PHARMACOLOGICAL EVALUATION OF ANTIFERTILITY EFFECT OF ETHANOLIC EXTRACT OF DAEMIA EXTensa LEAF ON MALE RATS

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ABSTRACT

The effect of 50% ethanol extract of D. extensa leaves on reproduction was studied on male rats. The study was divided into four groups of five animals each. The first group (I) received vehicle alone to serve as control. The second, third and fourth groups (II, III and IV) of animals were administered the leaf extract daily 50mg/kg body weight, p.o.100mg/kg body weight, p.o. and 200mg/kg body weight, p.o., respectively, for a period of 60 days. Significant decreases in weight of testes, epididymis, seminal vesicle and ventral prostate were observed. No morphological changes were observed in Sertoli cells. The body weights of all groups were elevated markedly. No alterations were recorded in any hematological parameters. It is concluded that the 50% ethanol extract of D. extensa leaf produced dose related effect on male reproduction without altering general body metabolism.

Keywords: D. extensa leaf; Anti-fertility; Male rats; 50% Ethanol extract; Reproduction; Hormone levels.

INTRODUCTION

D. extensa (Asclepiadaceae) is a perennial twining herb, foul-smelling when bruised; Stems bears milky juice and covered with longer stiff erect hairs 1mm; Leaves are thin, broadly ovate and heart-shaped 2-12 cm long, covered with soft hairs; Greenish yellow or dull white, sweet-scented flowers born in axillary, double white corona at the base of a staminal column, long-peduncled, umbellate or corymbose clusters tinged with purple; Fruits paired with follicles 5.8 cm long and 1 cm in diameter, reflexed, beak long, covered with soft spinous outgrowth and release many seeds with long white hairs when they split open. Seeds are densely velvety on both sides. The entire plant constitutes the drug and is used as a medicine.

The quest for the oral contraceptive agent that can control human fertility is as old as recorded history. Although a wide variety of synthetic contraceptive agents are available, these cannot be used continuously due to their severe side effects. Hence, people are looking back to age old tradition of using herbal medicines, which have minimum side effects. India in general and Western Ghats region in particular has enormous wealth of medicinal plants. Presently, a major programme on systematic investigation of medicinal plants for their phytochemical, biological and pharmacological properties, including antifertility properties, was undertaken in our laboratory. As part of this research programme, we present in this paper antifertility efficacy of leaves of the plant D. extensa. The herb is antiseptic, antispasmodic, carminative, diaphoretic, emmenagogue, sedative and stimulant. A tea made from the leaves has traditionally been used in the treatment of fevers, headaches, minor respiratory infections, digestive disorders, menstrual complaints and various minor ailments. It is occasionally used as a treatment for intestinal worms. The leaves of D. extensa are flavonoids alkaloids, terpenoids, tannins, steroids and carbohydrates. Although a large number of compounds have been isolated from various parts of D. extensa. Phytochemical studies have shown the presence of cardenolides, alkaloids, triterpenes (lupeol), saponins, steroidal compounds. The seeds of D. extensa contain uzarigenin, coroglaucigen, calactin, calotropin, other cardenolides and a bitter resin, Pergularin and have a cardiotoxic action. It has been suggested that the plant seed action on the uterus is similar to that of prostatin and is not inhibited by progestrone.

MATERIALS AND METHODS

Animal model

Twenty colony bred adult male albino rats, 3-5 months old and weighing between 175 and 250 gm with proved fertility were marked properly and housed two or three animals in polypropylene cages under controlled environmental conditions (12- light: 12 h dark). They were fed pelleted standard rat feed (Ashirwad Food, Chandigarh, India) supplemented with soaked gram and wheat and allowed free access to water.

Plant material

One collection of the D. extensa leaves from Indore, Madhya Pradesh India. The plant is authenticated and voucher specimen submitted in the department. The leaves were shade dried, powered and soxhlet with ethanol (50% v/v) at 55-60 °C for 36 h. The solvent was distilled off under petroleum ether, benzene, chloroform and acetone to remove impurities left, if any during extraction. The yields of petroleum ether, benzene, chloroform and acetone and ethanol was respectively 1.5%, 1.8%, 2.0%, 2.4%, 5.8% and 9.6%. Thus the resulting mass was dried under vacuum and kept at 4 °C.

