

## NOVEL ANTIBACTERIAL EFFECTS OF ALUMINA NANOPARTICLES ON *BACILLUS CEREUS* AND *BACILLUS SUBTILIS* IN COMPARISON WITH ANTIBIOTICS

<sup>a</sup>G. GEOPRINCY, <sup>b</sup>N. NAGENDHRA GANDHI, <sup>c</sup>\*S. RENGANATHAN

<sup>a,b,c</sup>Department of Chemical Engineering, Alagappa College of Technology, Anna University, Chennai 600025, India.  
Email: rengsah@rediffmail.com

Received: 2 Feb 2012, Revised and Accepted: 23 March 2012

### ABSTRACT

Alumina nanoparticles were synthesized through ball milling method, typically characterized and the antimicrobial activity was analysed against four major pathogenic strains such as *Bacillus cereus* MTCC 441, *Bacillus subtilis* MTCC 430, *Klebsiella pneumoniae* MTCC 109 and *Vibrio cholerae*. Upon characterization, the X-Ray Diffraction pattern defines the size distribution of the particle corresponding to the characteristic peaks obtained as 66.5 nm, 50.15 nm, 50.94 nm, 51.96 nm, 34.9 nm, 35.93 nm, 54.41 nm, 36.91 nm, 56.62 nm, 39.55 nm and 39.95 nm, respectively. The formation of spherical shaped nanoparticles was confirmed by Scanning Electron Microscopy. Also, the FTIR spectrum indicates the presence of significant functional groups such as O-H stretching, N-H amine group, a strong N-O nitro group, an alkene =C-H bending and a strong alkyl halide C-I stretch, respectively. In this investigation, alumina showed a characteristic inhibition zone of 12 mm diameter against *Bacillus subtilis* and 23 mm diameter against *Bacillus cereus* for 100 µg MIC (Minimum Inhibitory Concentration) of nanoparticles where as, no inhibitory effect was observed against *Klebsiella pneumoniae* and *Vibrio cholerae*. Also, the antimicrobial effects of alumina nanoparticles were compared with reference antibiotics like amoxicillin, chlormphenicol, erythromycin and rifamycin for MIC value of 100 µg.

**Keywords:** Alumina nanoparticles, Antibiotics, Antimicrobial activity, Pathogenic strains

### INTRODUCTION

Aluminium oxide is an electrical insulator posing relatively high thermal conductivity (30 Wm<sup>-1</sup>K<sup>-1</sup>). The most commonly available form was crystalline in nature, namely corundum or α-aluminium oxide. The hardness of the material makes it suitable for using as an abrasive. Aluminium oxide offers resistance to metallic aluminium, when exposed to weathering.

When metallic aluminum comes in contact with atmospheric oxygen, it becomes very reactive. A thin layer of alumina (4 nm thickness) forms in about 100 picoseconds on any exposed aluminium surface, which acts a protective covering for further oxidation.

It was well known that metal oxide nanoparticles exhibit antimicrobial properties. But, free radical scavenging property of the alumina particles may cause lowered antimicrobial activity and as a result even at higher concentration, the particles exhibit mild toxicity toward micro organisms<sup>1</sup>. The mechanism of inducing the oxidative stress makes the propensity for the nanoparticles to cross the cell barriers, enter the cells and interact with the sub cellular structures<sup>2</sup>.

The influence of four different preparation methods of C<sub>60</sub> fullerene (stirred C<sub>60</sub>, THF-C<sub>60</sub>, toluene-C<sub>60</sub>, and PVP-C<sub>60</sub>) on *Bacillus subtilis* exhibited relatively strong antibacterial activity ranging from 0.09 ± 0.01 mg/L- 0.7 ± 0.3 mg/L<sup>3</sup>. Smaller aggregates offered greater antibacterial activity, and the increase in toxicity was disproportionately higher than the associated increase in the surface area.

