SYNTHESIS AND EVALUATION OF SOME NEW 2-(5-(4-BENZAMIDOBENZYLIDENE)-2,4-DIOXOTHIAZOLIDIN-3-YL)ACETIC ACID ANALOGS AS ALDOSE REDUCTASE INHIBITORS

JYOTI PANDEY1,2, ARSHAD ALI2, ARUN KUMAR GUPTA3*

1School of Pharmacy, Suresh Gyan Vihar University, Jaipur, Rajasthan, India. 2Department of Pharmaceutical Chemistry, Smriti College of Pharmaceutical Sciences, Indore, Madhya Pradesh, India. 3Department of Pharmaceutical, RKDF Institute of Pharmaceutical Sciences, Indore, Madhya Pradesh, India. Email: arunkg73@gmail.com

Received: 06 April 2016, Revised and Accepted: 10 October 2016

ABSTRACT

Objective: Aldose reductase (ALR) enzyme plays a significant role in conversion of excess amount of glucose into sorbitol in diabetic condition, inhibitors of which decrease the secondary complication of diabetes mellitus. Scarce treatment of diabetic complications has motivated our interest in the search of new aldose reductase inhibitors (ARIs) endowed with more favorable biological properties.

Methods: Newer (4-(benzamidobenzylidene)-2,4-dioxothiazolidin-3-yl) acetic acid derivatives were synthesized, and these compounds were evaluated for their ARI and antidiabetic activity.

Results: ARI activity of synthesized compounds was found in the range of 57.8-71.9% at 5µg/mL. Similarly, synthesized compounds decrease blood glucose level in the range of 64.4-70.5 mg/dl at 15 mg/kg body weight.

Conclusion: (E)-2-(5-(4-(substituted benzamido)benzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid analogs shows comparable ARI as well as antidiabetic activity. These new class of compounds might be address the diabetic complications with safety.

Keywords: Aldose reductase inhibitors, Diabetes mellitus, N-acetic acid-2,4-thiazolidinediones.

INTRODUCTION

Diabetes mellitus (DM) have reached global pandemic proportions with India being designated "diabetes capital" of the world. DM is a multisystem disorder comprising metabolic and vascular abnormalities resulting from insulin deficiency, with or without insulin resistance. Diabetes is a prevalent, costly condition that causes significant illness, disability, and premature death. Insulin deficiency, in turn, leads to chronic hyperglycemia with disturbances of carbohydrate, fat, and protein metabolism. The diabetic complications and the United Kingdom prospective diabetes study demonstrated that strict and sustained control of glucose excursions through interventions including intensive insulin therapy reduces the risk of developing these complications in types 1 and 2 diabetes. However, relatively few diabetics have adopted this strict, physician monitored regimen, and this type of round-the-clock control is not practical for patients at large. During hyperglycemic event, the elevated glucose level enhances the activity of AR by increasing glucose flux through this pathway. Aldose reductase (ALR) contributes to the development of secondary diabetic complications. It is, therefore, believe to be a promising drug target. At present, there is no specific therapy available for diabetic complications. A metabolic approach is to control excess glucose flux in diabetic tissue through the first step of polyol pathway by aldose reductase inhibitors (ARIs) [1-5].

