

ANTIBACTERIAL AND PHYTOCHEMICAL STUDIES OF VARIOUS EXTRACTS OF ROOTS OF *DECALEPIS HAMILTONII* WIGHT AND ARN

M.DEVI^a AND P.LATHA^b

^a Department of Biochemistry, Muthayammal College of arts and science, Rasipuram - 637408, Namakkal, Tamil Nadu, ^bDepartment of Biochemistry and Molecular Biology, Pondicherry University, Puducherry - 605014, India. Email: kmdevi6@gmail.com

Received: 11 Jan 2012, Revised and Accepted: 28 Feb 2012

ABSTRACT

In the present study, the various root extracts of *Decalepis hamiltonii* were screened phytochemically for the presence of secondary metabolites and for in vitro antibacterial activity respectively. The in vitro antibacterial activity of the various extracts of *Decalepis hamiltonii* was studied against *Escherichia coli*, *Klebsiella pneumonia*, *Salmonella typhi*, *Proteus mirabilis*, *Vibrio cholera*, *Shigella sonnie*, *Serratias sp*, *Staphylococcus aureus* and *Bacillus subtilis* by disc diffusion method. Streptomycin and Gentamycin were used as standard reference drugs while DMSO was included as a solubilizing agent as well as a negative control in this study. All the extracts were found to possess different degrees of antibacterial activity except aqueous extract.

Keywords: *Decalepis hamiltonii*, Phytochemical analysis, Antibacterial activity, Disc diffusion method.

INTRODUCTION

Decalepis hamiltonii Wight and Arn (swallow root) is a monogeneric climbing shrub endemic to the Deccan peninsula. *Decalepis hamiltonii* Wight and Arn commonly called as maredu kummulu or barre sugandhi or maradu gaddalu or makali beru belonging to the family Asclepiadaceae. Its roots have been used in Ayurveda, the ancient Indian traditional systems of medicine to stimulate appetite, relieve flatulence and as a general tonic¹. It is also useful as a blood purifier, preservative and as a source of bioinsecticide for stored food grains^{2,3}. Its tubers are consumed as pickles and as a juice for its alleged health promoting properties. Earlier studies have shown that roots contain aldehyde, inositols, saponins, amyryns and lupeols^{4,5,6} as well as volatile compounds such as 2-hydroxy-4-methoxybenzaldehyde, vanillin, 2-phenyl ethyl alcohol, benzaldehyde and others⁷. The roots have also been used as a substitute for *Hemidesmus indicus* in ayurvedic preparations of ancient Indian medicine¹. It possess potent antioxidant properties⁸, antiulcer⁹, anti-inflammatory and antipyretic¹⁰, gastroprotective¹¹ activities. In addition 4-hydroxyisophthalic acid, 14-aminotetradecanoic acid, 4-(1-hydroxy-1-methylethyl)-1-methyl-1,2-cyclohexane diol, 2-(hydroxymethyl)-3-ethoxybenzaldehyde, 2,4,8-trihydroxybicyclo (3.2.1) octan-3-one, bis-2,3,4,6-galloyl- α/β -D-glucopyranoside, bornerol and ellagic acid have been identified in swallow root^{12,13}. The present study was to analyze the presence of phytochemical and to evaluate the antibacterial activity of various extracts of *Decalepis hamiltonii* against several Gram positive and Gram negative bacterial strains in vitro.

MATERIALS AND METHODS

Plant material

The roots were collected from herbal suppliers in Chennai, India. The root (plant material) was identified and authenticated at Plant Anatomy and Research Center, Chennai, Tamil Nadu, India.

Preparation of plant extract

The roots were air dried under shade and powdered to 40 meshes coarse powder and stored in airtight bottles. 100g of *Decalepis hamiltonii* root powder was subjected to successive extraction with different solvents in increasing polarity viz. petroleum ether, benzene, chloroform, ethyl acetate, acetone, methanol, ethanol and distilled water by using Soxhlet apparatus. The solvents were evaporated under reduced pressure and stored in desiccators at 4°C.

