

PRIMARY CLARITHROMYCIN RESISTENCE OF HELICOBACTER PYLORI BY FLUORESCENT IN SITU HYBRIDIZATION IN ALEPPO-SYRIA

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ABSTRACT

Helicobacter pylori is a human pathogen responsible for serious diseases including peptic ulcer disease and gastric cancer. The triple therapy included clarithromycin is the recommended first choice treatment, but primary clarithromycin resistance markedly reduces *H.pylori* eradication rate. The aim of this study to assess the prevalence of primary clarithromycin resistance in Aleppo of Syria by fluorescent in situ hybridization, which is a simple, rapid assay and permits detection of *H. pylori* and clarithromycin resistance simultaneously.

From September 2011 to May 2012, 130 *H.pylori* strains were isolated, among these, 16 strains were resistant to clarithromycin (12.3%). Of these, one were found to contain a mixed population of both sensitive and resistant *H.pylori*. Clarithromycin resistance rate was higher in normal endoscopic findings patients as compared to other findings patients (30.4% vs. 11.5%, $p=0.04$).

Based on the resistance rate we observed, it could be suggested that an empirical choice of a clarithromycin containing regimen based therapy could still remain the first-line therapeutic approach in clinical practice in Aleppo.

Keywords: *Helicobacter pylori*, Clarithromycin, Fluorescent in situ hybridization.

INTRODUCTION

Helicobacter pylori associated with many digestive disorders such as gastritis¹⁻², peptic ulcer disease³⁻⁴, gastric mucosa-associated lymphoid tissue (MALT) lymphoma⁵⁻⁶, and gastric adenocarcinoma⁷⁻⁹. The role of *H. pylori* in extra intestinal diseases has also been suggested.¹⁰⁻¹¹⁻¹²⁻¹³

Has proved that eradication of *H.pylori* enhances healing of peptic ulcers¹⁴⁻¹⁵ reduces its recurrence¹⁶⁻¹⁷, induces regression of preneoplastic lesions¹⁸ and decreases gastric cancer risk¹⁹⁻²⁰. According to the last Maastricht III consensus report, PPI- clarithromycin-amoxicillin or metronidazole is the recommended first choice treatment²¹. However, these treatment may fail for several reasons, particularly the growing resistance of *H.pylori* to clarithromycin.²²⁻²³ A major difference in eradication rates was found: 87.8% when strains were clarithromycin susceptible against 18.3% when strains were clarithromycin resistant.²⁴

It would be impractical and costly to test clarithromycin susceptibility for each individual seeking anti *H.pylori* treatment, so it is important for clinicians to know the local resistance rate. This is which is confirmed by Maastricht III Consensus report as recommended that the threshold of clarithromycin resistance at which this antibiotic should not be used, or clarithromycin susceptibility testing performed, is 15–20%.²¹

The most frequent point mutations responsible for clarithromycin resistance are A2142G, A2143G, and A2142C within the *H. pylori*

23S rRNA gene²⁵⁻²⁶, so that, molecular methods can be used for their identification, such as fluorescent in situ hybridization (FISH) which is a simple and rapid assay. Moreover it permits detection of *H. pylori* and clarithromycin resistance simultaneously.²⁷⁻²⁸

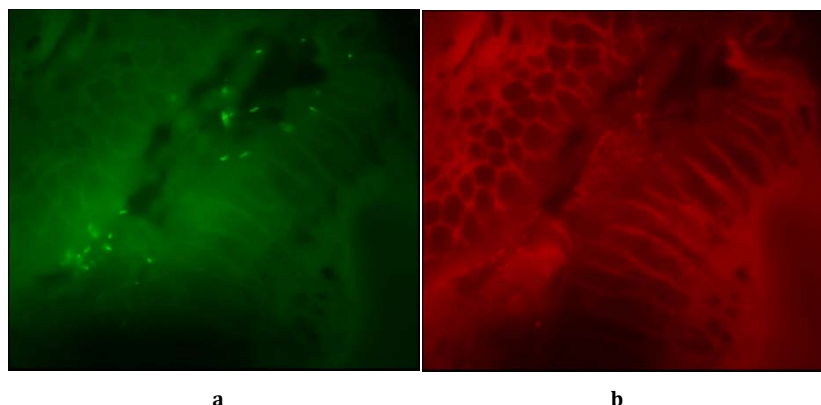
The aim of this study is to determine the primary clarithromycin resistance rate of *H.pylori* isolated from adults patients in the Aleppo city of Syria, by fluorescent in situ hybridization.

