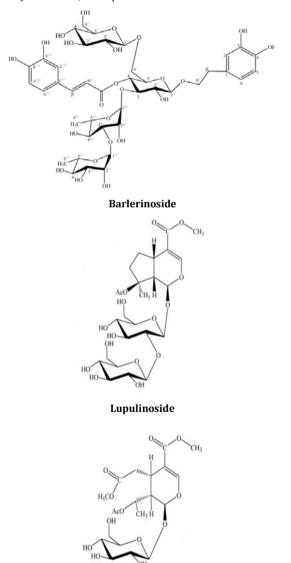
The inhibitory effects of a methanolic extract of *A. precatorius* seeds (5 and 20 mg/ml) on the motility of washed human spermatozoa was noticed. The extract caused a concentration-related impairment of percentage sperm motility. With the highest concentration tested (20 mg/ml), the onset of the antimotility action was almost immediate. In addition, this concentration impaired the functional integrity of the plasma membrane (hypoosmotic swelling test) and viability (nigrosin-eosin stain) of spermatozoa. In contrast, with a lower concentration (5.0 mg/ml), such effects were not evident. It is concluded that at the lower concentrations the antimotility action may result from a rise in intracellular calcium (not via influx) and/or a decline in cAMP content and/or enhanced generation of a reactive oxygen species<sup>32</sup>.

The ethanolic extract of *A. precatorius* seeds intraperitoneally administered with 20, 40 and 60 mg/kg doses for 20 days showed disrupted arrangement of seminiferous tubules, loosening of germinal epithelium and low counts of leydig cells, germ cells and sperm cells. Histomorphology of the epididymus showed a decrease in tubule size, epithelial height and a reduction in sperm number in the tubular lumen. Plasma testosterone levels decreased significantly with a higher dose (60 mg/kg) compared to controls. This suggests that *A. precatorius* seed extract with higher dose (60 mg/kg) tends to suppress spermatogenesis and is hence liable to cause infertility in male mice<sup>33</sup>.

## Barleria prionitis

*Barleria prionitis* L. (Family Acanthaceae) is commonly known as Vajradanti. In indigenous system of medicine in India, the aerial parts (stem, leave & flower) are used in fever, toothache, inflammation, as diuretic and gastrointestinal disorders; bark in whooping cough as an expectorant; the whole plant and especially the roots are used as tonic<sup>34</sup>. Leaves, stem and root of *B. prionitis* possess antibacterial and anti-inflammatory activities.

From the aerial parts of *B. prionitis*, one new phenylethanoid glycoside, barlerinoside along with six known iridoid glycosides, namely, shanzhiside methyl ester, 6-0-trans-p-coumaroyl-8-0-acetylshanzhiside methyl ester, barlerin, acetylbarlerin, 7-methoxydiderroside, and lupulinoside were isolated<sup>35</sup>.



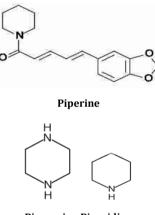
7-methoxydiderroside

Male rats treated with isolated fractions of the *B. prionitis* root methanolic extract (100 mg/kg for 60 days) showed a significant reduction on spermatogenesis without affecting general body metabolism. Sperm motility as well density in cauda epididymides was reduced significantly. The population of various spermatogenic cells such as primary spermatocytes, secondary spermatocytes and round spermatids were declined significantly in treated animals<sup>36</sup>.

Oral administration of root extract of *B. prionitis* L. to male rats (100 mg/rat per day) for the period of 60 days did not cause body weight loss. The root extract brought about an interference with spermatogenesis. The round spermatids were decreased by 73.6% (P< or =0.001). The extract reduced the fertility of male rats by 100%. Cross sectional surface area of Sertoli cells and mature Leydig cell numbers were significantly reduced (36.9%). Testicular glycogen contents were low. Antifertility effects of Barleria seemed to be mediated by disturbances in testicular somatic cells functions (Leydig and Sertoli cells) resulting in the physio-morphological events of spermatogenesis <sup>37</sup>.

## Piper nigrum

*Piper nigrum* L. commonly known as black pepper belongs to family Piperaceae. The fruits of *P. nigrum* are not only important as a spice or flavoring agent, but have also been prescribed for cholera, dyspepsia, diarrhea, various gastric ailments, and paralytic and arthritic disorders<sup>38</sup>. It mainly contains amide alkaloids, and piperine is the major active component<sup>39,40</sup>.



**Piperazine Piperidine** 

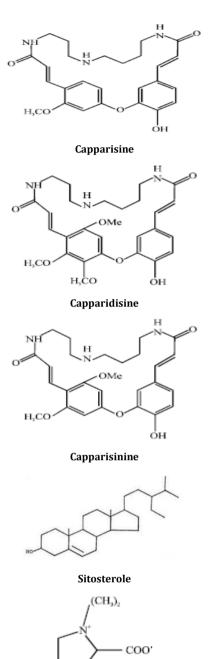
Oral administration of fruit powder of *P. nigrum* (25 and 100 mg/kg body weight/day for 20 and 90 days) to male mice of the Parkes (P) strain adversely affects sperm parameters and also caused marked alterations in male reproductive organs<sup>41</sup>.

Piperine (1-piperoylpiperidine) is an alkaloid present in the fruits of black pepper (*Piper nigrum*), long pepper (*Piper longum*) and other piper species. Piperine is the major pungent substance present in these plants and is commonly used as a spice all over the world for seasoning and flavoring food. The weights of the caput, corpus and cauda regions of the epididymis significantly decreased at dose of 100 mg/kg. Epididymal sperm count and motility decreased at 10 mg/kg and 100 mg/kg, and sperm viability decreased significantly at 100 mg/kg. Piperine could damage the epididymal environment and sperm function<sup>42</sup>.

## Capparis aphylla

*Capparis aphylla* (syn: *C. decidua*), family Capparidaceae, is commonly known as desert broom (Eng.); Swartstrom, Babejaanarm (Afr.); Sengam, Kuzhalaathondai (Tamil)<sup>43</sup>. The plants were used in several medicines such as anthelmenties, muscular injury, swelling, jaundice, appetizer, cardiac diseases, pyorrhea, cholera, dysentery, rheumatism, constipation, stomach disorder and skin diseases<sup>44</sup>.

It contains capparin, capparilin, capparinin, caparidisine, capparisine, capparisinine, sitosterole, i-stachydrin, n-pentocosane and n-triacontanol<sup>45,46</sup>.



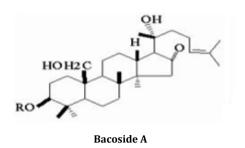
#### Stachydrine

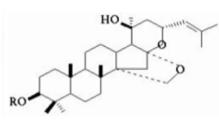
Ethanol extract of C. aphylla was evaluated for possible spermatotoxic effect in 90 days old male rat. The ethanol extract of C. aphylla at the doses of 50, 100 and 200 mg/kg of body weight when administered intra peritonially for 55 days revealed spermatotoxic effect in 90 days old male rat. The fertility of the treated rats was reduced drastically. The sperm concentration in the epididymis and sperm motility decreased, whereas sperm abnormalities increased in particular sperm abnormalities like flexed head, detached head and coiling of end tail. Thus C. aphylla treatment resulted in impairment of male fertility in the rat by affecting both spermatogenesis and cauda epididymal spermatozoa47.

#### Bacopa monnieri

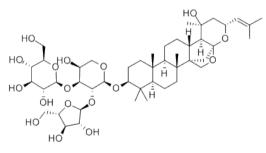
*Bacopa monnieri* L. (Family Scrophulariaceae) commonly known as Brahmi has been used in the Ayurvedic system of medicine for centuries<sup>48</sup>.

Main chemical components are saponins, bacosides, bacopasides, monnierin, brahmine, nicotine, herpestine and hersaponin<sup>49,50</sup>.

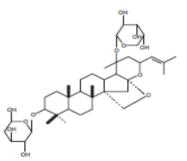




Bacoside A1



Bacopaside



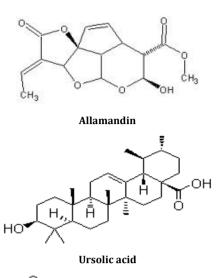
#### Bacosaponin

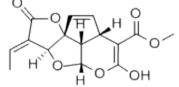
Oral administration of Brahmi (250 mg/kg body weight/day, for 28 and 56 days) to male mice of the Parkes (P) strain caused reduction in motility, viability, morphology, and number of spermatozoa in cauda epididymidis. Histologically, testes in mice treated with the plant extract showed alterations in the seminiferous tubules. These results thus suggest that Brahmi treatment causes suppression of spermatogenesis and fertility, without producing apparent toxic effects<sup>51</sup>.

