ISSN- 0975-1491 Vol 4, Issue 4, 2012

Review Article

LOCAL DRUG DELIVERY IN PERIODONTICS: A STRATEGIC INTERVENTION

DR. VIDYA DODWAD*, DR. SHUBHRA VAISH, DR. AAKRITI MAHAJAN, DR. MEHAK CHHOKRA

Received: 15 Jun 2012, Revised and Accepted: 1 July 2012

ABSTRACT

Periodontitis is an inflammatory disease involving supportive structures of the teeth which is seen universally in all groups, ethnicities, races and both genders. Various antibacterial agents have been used effectively in the management of periodontal infection. The effectiveness of mechanical debridement of plaque and repeated topical and systemic administration of antibacterial agents are limited due to the lack of accessibility to periodontopathic organisms in the periodontal pocket. Local delivery of antimicrobials has been investigated for the possibility of overcoming the limitations of conventional therapy. These products provide a long-term, effective treatment at the site of infection at much smaller doses. Prospective multicentre studies considering risk factors for disease progression have to be designed to identify patients who may benefit the most from Local drug delivery.

INTRODUCTION

Periodontal disease is a general term which encompasses several pathological conditions affecting the tooth supporting structures. It includes conditions such as chronic periodontitis, aggressive periodontitis, systemic disease-associated periodontitis and necrotizing periodontitis.¹ It has been well-established that periodontal disease is the result of a local bacterial infection with a pathogenic microflora within the periodontal pocket. The microflora found in periodontitis is complex and composed mainly of gram negative anaerobic bacteria.² Moreover; studies have shown that the various clinical forms of periodontitis are associated with different microbiota.³

Traditional therapies for periodontal disease have included mechanical debridement to disrupt the subgingival flora and provide clean, smooth and biologically compatible root surfaces. Unfortunately, in some instances, the complex anatomy of the root and the contours of the lesion may hamper the treatment and prevent sufficient reduction of the bacterial load to make the tooth surface biologically acceptable. In addition, control of supragingival plaque is essential in order to prevent recolonization of the subgingival area by periodontal pathogens.⁴ Indeed, several clinical studies have clearly indicated that scaling and root planing, in combination with optimal oral hygiene, result in an alteration of the subgingival plaque which is sufficient to stop periodontal destruction in most cases.⁵ It also has been shown that patients who fail to achieve acceptable plaque control during or after subgingival treatment frequently suffer from recurrent periodontitis. Thus, oral hygiene is of the utmost importance for the clinical outcome of nonsurgical as well as surgical treatment.

However, severe or aggressive forms of periodontitis in young subjects often cannot be arrested by mechanical treatment alone. Furthermore, there are some patients or sites where even repeated treatment fails to stop the disease. These are referred to as refractory subjects or non-responding sites.⁶ This could be related to the persistence of pathogens in the pocket after treatment or to the production by the bacteria of specific virulence factors interfering with the host defense (e.g. leukotoxin production, encapsulation, etc.). It also could be due to the recolonization of treated sites from bacterial reservoirs such as dentinal tubules and soft tissues.⁷ In this context, it is evident that antimicrobial agents are of great interest and may be valuable as adjuncts to mechanical therapy.⁸

Systemic Therapy - Drawbacks?

Systemically applied antimicrobials have been advocated for the treatment of severe forms of periodontitis. Early approaches to systemic antibiotics in periodontal therapy included mainly single drug therapies with Tetracyclines, Penicillins, Metronidazole or Clindamycin.⁹ However, in the early 1970s, concern emerged with respect to systemic antibiotic therapy for chronic infections such as

periodontal disease. Indeed, side effects including hypersensitivity, gastrointestinal intolerance and the development of bacterial resistance have been described. Some studies also reported poor results due to the fact that the active product could not achieve an adequate concentration at the site of action and/or due to the inability of the active product to be retained locally for a sufficient period of time.