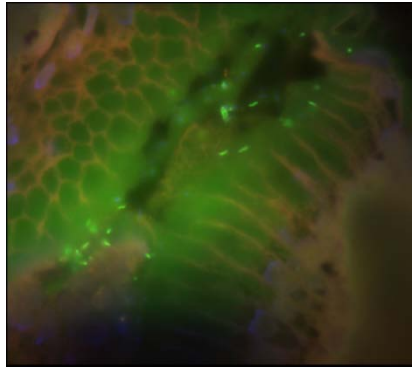
MATERIALS AND METHODS

From September 2011 to May 2012, gastric biopsy specimens were collected of adults patients (≥ 18 years old) who underwent upper gastrointestinal endoscopy for different indications at gastroenterology department in Aleppo University Hospital. All patients had never been treated for *H.pylori* infection, and the following data were also registered: age, gender, smoking habit, and endoscopic findings.

Fluorescent in situ hybridization

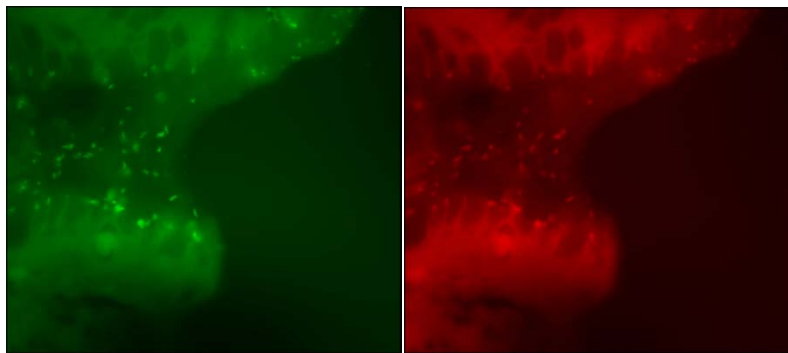
The biopsies were immediately fixed in formalin, embedded in paraffin, sectioned (4 μ m slice thickness) and dehydrated in an xylol and 96% ethanol. The deparaffinised, air-dried slides were incubated in the microwave oven at 400W for 10 min, this procedure greatly increases signal/noise ratio and renders largely insensitive to sample/fixation variations.





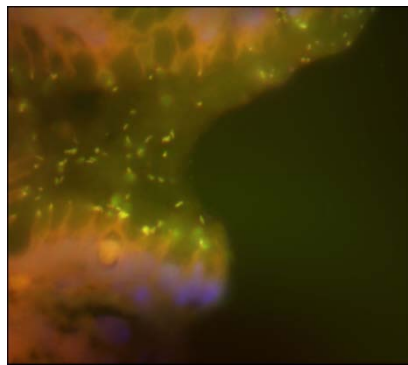
c

Fig. 1: It shows the detection of clarithromycin-sensitive *H. pylori* in the same section (a) green signal due to hybridization with *H. pylori*-specific, fluorescein-labelled probe (b) no red signal (c) green signal under dual band filter



a

b



c

Fig. 2: It shows the detection of clarithromycin-resistant *H. pylori* in the same section (a) green signal due to hybridization with *H. pylori*-specific, fluorescein-labelled probe (b) red signal due to hybridization with a Cy3-labelled mixture of probes specific for the most frequent mutations associated with clarithromycin resistance (c) yellow signal under dual band filter. Note the yellow appearance of resistant bacteria that results from an additive mixture of red and green fluorescence.

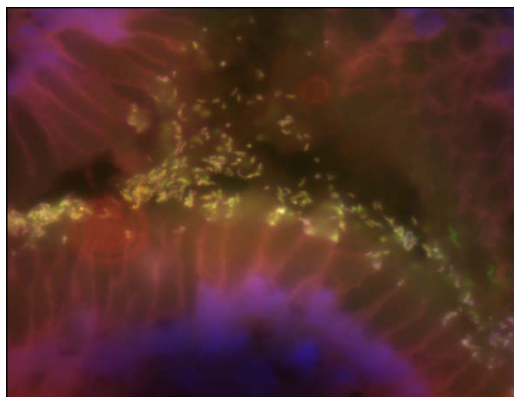


Fig. 3: It shows the detection of mixed population of both sensitive (green signal) and resistant (yellow signal) *H.pylori*