ROS-independent oxidative stress created by (nC<sub>60</sub>) fullerene favours changes in cell membrane potential accompanied by protein oxidation and thereby interfere with cellular respiration<sup>4</sup> which was different from the previously reported nanomaterial based antibacterial mechanisms that involves ROS generation (metal oxides)<sup>5</sup>.

Ions like Ag<sup>+</sup>, Cu<sup>2+</sup>, Fe<sup>3+</sup>, Zn<sup>2+</sup> and Nb<sup>5+</sup> dispersed in different matrices such as zeolites, apatite, phosphates, clays, titanium oxides and glasses have been employed in many applications<sup>6</sup>. Also, micro patterns fabricated with antimicrobial enzymes and alumina nanoparticles offering antimicrobial activity and abrasions resistant property were reported<sup>7</sup>. With respect to alumina nanoparticles, surface characteristics of the particles play a significant role in

phytotoxicity<sup>8</sup>. The volume of surface that comes in contact with microorganism is directly proportional to the amount of antimicrobial activity exhibited by the nanoparticle.

In this study we investigated, the antimicrobial properties of alumina nanoparticles synthesized by ball milling method using toluene as a solvent, rendered maximum inhibitory effects against four major pathogenic strains *Bacillus cereus*, *Bacillus subtilis*, *Vibrio cholerae* and *Klebsiella pneumoniae*.

### MATERIALS AND METHODS

#### Synthesis and characterization of alumina nanoparticles

##### Ball milling method

The zirconium balls were washed thoroughly using acetone and water. The washed balls were dried completely. Materials and balls were taken in the ratio of 1:10 (w/w). About 10 ml of Toluene (solvent) was added to the mixture. The mixture was set in a vial and ball milling was started fixing the speed at 200 rpm. Time was set for two hours and was continued for 8 hours and at every interval of 2 hours toluene was added. After 8 hours the balls and materials were separated. The material was subjected to drying in the oven at 60 °C overnight<sup>1</sup>. After drying, the material was placed in the furnace to sinter the material at 400°C.

The synthesized alumina nanoparticles were characterized by XRD analysis for predicting size and structure, SEM analysis to determine the shape and FTIR analysis for finding specific groups.

##### Antimicrobial Test

##### Test organisms used for the analysis

Clinical isolates of *Bacillus cereus*, *Bacillus subtilis*, *Klebsiella pneumoniae*, *Vibrio cholerae* were collected from KMCH, Coimbatore (India). Each test strain was inoculated in Mueller Hinton liquid medium (broth) and incubated in a temperature controlled shaker (120 rpm) at 30°C overnight. Antibiotics (amoxicillin, chlormphenicol, erythromycin and rifamycin) used for the analysis were purchased from Sigma Aldrich, Bangalore, India and Merck Limited, Mumbai, India.

##### Well Diffusion assay

16 petriplates containing Nutrient agar were prepared. 24 hours growing culture (*Bacillus subtilis* MTCC 441, *Bacillus cereus* MTCC

430, *Vibrio cholerae* and *Klebsiella pneumoniae* MTCC 109) were swabbed on it. Four plates for each organism were inoculated. Four wells (10mm diameter) were made by using cork borer. First well was loaded with 100 $\mu$ l of distilled water. Second well was loaded with 100 $\mu$ l of 1mg/ml concentration of alumina nanoparticles. Third well was loaded with 100 $\mu$ l of 1mg/ml of reference antibiotics (Amoxicillin, Chloramphenicol, Erythromycin and Rifamycin). The fourth well was loaded with 100 $\mu$ l final volume with equal amounts of alumina nanoparticle (0.5mg/ml) and antibiotic (0.5mg/ml) altogether. The plates were then incubated for 24 hours at 37 ° C. The zone of inhibition (in diameter) was measured.