# Allamanda cathartica

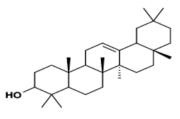
*Allamanda cathartica* Linn. (Apocyanaceae) is widely growing perennial shrub. The leaves are smooth and thick<sup>52</sup>. The roots are used against jaundice, complications with malaria and enlarged spleen in traditional medicine. The flowers act as a laxative. Moreover, yellow Allamanda has also antibiotic action against Staphylococcus<sup>53</sup>.

All parts of the plant contain allamandin, a toxic iridoid lactone. Leaves and stems yield ursolic acid,  $\beta$ -amyrin and  $\beta$ -sitosterol. Plumericin and isoplumericin are extracted from stem and rootbark, also from leaves and roots, besides plumieride and long chain esters<sup>54</sup>.





Plumericin



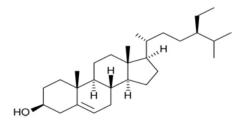


The oral administration of aqueous leaf extract of *A. cathartica* (150 mg/kg body weight/day for 14, 28 and 42 days) induces infertility and changes in various male reproductive endpoints in Parkes strain mice. Histologically, testes in extract-treated mice showed nonuniform degenerative changes in the seminiferous. The treatment also had adverse effects on motility, viability, morphology and on number of spermatozoa in the cauda epididymidis. Fertility of the extract-treated males was also suppressed<sup>55</sup>.

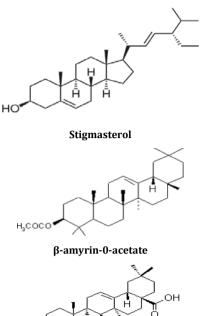
#### Dendrophthoe falcate

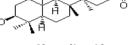
*Dendrophthoe falcate* (L.f.) Ettingsh. (known as mistletoe) is a perennial, climbing woody parasitic plant of the family Loranthaceae. In India, it is widely distributed and is commonly known as 'bandaa' and 'bandhulu'<sup>56</sup>. It is used ethnomedicinally for treating ulcers, asthma, impotence, paralysis, skin diseases, and wounds<sup>57</sup>.

Leaves contain flavonoids such as Quercetin, quercetrin;Tannins comprising of gallic and chebulinic acid. Young shoots contain nearly 10 percent tannins and the stem contains  $\beta$ -amyrin-0-acetate, oleonolic acid its methyl ester acetate,  $\beta$ -sitosterol and stigmasterol. Root contains Catechin and leucocynidin in the bark<sup>58</sup>.



 $\beta$  -Sitosterol





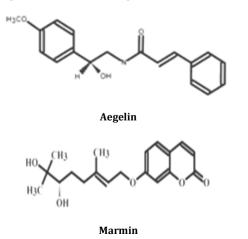
**Oleonolic** acid

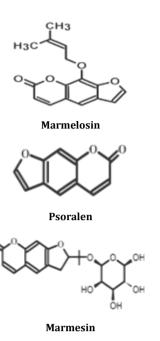
An oral administration of 70% methanolic extract of stem of *D. falcata* at a dose level of 100 mg/kg wt/day fed to male albino rats for 60 days did not decrease body weight, while the testes and epididymides weight were significantly reduced, and the seminal vesicles and ventral prostate also showed a significant reduction (P < 0.01). Treated animals showed a notable depression of spermatogenesis. The reduced sperm count and motility resulted in 100% negative fertility at 100 mg/kg dose level<sup>59</sup>.

# Aegle marmelos

*Aegle marmelos* (Linn), family Rutacae, commonly known as Bael, is a sacred tree for Hindu Religon<sup>60</sup>. Alcoholic extracts of the roots and fruits showed hypoglycemic and antidiabetic activity<sup>61, 62</sup>. With respect to clinical applications, it should be noted that the roots are astringent, bitter and febrifuge. They are useful in diarrhea, dysentery, dyspepsia and stomachalgia<sup>63</sup>.

Several chemical constituents have been isolated and from various parts of the bael tree. These include alkaloids, coumarins and steroids. The leaves contain skimianinc, sterol and aegelin. The active constituent of the fruit is marmolosin, which is identical to imperatorin. Odler coumarins contained in the fruits are altoimperatorin and B-sitosterol. Roots of the tree have been found to contain psoralin, xanthotoxin, scopoletin and tembamide<sup>64</sup>.



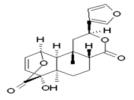


50 % ethanolic extract from the leaves of *A. marmelos* (AMLEt) (100, 200 and 300 mg(-1) kg (-1) day(-1) for each rat for 60 days) caused a reduction in weight of all the major accessory sex organs. There was a marked decline in motility and density of the sperm derived from cauda epididymis of the treated animals. *A. marmelos* reduced fertility of male rats by 100% at the 300-mg dose level. Serum testosterone levels also decreased significantly. Thus, the leaf extract of *A. marmelos* (AMLEt) suppresses fertility in male rats<sup>65</sup>. A dose related reduction in the testicular sperm count, epididymal sperm count and motility and abnormal sperm count was observed when the animals were administered the aqueous leaf extract (250mg/kg body wt.)<sup>66</sup>.

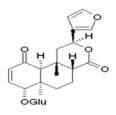
### Tinospora cordifolia

*Tinospora cordifolia* (Willd.) belongs to the Menispermaceae family and known as Gulancha in English, Guduchi in Sanskrit, and Giloya in Hindi<sup>67</sup>. It is reported to possess anti-spasmodic, antiinflammatory, anti-allergic, anti-diabetic, antioxidant properties<sup>68</sup>.

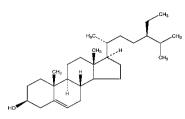
The chemical constituents reported from this shrub belong to different classes, such as alkaloids, diterpenoid lactones, glycosides, steroids, sesquiterpenoid, phenolics, aliphatic compounds and polysaccharides. Tinosporin,  $\beta$ -sitosterol, cordifol, columbin, chasmanthin, tinosporid, tinosporasid, cordifolid and palmarin are the main<sup>69, 70</sup>.







Tinosporid



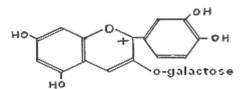
#### β-sitosterol

Oral administration of 70% methanolic extract of *T. cordifolia* stem to male rats at the dose level of 100 mg/rat/day for 60 days did not cause body weight loss but decreased the weight of testes, epididymis, seminal vesicle and ventral prostate in a significant manner. Sperm motility as well as sperm density were reduced significantly which resulted in reduction of male fertility by 100%. The stem extract brought about an interference with spermatogenesis. These results suggested antifertility effects of the stem extract of *T. cordifolia* in male rats<sup>71</sup>.

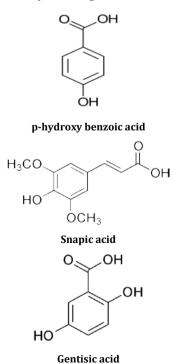
#### Martynia annua

*Martynia annua* L. (Family Martyniaccae), commonly known as scorpion (in Hindi, Bichchhu or Baghnukh), possess different medicinal properties. Fruit is used as anti-inflammatory. Leaves are antiseptic and are used in epilepsy. Roots used treatment of snake bite. Entire plant used to treat menstrual disorders. Dried entire plant has analgesic activity, anticonvulsant activity<sup>72</sup>.

Chemical examination of *M. annua* plant revealed the presence of alkaloid, glycosides, tannin, carbohydrates<sup>73</sup>, phenols, flavonoids, lcucanthocyanins<sup>74</sup>. Flowers contain cyanidin-3-galactoside<sup>75</sup> whilst p-hydroxy benzoic acid and snapic acid, and gentisic acid, respectively, are present in leaves and fruits, in addition to the p - hydroxy benzoic acid<sup>76</sup>.



Cyanidin-3-galactoside

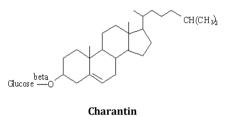


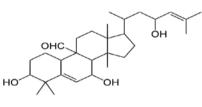
The 50% ethanol extract of *M. annua* L. root at dose level of 50 mg, 100 mg and 200 mg/kg body weight daily for a period of 60 days showed adverse effect on reproduction of male rats. Significant decrease in the weights of testes, epididymides, seminal vesicle and ventral prostate was noticed. There was a dose related reduction in the testicular sperm count, epididymal sperm count and motility. Significant reduction in serum concentration of luteinising hormone and testosterone was also observed. It is concluded that the 50% ethanol extract of *M. annua* root have dose related effects on male reproduction without altering general body metabolism<sup>77</sup>.

#### Momordica charantia

*Momordica charantia* Linn, belonging to the family of Cucurbitaceae, is an indigenous medicinal and vegetable plant found in the tropical and subtropical regions of the world and is commonly known as bitter gourd or bitter melon. *M. charantia* is one of the most promising plants for diabetes today<sup>78-80</sup>.