The hybridization were done at research laboratory in Faculty of medicine of Aleppo University using the commercially available test Bactfish™ *Helicobacter pylori* Combi kit. The probe for *H.pylori* identification was labeled with fluorescein (green signal) and the probes which detect tho most prevalent point mutations for clarithromycin resistance were labeled with fluorochrome Cy3 (red signal). Following hybridization for 90 min at 46°C, the sections were washed at 46°C for 2*15 min in wash buffer. The air dried sections were counterstained with 4',6'-diamidino-2-phenylindole (DAPI) which binds to the DNA of the bacteria. Slides were inspected with Zeiss Axioskope 40 microscope equipped with a standard fluorescence filters set. (figure1,2,3)

Statistical analysis

Logistic regression analysis was performed in order to identify possible risk factors (age, gender, endoscopic findings, smoking habit) for clarithromycin resistance. The odds ratio (OR) and 95% confidence intervals (CI) were calculated. P value of < 0.05 indicated significant differences. Statistical analysis was performed using a specific software (SPSS 13.0 for windows)

RESULTS

During the study period 130 *H.pylori* strains were isolated. The patients distributed according to the clinical and personal specifications as the following (table 1)

Table 1: Shows the clinical and personal specifications of the patients

	Number of patients
total	130
Age : 18-39	76
40-59	46
≥60	8
Sex : male	66
female	64
Smoking habit : smoking	43
no smoking	87
Endoscopic findings : gastritis	45
peptic ulcer	36
normal	23
others	26

The clarithromycin resistant strains were detected in 12.3% of patients (n=16). Of these, one were found to contain a mixed population of both sensitive and resistant *H.pylori*

Data for the analysis of risk factors associated with clarithromycin resistance are shown in table 2.

Table 2: Shows the association of risk factors with clarithromycin resistance rate

Risk factor	Clarithromycin resistance rate	OR (95% CI)	P value
Age : 18-39	14.4% (11/76)	1.2(0.14-10.76)	0.6
40-59	8.7% (4/46)	0.67(0.65-6.87)	
≥60	12.5% (1/8)	1.0 reference	
Sex : male	7.8% (5/66)	1.0 reference	0.1
female	17.2% (11/64)	2.5(0.83-7.76)	
Smoking habit :smoking	9.3% (4/43)	0.7(0.21-2.37)	0.5
no smoking	13.8% (12/87)	1.0 reference	
Endoscopic findings : gastritis	2.2% (1/45)	0.18(0.02-1.81)	0.04
peptic ulcer	13.9% (5/36)	1.24(0.27-5.7)	
normal	30.4% (7/23)	3.35(0.75-5.7)	
others	11.5% (3/26)	1.0 reference	

A distinctly higher prevalence of clarithromycin resistance was observed in normal endoscopic findings patients (30.4%) as compared to other findings patients (11.5%), the difference being statistically significant (OR: 3.35; 95% CI: 0.75 -5.7; P=0.04). Similarly clarithromycin resistance was observed in female (17.2%) was twice as much compared to male patients (7.8%), although the difference failed to reach a statistical significance (OR: 2.5; 95% CI: 0.83 -7.76; P=0.1).

The statistical analysis excluded any significant association between the clarithromycin resistance and both the age and smoking habit of patients

DISCUSSION

The method approved by the Clinical and Laboratory Standards Institute (CLSI) to test for clarithromycin susceptibility of *H.pylori* is agar dilution²⁹. Due to the complexities of this method, many studies proved an alternative reliable method to detect clarithromycin resistant *H.pylori* is fluorescent in situ hybridization (FISH)³⁰⁻³¹⁻³². The advantage of FISH method is the rapid detection of *H.pylori* and its susceptibility to clarithromycin. In addition, this technique is able to detect mixed populations of clarithromycin-sensitive and resistant organisms which is may explain some of clarithromycin based treatment failures that occur in persons infected with clarithromycin-sensitive isolates as determined by culture and agar dilution²⁴.

The primary clarithromycin resistance rate in the present study is 12.3%, which is similar to that was found in Tunisia (14.6%)³³, and Iran (14.3%)³⁴. This prevalence is higher than that was found in Brazil (8%)³⁵, and Malaysia (0%)³⁶, but is lower than that was detected in France (26%)³⁷, and Turkey (41.9%)³⁸. This difference in clarithromycin resistance rates between countries might be due to the different prescription and administration of this antibiotic. Therefore, the selection of a regimen among those recommended for *H.pylori* eradication, must consider the local data of antimicrobial resistance.