These drawbacks would be markedly reduced if antimicrobial agents applied locally could be used, although unwanted effects such as gastrointestinal disturbances and development of antibiotic resistance cannot be totally ruled out.¹¹ The local tissue concentration of a drug can be enhanced by incorporating the active agent into controlled release delivery systems to be placed directly in the periodontal pocket. Such systems may have applications where systemic drugs are currently used, for instance localized juvenile periodontitis, refractory periodontitis and periodontitis with secondary systemic involvement, e.g. HIV periodontitis.

Sustained local delivery systems might also be recommended for sites considered as difficult to instrument because of depth or anatomical complexity, for example in the case of furcation defects.¹²

However, studies suggest that these systems, used as adjuncts to scaling and root planing, give a slight advantage over mechanical treatment alone. On the other hand, few studies have shown promising results with local drug delivery systems on sites that responded poorly or showed recurrence after scaling and root planing.¹³ If such improvements are maintained for a long-term period, then these systems would be an interesting tool in the management of localized periodontal lesions.

Local Delivery Agents

The choice of the antimicrobial agents in periodontal diseases must be based on the bacterial etiology of the infection. Several antibiotics have been tested for their clinical and microbiological efficacy in periodontal diseases. It can be noted that only a limited number of antimicrobial agents have been used so far in formulations of local delivery systems.

There are distinct phases in a periodontal treatment plan where a dental practitioner can use a sustained release device. It can be used as an adjunct to scaling and root planing and for periodontal maintenance therapy. It can be safely used in medically compromised patients for whom surgery is not an option or those who refuse surgical treatment. It is highly contraindicated in patients with known hypersensitivity to the antimicrobial used as local drug and the delivery of antimicrobial using ultrasonic devices is contraindicated in asthmatics and infective conditions such as AIDS. Tuberculosis.

It has been observed that the local route of drug delivery can attain 100-fold higher concentrations of an antimicrobial agent in

subgingival sites compared with a systemic drug regimen. This reduces the total patient dose by over 400 fold thereby reducing the potential problems with the use of systemic antibiotic drug regimens and development of drug-resistant microbial populations at non oral body sites. 14 The only problem faced while using local drug delivery is the difficulty in placing therapeutic concentrations of the antimicrobial agent into deeper parts of periodontal pockets and furcation lesions. Lack of adequate manual dexterity, limited understanding of periodontal anatomy, and poor compliance limits the use of antimicrobial agents by patients as a part of their home self-care procedures. It also does

not markedly affect periodontal pathogens residing within adjacent gingival connective tissues and on extra-pocket oral surfaces (tongue, tonsils and buccal mucosa), which increases the risk of re-infection.

Various antimicrobial agents have been used and investigated as local drug delivery in the treatment of periodontal disease. These devices may be sustained release (drug delivery for less than 24 hrs) or controlled release (drug delivery for more than 24 hrs). Most important local drug delivery agents along with their commercial names have been listed in Table 1.

Table 1:

Agent	Type of delivery	Product available	Dosage form and concentration
Metronidazole Dental Gel	Sustained	Elyzol (25%)	biodegradable gel
Minocycline	Sustained	Dentomycin gel (2%)	Biodegradable gel
		Arestin (2%)	Biodegradable mix in syringe
Tetracycline Fiber	Controlled	Actisite (25% w/v tetracycline HCL)	Nonresorbable fiber
		Periodontal plus AB	Resorbable fiber
Chlorhexidine Chip	Controlled	Periochip (2.5mg)	Biodegradable device
		Periocol (2.5mg)	Biodegradable Gel
		Chlosite (1.5% CHX)	
Doxycycline Polymer	Controlled	Atridox (10%)	Bio degradable powder in syringe

Various agents used are

1. Tetracycline

Goodson et al in 1979 first proposed the concept of controlled delivery in the treatment of periodontitis. The first delivery devices involved hollow fibers of cellulose acetate filled with tetracycline. Tetracyclines are a group of closely related bacteriostatic antimicrobials. They have been frequently used in treating refractory periodontitis, including localized aggressive periodontitis. 15

