Academic Sciences

International Journal of Pharmacy and Pharmaceutical Sciences

ISSN- 0975-1491

Vol 5, Issue 1, 2013

Research Article

STUDY OF STABILITY OF GALANTAMINE PEPTIDE ESTERS AT ROOM TEMPERATURE AND DIFFERENT PH VALUES

DOBRINA TSVETKOVA¹, DANKA OBRESHKOVA¹, NIKOLAI DANCHEV²

¹Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Medical University – Sofia, ²Department of Pharmacology, Toxycology and Pharmacotherapy, Faculty of Pharmacy, Medical University – Sofia.

Received: 23 April 2012, Revised and Accepted: 05 Aug 2012

ABSTRACT

In current work are summarized the experimental results from the study of chemical stability of Galantamine peptide esters 3,4 – dichlorophenyl – Alanil – Leucil – Glycil – Galantamine (LEU – GAL) and 3,4 – dichlorophenyl – Alanil – Valil – Glycil – Galantamine (VAL – GAL) in different aqueous buffer solutions with pH = 2, pH = 7.4, pH = 9 at room temperature in period of 6 hours. The applied chromatographic TLC conditions were: stationary phase: TLC aluminum plates (Merck) precoated with silica gel $G_{60}F_{254}$; mobile phase: n – butanol: water = 30: 10 v/v; length run – 120 mm. Densitometric analysis was carried out in reflectance – absorbance mode at λ = 282 nm.

Linear regression analysis was employed to calculate the regression equations and the correlation coefficients. The obtained regression equations, showing the proportional accordance A = f(C) at pH = 2 were: y = 8388.x - 3.704, $R^2 = 0.998$ (LEU – GAL); y = 3021.x - 0.391, $R^2 = 0.999$ (VAL – GAL).

The examined Galantamine esters are resistant at room temperature to chemical hydrolysis in aqueous buffer solutions with pH = 2, pH = 7.4, pH = 9, which is proved by the following facts: 1) on chromatograms didn't exist spots with t_R , corresponded to t_R of the respective derivative: $t_R = 0.23$ (LEU – GAL); $t_R = 0.33$ (VAL – GAL); 2) the area of spots remains similar during 6 hour of experiment. At pH = 2 the content of LEU – GAL is in interval 10.22 mg – 10.58 mg and the quantity of VAL – GAL is in interval 9.07 mg – 11.05 mg.

Keywords: Galantamine, Peptide esters, Stability, pH.

INTRODUCTION

Alzheimer's disease is a progressive neurodegenerative disorder associated with loss of neurons in brain [1] and presence of excessive amounts of neurotic plaques containing amyloid β protein [2]. For therapy of neurodegenerative Alzheimer's disease are applied inhibitors of acethylcholinesterase [3]: Donepezil [4], Galantamine [4, 5], Rivastigmine [4, 6] and Tacrine [7]. Alzheimer's disease is pathologically characterized by amyloid plaques and neurofibrillary tangles [8]. In the brain of patients a major pathological alteration are senile plaques, composed of amyloid ß peptide (Aß) in length from Aß38 to Aß42 [9]. Aß is generated by the consecutive cuts of proteases β - and γ - secretase, which liberate the amyloidogenic peptide from its precursor, the ß amyloid precursor protein (APP) by endoproteolysis 10. ß secretase (BACE) mediates the N - terminal cleavage, producing a membrane - associated C - terminal fragment (CTFß) of ßAPP 11, 12. The latter is further processed to Aß by γ – secretases, which cleave within the single transmembrane region 13. Subsequent aggregation is thought to result in the formation of neurotoxic protofibrils and the deposition of amyloid plaques 14.

One of the most promising therapeutic strategies to slow progression of Alzheimer's disease pathology is the application of γ – secretase inhibitors, which reduce the amyloid A β 40/42 peptide (A β) production [15, 16] by blocking γ – secretase activity [17]. The first in vivo test of γ – secretase inhibitor is of dipeptide DAPT, which reduces β – amyloid peptide levels in brain 18.