Many studies which have assessed the correlation of clarithromycin resistance of *H.pylori* with risk factors have been controversial, for example, Wueppenhorst N³⁹ didn't find correlation of clarithromycin resistance with both the sex and disease status of patients , this findings contrast with the results were reported by De Francesco V⁴⁰ who disclosed a higher clarithromycin resistance rate for strains isolated from female and non ulcer dyspepsia patients, but failed identify as an independent risk factor for primary clarithromycin resistance. Whereas, Meyer JM⁴¹ found that clarithromycin resistance was significantly associated with female sex and older age. Anyway, both last studies failed to provide logical interpretation of high clarithromycin resistance rate in female.

The recent study found that the clarithromycin resistance rate was higher in normal endoscopic findings patients (30.4%) as compared to other findings patients (11.5%). Such a phenomenon could play a role in the efficacy of standard eradication therapy observed according to gastroduodenal pathology, the cure rate being generally lower in non-ulcer dyspepsia patients (i.e., dyspepsia patients with normal endoscopic findings)⁴²⁻⁴³

In conclusion, Based on the resistance rate we observed, it could be suggested that an empirical choice of a clarithromycin containing regimen based therapy could still remain the first-line therapeutic approach in clinical practice in Aleppo, especially in non normal endoscopic findings patients.