Fibers: The ACTISITE tetracycline fibres have been approved for the treatment of adult periodontitis both by the United States Food and Drug Administration (FDA) and by the European Union's regulatory agencies. These are non-resorbable biologically inert, generally considered as safe, plastic copolymer (ethylene and vinyl-acetate) loaded with 25% w/w tetracycline HCI powder packaged as a thread of 0.5 mm in diameter and 23 cm in length. It maintains constant concentrations of active drug in the crevicular fluid in excess of 1000 µg/mL for a period of 10 days. Following application of tetracycline fibres a definite reduction in the subgingival microbiota has been observed. 20

Recently bioresorbable tetracycline fiber has been developed with base of collagen film, which is commercially available as PERIODONTAL PLUS AB. It offers the advantage of no second appointment for removal as it biodegrades within 7 days.

Gel

Tetracycline-Serratiopeptidase-Containing Periodontal Gel Formulation has shown statistically significant results along with scaling and root planing.¹⁸

Bioerodible Injectable Poly (ortho ester) for Tetracycline Controlled Delivery formulations loaded with tetracycline 10% or 20% showed complete in vitro degradation concomitant with drug release.¹⁹

In general, the data indicates that tetracycline fibers used as a monotherapy without adjunctive scaling and root planing were effective at reducing probing depths, in gaining clinical attachment and reducing monitored bacteria^{21,22}, though the results of monotherapy were not significantly different when compared to scaling and root planing.²³

Tetracycline fibers have shown a better result when used as an adjunct to scaling and root planing in patients with refractory periodontitis than in patients with adult periodontitis.²⁴ Similarly, other investigations²² and several case reports²⁵ have indicated better results when combined therapy was used at non-responsive sites.

2. Subgingival Doxycycline

Doxycycline is a bacteriostatic agent and has the ability to downregulate MMP's a family of zinc dependent enzymes that are capable of degrading a variety extracellular matrix molecules including collagens.²⁸

The only FDA approved 10% doxycycline in a gel system ATRIDOX (42.5 mg doxycycline) is a subgingival controlled-release product composed of a 2 syringe mixing system.

Doxycycline levels in GCF peaked to 1,500 - 2000 μ g/mL in 2 hours following treatment with ATRIDOX. These levels remained above 1000 μ g/mL through 18 hours, at which time the levels began to decline gradually. Local levels of doxycycline have been found to remain well above the minimum inhibitory concentration for periodontal pathogens (6.0 μ g/mL) through Day 7. Approximately 95% of the polymer is bio absorbed or expelled from the pocket naturally within 28 days. ²⁹

Several studies have reported the efficacy of 10% doxycycline hyclate as a local delivery antimicrobial agent for attaining probing depth reduction and gaining clinical attachment.^{30,31}

3. Subgingival Minocycline

Local delivery of minocycline, a bacteriostatic antibiotic 15 has been tried clinically via in three different modes i.e. film, microspheres, and ointment, 32,33

Film

Ethyl cellulose film containing 30% of Minocycline were tested as sustained release devices. The results of this study indicated that the use of this device may cause complete eradication of pathogenic flora from the pocket after 14 days. $^{\rm 35}$

Microsphere

The FDA recently approved a new, locally delivered, sustained-release form of minocycline microspheres (ARESTIN) for subgingival placement. The 2% minocycline is encapsulated into bioresorbable microspheres ($20\text{-}60\mu\text{m}$ in diameter) in a gel carrier and has resorption time of 21 days. Gingival crevicular fluid hydrolyses the polymer and releases minocycline for a period of 14 days or longer before resorbing completely. 35

Ointment

Minocycline ointment is a bioabsorbable sustained delivery system consisting of 2% minocycline hydrochloride in a matrix of

hydroxyethyl-cellulose, aminoalkyl-methacrylate, triacetine and glycerine. Dentomycin (2% Minocycline gel) has received regulatory approval for the treatment of periodontitis in the European Union. The same product is available in Japan with the name Periocline. The concentration of minocycline in the periodontal pocket is about $1300\mu g/ml$, 1 hr after single topical application of 0.05~ml ointment (1mg of minocycline) and is reduced to $90\mu g/ml$ after 7 hrs. 34