Determination of acute toxicity (LD50)

The acute toxicity for ethanolic and aqueous extracts of D. extensa leaves were determined in albino mice, maintained under standard conditions. The animals were fasted overnight prior to the experiment; fixed dose method was adopted as per OECD Guideline No. 420. Animal ethical clearance for performing the experiments on animals was obtained from the Institutional Animal Ethical Committee (IAEC).

Treatment protocol

Animals were equally distributed into four treatment groups containing five in each.

Group I—Animals in this group were given vehicle (Sterile distilled water) alone p.o. for 60 days to serve as vehicle treated control.

Group II—Animals in this group were treated with D. extensa leaf (50% EtOH) extract at the dose of 50 mg/kg body weight/day; p.o. for 60 days.

Groups III—Animals in this groups received D. extensa leaf (50% EtOH) extract at the dose of 100 mg/kg body weight/day; p.o. for 60 days.

Group IV—Animals in this groups received D. extensa leaf (50% EtOH) extract at the dose of 200 mg/kg body weight/day; p.o. for 60 days.

A suspension of the extract was prepared in sterile distilled water (100 mg/ml) before administration. The required drug was administered orally with a glass syringe fitted with a feeding needle.
Sacrification schedule
Twenty-four hours after their last dose, the rats were weighed and sacrificed under light ether anesthesia.

Parameter
Sperm motility and count
Cauda epididymal sperm motility and count, and testicular sperm count (mature spermatozoa with head and tail) were made according to the procedure of Prasad et al. (1972). One hundred milligram of each tissue was minced in 1 ml of physiological saline. For sperm motility, one drop of evenly mixed sample was applied to a glass slide under the cover glass. The percent motility was determined by counting both motile and immotile spermatozoa per unit area. Cauda epididymal and testicular sperm count were also made expressed as million/mm$^3$ of suspension.

Fertility test
Male rats were introduced to female, 200-300 gm (male: female ratio, 1:2) at 17:00 h after 55 days of treatment. The successful mating was confirmed in the forthcoming morning from 56 to 61 day by vaginal plug and spermatozoa in the vaginal smear. The inseminated female were separated and allowed to deliver at term and the number of pups delivered and their characteristics were noted.

Body and organ weights
The initial and final body weights of the animals were recorded. The testes, epididymides, seminal vesicle and ventral prostate were dissected out, freed from adherent tissues and blood, and weighed to the nearest milligram.

Testicular histology
One (right) of the two testes of each animal was fixed in Bouin’s fixative solution, dehydrated in graded ethanol, cleared in xylene and embedded in paraffin wax. Section were cut at 5 microm, stained with Harris hematoxylin and eosin and observed under a light microscope.

Hematology
The counts of RBC and WBC, hemoglobin, hematocrit, and standard hematological indices (color index, mean corpuscular hemoglobin, mean corpuscular hemoglobin concentration and mean corpuscular volume) were determined from the blood collected directly from the heart at the time of scarification.

Statistical Analysis
Data are expressed as mean ± S.D. and analyzed for statistical significance by using one way analysis of variance (ANOVA). Results were considered significant at the $P \leq 0.05$ level.

RESULT AND DISCUSSION
Sperm motility and count
A significant decrease ($P \leq 0.01$) in percent cauda epididymal sperm motility was evident in groups II, III and IV animals when compared with group I animals. After 60 days of treatment only 37.5 ± 9.8, 32.5 ± 6.7 and 29.6 ± 9.8%, respectively, of spermatozoa versus 60.2 ± 2.5% of spermatozoa in group I was found to be motile. The sperm count from the cauda epididymis and testis were also diminished significantly in all treatment groups ($P \leq 0.01$) (Table 1).

Fertility
The number of fertile males decreased in all treatment groups, leaving 3, 2 and 1 still fertile 60 days of treatment, respectively, in groups II, III and IV. The ratio between delivered and inseminated female (5/10, 4/10 and 2/10 animals versus 10/10 animals in groups I), and the number of pups (50, 28 and 16 pups versus 90 pups in group I) dropped after 60 days of treatment. However, no significant difference was observed in the litter size of the female in any group. Spermatozoa with shortened and thinned flagella were present in the semen found in the vaginal smears of females, which were cohabited with the treated males. All delivered pups were normal healthy (Table 2).