REFERENCES

- Gao L, Weck MN, Michel A, Pawlita M, Brenner H. Association between chronic atrophic gastritis and serum antibodies to 15 *Helicobacter pylori* proteins measured by multiplex serology. *Cancer Res* 2009; 69(7), 2973-80.
- Zhou L, Sung JJ, Lin S, Jin Z, Ding S, Huang X, et al. A five-year follow-up study on the pathological changes of gastric mucosa after *H. pylori* eradication. *Chin Med J (Engl)* 2003; 116(1), 11-4.
- Kusters JG, van Vliet AH, Kuipers EJ. Pathogenesis of *Helicobacter pylori* infection. *Clin Microbiol Rev* 2006; 19(3), 449-90.
- Arkkila PE, Seppälä K, Kosunen TU, Sipponen P, Mäkinen J, Rautelin H, et al. *Helicobacter pylori* eradication as the sole treatment for gastric and duodenal ulcers. *Eur J Gastroenterol Hepatol* 2005; 17(1), 93-101.
- Ota H, Asano N. Crucial roles of *Helicobacter pylori* infection in the pathogenesis of gastric cancer and gastric mucosa-associated lymphoid tissue (MALT) lymphoma. *Rinsho Byori* 2009; 57(9), 861-9.
- Andriani A, Miedico A, Tedeschi L, Patti C, Di Raimondo F, Leone M, et al. Management and long-term follow-up of early stage *H. pylori*-associated gastric MALT-lymphoma in clinical practice: an Italian, multicentre study. *Dig Liver Dis*. 2009 Jul;41(7):467-73. Epub 2008 Oct 21.
- Fock KM, Ang TL. Epidemiology of *Helicobacter pylori* infection and gastric cancer in Asia. *J Gastroenterol Hepatol* 2010 ; 25(3),479-86.
- Herrera V, Parsonnet J. *Helicobacter pylori* and gastric adenocarcinoma. *Clin Microbiol Infect* 2009 ; 15(11),971-6.
- Wang C, Yuan Y, Hunt RH. The association between *Helicobacter pylori* infection and early gastric cancer: a meta-analysis. *Am J Gastroenterol* 2007 ; 102(8),1789-98.
- El-Mashad N, El-Emshtay WM, Arfat MS, Koura BA, Metwally SS. Relation of Cag-A-positive *Helicobacter pylori* strain and some inflammatory markers in patients with ischemic heart diseases. *Egypt J Immunol* 2009;16(1):39-47.
- Arnold DM, Bernotas A, Nazi I, Stasi R, Kuwana M, Liu Y, et al. Platelet count response to *H. pylori* treatment in patients with immune thrombocytopenic purpura with and without *H. pylori* infection: a systematic review. *Haematologica*. 2009 Jun;94(6):850-6
- Huang X, Qu X, Yan W, Huang Y, Cai M, Hu B, et al. Iron deficiency anaemia can be improved after eradication of *Helicobacter pylori*. *Postgrad Med J*. 2010 May;86(1015):272-8.
- Balach O. *Microbiology*. 2 nd ed. Aleppo: Aleppo University Publications; 2009.p.343-53.
- Di Mario F, Battaglia F, Dal Bò N, Leandro G, Benedetti E, Bottona E, et al. Cure of *Helicobacter pylori*-positive active duodenal ulcer patients: a double-blind, multicentre, 12-month study comparing a two-week dual vs a one-week triple therapy. *GISU (Interdisciplinary Group for Ulcer Study)*. *Dig Liver Dis*. 2000 Mar;32(2):108-15.
- Arkkila PE, Seppälä K, Kosunen TU, Haapiainen R, Kivilaakso E, Sipponen P, et al. Eradication of *Helicobacter pylori* improves the healing rate and reduces the relapse rate of nonbleeding ulcers in patients with bleeding peptic ulcer. *Am J Gastroenterol*. 2003 Oct;98(10):2149-56.
- Ford AC, Delaney BC, Forman D, Moayyedi P. Eradication therapy for peptic ulcer disease in *Helicobacter pylori* positive patients. *Cochrane Database Syst Rev*. 2006 Apr 19;(2):CD003840.
- Arkkila PE, Seppälä K, Kosunen TU, Sipponen P, Mäkinen J, Rautelin H, et al. *Helicobacter pylori* eradication as the sole treatment for gastric and duodenal ulcers. *Eur J Gastroenterol Hepatol*. 2005 Jan;17(1):93-101.
- Eusebi LH, Ceroni L, Laterza L, Kurelac I, Fuccio L. The association between *Helicobacter pylori* and gastric cancer: the role of infection's eradication. *Recent advances*. *Recenti Prog Med*. 2010 Feb;101(2):66-9. Review. Italian.
- Fuccio L, Zagari RM, Eusebi LH, Laterza L, Cennamo V, Ceroni L, et al. Meta-analysis: can *Helicobacter pylori* eradication treatment reduce the risk for gastric cancer? *Ann Intern Med*. 2009 Jul 21;151(2):121-8.
- Cheung TK, Wong BC. Treatment of *Helicobacter pylori* and prevention of gastric cancer. *J Dig Dis*. 2008 Feb;9(1):8-13.
- Malfertheiner P, Megraud F, O'Morain C, Bazzoli F, El-Omar E, Graham D, et al. Current concepts in the management of *Helicobacter pylori* infection: the Maastricht III Consensus Report. *Gut*. 2007 Jun;56(6):772-81.
- Santha sheela NB, Damodharan N, Surekha I, Srinivas rao T. Formulation and evaluation of clarithromycin gastroretentive dosage form. *Int J Pharmacy and Pharm Sci*. 2010 ;3(2),48-55.
- Sambathkumar R, Venkateswaramurthy N, Vijayabaskaran M, Perumal P. Formulation of clarithromycin loaded mucoadhesive microspheres by emulsification-internal gelation technique for anti-*Helicobacter pylori* therapy. *Int J Pharmacy and Pharm Sci*. 2011 ;2(3),173-177.
- Mégraud F. *H. pylori* antibiotic resistance: prevalence, importance, and advances in testing. *Gut*. 2004 ; 53(9),1374-84.
- Kim JM, Kim JS, Kim N, Kim YJ, Kim IY, Chee YJ, et al. Gene mutations of 23S rRNA associated with clarithromycin resistance in *Helicobacter pylori* strains isolated from Korean patients. *J Microbiol Biotechnol*. 2008 ; 18(9),1584-9.
- Ho SL, Tan EL, Sam CK, Goh KL. Clarithromycin resistance and point mutations in the 23S rRNA gene in *Helicobacter pylori* isolates from Malaysia. *J Dig Dis*. 2010 Apr;11(2):101-5.
- Moosavian M, Tajbakhsh S, Samarraf-Zadeh AR. Rapid detection of clarithromycin-resistant *Helicobacter pylori* in patients with dyspepsia by fluorescent in situ hybridization (FISH) compared with the E-test. *Ann Saudi Med*. 2007 Mar-Apr;27(2):84-8.
- Bakir Ozbey S, Ozakin C, Keskin M. Antibiotic resistance rates of *Helicobacter pylori* isolates and the comparison of E-test and fluorescent in situ hybridization methods for the detection of clarithromycin resistant strains. *Mikrobiyol Bul*. 2009 ; 43(2):227-34.
- clinical and laboratory standards institute. 2010. Performance standards for antimicrobial susceptibility testing ; twentieth informational supplement M100-S20.
- Cerqueira L, Fernandes RM, Ferreira RM, Carneiro F, Dinis-Ribeiro M, Figueiredo C, et al. PNA-FISH as a new diagnostic method for the determination of clarithromycin resistance of *Helicobacter pylori*. *BMC Microbiol*. 2011 May 14;11:101.
- Morris JM, Reasonover AL, Bruce MG, Bruden DL, McMahon BJ, Sacco FD, et al. Evaluation of seaFAST, a rapid fluorescent in situ hybridization test, for detection of *Helicobacter pylori* and resistance to clarithromycin in paraffin-embedded biopsy sections. *J Clin Microbiol*. 2005 Jul;43(7):3494-6.
- Jüttner S, Vieth M, Miehle S, Schneider-Brachert W, Kirsch C, Pfeuffer T, et al. Reliable detection of macrolide-resistant *Helicobacter pylori* via fluorescence in situ hybridization in formalin-fixed tissue. *Mod Pathol*. 2004 Jun;17(6):684-9.
- Ben Mansour K, Buruoca C, Zribi M, Masmoudi A, Karoui S, Kallel L, et al. Primary resistance to clarithromycin, metronidazole and amoxicillin of *Helicobacter pylori* isolated from Tunisian patients with peptic ulcers and gastritis: a prospective multicentre study. *Ann Clin Microbiol Antimicrob*. 2010 Aug 13;9:22.
- Shokrzadeh L, Jafari F, Dabiri H, Baghaei K, Zojaji H, Alizadeh AH, et al. Antibiotic susceptibility profile of *Helicobacter pylori* isolated from the dyspepsia patients in Tehran, Iran. *Saudi J Gastroenterol*. 2011 Jul-Aug;17(4):261-4.