## RESULTS

### X-Ray diffraction analysis

The scintillation detector present in the instrument moves through the required angle at specific counts and scans the sample with a start angle at 10 $^{\circ}$  and a stop angle at 70 $^{\circ}$ . The output was obtained in the form of a graph with 2 $\theta$  on x- axis and intensity on y-axis. The

obtained graph containing different peaks corresponding to different planes of the crystal was compared with the standard data in JCPDS tool. From the result obtained, the average size of the nanoparticle was calculated using Scherer's formula,

$$D = 0.9\lambda / \beta \cos\theta$$

Where,

$\lambda$  is wavelength of copper K $\alpha$  line (1.5406 Å)  $\theta$  is diffraction angle,  $\beta$  is full width at half maximum of peak (FWHM), and D is the average particle size. Depending upon the characteristic peaks obtained in Fig.1, the X-RD pattern defines the distribution of the size of the particle as 66.5 nm, 50.15 nm, 50.94 nm, 51.96 nm, 34.9 nm, 35.93 nm, 54.41 nm, 36.91 nm, 56.62 nm, 39.55 nm and 39.95 nm, respectively.

### Scanning Electron Microscopy Analysis

The scanning electron microscopy results clearly indicate the formation of spherical alumina nanoparticles with its size ranging between 100 nm to 200 nm is shown in the Fig.2.