AR catalyzes the nicotinamide adenine dinucleotide phosphate (NADPH) dependent reduction of glucose into sorbitol in the first step of the polyol pathway, which further dehydrogenases through an NAD+ dependent reaction into fructose [6,7]. The deprivation of NADPH and NAD+ and the intracellular accumulation of sorbitol result in biochemical imbalances which cause damage in target tissues. The oxidative stress triggered by the glucose oxidation process and increased ALR activity is considered to be a major mechanism responsible for the onset of diabetic complications [8,9]. A variety of ARIs have been reported; however, in clinical studies, many of them have exhibited low efficacy or a narrow spectrum of tissue activity, generally because of unfavorable pharmacokinetics, or have proved to produce toxic side-effects. At present, epalrestat is the only ARI available on the market [10-12]. Literature reveals that in the last few years, numerous 5-arylidene-2,4-thiazolidinediones derivatives produced appreciable ARI inhibition [13,14], but their effectiveness generally decreases in vivo, probably due to their poor penetrability to key target tissues, in particular, peripheral nerves [15-17]. Thus, the aim of this work to develop new ARIs, which improves physicochemical properties and better bioavailability. In this study, new active analogs of 5-arylidene-2,4-dioxothiazolidines were identified and synthesized as potent in vitro ARI. In particular, 5-benzylidene moieties bearing methyl, methoxy, chloro, and/or dichloro substituted benzamido derivatives were prepared, which is potentially able to enhance stability of enzyme-inhibitor complex.

METHODS

All the chemicals used in the synthesis of designed compounds were of synthetic grade, and they were procured from Loba, Highmedia, and E. Merck. Thin layer chromatographic method was used for monitoring the progress of the reactions, thin layer chromatography (TLC) was performed using silica gel-G on glass plate in different solvent systems. Iodine vapor and UV detector (long wavelength) were used as detecting agents. The purification of intermediates and final compounds was carried out through recrystallization and column chromatography technique. For the purpose of chromatography glass column (high 18" with internal diameter 20 mm), column grade silica gel mesh #240-400 as the stationary phase and appropriate solvent system as mobile phase were used. The melting points of synthesized compounds and intermediates were determined by open capillary method, which were uncorrected.

The absorption maxima (λmax) of the intermediate and synthesized compounds were determined on Shimadzu 1700 UV-visual
N (0.001 mol) was added. To the reaction mixture substituted benzoyl anhydrous dichloromethane. To this, a catalytic amount of triethylamine acetic acid (0.001 mol) was taken in round-bottomed flask containing substituted 2-(5-(4-benzamidobenzyl)-2,4-dioxothiazolidin-3-yl)acetic acid derivatives (G1-G6).

**Synthesis of substituted 2-(5-(4-benzamidobenzyl)-2,4-dioxothiazolidin-3-yl)acetic acid derivatives (G1-G6)**

The crude compound C (0.025 mol) and granulated tin (0.038 mol) were taken in round-bottomed flask equipped with reflux condenser. The crude compound C (0.025 mol) and thiazolidinedione-\text{-}acetic acid (0.025 mol) was taken in round-bottomed flask containing glacial acetic acid. To this, a catalytic amount of sodium acetate (0.080 g) was added. The reaction mixture was stirred and heated at 100-105°C for 10-12 hrs. Progress of the reaction mixture was checked through TLC. After completion of reaction, the mixture was kept aside for overnight at RT. The crystalline product was filtered, washed with cold acetic acid and used in next step (Scheme 1).

**Synthesis of substituted benzoyl chloride (F1-F6)**

Different substituted benzoic acid derivatives (0.01 mol) were refluxed with thionyl chloride for 3-4 hrs, and the reaction was monitored through TLC. After completion of reaction, the mixture was kept aside for overnight at RT. The crystalline product was filtered, washed with cold acetic acid and used in next step (Scheme 2).

**Synthesis of substituted 2-(5-(4-aminobenzyl)-2,4-dioxothiazolidin-3-yl)acetic acid derivatives (D)**

The crude compound C (0.025 mol) and granulated tin (0.038 mol) were taken in round-bottomed flask equipped with reflux condenser. The reaction mixture was stirred and heated at 100-105°C for 10-12 hrs. Progress of the reaction mixture was checked through TLC. After completion of reaction, the mixture was kept aside for overnight at RT. The crystalline product was filtered, washed with cold acetic acid and used in next step (Scheme 2).