Microorganisms used

The microbial strains were obtained from National Chemical Laboratory (NCL), Pune, India. The organisms were maintained on

nutrient agar (Hi Media, India) slope at 4°C and subculture before use. Among 9 microorganisms used, 7 Gram positive bacteria were *Escherichia coli*, *Klebsiella pneumonia*, *Salmonella typhi*, *Proteus mirabilis*, *Vibrio cholera*, *Shigella sonnie*, *Serratia sp* and 2 Gram negative bacteria were *Staphylococcus aureus* and *Bacillus subtilis*.

Preliminary phytochemical analysis

The qualitative chemical analysis of various extracts were carried out for the presence of alkaloids, flavanoids, saponins, steroids, glycosides, phenols, thiols and resins using the method adopted in similar surveys¹⁴.

Antibacterial activity

The antibacterial activity screening was performed by disc diffusion method¹⁵ for various extracts. The Mueller Hinton Agar (Hi Media) was used as bacteriological medium. Mueller Hinton Agar plates were prepared by pouring 15ml of molten media into the sterile petriplates. The plates were allowed to solidify for 15 minutes and 0.1% inoculum suspension was swabbed uniformly and inoculum was allowed to dry for 5 minutes. Under aseptic conditions, 6mm diameter (whatman no 1) filter paper disc were impregnated with 10 μ l (contains 5mg/ disc) of various extracts of *Decalepis hamiltonii* dissolved in DMSO. The discs were overlaid on MHA plates and incubated at 37°C for 24 hours. The diameter of zone of inhibition produced by the extracts was compared with standard drugs (10 μ g/ disc Gentamycin and 10 μ g/ disc Streptomycin). For each bacterial strain controls were maintained, where DMSO is used instead of extracts. The experiment was performed thrice to minimize the error and the mean values are presented and reported.

RESULTS AND DISCUSSION

Plants are important source of potentially useful structures for the development of new chemotherapeutic agents, the first step towards this goal is *in vitro* antibacterial activity. The extracts of higher plant can be very good sourcing of antibiotics against various bacterial pathogens¹⁶. The results of the phytochemical screening of the roots of *Decalepis hamiltonii* are presented in Table 1. The phytochemical analysis revealed the presence of flavanoids, saponins, tannins, steroids, cardiac glycosides. The antibacterial activity of extracts against 9 bacterial strains was presented in Table 2. All the extracts were found to possess different degrees of antibacterial activities except aqueous extract. Petroleum ether extract showed a broad spectrum antibacterial activities may be due to the presence of steroids and glycosides than other extracts. Methanolic extract showed antibacterial activity to all microorganisms except *Staphylococcus aureus*. Acetone extract showed antibacterial activity

except to *Shigella sonnei*. Some of the extracts were ineffective in this study do not possess antibiotic properties or the plant extracts may have antibacterial constituents just not in sufficient concentration so as to be effective.

Various workers have already shown that Gram positive bacteria are more susceptible towards plant extracts as compared to Gram negative bacteria^{17, 18}. These differences may be attributed to fact that the cell wall in Gram positive bacteria is of a single layer where as the Gram negative cell wall is multilayered structure¹⁹. Alternatively, the passage of the active compound through the Gram negative cell wall may be inhibited. Plant based antimicrobials have enormous therapeutic potential as they serve because of lesser side effects. Tannins are well known to possess general antimicrobial properties²⁰. Tannins are quite resistant to microbial attack and are known to inhibit the growth of some microorganisms. It is this antimicrobial effect of tannins that slow down the rate of biodegradation of soil organic matter. Antimicrobial agents can

damage pathogens in several ways. The major mode of actions is interference with cell wall synthesis, inhibition of protein synthesis, interference with nucleic acid synthesis, and inhibition of a metabolic pathway²¹.