Results have shown that the combination of ointment with scaling and root planing was significantly better than scaling and root planing alone in pockets $> 7 \text{mm}.^{34}$

4. Subgingival Chlorhexidine

The use of chlorhexidine as an antifungal and antibacterial agent has been well established. Chlorhexidine is being used in mouth rinses and is highly recommended in the hygiene phase of treatment as an adjunct to tooth brushing. The major application has been for the control of dental plaque and gingivitis. Its mechanism of action relates to reduction in pellicle formation, alteration of bacterial adherence to teeth and an alteration of bacterial cell walls causing lysis. Its antibacterial action is due to an increase of the cellular membrane permeability followed by the coagulation of intracellular cytoplasmic macromolecules. Because chlorhexidine is highly cationic, it exhibits high substantivity. The long term efficacy of chlorhexidine on the periodontal pocket flora is dependent on the duration of exposure. However, intracrevicular irrigation of the periodontal pocket with chlorhexidine has only a short lived effect on the pocket flora.⁶

Chlorhexidine is available in the form of mouthrinses, Gels, varnishes, and chip to be used as a local drug delivery agent for the treatment of periodontal diseases.

Periochip

2.5 mg Chlorhexidine Gluconate PerioChip, the controlled subgingival delivery of chlorhexidine gluconate, is a small, orangebrown, tombstone-shaped chip (4.0x 0.5x 0.35mm) in a biodegradable matrix of hydrolyzed gelatin and has been approved by FDA. Studies with PerioChip showed reduction in the numbers of the putative periodontopathic organisms *Porphyromonas gingivalis, Prevotella intermedia, Bacteroides forsythus,* and *Campylobacter rectus* after placement of the chip. No overgrowth of opportunistic organisms or other adverse changes in the oral microbial ecosystem were noted.

PerioChip releases chlorhexidine in vitro in a biphasic manner, initially releasing approximately 40% of the chlorhexidine within the first 24 hours, and then releasing the remaining chlorhexidine in an almost linear fashion for 7–10 days.

Several large clinical trials were completed which compared the efficacy of scaling/root planing and combined therapy employing Chlorhexidine chips. The differences between therapies were statistically significant, but may not be clinically relevant. Furthermore, the number of sites with probing depth reduction was greater with combined therapy. ^{36,37}

Periocol-CG

Periocol CG is prepared by incorporating 2.5mg chlorhexidine from a 20% chlorhexidine solution in collagen membrane. Size of the chip is $4x5\ mm$ and thickness is 0.25 - $0.32\ mm$ and $10\ mg$ wt.

Collagen is a natural protein, which is chemotactic for fibroblasts, enhances fibroblast attachment via its scaffold like fibrillar structure and stimulates platelet degranulation, thereby accelerating fibers and clot attachment. It has been shown to resorb after 30 days; however their coronal edge degrades within 10 days. 38

Chlo-Site

Chlo-Site is an agent containing 1.5% chlorhexidine of xanthan type (Ghimas Company, Italy). Xanthan gel is a saccharide polymer, which constitutes of a three-dimensional mesh mechanism, which is biocompatible with chlorhexidine.

The gel gets vanished from the pocket within l0-30 days of injection and effective concentration of chlorhexidine against microorganisms is established for at least 15 days in the region. Both chlorhexidine and gel matrix are mucoadhesive so that they stick inside the pockets and are not easily washed out by gingival fluid or saliva. It degrades spontaneously at the site of application, is well tolerated and is efficient in treatment of periodontal pockets & perimplantitis. 38