Bitter melon has some interesting biological and pharmacological activities, e.g. anticancer, antiviral, antibacterial, analgesic, antiinflammatory, hypotensive, anti-fertility, hepatotoxicity and antioxidant<sup>81-83</sup>. Fruit contains Momordicin, charantin, polypeptidep insulin and ascorbigen<sup>84</sup>.





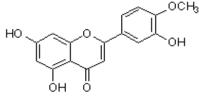
Momordicin

Petroleum ether, benzene and alcohol extracts of the seeds of *M. charantia* tested in rats at the dose level of 25 mg/100 g body weight for 35 days showed antispermatogenic activity as the number of spermatocytes, spermatids and spermatozoa was decreased. Increase in cholesterol level and Sudanophilic lipid accumulation indicates inhibition in the steroidogenesis. Out of the three extracts, the alcohol extract was more potent in its antispermatogenic, antisteroidogenic and androgenic activities<sup>85</sup>.

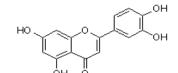
#### **Rosmarinus officinalis**

*Rosmarinus officinalis* L. (Labiatae) is an edible evergreen shrub native to the Mediterranean area. The leaves of the plant are commonly used as a spice and as a source of antioxidant compounds employed in food conservation<sup>86-89</sup>.

Phytochemical studies revealed the presence of several compounds in *R. officinalis* including phenolic diterpenes, diterpenoid quinines, flavonoids and essential oils<sup>90</sup>. Flavonoids have been shown to produce antiandrogenic activity and affect fertility in male dogs. Flavonoids include diosmetin, diosmin, genkwanin and derivatives, luteolin and derivatives, hispidulin, neptin, nepitrin and apigenin <sup>91</sup>.

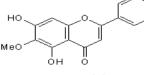


Diosmetin

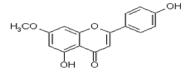




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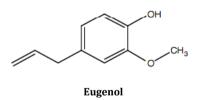
#### Genkwanin

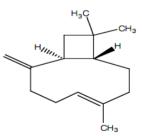
Ingestion of rosemary (*R. officinalis* L.) at levels of 250 and 500 mg/kg body wt for 63 days caused a significant decline in spermatogenesis in testes due to a decrease in the number of primary and secondary spermatocytes and spermatids is attributed to a significant decrease in testosterone. Sperm motility and density were also significantly decreased in the cauda epididymis and in the testes of rosemary-treated male rats<sup>92</sup>.

#### Syzygium aromaticum

*Syzygium aromaticum* L. commonly known as clove belongs to family Myrtaceae. It is used as a spice to **fld**d vor to exotic food preparations<sup>93</sup>.

Important constituents of clove oil include eugenol, betacaryophyllene and vanillin; crategolic acid; tannins, gallotannic acid, methyl salicylate (painkiller); the flavonoids eugenin, kaempferol, rhamnetin, and eugenitin; triterpenoids like oleanolic acid, stigmasterol and campesterol; and several sesquiterpenes<sup>94,95</sup>.





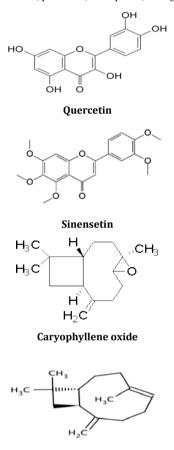
#### **β-caryophyllene**

The flower buds of *S. aromaticum* (clove), a common food flavor, have been used as indigenous medicine for the treatment of male sexual disorders in Asian countries. Oral exposure of hexane extract of flower buds of *S. aromaticum* in three doses (15mg, 30mg and 60mg/kg BW) for a single spermatogenic cycle (35 days) in Parkes (P) strain mice induced non-uniform degenerative changes in the seminiferous tubules associated with decrease in daily sperm production and depletion of round and elongated spermatids population<sup>96</sup>.

#### Chromolaena odoratum

*Chromolaena odorata* (Asteraceae) commonly known as Siam weed, is a fast-growing perennial and invasive weed. It has been reported to possess anti-inflammatory, antipyretic, antispasmodic activities<sup>97-99</sup>.

Terpenoids compounds were major components of *C. odorata* oil, such as trans-caryophyllene (16.58%), delta-cadinene (15.85%), alpha-copaene (11.58%), caryophyllene oxide (9.63%), germacrene-D (4.96%), and delta-humulene (4.32%). The leaves of this plant have be found to be a rich source of flavonoids which are quercetin, sinensetin, sakuranetin, padmatin, kaempferol, salvagenin<sup>100,101</sup>.



#### Trans-caryophyllene

Oral administration of aqueous extract of *C. odoratum* leaves (250 and 500 mg kg(-1) body weight) for 14 days in male albino rats revealed a significant reduction (P < 0.05) in testicular body weight ratio and histological examination revealed disruption in the arrangement of seminiferous tubules with no distinct basement membrane. These changes were accompanied by reduction in the number of spermatozoa. All these results indicated that aqueous extract of *C. odoratum* leaves possesses antiandrogenic property by interfering with steroidogenesis at the testicular level and this will adversely affect the functional capacity of the testes and the fertility of the animal<sup>102</sup>.

#### CONCLUSION

Plants have been a source of medicine in the past centuries and today scientists and the general public recognize their value as a source of new or complimentary medicinal products. Recently, wide array of research investigations highlight the potential health beneficial principles from phytal sources. Medicinal plants constitute one of the main sources of new pharmaceuticals and health care products.

There has been an increase in demand for the phytopharmaceuticals all over the world because of the fact that the allopathic drugs have more side effects. This review makes an

attempt to compile some of antifertility plants from Ayurveda as well as from foreign origin so as to give scientific account on usuage of anti- fertility plants. Various phytoconstituents like alkaloids, flavonoids, tannins, xanthones, triterpenes, quinones etc. were involved in anti- fertility activity. Although a number of plants have been reported to possess cent percent antifertility activity but till date these plants have not yet come up at the level of clinical trials. Standardization of methods, quality control, data on safety and efficacy need for proper understanding of the use of herbal medicines.

S. No	Name of plant	Vernacula r Name	Part used	Type of plant Extract	Dose	Duration	Animal model	Activities	References
1.	Abrus precatorius	Chirmi	Seed	Alcoholic extract	20 and 40 mg/kg	45 days	Rat	Antifertility effect	31
	precutorius		Seed	Ethanolic extract	20, 40  and  60  mg/kg	20 days	Mice	Antifertility effect	33
2.	Aegle marmelos	Bael	Leaf	50% ethanolic extract	100, 200 and 300 mg/kg b. wt./day	60 days	Rat	Antifertility effect	65
			Leaf	50% ethanolic extract	200 and 300 mg/kg b.wt./day	60 days	Rat	Antifertility effect	103
			Leaf	Aqueous extract	250mg/kg body wt. and 350mg/kg b wt	45 days	Rat	Antifertility effect	66
3.	Albizzia lebbeck	Siris	Pods	Methanolic extract	50, 100 and 200 mg/kg/day	60 days	Rat	Antifertility effect	104
			Bark	Methanolic extract	nig/kg/day 100 mg/rat/day	60 days	Rat	Antispermatogen ic and antiandrogenic activities	105
4.	Allamanda cathartica	Golden trumpet	Leaf	Aqueous extract	150 mg/kg b. wt./day	14, 28 and 42 days	Mice	Antifertility effect	55
5.	Allium sativum	Garlic	Bulb	Crude extract	5%, 10%, 15% and 30% crude garlic	30 days	Rat	Antispermatogen ic and antiandrogenic activities	106
			Bulb	Aqueous extract	500 and 1000 mg/kg/d	28 days	Rat	Antispermatogen ic Activity	107
6.	Aloe Barbadensis	Aloe Vera	Leaf	Aqueous extract	70 mg/kg and 100 mg/kg	56 days	Rat	Antifertility effect	108
7.	Amalakyadi churna			Ethanol extract	250 mg/kg and 400 mg/kg b. wt./day	30 days	Mice	Antifertility effect	109
8.	Anethum graveolens	Soya	Seeds	Aqueous extract	70 and 100 mg/kg	32 days	Rat	Antifertility effect	110
9	Andrographis paniculata	Kiryat	Leaves	Alcoholic extract	250 and 500 mg/kg b. wt.	30 and 60 days	Rat	Antifertility effect	111
10.	Austroplenckia populnea		Leaf	Hydro methanolic extract	500 mg/kg/day	70 days	Rat	Antifertility effect	112
11.	Azadirachta indica	Neem	Leaves	Dry powder	20, 40 and 60 mg/rat/day	24 days	Rat	Antispermatic activity	113
			Seeds	Aqueous extract	5mg/kg, 15mg/kg and 25mg/kg	6 days	Rat	Antifertility effect	114
			Leaves	Aqueous extract	100 mg/rat/day		Rat	Antispermatogen ic and antiandrogenic properties	115
			Seeds	Alcoholic extract	100 mg/kg	15 days	Mice	Antifertility effect	116
			Leaves	Aqueous extract	50, 100, and 200 mg/kg b. wt./day	28 days	Mice	Antispermatic activity	117
			Leaves		5.0%, 10.0% and 15.0% neem leaf meal	16 weeks	Rabbits	Antispermatogen ic effect	118
			Neem oil		0.6 and 1.2 mL of neem oil/animal	6 weeks	Rat	Structural changes	119