For stability analysis are applied the following methods: HPLC (Bendamustine hydrochloride) [19], HPTLC (Ezetimibe) [20].

The aim of current study is to investigate the chemical stability of new synthetized esters of Galantamine in different aqueous buffer solutions with pH = 2, pH = 7.4, pH = 9.

MATERIALS AND METHODS

MATERIALS

I) 3,4 – dichlorophenyl – Alanil – Leucil – Glycil – Galantamine (LEU – GAL) and 3,4 – dichlorophenyl – Alanil – Valil – Glycil – Galantamine (VAL – GAL) (syntetized from prof. Vesenkov from Department of

Organic Chemistry, University of Chemical Technology and Metallurgy) (Fig. 1.) [21].



Fig. 1: Structures of Galantamine peptide esters.

 $R = -CH(CH_3)_2 3,4 - dichloro - Alanil - Leucil - Glycil - Galantamine$

R = - CH₂CH(CH₃)₂3,4 - dichloro - Alanil - Valin - Glycil - Galantamine

II) Reagents with analytical grade quality: n – butanol, water, boric acid, 0.1 mol/l HCl, 0.1 M sodium hydroxide, 1 M sodium hydroxide, disodium hydrogen phosphate, sodium chloride, potassium chloride, potassium dihydrogen phosphate.

III) TLC plates: Silicagel G₆₀F₂₅₄, 20 cm x 20 cm (Merck).

METHOD: TLC – densitometry.

I. Instrumentation

The chromatographic procedure was carried out using 10 μl sample syringe (Hamilton, Bonaduz, Switzerland) and TLC densitometric scanner TR 541a, performed in the reflectance – absorbance mode at λ = 282 nm.

II. Chromatographic TLC conditions: stationary phase: precoated with Silicagel G₆₀F₂₅₄ TLC plates; mobile phase n – butanol: water = 30: 10 v/v; detection at $\lambda = 282 \text{ nm}$; length run – 120 mm.

III. Buffer preparation:

Buffer reagents were of reagent grade. Buffer solutions were prepared according to European Pharmacopoeia 5.0: 01/2005:40103 as follows:

1) Buffer solution pH 2.0: 4000200:

6.57 g of potassium chloride were dissolved in water R and to the obtained solution 119.0 ml 0.1 mol/l HCl were added. The mixture was diluted in volumetric flask to 1000.0 ml with water R.

2) Phosphate buffered saline pH 7.4: 4005000.

2.38~g of disodium hydrogen phosphate R, 0.19~g of potassium dihydrogen phosphate R and 8.0 g of sodium chloride R were dissolved in water R. The obtained solution was diluted in volumetric flask to 1000.0 ml with the same solvemt.

3) Buffer solution pH 9.0: 4007000.

Solution I. 6.18 g of boric acid R were dissolved in 0.1 M potassium chloride R and solution was diluted in volumetric flask to 1000.0 ml with the same solvent.

Solution II. 0.1 M sodium hydroxide.

Buffer solution pH 9.0 was prepared by mixing of 1000.0 ml Solution I and 420.0 ml Solution II.

4) Solutions with NaOH.

a) to 5 ml of 1 mol/l solution of 3,4 – dichlorophenyl – Alanil – Leucil – Glycil – Galanthamine (Mr = 672.2) were added 5 ml 1 mol/l NaOH

b) to 5 ml of 1 mol/l solution of 3,4 – dichlorophenyl – Alanil – Valil – Glycil – Galanthamine (Mr = 658.2) were added 5 ml 1 mol/l NaOH

IV. Preparation of solutions for linearity of 3,4 – dichlorophenyl – Alanil – Leucil – Glycil – Galantamine (LEU – GAL).

An accurately weighed quantity of LEU – GAL: 10 mg, 30 mg, 40 mg, 45 mg, 80 mg, 100 mg, 110 mg, 120 mg, 125 mg, 130 mg, 135 mg, 140 mg, 160 mg, 180 mg was dissolved in Buffer solution pH 2.0 to 1.0 ml to obtain solutions with concentration correspondingly: 1.10^{-1}

 2 g/ml; 3.10- 2 g/ml; 4.10- 2 g/ml, 4.5.10- 2 g/ml, 8.10- 2 g/ml, 1.10- 1 g/ml, 1.1.10- 1 g/ml, 1.2.10- 1 g/ml, 1.2.5.10- 1 g/ml, 1.3.10- 1 g/ml, 1.4.10- 1 g/ml, 1.6.10- 1 g/ml, 1.8.10- 1 g/ml.