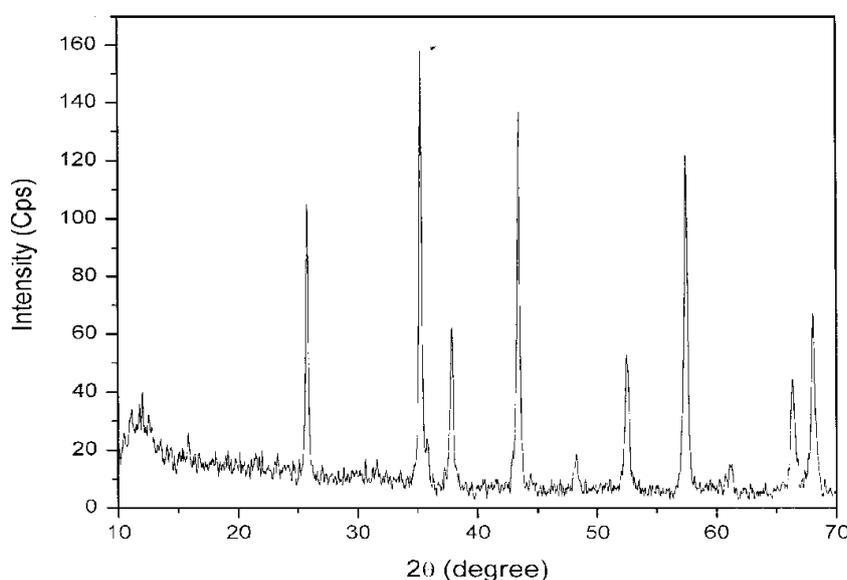


Fig.1: X-Ray Diffraction (X-RD)analysis pattern for alumina nanoparticles

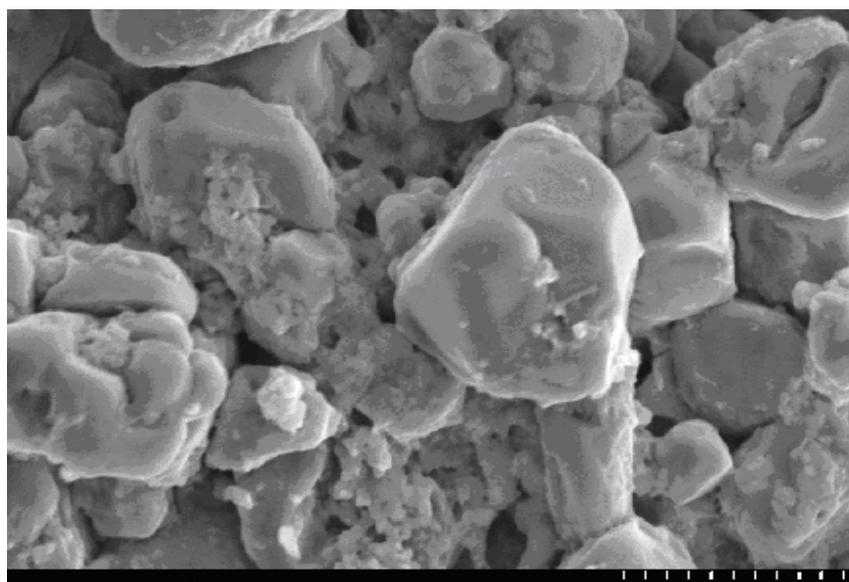
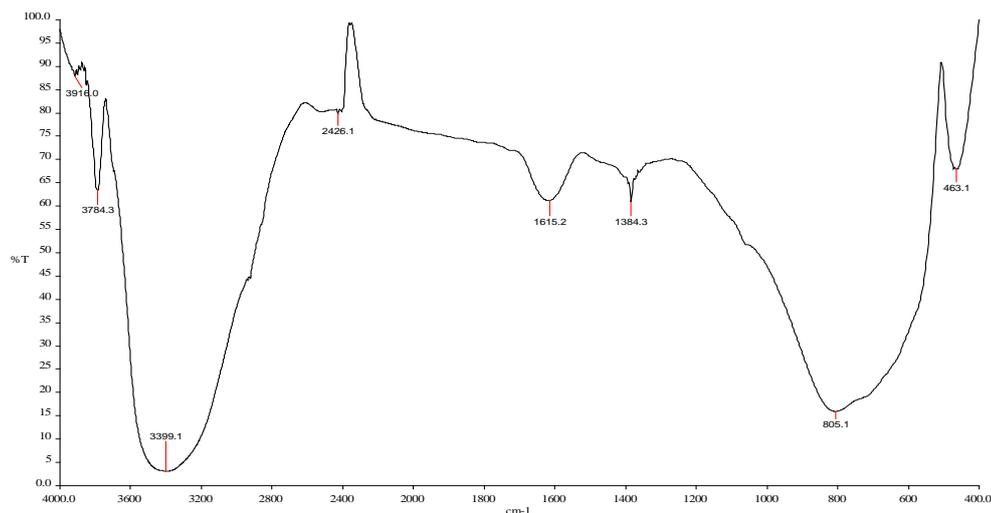


Fig. 2: Scanning Electron Microscopy (SEM) pattern depicting spherical shaped alumina nanoparticles



**Fig. 3: Fourier Transform Infra-red Spectroscopy (FTIR) analysis of alumina nanoparticles showing the presence of characteristic functional groups**

#### Fourier Transform Infra Red Spectroscopy Analysis

The FTIR spectrum of  $Al_2O_3$  nanoparticles is shown in Fig.3. The higher energy region peaks at  $3784\text{ cm}^{-1}$  and  $2426\text{ cm}^{-1}$  are assigned to O-H stretching of  $Al_2O_3$  nanoparticles. Also a stretch of N-H amine group was observed at  $3399\text{ cm}^{-1}$  and  $1615\text{ cm}^{-1}$  respectively. A strong stretch of N-O nitro group was observed at  $1384\text{ cm}^{-1}$ . An alkene =C-H bending was observed at  $805\text{ cm}^{-1}$ . Also a strong alkyl halide C-I stretch was observed at  $463\text{ cm}^{-1}$ . Upon verification with the present database, the spectrum matches with that of  $Al_2O_3$ .

#### Antimicrobial Activity

The antibacterial activity of alumina nanoparticles were tested against four major pathogenic strains *Bacillus cereus* MTCC 441, *Bacillus subtilis* MTCC 430, *Klebsiella pneumoniae* MTCC 109 and *Vibrio cholerae* and the zone of inhibition produced was illustrated in the Table.1, given below. Also the antimicrobial activities of nanoparticles were compared with the antimicrobial activity of antibiotics for the minimum inhibitory concentration value of  $100\mu\text{g}$ . The combined effects of nanoparticles and antibiotics were also analysed by well diffusion method.