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			Leaves	Aqueous extract	250 and 350 mg/kg body wt.	30 days	Rat	Spermicidal Activity	120
12.	Bacopa monnieri	Brahmi	Leaves	Dry powder	250 mg/kg body wt./day	28 and 56 days	Mice	Suppression of spermatogenesis and fertility	51
13.	Barleria prionitis	Vajradanti	Root	Methanolic extract	100 mg/kg	60 days	Rat	Antispermatogen ic Activity	36
			Root	Alcoholic extract	100 mg/kg	60 days	Rat	Antifertility effect	37
14.	Cannabis sativa	Ganja	Root	Alcoholic extract	20 mg/day	20 consecutive days	Rat	Antispermatogen ic activities	121
15.	Capparis aphylla	kair		Ethanolic extract	50, 100 and 200 mg/kg	55 days	Rat	Antispermatogen ic Activity	47
16.	Carica papaya	Рарауа	Seeds	Aqueous extract	50 and 100 mg/kg b.wt.	8 weeks	Rat	Antispermatogen ic properties	122
			Seeds	Alcoholic extract	0.5 mg/kg	7 days	Rat	Affects cauda epididymis	123
			Seeds	Chlorofor m extract	50 mg/kg	360 days	Monke y	Antispermatogen ic effect	124
			Seeds	Alcoholic extract	100, 200 and 300 mg/kg b.wt.	45 days	Mice	Spermicidal activity	125
			Seeds	Chlorofor m extract	10 mg/rat/day	150 days	Rat	Ultrastructural changes in the testis	126
			Leaves	Aqueous extract	500 mg/kg b.wt.	21 days	Rat	Antifertility effect	127
			Seeds	Alcoholic extract	50 and 200 mg/kg/day	1 and 8 weeks	Rat	Antifertility effect	128
			Seeds	Chlorofor m extract	20 and 50 mg/animal/da y	150 days	Rabbit	Spermicidal activity	129
17.	Chromolaena odoratum		Leaves	Aqueous extract	250 and 500 mg/kg b.wt.	14 days	Rat	Antiandrogenic effects	104
18	Citrullus colocynthis	Tumba	Root	50% ethanolic extract	50, 100 and 200 mg/kg b.wt./day	60 days	Rat	Antispermatogen ic effects	130
			Fruit	50% ethanolic extract	100 mg/kg/day	20, 40, and 60 days	Rat	Antispermatogen ic and antiandrogenic activities	131
19.	Colebrookia oppositifolia	Binda	Leaves	Ethanolic extract	100 and 200 mg/kg	8-10 weeks	Rat	Depression of spermatogenesis	132
20.	Crotalaria juncea	Indian Hemp	Seeds	Petroleum ether, benzene and ethanol extracts	25 mg/kg	30 days	Mice	Antispermatogen ic and antiandrogenic effects	133
21.	Curcuma longa	Haldi	Rhizom e Rhizom	Methanolic extract Aqueous	500 mg/kg/day 600 mg/kg b.	60 days 56 and 84	Rat Mice	Antifertility effect Antifertility	26 27
22.	Dendrophthoe	Banda	e Stem	extract 70%	wt./day 100 mg/kg	days 60 days	Rat	effect Depression of	59
	falcata	Janua	Juli	methanolic extract	wt/day	oo uuyo	mat	spermatogenesis	57
23.	Fadogia agrestis		Stem	Aqueous extract	18, 50 and 100 mg/kg b.wt.	28 days	Rat	Adverse effects on the male rat testicular function	134
24.	Juniperus phoenica	Phoenicean Juniper or Arâr	Cones	Ethanolic extract	intraperitoneal injections of 400 or 800 mg/kg	21 consecutive days	Rat	function Antifertility activity	135
25	Leptadenia hastata		Leaves and	Aqueous extract	100, 200, 400 and 800 mg/kg	60 days	Rat	Antispermatogen ic Activity	136
26	Madhuca Indica	Mahua	stem Leaves	Alcoholic extract	b. wt./day 200 mg/kg b. wt./day	20 days	Rat	Activity Antifertility effect	137
27.	Martynia annua	Bichchhu	Root	50% ethanol	50, 100 and 200 mg/kg b.	60 days	Rat	Antifertility effect	76

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20	Mantha	Decall	I 6	extract	wt./day	20.40 1.60	M	A	120
28.	Mentha arvensis	Pudhina	Leaf	Petroleum ether extract	10 and 20 mg/mouse/ day	20, 40 and 60 days	Mice	Antifertility property	138
29.	Momordica charantia	Karela	Seeds	Petroleum ether, benzene and alcohol extracts	25 mg/100 g body weight	35 days	Rat	Antispermatogen ic and androgenic activities	84
30.	Mondia whitei	Whites Ginger	Root bark	Aqueous extract	400 mg/kg/day	55 days	Rat	Antispermatogen ic and antifertility activities	139
				Hexane extract	500 and 1000 mg/kg b.wt.	30 days	Rat	Antifertility activities	140
31.	Morinda lucida	Indian mulberry	Leaf	Leaf extract	400 mg/(kg·d)	13 weeks	Rat	Antispermatogen ic properties	141
32	Mucuna Urens		seed	Ethanol extract	70 mg/kg, 140 mg/kg, 210 mg/kg	14 days	Rat	antispermatogen ic activity	142
33	Ocimum sanctum	Tulsi	leaves	Benzene extract	300 mg/kg b. wt.	Fourty eight hours after the injection the rats were sacrificed	Rat	Antifertility property	143
			Leaf	Aqueous crude extract	11, 22, 44 and 88 mg/kg	1, 2 and 4 weeks	Mice	Antifertility effects	144
			Fresh Leaves	CALLACT	2 g of fresh leaves per rabbit	30 days	Rabbit	Antifertility effects	145
			leaves	Benzene extract	250 mg/kg b. wt./day	48 days	Rat	Antispermatogen ic and anti- androgenic property	12
34.	Piper betle	Pan	Leaf- stalk	Alcoholic extract	500 and 1000 mg/kg b. wt.	60 days	Mice	Antifertility effect	146
35.	Piper nigrum	Long pepper	Fruit	Dry powder	25 and 100 mg/kg	20 and 90 days	Mice	Antispermatogen ic and antifertility activity	41
36.	Quassia amara	Surinam wood	Bark	Chlorofor m extracts	different dilutions	Single daily intramuscula r injections of the extract for 15 days	Rat	Antifertility effect	147
37.	Rosmarinus officinalis	Rosemary	Fruit	Mehanolic extract	250 and 500 mg/kg b. wt.	63 days	Rat	Antispermatogen ic Activity	94
38.	Ruta graveolens	Sadab	Leaf	Alcoholic extract	20 mg/day	20 consecutive days	Rat	Antispermatogen ic activities	121
39.	Sapindus emarginatus	Ritha		Alcoholic extract	50 mg/day/rat	60 days	Rat	Antifertility activity	148
40.	Sarcostemma acidum	Somlata	Stem	70% methanolic extract	50 and 100 mg/kg/day	60 days	Rat	Arrest of spermatogenesis	149
41.	Syzygium aromaticum	Clove	Flower buds	Hexane extract	15mg, 30mg, and 60mg/kg b.wt.	35 days	Mice	Degenerative changes in the seminiferous tubules	98
42.	Tecoma stans	Piliya	Leaves	Ethanolic extract	500 mg/day/rat	60 days	Rat	Antispermatogen ic properties	150
43	Terminalia bellirica	Harad	Fruit	Alcoholic extracts	50 mg/day/rat	60 days	Rat	Antifertility effect	149
			Bark	Benzene and ethanol extract	10mg and 25mg/100g body weight of benzene and ethanol extracts	50 days	Rat	Structural and functional alteration	151
44	Thevetia peruviana	Yellow Oleander	stem bark	Methanol extract	100 mg/rat/day	60 days	Rat	antispermatogen ic activity	152

45.	Tinospora cordifolia	Neem giloy	Stem	70% methanolic extract	100 mg/rat/day	60 days	Rat	Antifertility effect	70
46	Trachyspermu m ammi	Ajvain	Fruit	Ethanolic extract	100, 200 and 400mg/kg	60 days	Rat	Antifertility effect	153
47.	Trigonella foenum- graecum	Fenugreek (Methi)	Seeds	Dry powder	feeding diets containing 30% fenugreek seeds		Rabbit	Antifertility activity	154
48	Zizyphus mauritiana	Ber	Bark	Aqueous, methanolic and saponin extracts	0.1mg/ml and 0.5mg/ml		Human	spermicidal property	155

