

## ANTIMICROBIAL ACTIVITY OF SELECTED INDIAN FOLK MEDICINAL PLANTS: *MYRISTICA FATUA*, *ALSTONIA BOONEI*, *HELICTERES ISORA*, *VITEX ALTISSIMA*, AND *ATALANTIA RACEMOSA*

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### ABSTRACT

**Objective:** To determine antimicrobial activity of methanol, ethyl acetate, and acetone extracts (AEs) of *Myristica fatua*, *Alstonia boonei*, *Helicteres isora*, *Vitex altissima*, and *Atalantia racemosa* against different species of pathogens, *Streptococcus faecalis*, *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Bacillus subtilis*, *Staphylococcus aureus*, and *Candida albicans*.

**Methods:** Antimicrobial activity of various plant extracts was measured by agar well diffusion method.

**Results:** AE of *A. boonei* showed the highest inhibitory effect against *E. coli* (20.83±0.32 mm) and *S. faecalis* (19.00±1.00 mm). All the extracts of *H. isora* leaves showed different zone of inhibition observed in all the tested pathogens ranges between 8.13±1.53 and 15.25±1.23 mm. Ethyl acetate extract of *V. altissima* showed highest activity against *B. subtilis* (19.67±1.53 mm). Methanol and acetone leaves extracts of *A. racemosa* have good fungal activity against the *C. albicans* (19.33±1.26-16.00±1.00 mm). Methanol extract of *M. fatua* showed high antimicrobial activity against *P. aeruginosa* (15.10±0.17 mm) and *B. subtilis* (14.23±0.21 mm).

**Conclusion:** The results from the study suggest that the leaves of *M. fatua*, *A. boonei*, *H. isora*, *V. altissima*, and *A. racemosa* showed good antimicrobial activity against the different pathogens. They are used as the alternative source for the control and treatment of microbial infections.

**Keywords:** Antimicrobial activity, Leaves extracts, Well diffusion method, Pathogenic strains.

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### INTRODUCTION

The use of medicinal plants is a very important therapeutic resource for the treatment of diseases and the use of traditional medicine has expanded globally and is gaining popularity. It has continued to be used not only for primary health care of the in developing countries [1]. The most important of these bioactive compounds are flavonoids, stilbenes, and alkaloids are used antimicrobial activity [2].

*Vitex altissima* L.f. (Syn.: *Vitex appendiculata* Rottler; *Vitex zeylanica* Turcz., nom. illeg.) belongs to the family Lamiaceae which includes approximately 3500 species in 220 genera, distributed worldwide, but mostly in the Mediterranean region and South West Asia. The largest gener are (*Salvia* 900), (*Scutellaria* 360), (*Stachys* 300), (*Teucrium* 250), and *Vitex* 250), it is also used in stomatitis, cardiac diseases, anorexia, blindness, leprosy, worm infestation, rheumatic swellings, and chest pain [3]. It also has anti-inflammatory (Sridhar *et al.*, 2004) and antioxidant properties [4].

The plant *Atalantia racemosa* Wight, belonging to the family Rutaceae is commonly called as Kattu naragam in Tamil. It is widely distributed Peninsular India, and Western Ghats - South, Central, and south Maharashtra Sahyadris. *A. racemosa* leaves decotion is used in the treatment of bronchitis, asthma and cough, bronchi and blood purifier [5]. The plant *A. racemosa* is routinely used by tribes around Tamil Nadu to alleviate various diseases [6-8]. In this plant identified important bioactives are campesterol and stigmaterol from fruit extract [9].

*Helicteres isora* L. is belongs to family Sterculiaceae is a sub-deciduous shrub. The species is native to Asia and Australia [10]. It occurs,

throughout India, from Jamuna eastwards to Nepal, Bihar and Bengal and southern India and Andaman Islands. The fruits of *H. isora* is commonly used to the astringent, acrid, refrigerant, demulcent, constipating, stomachic, vermifuge, vulnerary, hemostatic and urinary astringent. Colic, flatulence, diarrhea, dysentery, verminosis, wounds, ulcers, hemorrhages, epistaxis and diabetes [11]. In this plant shows number of bioactive compounds such as saponins, tannins, anthraquinones, alkaloids, triterpens, flavanoids, glycosides, reduced sugar, and phlobatannins [12].

*Alstonia boonei* De Wild. belongs to the family Apocynaceae. It is important medicinal plant used in various purposes to anti-inflammatory, analgesic, and antipyretic activities in Africa [13]. The crucial medicinal plants of genus *Alstonia* includes *Alstonia scholaris*, *Alstonia congensis*, and *Alstonia macrophylla* which have proved to be useful in various diseases by different parts of this plants and it is distributed throughout the tropical and the rain forest of west and Central Africa (Oleiver-Bever, 1986) [13-15]. *A. boonei* shows medicinal effective phytoconstituents are alkaloids and tannins [16].