**Synthesis of substituted benzoyl chloride (F1-F6)**

Different substituted benzoic acid derivatives (0.01 mol) were refluxed with thionyl chloride for 3-4 hrs, and the reaction was monitored through TLC. After completion of reaction, the mixture was kept aside for overnight at RT. The crystalline product was filtered, washed with cold acetic acid and used in next step (Scheme 2).

**General method of synthesis**

**Synthesis of (E)-2-(5-(4-nitrobenzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid derivatives (C)**

Equimolar concentration of derivative of 4-nitro benzoaldehyde (0.025 mol) and thiazolidinedione-N-acetic acid (0.025 mol) was taken in round-bottomed flask containing glacial acetic acid. To this, a catalytic amount of sodium acetate (0.080 g) was added. The reaction mixture was stirred and heated at 100-105°C for 10-12 hrs. Progress of the reaction mixture was checked through TLC. After completion of reaction, the mixture was kept aside for overnight at RT. The crystalline product was filtered, washed with cold acetic acid and used in next step (Scheme 1).

**Synthesis of 2-(5-(4-aminobenzyl)-2,4-dioxothiazolidin-3-yl)acetic acid derivatives (D)**

The crude compound C (0.025 mol) and granulated tin (0.038 mol) were taken in round-bottomed flask equipped with reflux condenser. The reaction mixture was stirred and heated at 100-105°C for 10-12 hrs. Progress of the reaction mixture was checked through TLC. After completion of reaction, the mixture was kept aside for overnight at RT. The crystalline product was filtered, washed with cold acetic acid and used in next step (Scheme 2).

**Synthesis of substituted benzoyl chloride (F1-F6)**

Different substituted benzoic acid derivatives (0.01 mol) were refluxed with thionyl chloride for 3-4 hrs, and the reaction was monitored through TLC. After completion of reaction, the mixture was kept aside for overnight at RT. The crystalline product was filtered, washed with cold acetic acid and used in next step (Scheme 2).

**Synthesis of substituted 2-(5-(4-benzamidobenzyl)-2,4-dioxothiazolidin-3-yl)acetic acid derivatives (G1-G6)**

Substituted 2-(5-(4-benzamidobenzyl)-2,4-dioxothiazolidin-3-yl)acetic acid (0.001 mol) was taken in round-bottomed flask containing anhydrous dichloromethane. To this, a catalytic amount of triethylamine (0.001 mol) was added. To the reaction mixture substituted benzylic chloride was added slowly with constant stirring. The progress of the reaction mixture was checked through TLC. After evaporation of the solvent under reduced pressure, the crude solid was purified by column chromatography (Scheme 3).

**Biological evaluation of thiazolidinedione analogs**

**In vitro biological evaluation**

**Enzyme preparation**

A purified goat lens extract was prepared in accordance with the method of Hayman and Kinoshita with slight modifications [18]. Lenses were quickly removed from goat eye following euthanasia and homogenized (Glass-Potter) in 5 volume of cold deionized water. The homogenate was centrifuged at 10,400 rpm at 0-4°C for 30 minutes. Saturated ammonium sulfate solution was added to the supernatant fraction to form a 40% solution, which was stirred for 30 minutes at 0-4°C and then centrifuged at 10,400 rpm for 20 minutes. Following this procedure, the recovered supernatant was subsequently fractionated with saturated ammonium sulfate solution using first a 50%, and then a 75% salt saturation. The precipitate recovered from the 75% saturated fraction, possessing ALR2 activity, was redisolved in 0.05 M NaCl and dialyzed overnight in 0.05 M NaCl. The dialyzed material was used for the enzymatic assay.