### REFERENCES

- Thakur DS, Kumar P, Kujur A, Kumar P, Kumar R. Contribution of Male Contraception in World Population. J Pharm Sci & Res 2010; 2(7):384-93.
- Dehghan MH, Martin T, Dehghanan R. Antifertility effect of Iranian neem seed alcoholic extract on epididymal sperm of mice. Iranian Journal of Reproductive Medicine 2005; 3(2):83-89.
- Gupta RS, Sharma R. A review on medicinal plants exhibiting antifertility activity in males. Natural Product Radiance 2006; 5(5):389-410.
- Hoesla CE, Saadb F, Pöppela M, Altwein JE. Reversible, Non-Barrier Male Contraception: Status and Prospects. Eur Urol 2005 Nov; 48(5):712-22.
- Montaserti A, Pourheydar M, Khazaei M, Ghorbani R. Antifertility effects of *Physalis alkekengi* alcoholic extract in female rat. Iranian Journal of Reproductive Medicine 2007; 5(1):13-16.
- Mishra N, Joshi S, Tondon VL, Munjal A. Evaluation of Antifertility potential of aqueous extract of *Bougainvillea* spectabilis leaves in Swiss albino mice. Int J Pharm Sci Drug Res 2009; 1(1):19-23.
- El-Kashoury AA, Salama AF, Selim AI, Mohamed RA. Animal Model Study of Reproductive Toxicity of the Chronic Exposure of Dicofol. Life Science Journal 2009; 6(3):1-18.
- Beckman LJ, Harvey SM. Factors affecting the consistent use of barrier methods of contraception. Obstet Gynecol 1996; 88(3 Suppl):65S-71S.
- Moore PJ, Adler NE, Kegeles SM. Adolescents and the contraceptive pill: the impact of beliefs on intentions and use. Obstet Gynecol 1996; 88(3 Suppl):48S-56S.
- 10. Wu FCW. Male contraception. Baillière's Clinical Obstetrics and Gynaecology 1996; 10(1):1-23.
- 11. Jensen JT. Male contraception. Curr Womens Health Rep 2002; 2(5):338-45.
- Aladakatti RH, Mukhtar A, Nazeer AR, Ghodesawar MG. Effect of benzene leaf extract of *Ocimum sanctum* on testis and spermatogenic pattern in albino rats. International Journal of Current Research 2010; 5:22-29.
- 13. Zha SW, Zha J, Huang YF. Male antifertility drugs and cell apoptosis. Zhonghua Nan Ke Xue 2008; 14(1):75-8.
- 14. Garg S, Talwar GP, Upadhyay SN. Immunocontraceptive activity guided fractionation and characterization of active constituents of neem (*Azadirachta indica*) seed extracts. J Ethnopharmacol 1998; 60(3):235-46.
- 15. Hiremath SP, Rudresh K, Badani S, Patil SB, Patil SR. Post-coital antifertility activity of *Acalypha indica* L. J Ethnopharmacol 1999; 67(3):253-8.
- Hiremath SP, Badani S, Hunasagatta SK, Patil SB. Antifertility and hormonal properties of flavones of *Striga orobanchioides*. Eur J Pharmacol. 2000; 391(1-2):193-7.
- 17. Ghosh D, Jana D, Debnath JM. Effects of leaf extract of *Stephania hernandifolia* on testicular gametogenesis and androgenesis in albino rats: a dose-dependent response study. Contraception 2002; 65(5):379-84.
- Remya M, Sharma RC, Deepali M, Sakshi B, Nilesh P, Tharini S. In vitro effects of *Aegle marmelos* on human sperm Vitality. Biomedicine 2009; 29(2):183-85.

- 19. WHO, 2000. Reproductive health research at WHO: a new beginning, Biennial Report 1998-99, Special Programme of Research, Development and Research Training in Human Reproduction, World Health Organization, Geneva.
- Reddy CM, Murthy DRK, Paril SB. Antispermatogenic and androgenic activities of various extracts of *Hibiscus rosasinesis* in albino mice. Indian J Exp Biol 1997; 35(11):1170-4.
- 21. Yadav R, Jain GC. Antifertility effect and hormonal profile of petroleum ether extract of seeds of *Cassia fistula* in female rats. International Journal of Pharm Tech Research 2009; 1(3):438-44.
- Lobo R, Prabhu K, Shirwaikar A, Shirwaikar A, Ballal M. Formulation and Evaluation of Antiseptic Activity of the Herbal Cream Containing *Curcuma longa* and Tea Tree Oil. Journal of Biologically Active Products from Nature (JBAPN) 2011; 1(1):27-32.
- 23. Bernard GT, Esteban P, Christopher JS, Turmerones: Isolation from Turmeric and their Structure Determination, Chem Commun, 1982; 6:363
- Liao S, Lin J, Dang MT, Zhang H, Kao YH, Fukuchi J, Hiipakka RA. Growth suppression of hamster flank organs by topical application of catechins, alizarin, curcumin, and myristoleic acid. Arch Dermatol Res 2001; 293(4):200-5.
- 25. Rithaporn T, Monga M, Rajasekharan M. Curcumin: a potential vaginal contraceptive. Contraception 2003; 68(3):219-23.
- 26. Purohit A, Bhagat M. Contraceptive effect of *Curcuma longa* (L.) in male albino rat. Asian J Androl 2004; 6(1):71-4.
- Mishra RK, Singh SK. Reversible antifertility effect of aqueous rhizome extract of *Curcuma longa* L. in male laboratory mice. Contraception 2009; 79(6):479-87.
- Sudaroli M, Chatterjee TK. Evaluation of red and white seed extracts of *Abrus precatorius* Linn. against freund's complete adjuvant induced arthritis in rats. Journal of Medicinal Plants Research 2007; 1(4):086-094.
- Khare CP. Encyclopedia of Indian Medicinal Plants. Rational Western therapy, Ayurvedic and other traditional usage Botany, Springer, New York, pp. 3-5, 2004.
- Siddiqui S, Siddiqui BS, Naim Z. Studies in the steroidal constituents of the seeds of *Abrus precatorius* Linn. (scarlet variety). Pakistan Journal of Scientific and Industrial Research 1978; 21(5-6):158-161.
- Bhatt N, Chawla SL, Rao MV. Contraception evaluation of seed extracts of *Abrus precatorius* L. in male albino rats (*Mus musculus*). J Herb Med Toxicol 2007; 1:45-48.
- Ratnasoorya WD, Amarasekera AS, Parera NSD, Premakumara GAS. Sperm anti-motility properties of seed extract of *Abrus* precatorius. J Ethanopharmacol 1991; 38:85-90.
- 33. Jahan S, Saeed N, Ijlal F, Khan MA, Ahmad M, Zafar M et al. Histomorphological study to evaluate anti-fertility effect of *Abrus precatorius* L. in adult male mice; Journal of Medicinal Plants Research 2009; 3(12):1021-1028.
- Chavan CB, Hogade MG, Bhinge SD, Kumbhar M, Tamboli A. In vitro anthelmintic activity of fruit extract of *Barleria prionitis* linn. against *Pheretima posthuma*. International Journal of Pharmacy and Pharmaceutical Sciences 2010; 2(3):49-50.
- 35. Ata A, Kalhari KA, Samarasekera R. Chemical constituents of *Barleria prionitis* and their enzyme inhibitory and free radical

scavenging activities; Phytochemistry Letters 2009; 2(1):37-40.