*Myristica fatua* Houtt. is commonly known as Kottapannu is belongs to the family Myristicaceae. It is fresh water swampy evergreen forests grow up to 600 m. It is distributed in Southern Western Ghats and Endemic to the Western Ghats - Agasthyamalai (West) and Central Malanad. In this plant, commonly used to the antimycobacterial and antiparasitics [17].

This paper mainly focused on the antimicrobial activity of five different medicinal plants against selected bacteria and fungus. These medicinal plants are used in various folk medicines by the local communities in India.

## METHODS

### Collection of plant material

The fresh plants were collected from Sadhuragiri hills, Virudhunagar District, Tamil Nadu, and India. Taxonomic identification of the plants was carried out with the help of Dr. R. Ramasubbu, Assistant Professor, Ganhighram Rural Institute - Deemed University, Gandhigram, Tamil Nadu, India.

### Preparation of leaves extracts

The dried leaves extracts were prepared by sequential extraction method using three organic solvents in the basis of the polarity of solvents (acetone, ethyl acetate, and methanol). 30 g of dried leaf powder sample was taken in a conical flask and 200 ml of acetone was added. The conical flask was kept on mechanical shaker for 24 hrs, after that the extract was filtered through Whatman filter paper 1 and the pellet was allowed for drying and this pellet was used for the next solvent extraction (ethyl acetate and methanol). The dried extract was recovered and stored in refrigerator for further analysis.

### Extract recovery percentage

After drying the respective extracts under oven temperature at 40°C, the percentage of extracts yield was calculated using the following equation.

$$\% \text{Yield} = \frac{\text{Extract} + \text{container (g)} - \text{Empty container (g)}}{\text{Sample weight (g)}} \times 100$$

### Antimicrobial activity

*Staphylococcus aureus*, *Streptococcus faecalis*, *Bacillus subtilis*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, and fungus *Candida albicans*. All samples were procured from Department of Biology, Gandhigram Rural Institute - Deemed University, Gandhigram.

The test organisms were maintained on nutrient agar slant and kept in a refrigerator at 4°C. 100 ml aliquots of nutrient broth were inoculated with the culture of test microorganisms using a loop and then incubated at 37°C for 24 hrs.

Antimicrobial activities of methanol, ethyl acetate, and acetone fractions of plants were carried out using the agar well-diffusion method. Mueller-Hinton agar medium (MHA) was used for antimicrobial susceptibility tests. The MHA medium was prepared by pouring 20 ml of molten media into sterile Petri plates. The plates were allowed to solidify, and 0.2 ml of an overnight broth culture of test microorganisms was added to 20 ml of cooled molten agar on the medium and allowed to dry for 5 minutes. For agar well diffusion method, four equidistant wells (6 mm in diameter) were cut from the agar with the help of a cork-borer. 40 µl of leaves extracts (methanol, ethyl acetate, and acetone extracts [AEs]) containing (4 mg) concentration was loaded on 6 mm well. The standard antibiotic solution gentamicin (10 µg) was placed on the surface of the plates. The plates were kept for incubation for 24 hrs at 37°C. The zone of inhibition was measured around the well-containing samples and standard. The experiments were performed in triplicates.

### Statistical analysis

All the data were reported as mean±standard deviation of three replicates. Statistical analysis was performed using Microsoft Excel.

## RESULTS

### Extract recovery percentage

The extractive values of five different plants in different solvent extracts are given in Table 1. The highest extractive yield was found in the *A. boonei* in methanol extract (ME) and followed ethyl acetate and AE.

### Antimicrobial activity

#### *A. boonei*

Antimicrobial activity of *A. boonei* all tested extracts were activity screened against all tested microorganisms. The results of the zone

of inhibition are summarized in Fig. 1. The AEs showed the inhibitory effect against *E. coli* (21.00±1.00 mm) and *S. faecalis* (19.00±1.00 mm), with compared to positive control. ME shows maximal inhibitory effect against all the test organisms. Ethyl acetate extract (EAE) of *A. boonei* showed significant antimicrobial activity against *B. subtilis* (17.67±0.47 mm). ME of *A. boonei* leaf showed similar inhibition zones were observed against *K. pneumoniae* and *P. aeruginosa* (13.67±0.47 mm). As a result of this plant, extract has high effect of antimicrobial activity.