35. Eisig JN, Silva FM, Barbuti RC, Navarro-Rodriguez T, Moraes-Filho JP, Pedrazzoli Jr J. Helicobacter pylori antibiotic resistance in Brazil: clarithromycin is still a good option. *Arq Gastroenterol*. 2011 Oct-Dec;48(4):261-4.
36. Goh KL, Navaratnam P. High Helicobacter pylori resistance to metronidazole but zero or low resistance to clarithromycin, levofloxacin, and other antibiotics in Malaysia. *Helicobacter*. 2011 Jun;16(3):241-5.
37. Raymond J, Lamarque D, Kalach N, Chaussade S, Burucoa C. High level of antimicrobial resistance in French Helicobacter pylori isolates. *Helicobacter*. 2010 Feb;15(1):21-7.
38. Bakir Ozbey S, Ozakin C, Keskin M. Antibiotic resistance rates of Helicobacter pylori isolates and the comparison of E-test and fluorescent in situ hybridization methods for the detection of clarithromycin resistant strains. *Mikrobiyol Bul*. 2009 Apr;43(2):227-34.
39. Wueppenhorst N, Stueger HP, Kist M, Glocker E. Identification and molecular characterization of triple- and quadruple-resistant Helicobacter pylori clinical isolates in Germany. *J Antimicrob Chemother*. 2009 Apr;63(4):648-53. Epub 2009 Feb 3.
40. De Francesco V, Giorgio F, Ierardi E, Zotti M, Neri M, Milano A, et al. Primary clarithromycin resistance in Helicobacter pylori: the Multicentric Italian Clarithromycin Resistance Observational (MICRO) study. *J Gastrointest Liver Dis*. 2011 Sep;20(3):235-9.
41. Meyer JM, Silliman NP, Wang W, Siepmann NY, Sugg JE, Morris D, et al. Risk factors for Helicobacter pylori resistance in the United States: the surveillance of H. pylori antimicrobial resistance partnership (SHARP) study, 1993-1999. *Ann Intern Med*. 2002 Jan 1;136(1):13-24.
42. Wong WM, Xiao SD, Hu PJ, Wang WH, Gu Q, Huang JQ, et al. Standard treatment for Helicobacter pylori infection is suboptimal in non-ulcer dyspepsia compared with duodenal ulcer in Chinese. *Aliment Pharmacol Ther*. 2005 Jan 1;21(1):73-81.
43. Gisbert JP, Marcos S, Gisbert JL, Pajares JM. Helicobacter pylori eradication therapy is more effective in peptic ulcer than in non-ulcer dyspepsia. *Eur J Gastroenterol Hepatol*. 2001 Nov;13(11):1303-7.