Body and organ weights
The final body weight of all groups increased markedly when compared with their respective initial body weights ($P \leq 0.01$). The final weights of group III (100mg/kg b. wt, po) and group IV (200mg/kg b. wt, po) significantly increased when compared with the final body weight of group I (vehicle treated control) animals ($P \leq 0.01$). A great decline in the weights of testes, epididymides, seminal vesicle and ventral prostate (expressed in mg/100 gm of body weight) were observed in all treatment groups when compared with group I animals (Table 3).

Histopathology of testis
The testes of group I (vehicle treated control) animals showed normal features with successive stage of transformation of the seminiferous epithelium into spermatozoa. Leydig cells were situated in between the tubules (Fig. 1). Histopathological examination of the testis after 60 days of treatment showed a clear correlation between the dose and the severity of lesion of the seminiferous epithelium. In rats treated with 50 mg/kg, po (group II), some lesion were observed and affected only a few tubules (Fig. 2). The dose of 100 mg/kg, po (group III) produced diffuse alteration of the tubules (Fig. 3). In rates treated with 200 mg/kg, po (group IV) almost all tubules were affected (Fig. 4). Since the differences among the doses were more quantitative than qualitative, only a general description of the findings related to all treatment groups is given. The seminiferous tubules to appear reduced in size with a frequently filled eosinophilic material but with normal lamina propria. In general, diminished spermatogenesis was evident at secondary spermatocyte stage. Pachyteme spermatocytes were undergoing degeneration. Disorganization and sloughing of immature germ cells were visible. The nuclei became pyknotic. Leydig cells revealed sign of atrophy. Contrary to this, no morphological changes were observed in the Sertoli cells.

The drug treated male rats clearly indicates that the drug caused structural and functional alteration in testes, epididymides and seminal vesicle. Interestingly, it was observed that sperm morphology remained altered in placebo - as well as drug treated animals. Depletion of sperm count in the drug treated animals suggests alteration in sperm production in the testes. Decrease in sperm motility suggests alteration of sperm maturation in the epididymides.

Hematology
Hematological parameter, RBC and WBC counts, hemoglobin, hematocrit and standard hematological indices varied within the control range. No drug-related effects on any of these parameters were observed in any groups when compared with vehicle treated control group (group I).

The present data shows that the 50% ethanol extract of D. extensa leaf suppresses testicular and epididymal sperm counts and causes lesions on the seminiferous tubules related to the dose. However, the FSH levels remained unaltered. It is a well known and widely accepted concept that LH is basically responsible for testosterone production. It is probable that the primary step in the mechanism of the effect on testis induced by the D. extensa extract was the suppression of LH. At the testicular level, the absence of stimulation by LH would secondarily cause Leydig cell dysfunction, thereby resulting in decline of testosterone secretion which is responsible for diminished spermatogenesis and hence, reduction in sperm counts. It is known that the structure and function of the epididymis are dependent on androgens. The impaired epididymal function may also be due to reduced activity of the testis which affects the normal passage of testicular fluid into the epididymis. This is also confirmed by reduced epididymal weight.
Fig. 1: Photomicrograph of testis of a rat of group I (vehicle treated control) showing normal features with successive stage of transformation of seminiferous epithelium to spermatozoa.

Fig. 2: Photomicrograph of an affected region of testis of a rat of group II (50 mg/kg body weight, po) after 60 days of treatment. None disorganized germinal epithelium.

Fig. 3: Photomicrograph of testis of a rat of group III (100 mg/kg body weight, po) after 60 days of treatment showing cellular damage in tubular elements.

Fig. 4: Photomicrograph of testis of a rat of group IV (200 mg/kg body weight, po) after 60 days of treatment. Note the arrest of spermatogenesis.
Table 1: Sperm characteristic after 60 days of treatment with 50% ethanol extract of D. extensa leaves on male rats.