#### *A. racemosa*

Antimicrobial activity of different leaf extract of *A. racemosa* was assayed against various bacterial and fungal pathogens by agar well diffusion method showed in Fig. 2. The result revealed that all the extracts were found to be significant against all the bacterial and fungal pathogens. It was observed that all the three solvent extracts showed prominent antimicrobial activity between 9.50±0.29 and 20.67±0.76 mm against most of the microorganisms used in the study. The ME of *A. racemosa* showed maximum antimicrobial potential followed by acetone and EAE. The ME of *A. racemosa* was most sensitive against *K. pneumoniae* (20.67±0.76 mm), and least activity was observed against *S. aureus* and *P. aeruginosa* (9.67±0.76 mm). AE was more effective against *C. albicans* (16.00±1.00 mm) than compared to other studied cultures. The EAE of *A. racemosa* showed exhibited zone of inhibition against *E. coli* (12.50±0.29 mm) followed by *S. faecalis* (11.50±0.29 mm), *B. subtilis* and *C. albicans* (11.33±0.63 mm), *K. pneumoniae* and *P. aeruginosa* (10.67±0.54 mm), and *S. aureus* (9.50±0.29 mm).

#### *H. isora*

Different solvent extracts of *H. isora* showed significant antimicrobial activity against all the seven microbial strains in Fig. 3. The AE demonstrated the highest activity followed by the methanol and EAE. The EAE showed the highest activity against *S. faecalis* (14.00±0.00 mm) and minimal activity against *K. pneumoniae* (8.13±0.23 mm). A significant growth inhibition was observed AE against *S. faecalis* (17.00±0.00 mm),

Table 1: Extractive value of plant extracts

| Plants              | ME (% w/w) | EAE (% w/w) | AE (% w/w) |
|---------------------|------------|-------------|------------|
| <i>A. boonei</i>    | 11.33      | 9.66        | 8.06       |
| <i>A. racemosa</i>  | 8.44       | 2.66        | 5.75       |
| <i>H. isora</i>     | 10.03      | 7.75        | 4.96       |
| <i>M. fatua</i>     | 6.03       | 4.16        | 2.66       |
| <i>V. altissima</i> | 9.33       | 8           | 2.83       |

*A. boonei*: *Alstonia boonei*, *A. racemosa*: *Atalantia racemosa*, *H. isora*: *Helicteres isora*, *M. fatua*: *Myristica fatua*, *V. altissima*: *Vitex altissima*, ME: Methanol extract, EAE: Ethyl acetate extract, AE: Acetone extract

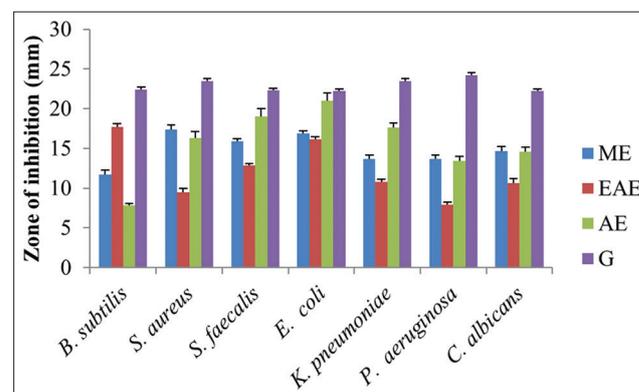
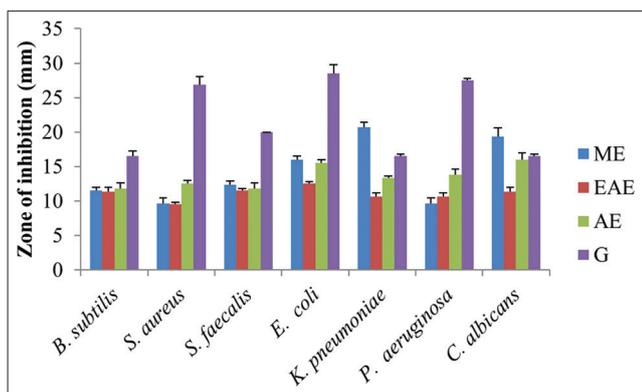
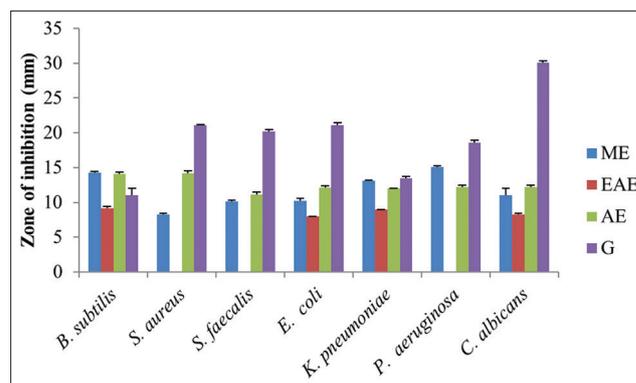