Alumina showed a characteristic inhibition zone of 23 mm diameter against *Bacillus cereus* (Fig.4) and 12 mm diameter against *Bacillus subtilis* (Fig.5) for  $100\mu\text{g}$  MIC (Minimum Inhibitory Concentration) of nanoparticles. However, Alumina nanoparticles showed a mild growth-inhibitory effect on *E. coli*, only at very high concentration of  $1000\mu\text{g}$  [1]. This focuses the sensitivity of the nanoparticles towards

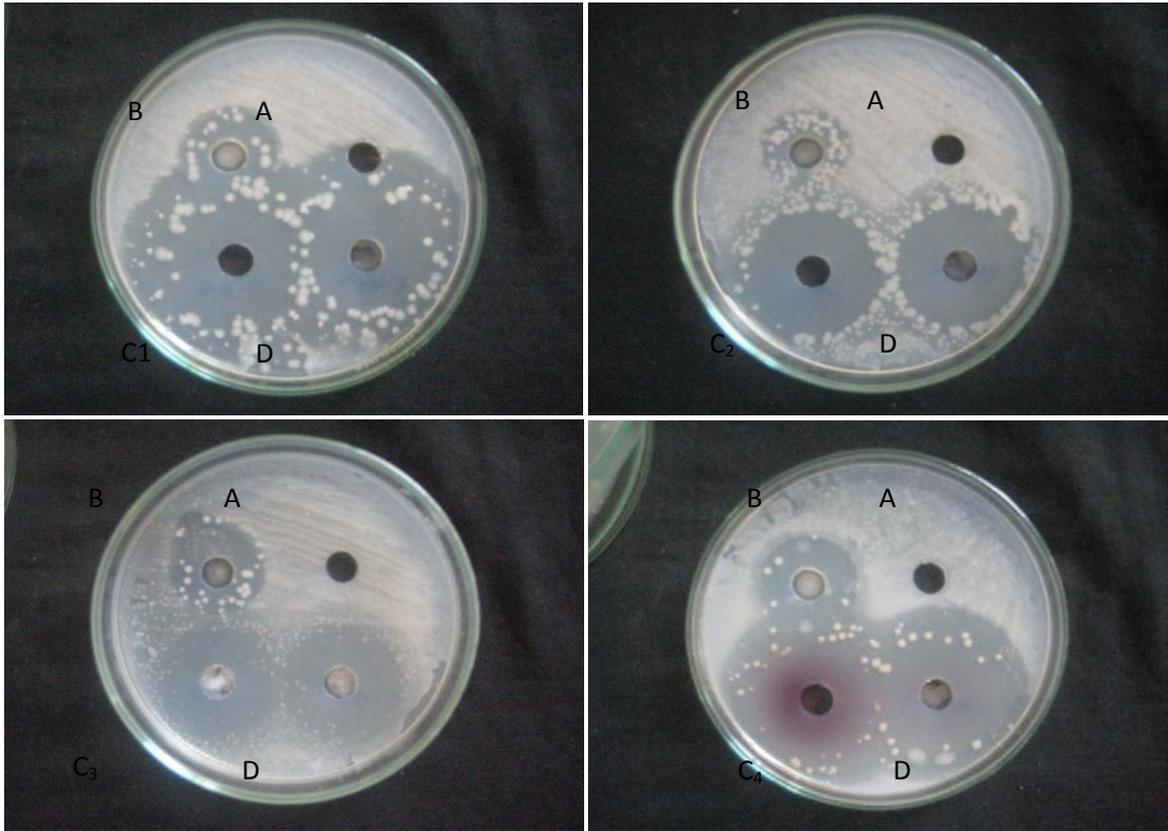
different organisms and strains. Though the alumina nanoparticles exhibited a pronounced antibacterial effect over *Bacillus subtilis* and *Bacillus cereus*, no inhibitory effect was observed against *Klebsiella pneumoniae* and *Vibrio cholerae*. It was inferred that both nanoparticles and bulk counterparts (metal particle more than nanosize) were seems to be toxic in inhibiting the growth of microbes, especially as stated in the case of nematodes. It was noticed from the literature that the toxicity of the nanoparticle and the oxide solubility influences the toxicity of alumina nanoparticles<sup>9</sup>.