V. Preparation of solutions for linearity of 3,4 – dichlorophenyl – Alanil – Valil – Glycil – Galantamine (VAL – GAL).

An accurately weighed quantity of VAL – GAL: 10 mg, 25 mg, 90 mg, 125 mg, 130 mg, 135 mg, 350 mg, 410 mg, 430 mg, 470 mg, 500 mg was dissolved in Buffer solution pH 2.0 to 1.0 ml to obtain solutions with concentration correspondingly: 1.10^{-2} g/ml; $2.5.10^{-2}$ g/ml; 9.10^{-2} g/ml, $1.25.10^{-1}$ g/ml, $1.3.10^{-1}$ g/ml, $1.35.10^{-1}$ g/ml, $3.5.10^{-1}$ g/ml, $4.1.10^{-1}$ g/ml, $4.3.10^{-1}$ g/ml, $4.7.10^{-1}$ g/ml, 5.10^{-1} g/ml.

VI. Preparation of solutions of Galantamine esters in buffers with pH = 2, pH = 7.4, pH = 9.

An accurately weighed quantity of 1 mg respectively of LEU – GAL and VAL – GAL was dissolved separately in buffer solution with pH = 2.0, pH = 7.4, pH = 9.0 to obtain concentration of the examined compounds: 1 mg/1ml. Samples of 10 μ l from the investigated solutions were taken at every 30 min. during interval of 6 hour and chromatograms were recorded on TLC plates Silicagel G₆₀F₂₅₄ and mobile phases: n – butanol: water = 30: 10 v/v. The chromatograms were scanned in reflectance – absorbance mode at $\lambda = 282$ nm.

RESULTS AND DISCUSSION

I. Linearity.

For the investigation of analytical parameter linearity were prepared solutions with inecreasing concentration of respective compound and were analyzed separately by the written TLC densitometric method. The proportional accordance between the spot area (A) and concentration (C) in g/ml is found and the results are shown on Table 1.

	fable 1: Results for anal	ytical patameter linearity	y for LEU – GAL and VAL – GAL.
--	---------------------------	----------------------------	--------------------------------

N:	3,4 – dichlorophen	yl – Alanil – Leucil – Glycil – Galantamine	3,4 - dichlorophenyl - Alanil - Valil - Glycil - Galantamine		
1.	[g/ml]	Spot area (A)	[g/ml]	Spot area (A)	
2.	1.10-2	83	1.10-2	30	
3.	3.10 ⁻²	265	2.5.10-2	74	
4.	4.10-2	326	9.10-2	270	
5	4.5.10 ⁻²	365	1.25.10-1	379	
6.	8.10-2	670	1.3.10-1	391	
7.	1.10-1	822	1.35.10-1	404	
8.	1.1.10-1	908	$3.5.10^{-1}$	1078	
9.	1.2.10-1	993	4.1.10-1	1235	
10.	1.25.10-1	1034	4.3.10-1	1292	
11.	1.3.10 ⁻²	1089	4.7.10-1	1420	
12.	1.35.10-2	1124	5.10-1	1506	
13.	1.4.10-2	1220			
14.	1.6.10-2	1331			
15.	1.8.10-2	1504			

Linear regression analysis was employed to calculate the regression equations and the correlation coefficients. The obtained regression equations, showing the proportional accordance A = f (C) are: y = 8388.x - 3.704, R²= 0.998 (LEU – GAL); y = 3021.x - 0.391, R²= 0.999 (VAL – GAL).

On Fig.1. (LEU – GAL) and Fig. 2. (VAL – GAL) are illustrated the calibration curves for linearity.