5. Subgingival Metronidazole

Among the antibiotics that have been considered for periodontal treatment, Metronidazole has often been chosen because of its selective efficacy against obligate anaerobes. It acts by inhibiting DNA synthesis. It is known to convert into a reactive reduced form and affects specifically anaerobic rods and spirochetes in subgingival microflora. After application of Elyzol 25% dental gel, Metronidazole concentrations of above 100 μ/ml were measurable in the periodontal pocket for at least 8 hours and concentrations above 1 μ/ml were found at 36 hours. 39

A topical medication ELYZOL contains an oil-based metronidazole 25% dental gel (glyceryl mono-oleate and sesame oil). It is applied in viscous consistency to the pocket, where it is liquidized by the body heat and then hardens again, forming crystals in contact with water. When metronidazole gel plus scaling and root planing were compared to root planing alone, the results have not been consistent. 39 One investigation suggested that there was a better result over a 9-month observation period when combined therapy was employed for probing depth reduction. 40

Future trends in local drug delivery

Eradication of microorganisms from the periodontal pocket is the most important step in treating periodontitis. The limitations of mouth rinsing and irrigation have prompted research for the development of alternative delivery systems. The requirements for treating periodontal disease include a means for targeting an anti-infective agent to infection sites and sustaining its localized concentration at effective levels for a sufficient time while concurrently evoking minimal or no side effects.⁴⁴ Various newer agents are being investigated in the field of local drug delivery to ensure maximum benefit.

Clarithromycin gel

A study has been conducted to investigate the adjunctive effects of subgingivally delivered $0.5\,\%$ clarithromycin as an adjunct to scaling and root planing for treating chronic periodontitis smoker subjects. It was observed that the adjunctive use of 0.5% clarithromycin as a controlled drug delivery system enhanced the clinical outcome. At the end of 6 months, the mean GI, PI, SBI, PPD, CAL for the clarithromycin group was significantly reduced. This product is still under investigation and yet to be patented.

2. Herbal products

Various herbal formulations like aloe vera, neem, tulsi, propolis, cocoa husk, pomegranate, cranberry etc. are being used widely these days. 46,47 These products have shown promising results with no side effects and are economical as well.

- 3. Okade and co-workers developed a new sub gingival release delivery system (PT-01) containing Ofloxacin for sub gingival therapy. The PT-01 was found to be effective in the reduction of supra gingival plaque, reduction in the plaque index, reduction in bleeding on probing. 48
- 4. Fibroblast growth factor would be a very efficacious introduction in local drug delivery. To regenerate periodontal tissues, a sandwich membrane composed of a collagen sponge scaffold and gelatin microspheres containing basic fibroblast growth factor (bFGF) in a controlled-release system was developed. This sandwich membrane induced successful regeneration of the periodontal tissues in a short period of time (4weeks).
- 5. Colloidal drug carriers include micelles, emulsions, liposomes and nanoparticles (nanospheres and nanocapsules). It is noteworthy

that only liposomes and nanoparticles have been largely exploited for drug delivery because the methods of preparation are generally simple and easy to scale-up. The aim of using colloidal carriers is generally, to increase the specificity towards cells or tissues, to improve the bioavailability of drugs by increasing their diffusion through biological membranes and/or to protect them against enzyme inactivation.⁵⁰

Nanoparticles, owing to their small size, penetrate regions that may be inaccessible to other delivery systems. These systems reduce the frequency of administration and further provide a uniform distribution of the active agent over an extended period of time. Three preliminary studies have been conducted to assess the efficacy of nanoparticles in periodontal drug delivery.

- a) Dung et al used Antisense oligonucleotide- loaded chitosantripolyphosphate (TPP) nanoparticles and showed the sustained release of oligonucleotides which is suitable for the local therapeutic application in periodontal diseases.³⁵
- b) Pinon et al conducted a preliminary in vivo study in dogs with induced periodontal defects using Triclosan-loaded polymeric (PLGA, PLA and cellulose acetate phthalate) nanoparticles and suggested that triclosan-loaded nanoparticles penetrate through the junctional epithelium.³⁵
- c) Moulari et. al, investigated the in vitro bactericidal activity of the Harungana madagascariensis leaf extract (HLE) on the oral bacterial strains largely implicated in dental caries and gingivitis infections. HLE-loaded PLGA nanoparticles were prepared using interfacial polymer deposition following the solvent diffusion method. Incorporation of the HLE into a colloidal carrier improved its antibacterial performance and diminution of the bactericidal concentration was observed. ³⁵