Also, the antimicrobial effects of alumina nanoparticles were compared with reference antibiotics like amoxicillin, chloramphenicol, erythromycin and rifamycin for MIC value of  $100\mu\text{g}$ . Against *Bacillus subtilis*, all the four broad spectrum antibiotics showed typical zone of inhibition whose diameter was double the time when compared with alumina nanoparticles (Fig.5). With respect to *Bacillus cereus*, antibiotics expressed sensitivity nearly equal to that of alumina nanoparticles. In spite of which no effect was observed for alumina nanoparticles against *Klebsiella pneumoniae* and *Vibrio Cholerae*.

No characteristic enhancement in inhibitory zone area was observed for the assay, when alumina nanoparticles and antibiotics are given together. Similar type of analysis was performed with silver nanoparticles combined with and without antibiotics. This analysis inferred a characteristic increase in the fold area over the inhibitory zone when silver nanoparticles and antibiotics are given together against test organisms like *Staphylococcus aureus* and *Escherichia coli*<sup>10</sup>.

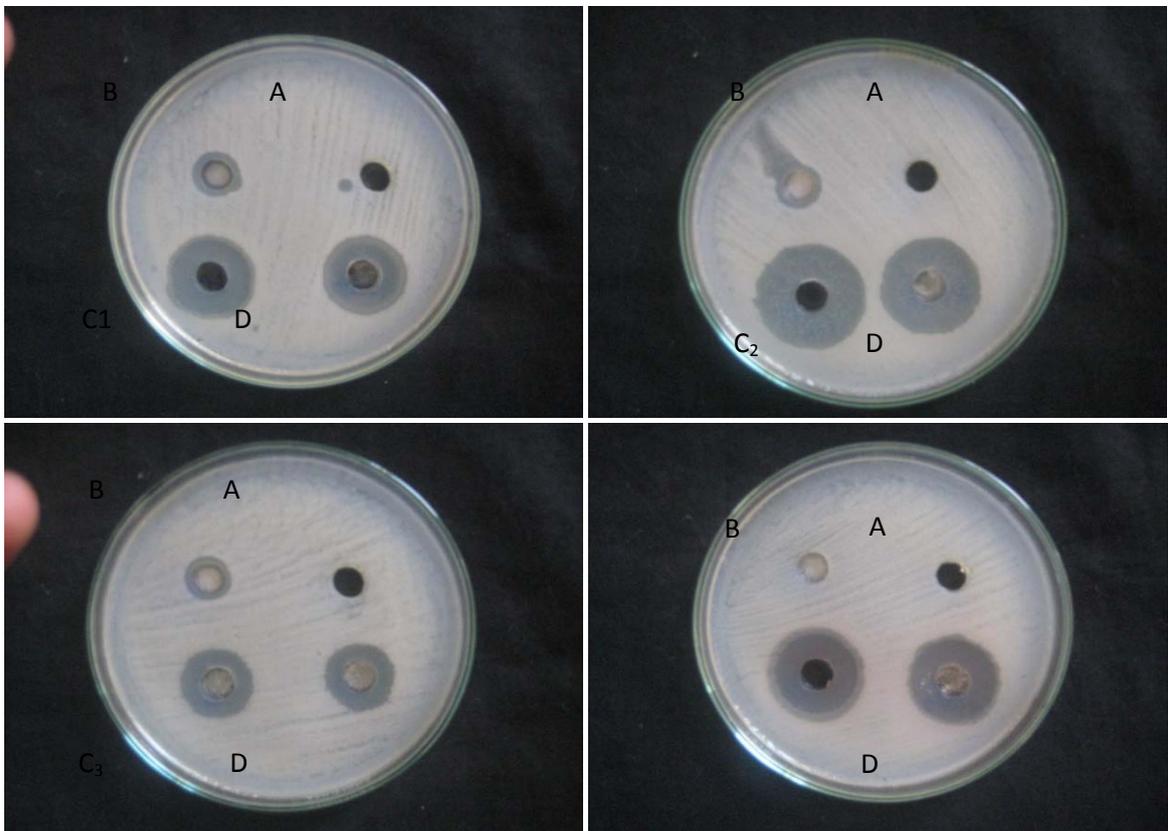
**Table 1: Zone of inhibition produced by alumina nanoparticles, reference antibiotics and nanoparticle with antibiotics**