The potential for developing antimicrobials from higher plants appears rewarding as it will lead to the development of a phytomedicine to act against microbes. Plant - based antimicrobials have enormous therapeutic potential as they can serve the purpose with lesser side effects that are often associated with synthetic antimicrobials²². However, the present study of in vitro antimicrobial evaluation of *Decalepis hamiltonii* forms a primary platform for further photochemical and pharmacological studies. The result of present study supports the traditional usage of the studied *Decalepis hamiltonii* and suggests that some of the plant extracts possess compounds with antimicrobial properties that can be used as antimicrobial agents in new drugs for the therapy of infectious disease caused by pathogens.

Table 1: Photochemical screening for various extracts of root of *Decalepis hamiltonii*

Secondary metabolites	PE	B	C	E	A	M	Et	Aq
Alkaloids	-	-	-	-	-	-	-	-
Flavanoids	-	-	-	+	++	+++	++	-
Glycosides	+	-	-	+	+	+	+	-
Steroids	+	+	+	+	+	++	+	-
Saponins	+	-	-	-	+	+	-	-
Tannins	+	-	-	+	+++	+++	++	-
Phenols	+	-	-	+	+	+	-	-
Resins	-	-	-	+	+	+	-	-
Thiols	-	-	-	+	-	+	-	-
Carbohydrates	++	-	+	+	+	++	++	-

PE - Petroleum ether B - Benzene C - Chloroform E - Ethyl acetate; A - Acetone M - Methanol Et - Ethanol Aq - Aqueous

Table 2: Antibacterial activity of various extracts of root of *Decalepis hamiltonii* against various bacterial strains by disc diffusion method

Microorganisms	PE	B	C	EA	A	M	Et	Aq	S	G	DMSO
Staphylococcus aureus	25	12	9	10	12	20	15	-	21	20	-
Bacillus subtilis	20	11	13	8	12	15	12	-	16	16	-
Escherichia coli	21	8	8	8	14	15	14	-	11	11	-
Salmonella typhi	20	10	9	8	12	16	10	-	15	10	-
Klebsiella pneumonia	18	-	10	10	12	11	-	-	14	13	-
Proteus mirabilis	23	8	8	-	18	14	12	-	16	15	-
Vibrio cholera	22	8	9	8	14	12	11	-	11	12	-
Shigella sonnei	17	8	9	9	12	14	12	-	-	-	-
Serratia spp	17	-	-	-	-	-	-	-	20	17	-

Values include cup border diameter (6mm)

Values are mean of three replicates

PE - Petroleum ether B - Benzene C - Chloroform E - Ethyl acetate; A - Acetone M - Methanol Et - Ethanol Aq - Aqueous; S - Streptomycin G - Gentamycin

ACKNOWLEDGEMENT

We thank the department of Microbiology and Biochemistry of Muthayammal College of Arts and Science, Rasipuram, Tamil Nadu, India for their encouragement and support technical support in testing the extracts for activity.

REFERENCES

- Nayar RC, Shetty JKP, Mary Z, Yoganarshimhan. Pharmacognostical studies on the root of *Decalepis hamiltonii* Wt and Arn and comparison with *Hemidesmus indicus* (L) R.Br. Proceedings of Indian Academy of Science 1978; 87: 37-48.
- George J, Pereira J, Divakar S, Udaysankar K, Ravishankar GA. A method for the preparation of active fraction from the root of *Decalepis hamiltonii*, useful as bioinsecticide, Indian Patent No. 1301/Dec/ 98, 1998.
- George J, Pereria J, Divakar S, Udaysankar K, Ravishankar GA. Current Science 1999; 77: 501-502.
- Murti PBR, Sheshadiri TR. A study of the chemical compounds of *Decalepis hamiltonii*. Proceedings of Indian Academy of Science 1940; 13: 221-232.
- Murti PBR, Sheshadiri TR. A study of the chemical compounds of *Decalepis hamiltonii*. Proceedings of Indian Academy of Science 1941; 13: 339-403.
- Murti PBR, Sheshadiri TR. A study of the chemical compounds of *Decalepis hamiltonii*. Proceedings of Indian Academy of Science 1941; 14: 93-99.
- Nagarajan S, Rao L J N, Gurudutt KN. Chemical composition of the volatile of *Decalepis hamiltonii* (Wright and Arn). Flavor and Fragrance journal 2001; 16: 27-29.
- Srivastava A, Shereen Harish R, Shivanandappa T. Antioxidant activity of the roots of *Decalepis hamiltonii* (Wright and Arn). LWT- Food Science and Technology 2006; 39: 1059-1065.