CONCLUSION

Current data suggest that local delivery of antimicrobials into a periodontal pocket can improve the periodontal health. However these drug systems do not provide a superior result when compared to scaling and root planing. Thus the benefits of using these systems as a monotherapy are questionable. In conjunction with scaling and root planing, the adjunctive use of local drug delivery may enhance the results in sites that don't respond to conventional therapy. A few localised persistent lesions in otherwise well controlled patients may offer the greatest potential for success with this treatment modality. Prudent administration of antimicrobial agents following judicious pharmacologic principles will preclude the abuse of chemotherapeutic agents and reduce the potential of developing or selecting drug resistant bacterial strains. Local drug delivery systems with controlled release properties have the potential to be used as a therapeutic component in the management of periodontal diseases. However, additional randomized, controlled studies are needed to help delineate the types of lesions, periodontal diseases, or specific situations where local delivery systems would be most

It can be concluded that the adjunctive use of local drug delivery may provide a defined but limited beneficial response. However the magnitude of change anticipated by combined therapy must be interpreted in light of the severity of the defects being treated. Therefore the clinician will need to make decisions based on the desired outcomes of the therapy.

REFERENCES

- Highfield J. Diagnosis and classification of periodontal disease Aust Dental J: 2009:54(s1); S11–S26.
- Haffajee AD, Socransky SS. Microbial etiological agents of destructive periodontal disease. Periodontol 2000:1994; 78-
- 3. Moore JWEC, Moore LVH. The bacteria of periodontal diseases, Periodontol 2000:1994: 66-77.
- Baehni PC: Supportive care of the periodontal patient. Curr Opin Periodontol; 1997: 151-157.
- 5. Suomi, JD, West JD, Chang JJ, McClendon BJ. The effect of controlled oral hygiene procedures on the progression of