Organisms	Antibiotics used	Control $\pm$ 2mm (diameter)	Alumina NP ( $100\mu\text{g}$ ) $\pm$ 2 mm (diameter)	Antibiotics ( $100\mu\text{g}$ ) $\pm$ 2mm (diameter)	Alumina NP + Antibiotics ( $100\mu\text{g}$ ) $\pm$ 2mm (diameter)
<i>Bacillus subtilis</i>	Amoxicillin	-	12	21	22
	Chloramphenicol	-	12	26	26
	Erythromycin	-	11	18	19
	Rifamycin	-	-	21	23
<i>Bacillus cereus</i>	Amoxicillin	-	24	38	40
	Chloramphenicol	-	20	33	33
	Erythromycin	-	23	35	35
	Rifamycin	-	23	40	41
<i>Vibrio cholerae</i>	Amoxicillin	-	-	28	28
	Chloramphenicol	-	-	26	27
	Erythromycin	-	-	-	-
	Rifamycin	-	-	20	20
<i>Klebsiella pneumoniae</i>	Amoxicillin	-	-	18	18
	Chloramphenicol	-	-	22	24
	Erythromycin	-	-	13	13
	Rifamycin	-	-	23	22



**Fig. 4: zone of inhibition formed against *Bacillus cereus***

A - Control; B - Nanoparticle; C<sub>1</sub> - Amoxicillin; C<sub>2</sub> - Chloramphenicol; C<sub>3</sub> - Erythromycin; C<sub>4</sub> - Rifamycin; D - Nanoparticles + Antibiotics



**Fig. 5: zone of inhibition formed against *Bacillus subtilis***

A - Control; B - Nanoparticle; C<sub>1</sub> - Amoxicillin; C<sub>2</sub> - Chloramphenicol; C<sub>3</sub> - Erythromycin; C<sub>4</sub> - Rifamycin; D - Nanoparticles + Antibiotics

## DISCUSSION

From the above experimental analysis, it was concluded that, the alumina nanoparticles synthesized through ball milling method conferred uniform size distribution ranging between 50 nm to 60 nm, which was evident from the X-RD results. Besides, the spherical shaped alumina nanoparticles analysed through scanning electron microscopy (SEM) inculcates the significance of the surface morphology of the nanoparticle participating in the cellular interaction i.e, the number of active sites coming in contact with the cells which render cytotoxic effect<sup>8</sup>. The presence of high energy bands and stretches strengthening the formation of nanoparticles was confirmed by FTIR analysis. With respect to the context of antimicrobial activity, alumina nanoparticles produced 13 mm and 23mm diameter of inhibition zone (Table 1) against *Bacillus cereus* and *Bacillus subtilis*, respectively. This infers the sensitivity of the microbial species towards the nanoparticles for 100µg MIC (Minimum Inhibitory Concentration). But, no inhibitory effect was found with *Vibrio cholerae* and *Klebsiella pneumoniae*.