On Table 2. (LEU – GAL) and Table 3. (VAL – GAL) are presented the results from 6 h, t = 20 °C at pH = 2 for: 1) spot area (A): $A_{LEU} - GAL$, $A_{VAL} - GAL$; 2) quantity of compounds [mg/ml]: [LEU – GAL], [VAL – GAL]; 3) Chauvenet's criterion for spot area (UA): U $A_{LEU} - GAL$, U $A_{VAL} - GAL$; 4) Chauvenet's criterion for content of compounds (UC): U[LEU – GAL], U[VAL – GAL], U[VAL – GAL].

The results during 6 h of the experiment show that the examined Galantamine esters are resistant at room temperature to chemical hydrolysis in aqueous buffer solutions with pH = 2, pH = 7.4, pH = 9, selected as model of pH of the stomach, blood and intestine. The chemical stability of derivatives is proved by the following facts: 1) on chromatograms didn't exist spots with t_{R_0} corresponded to t_R of the

respective derivative: $t_R = 0.23$ [LEU – GAL]; $t_R = 0.33$ [LEU – GAL]; 2) the area of spots remains similar during 6 hour of experiment.

For all of the obtained results for spot area and for content of LEU – GAL and VAL – GAL is necessary to estimate the Chauvenet's criterion (U), because when U for one value is higher than the relevant standard criterion (USt), the result must be removed as unexpected. The relations: U A_{LEU} - GAL 2.03, U [LEU – GAL] < 2.03 (Table 2.) and U A_{VAL} - G_{AL} < 2.03, U [VAL – GAL] < 2.03 (Table 3.) show, that all experimental results for UA and UC are lower, than standard requirement: Umax = 2.03 (n = 13), and it isn't necessary to remove data for A and C. For the calculation of sample standard deviation (SD) is applied the Bessel's correction, in which the denominator N – 1 (degrees of freedom) is used instead of N and in this case (S)²is an unbiased estimator for (SD)².

On Table 4. (LEU – GAL) and Table 5. (VAL – GAL) are presented the results from 6 h, t = 20 °C at pH = 7.4, pH = 9 and solutions mol/mol with NaOH for: 1) spot area (A): A_{LEU} – G_{AL} , A_{VAL} – G_{AL} ; 2) Chauvenet's criterion for spot area (UA): U A_{LEU} – G_{AL} , U A_{VAL} – G_{AL} . All values of UA are lower, than standard requirement: Umax = 2.03 (n = 13).



Fig. 2: Calibration curve for linearity of VAL – GAL at pH =2.

Table 2. Snot area and	contont [mg/m]]	of LELL CAL for 6	h + - 20 of a + nU - 2
Table 2: Spot area and	content [mg/mi]	OI LEU - GAL IOF O	$\Pi_{1} \iota = 20 \circ \iota a \iota p \Pi = 2$
	L 0/ 1		, 1

T [min.]	Spot area (A)		Content (C) [mg/ml]		
	ALEU - GAL	U ALEU - GAL	[LEU – GAL]	U [LEU – GAL]	
0	84	0.79	10.46	0.77	
30	83	0.14	10.34	0.15	
60	84	0.79	10.46	0.77	
90	84	0.79	10.46	0.77	
120	82	1.07	10.22	1.08	
150	83	0.14	10.34	0.15	
180	82	1.07	10.22	1.08	
210	82	1.07	10.22	1.08	
240	84	0.79	10.46	0.77	
270	82	1.07	10.22	1.08	
300	84	0.79	10.46	0.77	
330	82	1.07	10.22	1.08	
360	85	1.73	10.58	1.69	
X	83.15		10.36		
SD	1.07		0.13		