- periodontal disease in adults: radiographic findings. J Periodontol 1971; 42: 562-564.
- Abdellaouia KS, Castionib NV, Gurnya R. Local delivery of antimicrobial agents for the treatment of periodontal diseases. Eur J Pharm Biopharm; 2000: 83-99.
- Adriens PA, Boever JA, Loesche WJ. Bacterial invasion in root cementum and radicular dentin of periodontally diseased teeth in humans. J Periodontol:1988; 59: 222-230.
- Gordon J, Walker C, Lamster I, West T, Socransky S, Seiger M, Fasciano R, Efficacy of Clindamycin Hydrochloride in refractory periodontitis-12-month results. J Periodontol 1985; 56: 75-80.
- Slots J. Selection of antimicrobials agents in periodontal therapy. J Periodont Res 2002; 37:389-398.
- Mombelli A, Van Winkelhoff AJ. The systemic use of antibiotics in periodontal therapy, in: N.P. Lang, T. Karring, J. Lindhe (Eds.), Proceedings of the Second European Workshop on Periodontology, Quintessence, London, 1997, pp. 38-77.
- 11. Vandekerckhove BNA, Quirynen M, Van Steenberghe D. The use of Tetracycline-containing controlled release fibers in the treatment of refractory periodontitis: J Periodontol 1997; 68:353-361.
- 12. Needleman IG. Controlled drug release in Periodontics: A review of new therapies. Br Dent | 1991; 170: 405-408.
- Noyan U, Yilmaz S, Kuru B, Kadir T, Acar O, Buget E. A clinical and microbiological evaluation of systemic and local metronidazole delivery in adult periodontitis patients. J Clin Periodontol 1997; 24: 158-165.
- 14. Goodson J. Antimicrobial strategies for treatment of periodontal diseases. Periodontol 2000: 1994; 5:142-168.
- Position paper: The role of controlled drug delivery for periodontitis. J Periodontol 2000; 71:125-140.
- Goodson JM, Offenbacher S, Farr DH, Hogan PE. Periodontal disease treatment by local drug delivery. J Periodontol 1985; 4: 265-272.
- Maurizio S, Tonetti. The topical use of antibiotics in periodontal pockets., in: N.P. Lang, T. Karring, J. Lindhe (Eds.), Proceedings of the Second European Workshop on Periodontology, Quintessence, London, 1997, pp. 109-132.
- Maheshwari M, Miglani G, Mali A, Paradkar A, Yamamura S, Kadam S. Development of Tetracycline-Serratiopeptidase-Containing Periodontal Gel: Formulation and Preliminary Clinical Study; AAPS Pharm Sci Tech 2006; 7 (3) Article 76:E162-171.
- 19. Schwach K, Loup PJ, Castioni N, Mombelli A, Baehni P, Barr J et al. Bioerodible Injectable Poly(ortho ester) for Tetracycline Controlled Delivery to Periodontal Pockets: Preliminary Trial in Humans; AAPS Pharm Sci 2002; 4 (4) article 20:1-7.
- Lowenguth RA, Chin I, Caton JG, Cobb CM, Drisko CL, Killoy WJ et al. Evaluation of periodontal treatment using controlled release tetracycline fibers. J Periodonol 1995; 66:700-707.
- Goodson JM, Hogan R Dunham S. Clinical responses following periodontal treatment by local drug delivery. J Periodontol 1985; 56:81-87
- 22. Sanz M, Serrano C, Garcia C, Echievarria C, O'Connor A. Clinical and microbiological efficacy of tetracycline fiber therapy in relapsing periodontal sites during supportive periodontal therapy. J Dent Res 1997; 76(Spec. Issue): 153 (Abstr. 1116).
- Drisko CL, Cobb CM, Killoy WJ, Michalowicz BS, Pihlstrom BL, Lowenguth RA et al.. Evaluation of periodontal treatments using controlled-release tetracycline fibers. Clinical response. J Periodontol 1995; 66:692-699.
- 24. Newman MG, Kornman KS, Doherty FM. A 6-month multicentre evaluation of adjunctive tetracycline fiber therapy used in conjunction with scaling and root planing in maintenance patients: Clinical results. J Periodontol 1994; 65:685-691.
- Kerry G. Tetracycline-loaded fibers as adjunctive treatment in periodontal disease, J Am Dent Assoc 1994; 125:1199-1203.
- Michalowicz BS, Pihlstrom BL, Drisko CL, Cobb CM, Killoy WJ, Caton JG et al. Evaluation of periodontal treatments using controlled-release tetracycline fibers: Maintenance response. J Periodontol 1995; 66:708-715.
- 27. Killoy WJ, Rapley JW, Drisko CL. Single and repeated tetracycline fiber treatment vs periodontal maintenance. J Dent Res 1996; 75(Spec. Issue):153 (Abstr. 1136).