Not only the antimicrobial property of alumina nanoparticles were analysed, but also the inhibitory effects of reference antibiotics were analysed. The formation inhibition zone infers the antibacterial properties of the particles used. Similar types of activity were reported on antibacterial effects of *Withania somnifera L.*, *Euphorbia hirta L.* and *Trianthema decandra L.*<sup>11, 12</sup>. Antibiotics offered typical antimicrobial effect for all the four pathogens (Table 1). Further, any enhancement in zone area was tested for the combination of nanoparticles and antibiotics. No enhancement in inhibition zone was observed.

## CONCLUSION

Based on these results obtained, three major points were predicted as follows,

1. Nanoparticles, metals and metal oxides are not showing the same sensitivity to all the microbial species and strains (Table 1).
2. The solubility of metal oxides influences the toxicity of nanoparticles. The concentration of the particle plays another significant role in the determination of antimicrobial activity.
3. The volume (or) the surface area of the nanoparticles that comes in contact with cells is directly proportional to the amount of antimicrobial activity offered by the particle.

## REFERENCES

1. Sadiq, Mohammed I, Chowdhury, Basudev, Chandrasekaran, Natarajan et al, Antimicrobial Sensitivity of Escherichia coli to Alumina Nanoparticles. *Nanomedicine* 2009; 5: 282 – 286.
2. Gunter Oberdorster, Vicki Stone, Ken Donaldson, Toxicology of Nanoparticles: A Historical Perspective. *Nanotoxicology* 2007; 1: 2-25.
3. Vijendra kumar mishra and Ashok kumar, Impact of the Metal Nanoparticles on the Plant Growth Promoting Rhizobacteria. *Dig J Nanomater Bios* 2009 ; 3 : 587-592.
4. Delina Y. Lyon, Pedro J.J Alvarez, Fullerene Water Suspension (Nc) exerts Antibacterial effects through ROS – independent Protein Oxidation. *Environ.Sci.Technol* 2008; 21: 8127-8132.
5. Roberta Brayner, Roselyn Ferrari illiou, Nicholas brivois, Shakib djediat, Djediat S, Marc F. Benedetti, Toxicological Impact Studies Based On Escherichia Coli Bacteria in ultrafine ZnO Nanoparticles Colloidal Medium. *Nano Lett* 2006; 6: 866–870
6. Jia husheng, Hou wensheng, Wei Liqiao, Xu bingshe, Liu xuguang, Antibacterial Nanomedicine: A Future Perspective. *Dent Mater* 2008; 24: 244–249.
7. Laura Treccani, Marcus Maiwald, Volker Zöllmer, Matthias Busse, Georg Grathwohl, Kurosch Rezwan, Antibacterial and Abrasion Resistant Alumina Micropatterns. *Adv Eng Mater* 2009; 7: B61–B66.
8. Ling yang, Daniel j. watts, Particle Surface Characteristics Play an Important Role in the Phytotoxicity of Alumina Nanoparticles. *Toxicology letters* 2005; 158: 122-132.
9. Huanhua Wang, Robert L. Wick, Baoshan Xing, *Environ Pollut* 2005; 157: 1171 -1177.
10. Zaheer Ahmed, Yaping Wang, Qiaoling cheng, M.Imran, Lactobacillus acidophilus Bacteriocin, from the Production to their application: An Overview. *Afr Js Biotechnol* 2010; 9: 2843 - 2850.
11. Geeta singh and Padma kumar, Antibacterial potential of alkaloids of *Withania somnifera L.* and *Euphorbia hirta L.* *Int J Pharm Pharm Sci* 2012; 4:79 – 81.
12. Sukantha.T.A, Shubashini K, Sripathi, Ravindran.N.T, Balashanmugam P, Antioxidant and antibacterial activities of *Trianthema decandra L.* *Int J Pharm Pharm Sci* 2012; 4:410 - 413.