**Enzymatic assay**

ALR2 activity was assayed at 30°C in a reaction mixture containing 0.75 mL of 10 mM D,L-glyceraldehyde, 0.5 mL of 0.104 mM NADPH, 0.75 mL of 0.1 M sodium phosphate buffer (pH=6.2), 0.3 mL of enzyme extract, and 0.7 mL of deionized water in a total volume of 3 mL. All the above reagents, except D,L-glyceraldehyde, were incubated at 30°C for 10 minutes; the substrate was then added to start the reaction, which was monitored for 5 minutes. Enzyme activity was calibrated by diluting the enzymatic solution to obtain an average reaction rate of 0.011±0.0010 absorbance units/minute for the sample. AR percentage inhibitory activity of the synthesized compounds (15 µL, 5 µg/mL) was determined using same procedure.

**In vivo biological evaluation**

**Induction of noninsulin dependent DM**

The acclimatized animals were kept fasting for 24 hrs with water ad-libitm and 1% W/V in saline of alloxan monohydrate (120 mg/kg i.p) was administrated. After 1 hr of alloxan administration, the animals were given ad-libitm. A 5% dextrose solution was given in feeding bottle for a day to overcome the early hypoglycemic phase. The blood glucose regulator was monitored after alloxination by withdrawing a drop of blood from the tail vein by Tail tipping method. The blood was dropped on the dextrostix reagent pad. The strip was inserted into microprocessor digital blood glucometer and readings were noted.
Experimental design
Antidiabetic activity of the synthesized compounds was carried out in the group of 6 rats. Group 1 for diabetic control (Alloxan induced), Group 2 for reference standard (Rosiglitazone 4 mg/kg body weight) and Group 3-8 for synthesized compounds (15 mg/kg body weight for acute study). Antidiabetic activity of the synthesized compounds was tested using alloxan induced diabetic model in albino rat. The dose of the synthesized compounds (15 mg/kg body weight) and rosiglitazone (4 mg/kg body weight) were administered orally in 2% acacia. The blood glucose level was monitored at different times 0, 1, 3, and 6 hrs, respectively.

RESULT AND DISCUSSION
The analytical data of synthesized compounds are as follows:

(E)-2-(5-(4-benzamidobenzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid G1
Practical yield and M.P. of the compound was found to be 60% and 230-240°C, respectively. The λ_max of the compound was determined in methanol and it was found to be 350 nm. G1 is soluble in DMSO, dimethylformamide (DMF), ethanol, acetone, methanol, and shows Rf value 0.50 in n-hexane:ethyl acetate (3:2) solvent system. IR (cm⁻¹) 2998 (C-H), 1595 and 1505 (aromatic C=C), 1160 (C-O), 1671 (amide C=O), 3397 (O-H); 1H NMR (CDCl₃) (d) 11.12 (s, 1H, O-H), 8.10 (s, 1H, amide NH), 7.55-7.81 (m, 5H, arom. CH; 1H, ethylene CH), 7.28-7.44 (m, 4H, arom. CH), 3.88 (s, 2H, methylene); MS (ESI+): 439.7 [M+Na]+.

(E)-2-(5-(4-benzamidobenzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid G2
Practical yield and M.P. of the compound was found to be 72% and 230°C, respectively. The λ_max of the compound was determined in methanol and it was found to be 356 nm. G2 is soluble in DMSO, DMF, ethanol, acetone, methanol, and shows Rf value 0.52 in n-hexane:ethyl acetate (3:2) solvent system. IR (cm⁻¹) 2984 (C-H), 1597 and 1508 (aromatic C=C), 1174 (C-O), 1666 (amide C=O), 3390 (O-H), 759 (C-Cl); 1H NMR (DMSO) (d) 11.00 (s, 1H, OH), 8.03 (s, 1H, amide NH), 7.71-7.28 (m, 8H, arom. CH; 1H, ethylene CH), 3.91 (s, 2H, methylene); 2.39 (s, 3H, Methyl CH); MS (ESI+): 419.1 [M+Na]+.