- Preshow PM. Host response modulation in Periodontics. Periodontol 2000:2008; 48:92-110.
- Polson AM, Garrett S, Stoller NH, Bandt CL, Hanes PJ, Killoy WJ et al. Multi-center comparative evaluation of subgingivally delivered sanguinarine and doxycycline in the treatment of Periodontitis. II. Clinical results. J Periodontol 1997; 68:119-126.
- Borden LC, Walker CB, Stone C, Mankodi S., Godowski K.C., Southard H. Microbiota effects following sustained release subgingival delivery of doxycycline. J Dent Res 1997; 76(Spec. Issue): 153 (Abstr. 1114).
- 31. Garrett S, Adams D, Bandt C, Beiswanger B, Bogle BC, Caton J et al. Two multicenter clinical trials of subgingival doxycycline in the treatment of Periodontitis. J Dent Res 1997; 76(Spec. Issue):153 (Abstr. 1113).
- 32. Akula S, Chava V. Minocyclines in periodontal therapy; J Indian Soc Periodontol 2000; 3: 49-51.
- 33. Vandekerckhove BN, Quirynen M, V Steenberghe. The use of locally delivered minocycline in the treatment of chronic periodontitis- A review of the literature. J Clin Periodontol 1998; 25: 964-968.
- Van Steenebergh D, Bercy P, Kohl J. Subgingival Minocycline Hydrochloride ointment in moderate to severe chronic adult periodontitis a randomised double blind, vehicle controlled multicenter study. J Periodontol 1993; 64:637-644.
- 35. Pragati S, Ashok S, Kuldeep S. Recent advances in periodontal drug delivery systems. Int J Drug Del 2009; 1: 1-14.
- 36. Jeffcoat M, Bray KS, Cianico SG, Dentino AR, Fine DH, Gordon JM, et al. Adjunctive use of a subgingival controlled-release Chlorhexidine chip reduces probing depth and improves attachment level compared with scaling and root planing alone. J Periodontol 1998; 69:989-997.
- Soskolne WA, Heasman PA, Stabholz A, Smart GJ, Palmer M, Flashner M, et al. Sustained local delivery of Chlorhexidine in the treatment of Periodontitis: a multicenter study. J Periodontol 1997; 68:32-38.
- NandaKumar P.K. Local Drug Delivery-Periocol in Periodontics. Trends Biomater Artif Organs 2006; 19(2);74-80.

- 39. Chhina K, Bhatnagar R. Local drug delivery. Indian J Dent Sci 2012; 4: 166-69.
- Greenstein G, Polson A. The Role of Local Drug Delivery in the Management of Periodontal Diseases: A Comprehensive Review. J Periodontol 1998; 69:507-520.
- 41. Sapna N, Vandana KL. Evaluation of Hyaluronan gel (Gengigel®) as a topical applicant in the treatment of gingivitis. J Investig Clin Den 2011; 3: 162–170.
- 42. AL-Bayaty1 FH, Taiyeb-Ali TB, Abdulla MA, Mahmud ZB. Antibacterial effects of Oradex, Gengigel and Salviathymol-n mouthwash on dental biofilm bacteria. Afr J Microbiol Res 2011;5(6): 636-642.
- 43. Dudic VB, Lang NP, Mombelli A. Microbiological and clinical effects of an antiseptic dental varnish after mechanical periodontal therapy. J Clin Periodontol. 1999; 26:341-6.
- 44. Venkatesh A, Ramamurthy J. Local drug delivery systems in the treatment of periodontitis An Overview. Int J Pharm Pharm Sci, 2012; 4: 30-37.
- 45. Agarwal E, Pradeep AR, Bajaj P, Naik SB. Efficacy of local drug delivery of 0.5 % clarithromycin gel as an adjunct to non-surgical periodontal therapy in the treatment of chronic periodontitis subjects among smokers A randomized controlled clinical trial. DOI: 10.1902/jop.2012.110600.
- 46. Kukreja BJ, Dodwad V. Herbal Mouthwashes A Gift Of Nature: Int J Pharm Bio Sci 2012; 3: 46-52.
- Bansal S, Rastogi S, Bajpai M. Mechanical, Chemical and Herbal Aspects of Periodontitis: A Review. Int J Pharm Sci Res 2012; 3(5): 1260-1267.
- 48. Shah N. Local delivery of antimicrobial agents in periodontal therapy. I Indian Soc Periodontol 1999; 2:26-29.
- Nakahara T, Nakamura T, Kobayashi E, Inoue M, Shigeno K et.al. Novel Approach to Regeneration of Periodontal Tissues Based on *in Situ* Tissue Engineering: Effects of Controlled Release of Basic Fibroblast Growth Factor from a Sandwich Membrane. Tissue Engineering 2003, 9(1): 153-162.
- Prajapati J, Patel H, Agrawal YK. Targeted drug delivery for central nervous system: a review. Int J Pharm Pharm Sci 2012; 4(3): 32-38.