N:	Т	pH = 2		pH = 2		
	[min.]	Spot area		Content [mg/ml]		
		AVAL - GAL	U Aval - gal	[VAL – GAL]	U [VAL – GAL]	
1.	0	30	0.36	10.06	0.36	
2.	30	27	1.20	9.07	1.19	
3.	60	29	0.16	9.73	0.16	
4.	90	28	0.68	9.40	0.67	
5.	120	28	0.68	9.40	0.67	
6.	150	28	0.68	9.40	0.67	
7.	180	31	0.88	10.39	0.88	
8.	210	29	0.16	9.73	0.16	
9.	240	27	1.20	9.07	1.19	
10.	270	28	0.68	9.40	0.67	
11.	300	32	1.39	10.72	1.39	
12.	330	33	1.91	11.05	1.91	
13.	360	31	0.88	10.39	0.88	
14.	X	29.31		9.83		
15.	SD	1.93		0.64		

Table 3: Spot area and content [mg/ml] of VAL – GAL for 6 h, t = 20 °C at pH = 2.

Table 4: Spot area for LEU – GAL for 6 h, t = 20 °C at different pH values.

Т	pH = 7.4		pH = 9		mol/mol with Na	ОН	
[min.]	Spot area		Spot area		Spot area		
	A LEU - GAL	U ALEU - GAL	A LEU - GAL	U Aleu - gal	Aleu - gal	U Aleu - gal	
0	23	0.17	31	0.5	29	0.57	
30	27	1.35	31	0.5	32	1.74	
60	26	0.97	34	0.04	27	0.21	
90	27	1.35	41	1.32	31	1.35	
120	21	0.94	25	1.6	26	0.6	
150	22	0.56	36	0.41	26	0.6	
180	22	0.56	29	0.87	25	0.99	
210	27	1.35	35	0.22	25	0.99	
240	20	1.32	44	1.86	24	1.38	
270	20	1.32	26	1.42	28	0.18	
300	22	0.56	36	0.41	31	1.35	
330	23	0.17	38	0.77	26	0.6	
360	25	0.59	33	0.14	28	0.18	
X	23.46		33.77		27.54		
SD	2.63		5.49		2.57		

Table 5: Spot area for VAL - GAL for 6 h, t = 20 °C at different pH values.

Т	pH = 7.4		pH = 9	pH = 9 Spot area		mol/mol with NaOH	
[min.]	Spot area		Spot area				
	A VAL - GAL	U Aval - gal	Aval - gal	U Aval - gal	Aval - Gal	U Aval - gal	
0	46	1.85	45	0.08	132	1.20	
30	55	0.05	37	1.86	129	0.55	
60	53	0.37	44	0.31	127	0.12	
90	55	0.05	49	0.8	130	0.76	
120	58	0.68	47	0.36	127	0.12	
150	52	0.58	40	1.20	124	0.53	
180	62	1.52	51	1.25	120	1.40	
210	52	0.58	46	0.14	130	0.76	
240	50	1.0	53	1.69	121	1.18	
270	54	0.16	47	0.36	121	1.18	
300	53	0.37	47	0.36	135	1.84	
330	63	1.73	44	0.31	125	0.32	
360	59	0.89	40	1.20	123	0.75	
X	54.77		45.38		126.46		
SD	4.75		4.5		4.63		

CONCLUSION

The examined Galantamine peptide esters 3,4 – dichlorophenyl – Alanil – Leucil – Glycil – Galantamine and 3,4 – dichlorophenyl – Alanil – Valil – Glycil – Galantamine are resistant at different aqueous buffer solutions with pH = 2, pH = 7.4, pH = 9 at room temperature in period of 6 hours. At pH = 2 content of LEU – GAL is

in interval 10.22 – 10.58 mg and quantity of VAL – GAL is in interval 9.07 mg – 11.05 mg.

ACKNOWLEDGEMENTS

This article was prepared with the financial support from Grant project N:14/2012 (Contract N:26/17.07.2012) – Medical University – Sofia, Bulgaria.

The authors would like to thank to prof. Ljubomir Vesenkov from the Department of Organic Chemistry, University of Chemical Technology and Metallurgy – Sofia, for the providing of substances of Galantamine peptide esters.