(E)-2-(5-(4-(2-chlorobenzamido)benzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid G3
Practical yield and M.P. of the compound was found to be 52.6% and 235-237°C, respectively. The λ_max of the compound was determined in methanol and it was found to be 348.5 nm. G3 is soluble in DMSO, DMF, ethanol, acetone, methanol, and shows Rf value 0.48 in n-hexane:ethyl acetate (3:2) solvent system. IR (cm⁻¹) 2994 (C-H), 1587 and 1505 (aromatic C=C), 1277 (C=O), 1672 (amide C=O), 1600 (O-H); 769 (C=C); 1H NMR (CDCl₃) (d) 11.04 (s, 1H, OH), 8.02 (s, 1H, amide NH), 7.59-7.28 (m, 4H, arom. CH; 1H, ethylene CH), 7.89-7.61 (m, 4H, arom. CH), 3.91 (s, 2H, methylene); MS (ESI+): 439.7 [M+Na]+.

(E)-2-(5-(4-(3,4-dichlorobenzamido)benzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid G4
Practical yield and M.P. of the compound was found to be 53% and 225-227°C (D), respectively. The λ_max of the compound was determined in methanol and it was found to be 335 nm. G4 is soluble in DMSO, DMF, ethanol, acetone, methanol, and shows Rf value 0.52 in n-hexane:ethyl acetate (3:2) solvent system. IR (cm⁻¹) 2984 (C-H), 1597 and 1508 (aromatic C=C), 1267 (C-O), 1718 (C=O), 3390 (O-H), 759 (C=C); 1H NMR (DMSO) (d) 11.04 (s, 1H, OH), 7.61-7.29 (m, 4H, arom. CH; 1H, ethylene CH), 8.05-7.71 (m, 4H, arom. CH; 1H, ethylene CH), 3.91 (s, 2H, methylene); MS (ESI+): 473.1 [M+Na]+.

(E)-2-(5-(4-(2-methylbenzamido)benzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid G5
Practical yield and M.P. of the compound was found to be 56% and 230-232°C (D), respectively. The λ_max of the compound was determined in methanol and it was found to be 339 nm. G5 is soluble in DMSO, DMF, ethanol, acetone, methanol, and shows Rf value 0.52 in n-hexane:ethyl acetate (3:2) solvent system. IR (cm⁻¹) 2965 (C-H), 1603 and 1505 (aromatic C=C), 1174 (C-O), 1666 (amide C=O), 3314 (O-H); 1H NMR (DMSO) (d) 1.10 (s, 3H, CH₃), 8.03 (s, 1H, amide NH), 7.71-7.28 (m, 8H, arom. CH; 1H, ethylene CH), 3.91 (s, 2H, methylene); 2.39 (s, 3H, Methyl CH), MS (ESI+): 419.1 [M+Na]+.

(E)-2-(5-(4-(4-methoxybenzamido)benzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid G6
Practical yield and M.P. of the compound was found to be 70.5% and 235°C (D), respectively. The λ_max of the compound was determined in methanol and it was found to be 347 nm. G6 is soluble in DMSO, DMF, ethanol, acetone, methanol, and shows Rf value 0.44 in n-hexane:ethyl acetate (3:2) solvent system. IR (cm⁻¹) 3077 (C-H), 1593 and 1509 (aromatic C=C), 1165 (C-O), 1677 (amide C=O), 3440 (O-H); 1H NMR (DMSO) (d) 1.10 (s, 3H, CH₃), 8.03 (s, 1H, amide NH), 7.81-7.08 (m, 8H, arom. CH; 1H, ethylene CH), 3.91 (s, 2H, methylene); 3.82 (s, 3H, Methoxy CH); MS (ESI+): 435.2 [M+Na]+.

The structure of synthesized compounds was supported by ¹H NMR, MASS spectral data, and IR findings. All spectral data were in accordance with assumed structures. Spectral study revealed that synthesized compound having anticipated structure (Table 1).