- Verma PK, Sharma A, Joshi SC, Gupta RS, Dixit VP. Effect of isolated fractions of *Barleria prionitis* root methanolic extract on reproductive function of male rats: preliminary study. Fitoterapia 2005; 76:428-32.
- Gupta RS, Kumar P, Dixit VP, Dobhal MP. Antifertility studies of the root extract of the *Barleria prionitis* Linn in male albino rats with special reference to testicular cell population dynamics. J Ethnopharmacol 2000; 70(2):111-7.
- Park IK, Lee SG, Shin SC, Park JD, Ahn YJ. Larvicidal Activity of Isobutylamides Identified in *Piper nigrum* fruits against three mosquito species. J Agric Food Chem 2002; 50(7):1866-70.
- Parmar VS, Jain SC, Bisht KS, Jain R, Taneja P, Jha A. Phytochemistry of the Genus Piper. Phytochemisrry 1997; 46(4):591-673.
- Srinivas PV, Rao JM. Isopiperolein B: an alkamide from *Piper nigrum*. Phytochemistry 1999; 52:957-958.
- 41. Mishra RK, Singh SK. Antispermatogenic and antifertility effects of fruits of *Piper nigrum* L. in male mice. Indian Journal of Experimental Biology 2009; 47:706-712.
- 42. D'cruz SC, Mathur PP. Effect of piperine on the epididymis of adult male rats. Asian J Androl 2005; 7(4):363–368.
- Revathi P, Vani B, Sarathchandiran I, Kadalmani B, Shyam KP, Palnivel K. Reproductive toxicity of *Capparis aphylla* (Roth.) in male albino rats. Int J Pharm Biomed Res 2010; 1(3):102-112.
- 44. Mishra SN, Tomar PC, Lakra N. Medicinal and food value of Capparis—a harsh terrain plant. Indian J Traditional Knowledge 2007; 6(1):230-238.
- 45. Gupta AK, Tondon N, Sharma M. Quality Standards of Indian Medicinal Plant Medicinal Plants Unit: Published by Indian Council of Medical Research, New Delhi, 2008; 3: 99-105.
- 46. Singh P, Mishra G, Sangeeta, Srivastava S, Jha KK, Khosa RL. Traditional uses, phytochemistry and pharmacological properties of *Capparis decidua* : An Overview. Der Pharmacia Lettre 2011; 3(2):71-82
- 47. Sarathchandiran I, Manavalan R, Akbarsha MA, Kadalmani B, Karar PK. Studies on spermatotoxic effect of ethanolic extract of *Capparis aphylla* (Roth). J Boil Sci 2007; 7:544-548.
- Debnath M, Malik CP, Bisen PS. Micropropagation: A Tool for the Production of High Quality Plant-based Medicines. Current Pharmaceut Biotechnol 2006; 7:33-4.
- Chakravarty AK, Garai S, Masuda K, Nakane T, Kawahara N. Bacopasides III-V: Three new triterpenoid glycosides from *Bacopa monniera*. Chem Pharm Bull 2003; 51:215-217.
- 50. Russo A, Borrelli F. *Bacopa monniera*, a reputed nootropic plant: an overview; Phytomedicine 2005; 12:305–317.
- 51. Singh A, Singh SK. Evaluation of antifertility potential of Brahmi in male mouse. Contraception 2009; 79(1):71-9
- Islam MR, Ahamed R, Rahman MO, Akbar MA, Al-Amin M, Alam KD *et al.* In Vitro Antimicrobial Activities of Four Medicinally Important Plants in Bangladesh. European Journal of Scientific Research 2010; 39(2):199-206.
- Nayak S, Nalabothu P, Sandiford S, Bhogadi V, Adogwa A. Evaluation of wound healing activity of *Allamanda cathartica*. L. and *Laurus nobilis*. L. extracts on rats BMC Complementary and Alternative Medicine 2006, 6:12; 1-6.
- Nithya K, Muthumary J. Bioactive Metabolite Produced By Phomopsis Sp., An Endophytic Fungus in *Allamanda Cathartica* Linn. Recent Research in Science and Technology 2011; 3(3):44-48.
- 55. Singh A, Singh SK. Reversible antifertility effect of aqueous leaf extract of *Allamanda cathartica* L. in male laboratory mice. Andrologia 2008; 40(6):337-45.
- Dashora N, Sodde V, Bhagat J, Prabhu KS, Lobo R. Antitumor Activity of *Dendrophthoe falcata* against Ehrlich Ascites Carcinoma in Swiss Albino Mice. Pharmaceutical Crops 2011; 2:1-7.
- 57. Pattanayaka SP, Sunitab P. Wound healing, anti-microbial and antioxidant potential of *Dendrophthoe falcata* (L.f) Ettingsh. Journal of Ethnopharmacology 2008; 120(2):241-247.
- 58. Manthri S, Kota CS, Talluri M. Pharmacognostic, phytochemical and pharmacological review of *Dendrophthoe falcata*. J Phytol 2011, 3(3):18-25.

- Gupta RS, Kachhawa JB, Sharma A. Effect of methanolic extract of *Dendrophthoe falcata* Stem on Reproductive Function of Male Albino Rats. J Herb Pharmacother 2007; 7(2):1-13.
- Kumar R, Kumar A, Prasa CS, Dubey NK, Samant R. Insecticidal activity of *Aegle marmelos* (L.) Correa essential oil against four stored grain insect pests. Internet Journal of Food Safety 2008; 10:39-4.
- Kamalakkannan N, Rajadurai M, Prince PS. Effect of *Aegle marmelos* fruits on normal and streptozotocin-diabetic Wistar rats. J Med Food 2003; 6:93-98.
- 62. Sabu MC, Kuttan R. Antidiabetic activity of *Aegle marmelos* and its relationship with its antioxidant properties. Indian J Physiol Pharmacol 2004; 48: 81-88.
- 63. Shoba FG, Thomas M. Study of anti-diarrheal activity of four plants in castor oil-induced diarrhea. J Ethnopharmacol 2001; 76:73-76.
- Chatterjee A. Isolation of allo-imperatorin and β-sitosterol from the fruits of *Aegle marmelos* Correa. Journal of the Indian Chemical Society 1957; 34(3):228–230.
- 65. Chauhan A, Agarwal M, Kushwaha S, Mutreja A. Antifertility studies of *Aegle marmelos* Corr., an Indian medicinal plant on male albino rats. Egyptian Journal of Biology 2008; 10:28-35.
- 66. Sathiyaraj K, Sivaraj A, Madhumitha G, Kumar PV, Saral AM, Devi K, Kumar BS. Antifertility effect of aqueous leaf extract of *Aegle Marmelos* on male albino rats. International Journal of Current Pharmaceutical Research 2010; 2(1):26-29.
- Sivakumar V, Dhana Rajan MS, Riyazullah MS. Preliminary phytochemical screening and evaluation of free radical scavenging activity of Tinospora cordifolia. International Journal of Pharmacy and Pharmaceutical Sciences 2010; 2(4):186-188.
- Premanath R, Lakshmidevi N. Studies on Anti-oxidant activity of *Tinospora cordifolia* (Miers.) Leaves using in vitro models. Journal of American Science 2010; 6(10):736-743.
- Singh SS, Pandey SC, Srivastava S, Gupta VS, Patro B, Ghosh AC. Chemistry and medicinal properties of *Tinospora cordifolia* (Guduchi). Indian Journal of Pharmacology 2003; 35:83-91.
- Upadhyay AK, Kumar K, Kumar A, Mishra HS. *Tinospora* cordifolia (Willd.) Hook. f. and Thoms. (Guduchi) – validation of the Ayurvedic pharmacology through experimental and clinical studies. Int J Ayurveda Res 2010; 1(2):112–121.
- Gupta RS, Sharma A. Antifertility effect of *Tinospora cordifolia* (Willd.) stem extract in male rats. Indian J. Exp. Biol. 2003; 41(8): 885-9.
- Nirmal SA, Nikalje AG, Jadhav RS, Tambe VD. Anthelmintic activity of *Martynia annua* roots. Indian Drugs 2007; 44:772-3.
- Pillai TSK, Thampi PP, Verma KC. Preliminary pharmacognostiol study of *Martynia diandiae* Glox. Madha Bharti Journal 1964; 29:11-13.
- 74. Parvati A, Narayana IL. Chemotaxonomy of a few Pedaliaceae. Current Science 1978; 47:282-83.
- Tiwari KP, Massod M, Rathore YKS, Minocha PK. Study of anthocyanin from the flowers of some medicinal plants. Vijana Parishad Anusandhan Patrika 1978; 21:277.
- 76. Das VSR, Rao KN, Rao JVS. Phenolic acid in some members of Pedaliaceae. Current Science 1966; 35:160.
- 77. Mali PC, Ansari AS, Chaturvedi M. Antifertility effect of chronically administered *Martynia annua* root extract on male rats. J Ethnopharmacol 2002; 82(2-3):61-7.
- Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TPA. Indian herbs and herbal drugs used for the treatment of diabetes. J Clin Biochemistry Nutri 2007; 40:163-173.
- Cefalu WT, Ye J, Wang ZQ. Efficacy of dietary supplementation with botanicals on carbohydrate metabolism in humans. Endocrine, Metabolic & Immune Disorders - Drug Targets 2008; 8:78-81.
- Nahas R, Moher M. Complementary and alternative medicine for the treatment of type 2 diabetes. Can Fam Physician 2009; 55:591-596.
- 81. Scartezzini P, Speroni E. Review on some plants of Indian traditional medicine with antioxidant activity. J Ethnopharmacol 2000; 71:23-43.