9. Yogender N, Jayaram S, Harish Nayaka MA, Lakshman, Dharmesh SM. Gastroprotective effect of Decalepis hamiltonii extract: Possible involvement of H⁺-K⁺-ATPase inhibition and antioxidative mechanism. *J Ethnopharmacol* 2007; 112: 173-179.
10. Lakshman K, Yoganasimhan SN, Shivaprasad HN, Jaiprakash B, Mohan S. Anti-inflammatory and antipyretic activity of Decalepis hamiltonii root extract. *Pharmaceutical Biology* 2006; 44(2): 127-129.
11. Naik Y, Jayaram S, Harish Nayaka MA, Lakshman, Dharmesh SM. Gastroprotective effect of swallow root (Decalepis hamiltonii) extract: possible involvement of H⁺-K⁺-ATPase inhibition and antioxidative mechanism. *J Ethnopharmacol* 2007; 112(1):173-9.
12. Srivastava A, Harish R, Shivanandappa T. Novel antioxidant compounds from the aqueous extract of roots of Decalepis hamiltonii (Wt and Arn) and their inhibitory effect on LDL oxidation. *J of Agricultural and Food Chemistry* 2006; 54: 790-795.
13. Srivastava A, Rao LJ, Shivanandappa T. Isolation of ellagic acid from the aqueous extract of roots of Decalepis hamiltonii. Antioxidant and cytoprotective activity. *Food Chemistry* 2007; 103: 224-233.
14. Harbone JB. *Phytochemical methods*. Chapman and Hall Ltd, London; 1973: p. 41-48.
15. Bassler AW, Kirby WMM, Sherris JC, Turek M. Antibiotic susceptibility testing by a standardized single disc method. *Am J Clin Pathol* 1966; 45: 493-496.
16. Sowmya G, Shetty, Vinayachandra, Hidayathulla S, Chandrashekar KR. Antimicrobial activity and Photochemical screening of Pterospermum reticulatum wight & arn. *Int J Pharm Pharm Sci* 2011; 3(5): 35-37.
17. Lin J, Opoku AR, Van Staden J. Preliminary screening of some traditional Zulu medicinal plants for anti-inflammatory and antimicrobial activity. *J Ethnopharmacol* 1999; 68: 267-274.
18. Parekh J, Chanda S. In vitro antimicrobial activities of extract of Launaea procumbens Roxb. (Labiatae), Vitis vinifera (Vitaceae) and Cyperus rotundus (Cyperaceae). *Afr J Biomedical Res* 2006; 9: 89-93.
19. Yao J, Moellering R. Antibacterial agents. In: *Manual of Clinical Microbiology*, Murray P, Baron E, Pfaller M, Tenover F, Tenover R (Eds), ASM, Washington DC, pp 1281-1290.
20. Murugan S, Uma Devi P, Kannika Parameswari N, Mani KR. Antimicrobial activity of Syzygium jambos against selected human pathogens. *Int J Pharm Pharm Sci* 2011; 3(2): 44-47.
21. Laxmi A, Siddhartha S, Archana M. Antimicrobial screening of methanol and aqueous extracts Swertia chirata. *Int J Pharm Pharm Sci* 2011; 3(4): 142-146.
22. Iwu MW, Duncan AR, Okunji CO. New antimicrobials of plant origin. In: Janick J ed. *Perspectives on New Crops and New Uses*. Alexandria, VA: ASHS Press; 1999: p 457- 462.