ARI
All the synthesized 2,4-dioxothiazolidin-3-yl acetic acid derivatives were evaluated for their ability to inhibit the in vitro reduction of DL-glyceraldehydes by partially purified ALR from goat lenses; sorbinil was used as a reference drug (Table 2 and Fig. 1). Inhibitory data indicate that 2-substitution on benzamido moiety shows more potent inhibitory activity as compare to un-substituted, 4-substituted or 3,4 di-substituted analogs.

In vivo biological evaluation
All the synthesized thiiazolidinedione also evaluated for their antidiabetic activity using rosiglitazone as reference drug. The decrease in blood glucose level against each compound is shown in Table 3 and 4.
This study has resulted in the identification of (E)-2-(5-(4-(substituted benzamido)benzylidene)-2,4-dioxothiazolidin-3-yl)acetic acid analogs as ARI and antidiabetic agents. These new class of compounds might address the diabetic complications with safety.

REFERENCES


Table 1: Structure, molecular weight, percentage yield, and retardation factor ($R_f$) of synthesized N-acetic acid thiazolidinedione analogs

<table>
<thead>
<tr>
<th>Compound no.</th>
<th>R</th>
<th>MW</th>
<th>% Yield</th>
<th>$R_f$ value</th>
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<tr>
<td>G1</td>
<td>H</td>
<td>383.37</td>
<td>60</td>
<td>0.50</td>
</tr>
<tr>
<td>G2</td>
<td>2-Cl</td>
<td>397.4</td>
<td>55</td>
<td>0.55</td>
</tr>
<tr>
<td>G3</td>
<td>4-Cl</td>
<td>452.26</td>
<td>52.6</td>
<td>0.48</td>
</tr>
<tr>
<td>G4</td>
<td>3,4-dichloro</td>
<td>417.62</td>
<td>72</td>
<td>0.52</td>
</tr>
<tr>
<td>G5</td>
<td>2-CH$_3$</td>
<td>397.4</td>
<td>56</td>
<td>0.52</td>
</tr>
<tr>
<td>G6</td>
<td>4-OCH$_3$</td>
<td>413.4</td>
<td>70.5</td>
<td>0.44</td>
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Table 2: Aldose reductase percentage inhibitory activity of synthesized N-acetic acid-2,4-thiazolidinedione analogs

<table>
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<tr>
<th>Compound</th>
<th>% Inhibition ($\pm$standard)</th>
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<tr>
<td>Standard (Sorbinil)</td>
<td>70±1.1</td>
</tr>
<tr>
<td>G1</td>
<td>57.87±8.25</td>
</tr>
<tr>
<td>G2</td>
<td>61.93±6.42</td>
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<tr>
<td>G3</td>
<td>52.97±6.72</td>
</tr>
<tr>
<td>G4</td>
<td>47.57±7.87</td>
</tr>
<tr>
<td>G5</td>
<td>71.96±1.98</td>
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<tr>
<td>G6</td>
<td>67.46±3.45</td>
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<table>
<thead>
<tr>
<th>Compound</th>
<th>Decrease in blood glucose level mg/dl $\pm$SEM</th>
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<tbody>
<tr>
<td></td>
<td>$^1$ hr</td>
</tr>
<tr>
<td>Control</td>
<td>3.81±2.40</td>
</tr>
<tr>
<td>Standard (roziglitazone)</td>
<td>23.31±21.08</td>
</tr>
<tr>
<td>G1</td>
<td>17.25±3.91*</td>
</tr>
<tr>
<td>G2</td>
<td>28.83±2.46**</td>
</tr>
<tr>
<td>G3</td>
<td>27.80±4.08**</td>
</tr>
<tr>
<td>G4</td>
<td>22.77±2.78**</td>
</tr>
<tr>
<td>G5</td>
<td>23.99±1.72**</td>
</tr>
<tr>
<td>G6</td>
<td>20.50±3.15**</td>
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</tbody>
</table>

$^*$Mean±SEM (n=6); $^p<0.05$; $^{**}p<0.01$. SEM: Standard error of mean.