REFERENSES

- 1. Yadav YC, Jain A, Deb L. A review: neuropharmacological screening techniques for pharmaceuticals. International Journal of Pharmacy and Pharmaceutical Sciences 2010;2,Suppl.2:10-4.
- Kulkarni PD, Ghaisas MM, Chivate ND, Sankpal PS. Memoryenchancing activity of Cissampelos pariela in mice. International Journal of Pharmacy and Pharmaceutical Sciences 2011;3(2):206-11.
- 3. Villarroya M, Garsia AG, Marco JL. New classes of AChE inhibitors with additional pharmacological effects of interest for the treatment of Alzheimer's disease. Curr Pharm Des 2004;10(25):3177-84.
- Van Dam D, Abramowski D, Staufenbiel M. Symptomatic effect of donepezil, rivastigmine, galantamine and memantine on cognitive deficits in the APP23 model. Psychopharmacology (Berl.) 2005;180(1):177-90.
- 5. Fulton B, Benfield P. Galanthamine. Drugs Aging 1996;9(1):60-5.
- Birks J, Grimley EJ, Iakovidou V, Tsolaki M, Holt FE. Rivastigmine for Alzheimer's disease. Cochrane Database Syst Rev 2009;2:CD001191.
- Qizilbash N, Birks J, Lopez Arrieta J, Lewington S, Szeto S. Tacrine for Alzheimer's disease. Cochrane Database Syst Rev 2007;3:CD000202.
- Goedert M, Spillantini MG. A century of Alzheimer's disease. Science 2006;314:777-81.
- 9. Selkoe DJ. Translating cell biology into therapeutic advances in Alzheimer's disease. Nature 1999;399:A23-A31.
- 10. Esler WP, Wolfe MS. A portrait of Alzheimer secretases new features and familiar faces. Science 2001;293:1449-54.

- 11. Vassar R, Citron M. A ß generating enzymes: recent advances in ß and γ secretase research. Neuron 2000;27:419-22.
- Yan R, Bienkowski MJ, Shuck ME, Miao H, Tory MC, Pauley AM et al. Membrane – anchored aspartylprotease with Alzheimer's disease beta – secretase activity. Nature 1999;402:533-37.
- 13. Tsai J Y, Wolf MS, Xia W. The Search for γ Secretase and Development of Inhibitors. Current Medicinal Chemistry 2002;9(11):1087-106.
- Walsh DM, Lomakin A, Benedek GB, Condron MM, Teplow DB. Amyloid β –protein fibrillogenesis. Detection of a protofibrillar intermediate. J Biol Chem 1997;272(35):22364-72.
- 15. Petit A, Bihel F, da Costa AC, Pourquie O, Checler F, Kraus JL. New protease inhibitors prevent γ – secretase – mediated production of A β 40/42 without affecting Notch cleavage. Nat Cell Biol 2001;3:507-11.
- Siemers E, Skinner M, Dean RA, Gonzales C, Satterwhite J, Farlow M et al. Safety, tolerability, and changes in amyloid beta concentrations after administration of a gamma – secretase inhibitor in volunteers. Clin Neuropharmacol 2005;28(3):126-32.
- 17. Geling A, Steiner H, Willem M, Bally Cuif L, Haass C. A γ secretase inhibitor blocks Notch signaling in vivo and causes a severe neurogenic phenotype in zebrafish. EMBO Reports 2002;3(7):688-94.
- 18. Dovey HF, John V, Anderson JP, Chen LZ, de Saint AP, Fang LY et al. Functional γ secretase inhibitors reduce ß amyloid peptide levels in brain. J Neurochem 2001;76(1):173-81.
- Pencheva I, Bogomilova A, Obreshkova D, Troev K. HPLC study on the stability of bendamustine hydrochloride immobilized onto polyphosphoesters. J Pharm Biomed 2008;48:1143-50.
- Mahadik MV, Dhaneshwar SR. Application of a stability indicating HPTLC method for the quantitative determination of ezetimibe in pharmaceutical dosage forms. Asian J of Pharm Sci 2007;2(5):182-90.
- Vezenkov L, Georgieva M, Danalev D, Ivanov Tch, Ivanova G. Synthesis and characterization of new Galanthamine derivatives comprising peptide moiety. Protein and Peptide Letters 2009;16(9):1024-28.