- 82. Grover JK, Yadav SP. Pharmacological actions and potential uses of *Momordica charantia*: a review. J Ethnopharmacol 2004; 93:123-132.
- 83. Beloin N, Gbeassor M, Akpagana K, Hudson J, de Soussa K, Koumaglo K *et al.* Ethnomedicinal uses of *Momordica charantia* (Cucurbitaceae) in Togo and relation to its phytochemistry and biological activity. J Ethnopharmacol 2005; 96:49-55.
- 84. Lotlikar MM, Rao MRR. Pharmacology of a hypoglycaemic principle isolated from the fruits of *Momordica charantia* Linn. Indian Journal of Pharmacy 1966; 28:129-133
- Naseem MZ, Patil SR, Patil SR, Ravindra, Patil RS. Antispermatogenic and androgenic activities of *Momordica charantia* (Karela) in albino rats. J Ethnopharmacol 1998; 61(1):9-16.
- Altinier G, Sosa S, Aquino RP, Mencherini T, Loggia RD, Tubaro A. Characterization of topical antiinflammatory compounds in *Rosmarinus officinalis* L. J Agric Food Chem 2007; 55:1718-172.
- 87. Shahidi F. Antioxidants in food and food antioxidants. Nahrung 2000; 44:158-163.
- Masuda T, Inaba Y, Takeda Y. Antioxidant mechanism of carnosic acid: structural identification of two oxidation products. J Agric Food Chem 2001; 49:5560-5565.
- Leal PF, Braga ME, Sato DN, Carvalho JE, Marques MO, Meireles MA. Functional properties of spice extracts obtained via supercritical fluid extraction. J Agric Food Chem 2003; 51:2520-2525.
- Sá, RCS, Leite MN, Oliveira LEG, Toledo MM, Greggio TC, Guerra MO. Preliminary assessment of *Rosmarinus officinalis* toxicity on male wistar rats' organs and reproductive system. Brazilian Journal of Pharmacognosy 2006; 16:324-332.
- 91. Bhargava SK. Antiandrogenic effects of a flavonoid rich fraction of *Vitex negundo* seeds: a histological and biochemical study in dogs. J Ethnopharmacol 1989; 27:327-339.
- Nusier MK, Bataineh HN, Daradkah HM. Adverse Effects of Rosemary (*Rosmarinus officinalis* L.) on Reproductive Function in Adult Male Rats. Exp Biol Med (Maywood). 2007; 232(6):809-13.
- Banerjee S, Panda CK, Das S. Clove (*Syzygium aromaticum* L.), a potential chemopreventive agent for lung cancer. Carcinogenesis 2006; 27(8):1645-1654.
- Jirovetz L, Buchbauer G, Stoilova I, Stoyanova A, Krastanov A, Schmidt E. Chemical Composition and Antioxidant Properties of Clove Leaf Essential Oil. J Agric Food Chem 2006; 54(17):6303-6307.
- Bhuiyan NI, Begum J, Nandi NC, Akter F. Constituents of the essential oil from leaves and buds of clove (*Syzigium caryophyllatum* (L.) Alston); African Journal of Plant Science 2010; 4(11):451-454.
- 96. Mishra RK, Singh SK. Safety assessment of *Syzygium aromaticum* flower bud (clove) extract with respect to testicular function in mice. Food Chem Toxicol 2008; 46(10):3333-8.
- Oludare BT, Olumayokun AO, Olufunmilola OS, Makinde JM. Anti-inflammatory, antipyretic and antispasmodic properties of *Chromolaena odorata*. Pharmaceutical Biology 2000; 38(5):367-370.
- Owoyele BV, Soladoye AO. Analgesic and anti-inflammatory activity of ethanolic extract of *Chromolaena odorata* leaves. Recent Progr Med Plants 2007; 18:397-406.
- 99. Rao KS, Chaudhury PK, Pradhan A. Evaluation of anti-oxidant activities and total phenolic content of *Chromolaena odorata*. Food and Chemical Toxicology 2010; 48(2):729-73.
- 100. Ling B, Zhang M, Kong C, Pang X, Liang G. Chemical composition of volatile oil from *Chromolaena odorata* and its effect on plant, fungi and insect growth. Ying Yong Sheng Tai Xue Bao 2003; 14(5):744-6.
- 101. Igboh MN, Ikewuchi CJ, Ikewuchi CC. Chemical Profile of *Chromolaena odorata* L. (King and Robinson) Leaves; Pakistan Journal of Nutrition 2009; 8(5):521-524.
- 102. Yakubu MT, Akanji MA, Oladiji AT. Evaluation of antiandrogenic potentials of aqueous extract of *Chromolaena odoratum* (L.) K. R. leaves in male rats. Andrologia 2007; 39(6):235-43.

- 103. Chauhan A, Agarwala M. Reversible Changes in the Antifertility Induced by *Aegle marmelos* in Male Albino Rats. Systems Biology in Reproductive Medicine 2008; 54(6):240-246.
- 104. Gupta RS, Kachhawa JB, Chaudhary R. Antifertility effects of methanolic pod extract of *Albizzia lebbeck* (L.) Benth in male rats. Asian J Androl 2004; 6(2):155-9.
- 105. Gupta RS, Kachhawa JB, Chaudhary R. Antispermatogenic, antiandrogenic activities of *Albizia lebbeck* (L.) Benth bark extract in male albino rats. Phytomedicine 2006; 13(4):277-83.
- 106. Hammami I, Amara S, Benahmed M, El May MV, Mauduit C. Chronic crude garlic-feeding modified adult male rat testicular markers: mechanisms of action. Reprod Biol Endocrinol 2009; 7:65.
- 107. Omotoso GO, Oyewopo AO, Kadir RE, Olawuyi ST, Jimoh AAG. Effects of aqueous extract of *Allium sativum* (garlic) on semen parameters in wistar rats. The Internet Journal of Urology 2010; 7(2).
- 108. Oyewopo AO, Oremosu AA, Akang EN, Noronha CC, Okanlawon AO. Effects of aloe vera (*Aloe barbadensis*) aqueous leaf extract on testicular weight, sperm count and motility of adult male sprague-dawley rats. Journal of American Science 2011; 7(4):31-34.
- 109. Seetharam YN, Sujeeth H, Jyothishwaran G, Barad A, Sharanabasappa G, Umareddy B, et al. Antifertility effect of ethanolic extract of Amalakyadi churna in male albino mice. Afr J Med Sci 2003; 5(3):247-50.
- 110. Malihezaman M, Sara P. Effects of aqueous extract of Anethum graveolens on male reproductive system of rats. J Boil Sci 2007; 7:815-818.
- 111. Kazmi N, Pandey SK. Comparative histopathologicaal studis with the effects of Clerodendron siphonanthus (R.Br.) and Andrographis paniculata (Nees.) on reproductive organs of male albino rats. J Ecophysiol Occup Hlth 2009; 9:131-135.
- 112. Mazaro R, Di Stasi L, De Grava Kempinas W. Effects of the hydromethanolic extract of *Austroplenckia populnea* (Celastraceae) on reproductive parameters of male rats. Contraception 2002; 66(3):205-9.
- 113. Shaikh PD, Manivannan B, Pathan KM, Kasturi M, Ahamed RN. Antispermatic activity of *Azadirachta Indica* leaves in albino rats. Current Science 1993; 64(9): 688-701.
- 114. Mahmoudi MM, Morowati M, Ghazi Khansari M, Nasrollazadeh B, Minaie B. Sterility effects of Neem (*Azadirachta indica*) extract on male rat. J Reprod Infertil 2002; 3(2):4-13.
- 115. Kasturi M, Ahamed RN, Pathan KM, Manivannan B, Aladakatti RH. Ultrastructural changes induced by leaves of *Azadirachta indica* (Neem) in the testis of albino rats. J Basic Clin Physiol Pharmacol 2002; 13(4):311-28.
- 116. Dehghan MH, Martin T, Dehghanan R. Antifertility effect of Iranian neem seed alcoholic extract on epididymal sperm of mice. Iranian Journal of Reproductive Medicine 2005; 3(2): 83-89.
- 117. Mishra RK, Singh SK. Effect of aqueous leaf extract of *Azadirachta indica* on the reproductive organs in male mice. Indian journal of experimental biology 2005; 43(11):1093-1103.
- 118. Ifeanyi PO, Ifeanyi CO, Michael UI. Semen quality characteristics, reaction time, testis weight and seminiferous tubule diameter of buck rabbits fed neem (*Azadirachta indica* A. Juss) leaf meal based diets. Iranian Journal of Reproductive Medicine 2009; 7(1):23-28.
- 119. Shaikh MA, Naqvi SNH, Chaudhry MZ. Effect of neem oil on the structure and function of the mature male albino rat testes. Braz J Morphol Sci 2009; 26(1):49-54.
- 120. Sathiyaraj K, Sivaraj A, Vinoth Kumar P, Devi K, Senthil Kumar B. Spermicidal Activity of *Azadirachta indica* (Neem) Aqueous Leaf Extract on Male Albino Rats. International Journal of PharmTech Research 2010; 2(1):588-591.
- 121. Sailani MR, Moeini H. Effect of *Ruta graveolens* and *Cannabis sativa* alcoholic extract on spermatogenesis in the adult wistar male rats. Indian J Urol 2007; 23:257-60
- 122. Udoh P, Kehinde A. Studies on antifertility effect of pawpaw seeds (*Carica papaya*) on the gonads of male albino rats. Phytotherapy Research 1999; 13:226-228.