<table>
<thead>
<tr>
<th>S. No.</th>
<th>Treatment group</th>
<th>Testes</th>
<th>Epididymis</th>
<th>Cauda epididymis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>count (%)</td>
<td>count (million/mm³)</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Group I</td>
<td>60.2 ± 2.8</td>
<td>4.8 ± 0.2</td>
<td>50.0 ± 3.2</td>
</tr>
<tr>
<td>2</td>
<td>Group II</td>
<td>37.5 ± 9.8&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.6 ± 0.8&lt;sup&gt;b&lt;/sup&gt;</td>
<td>20.2 ± 7.0&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>3</td>
<td>Group III</td>
<td>32.5 ± 6.7&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.1 ± 0.5&lt;sup&gt;a&lt;/sup&gt;</td>
<td>16.0 ± 1.5&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>4</td>
<td>Group IV</td>
<td>29.4 ± 9.8&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.9 ± 0.4&lt;sup&gt;a&lt;/sup&gt;</td>
<td>11.0 ± 2.4&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

Data are expressed as mean ± S.D., n = 5

<sup>a</sup>P < 0.01 Compared with corresponding group I

Table 2: Fertility of male rats after 60 days of treatment with 50% ethanol extract of D. extensa leaf (Male: Female ratio, 1: 2.)

<table>
<thead>
<tr>
<th>S. No.</th>
<th>Treatment groups</th>
<th>No. of fertile males/no. of treated males</th>
<th>No. of females delivered/no. of inseminated female</th>
<th>Total no. of pops</th>
<th>Litter size ± S.D.</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Group I</td>
<td>5/5</td>
<td>10/10</td>
<td>90</td>
<td>8.5 ± 0.80</td>
</tr>
<tr>
<td>2</td>
<td>Group II</td>
<td>3/5</td>
<td>05/10</td>
<td>50</td>
<td>10.6 ± 1.05</td>
</tr>
<tr>
<td>3</td>
<td>Group III</td>
<td>2/5</td>
<td>04/10</td>
<td>28</td>
<td>6.5 ± 1.10</td>
</tr>
<tr>
<td>4</td>
<td>Group IV</td>
<td>1/5</td>
<td>02/10</td>
<td>16</td>
<td>7.0 ± 1.20</td>
</tr>
</tbody>
</table>

n = 5 (male), n = 10 (female), compared with group I

Table 3: Body and organ weights after 60 days of treatment with 50% ethanol extract of D. extensa leaf on male rats

<table>
<thead>
<tr>
<th>S. No.</th>
<th>Treatment groups</th>
<th>Body weight (gm)</th>
<th>Testes*</th>
<th>Epididymis*</th>
<th>Ventral Prostate*</th>
<th>Seminal vesicle*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Initial</td>
<td>Final</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Group I</td>
<td>21.50 ± 10.2</td>
<td>285.2 ± 16.0&lt;sup&gt;b&lt;/sup&gt;</td>
<td>131.0 ± 16.0</td>
<td>447.4 ± 9.4</td>
<td>270.2 ± 86.0</td>
</tr>
<tr>
<td>2</td>
<td>Group II</td>
<td>214.0 ± 34.2</td>
<td>283.2 ± 33.3&lt;sup&gt;b&lt;/sup&gt;</td>
<td>908.3 ± 105.6&lt;sup&gt;c&lt;/sup&gt;</td>
<td>333.8 ± 11.8</td>
<td>122.2 ± 8.2</td>
</tr>
<tr>
<td>3</td>
<td>Group III</td>
<td>215.0 ± 12.5</td>
<td>335.5 ± 46.6&lt;sup&gt;bc&lt;/sup&gt;</td>
<td>800.2 ± 150.4&lt;sup&gt;c&lt;/sup&gt;</td>
<td>308.5 ± 180.0&lt;sup&gt;c&lt;/sup&gt;</td>
<td>111.8 ± 7.9&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>4</td>
<td>Group IV</td>
<td>226.0 ± 16.2</td>
<td>326.0 ± 21.8&lt;sup&gt;bc&lt;/sup&gt;</td>
<td>772.4 ± 104.9&lt;sup&gt;c&lt;/sup&gt;</td>
<td>297.4 ± 11.4&lt;sup&gt;c&lt;/sup&gt;</td>
<td>110.0 ± 7.9&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

Data are expressed as mean ± S.D.

<sup>a</sup>mg/100 gm of body weight; <sup>b</sup>P < 0.01 compared with corresponding initial body weight; <sup>c</sup>P < 0.01 Compared with corresponding group

CONCLUSION

In conclusion, the oral administration of 50% ethanol extract of D. extensa leaf to male rats produced dose related effect on reproduction. Further long term studies are in progress for the evaluation of complete and reversible fertility with this extract and also other extracts of this important plant.

REFERENCES