- 123. Verma RJ, Chinoy NJ. Effect of papaya seed extract on contractile response of cauda epididymal tubules. Asian J Androl 2002; 4:77-78.
- 124. Lohiya NK, Manivannan B, Mishra PK, Pathak N, Sriram S, Bhande SS, *et al.* Chloroform extract of *Carica papaya* seeds induces long-term reversible azoospermia in langur monkey. Asian Journal of Adrology 2002; 4:17-26.
- 125. Verma RJ, Nambiar D, Chinoy NJ. Toxicological effects of *Carica papaya* seed extract on spermatozoa of mice. Journal of Applied Toxicology 2006; 26:533-535.
- 126. Manivannan B, Mishra PK, Pathak N, Sriram S, Bhande SS, Panneerdoss S, *et al.* Ultrastructural changes in the testis and epididymis of rats following treatment with the benzene chromatographic fraction of the chloroform extract of the seeds of *Carica papaya*. Phytotherapy Research 2004; 18:285-289.
- 127. Oyekunle OA, Omope MM. Evaluation of andrological indices and testicular histology following chronic administration of aqueous extract of *Carica papaya* leaf in Wistar rat. African Journal of Pharmacy and Pharmacology 2010; 4(5):252-255.
- 128. Udoh P, Essien I, Udoh F. Effects of *Carica papaya* (paw paw) seeds extract on the morphology of pituitary–gonadal axis of male Wistar rats. Phytotherapy Research 2005; 19:1065-1068.
- 129. Lohiya NK, Mishra PK, Pathak N, Manivannan B, Jain SC. Reversible zoospermia by oral administration of the benzene chromatographic fraction of the chloroform extract of the seeds of *Carica papaya* in rabbits. Advances in contraception 1999; 15:141-161.
- 130. Mali PC, Chaturvedi M, Ansari AS, Dixit VP. Antispermatogenic Effects of an Ethanol Extract of *Citrullus colocynthis* Root in Male Albino Rats. Informa Healthcare 2001; 39(2):113-119.
- 131. Chaturvedi M, Mali PC, Ansari AS. Induction of reversible antifertility with a crude ethanol extract of *Citrullus colocynthis* Schrad fruit in male rats. Pharmacology 2003; 68(1):38-48.
- 132. Gupta RS, Yadav RK, Dixit VP, Dobhal MP. Antifertility studies of *Colebrookia oppositifolia* leaf extract in male rats with special reference to testicular cell population dynamics. Fitoterapia 2001; 72(3):236-45.
- 133. Vijaykumar B, Sangamma I, Sharanabasappa A, Patil SB. Antispermatogenic and hormonal effects of *Crotalaria juncea* Linn. seed extracts in male mice. Asian J Androl 2004; 6(1):67-70.
- 134. Yakubu MT, Akanji MA, Oladiji AT. Effects of oral administration of aqueous extract of *Fadogia agrestis* (Schweinf. Ex Hiern) stem on some testicular function indices of male rats. J Ethnopharmacol 2008; 115(2):288-92.
- 135. Shkukani HG, Salhab AS, Disi AM, Shomaf MS, Al Quadan F. Antifertility Effect of Ethanolic Extract of *Juniperus phoenica*(L.) in Male Albino Rats. J Herb Pharmacother 2008; 7(3/4):179-189.
- 136. Bayala B, Telefo PB, Bassole IHN, Tamboura HH, Belemtougri RG, Sawadogo L, *et al.* Anti-spermatogenic activity of *Leptadenia hastata* (Pers.) Decne leaf stems aqueous extracts in male wistar rats. Journal of Pharmacology and Toxicology 2011; 6(4):391-99.
- 137. Shivabasavaiah, Krishna Ram H, Pavana T, Ramyashree M, Ramya MC, Manjunath R. Antifertility Effects of *Madhuca Indica* Leaves in male swiss albino rats. Journal of Pharmacy Research 2011; 4(2):323-326.
- 138. Sharma N, Jocob D. Antifertility investigation and toxicological screening of the petroleum ether extract of the leaves of *Mentha arvensis* L. in male albino mice. J Ethnopharmacol 2001; 75(1):5-12.
- 139. Watcho P, Kamtchouing P, Sokeng S, Moundipa PF, Tantchou J, Essame JL, *et al.* Reversible antispermatogenic and antifertility

activities of *Mondia whitei* Linn. in male albino rat. Phytother Res 2001; 15(3):26-29.

- 140. Watcho, P., Donfack MM, Zelefack F, Nguelefack TB, Wansi S, Ngoula F, et al. Effects of the hexane extract of Mondia whitei on the reproductive organs of male rat. Afr J Trad Comp Alt Med 2005; 2(3):302-311.
- 141. Raji Y, Akinsomisoye O, Salman TM. Antispermatogenic activity of *Morinda lucida* extract in male Rats. Asian J Androl 2005; 7:405-410.
- 142. Etta HE, Bassey UP, Eneobong EE, Okon OB. Antispermatogenic Effects of Ethanol Extract of *Mucuna Urens*. Journal of Reproduction and Contraception 2009; 20(3):161-168.
- 143. Reghunandanan R, Sood S, Reghunandanan V, Mehta RM, Singh GP. Effect of *Ocimum sanctum* linn (tulsi) extract on testicular function. Indian J Med Sci 1995; 49:83-7.
- 144. Obianime AW, Aprioku JS, Esomonu CTO. Antifertility effects of aqueous crude extract of *Ocimum gratissimum* L. leaves in male mice. Journal of Medicinal Plants Research 2010; 4(9):809-816.
- 145. Sethi J, Yadav M, Sood S, Dahiya K, Singh V. Effect of tulsi (*Ocimum Sanctum* Linn.) on sperm count and reproductive hormones in male albino rabbits. Int J Ayurveda Res 2010; 1:208-10
- 146. Sarkar M, Gangopadhyay P, Basak B, Chakrabarty K, Banerji J, Adhikary P, *et al.* The reversible antifertility effect of *Piper betle* Linn. on Swiss albino male mice. Contraception 2000; 62(5):271-4.
- 147. Parveen S, Das S, Kundra CP, Pereira BM. A comprehensive evaluation of the reproductive toxicity of *Quassia amara* in male rats. Reprod Toxicol 2003; 17(1):45-50.
- 148. Venkatesh V, Sharma JD. A Comparative Study of Effect of Alcoholic Extracts of *Sapindus emarginatus, Terminalia belerica, Cuminum cyminum* and *Allium cepa* on Reproductive Organs of Male Albino Rats. Asian J Exp Sci 2002; 16:51-63
- 149. Verma PK, Sharma A, Mathur A, Sharma P, Gupta RS, Joshi SC, *et al.* Effect of *Sarcostemma acidum* stem extract on spermatogenesis in male albino rats. Asian J Androl 2002; 4(1):43-7.
- 150. Mathur N, Jain GC, Pandey G. Effect of *Tecoma stans* leaves on the reproductive system of male albino rats. Int J Pharmacol 2010; 6(2):152-156.
- 151. Patil SJ, Satishagouda S, Vishwanatha T, Patil SB. Effect of *Terminalia bellirica* barks extracts on activities of accessory reproductive ducts in male rats. International Journal of Pharmaceutical Sciences Review and Research 2010; 1(2):75-79.
- 152. Gupta R, Kachhawa JBS, Gupta RS, Sharma AK, Sharma MC, Dobhal MP. Phytochemical evaluation and antispermatogenic activity of *Thevetia peruviana* methanol extract in male albino rats. Human Fertility 2011; 14(1):53-59.
- 153. Surendra Kumar M, Reddy R, Manasa G, Vanaja P, Sirisha G, Astalakshmi N. Antifertility Effect Of *Trachyspermum ammi* (Linn) Sprague Fruits On Male Rats. International Journal of Pharmaceutical & Biological Archives 2011; 2(2):705-709.
- 154. Kassem A, Al-Aghbari A, AL-Habori M, Al-Mamary M. Evaluation of the potential antifertility effect of fenugreek seeds in male and female rabbits. Contraception 2006; 73(3):301-6.
- 155. Dubey R, Dubey K, Sridhar C, Jayaveera KN. Sperm immobilization activity of aqueous, methanolic and saponins extract of bark of *Ziziphus Mauritiana*. Der Pharmacia Sinica 2011; 2(2):11-16.