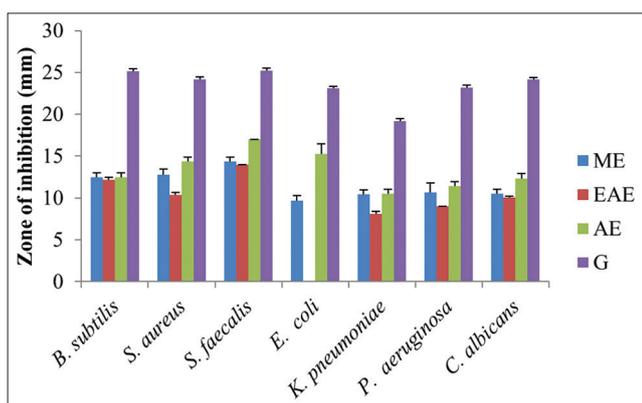
Fig. 1: Antimicrobial activity of *Alstonia boonei* leaf extracts. Each value represents mean±standard deviation of three replicates. ME - Methanol extract, EAE - Ethyl acetate extract, AE - Acetone extract, G - Gentamicin, plant extracts - 4 mg (0 - No inhibition)



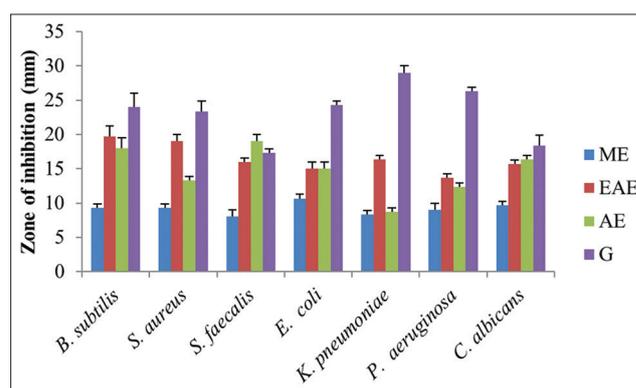
**Fig. 2: Antimicrobial activity of *Atalantia racemosa* leaf extracts.** Each value represents mean±standard deviation of three replicates. ME - Methanol extract, EAE - Ethyl acetate extract, AE - Acetone extract, G - Gentamicin, plant extracts - 4 mg (0 - No inhibition)



**Fig. 4: Antimicrobial activity of *Myristica fatua* leaf extracts.** Each value represents mean±standard deviation of three replicates. ME - Methanol extract, EAE - Ethyl acetate extract, AE - Acetone extract, G - Gentamicin, plant extracts - 4 mg (0 - No inhibition)



**Fig. 3: Antimicrobial activity of *Helicteres isora* leaf extracts.** Each value represents mean±standard deviation of three replicates. ME - Methanol extract, EAE - Ethyl acetate extract, AE - Acetone extract, G - Gentamicin, plant extracts - 4 mg (0 - No inhibition)



**Fig. 5: Antimicrobial activity of *Vitex altissima* leaf extracts.** Each value represents mean±standard deviation of three replicates. ME - Methanol extract, EAE - Ethyl acetate extract, AE - Acetone extract, G - Gentamicin, plant extracts - 4 mg (0 - No inhibition)

*E. coli* (15.25±1.23 mm) and *S. aureus* (14.33±0.58 mm). There is no any activity of EAE against *E. coli* (0.00±0.00 mm). They results MEs showed poor response against *E. coli* (9.67±0.58 mm).

**M. fatua**

*M. fatua* showed broad spectrum of antimicrobial activity in all tested microorganisms. These results suggest that all the three solvent extracts possess antibacterial and antifungal activity against such as *E. coli*, *P. aeruginosa*, *S. aureus*, *B. subtilis*, *K. pneumoniae*, *S. faecalis* and *C. albicans* showed in Fig. 4. Different solvent extracts of *Myristica fragrans* showed significant activity against all the seven microbial strains. The ME exhibited highest activity against *P. aeruginosa* with the diameter of zone of inhibition of 15.10±0.17 mm and *B. subtilis* (14.23±0.21 mm) respectively, and least activity was noted against *S. aureus* (8.23±0.25 mm). AE more potent effect against *P. aeruginosa* (12.17±0.29 mm) and least activity against *S. faecalis* (11.17±0.29 mm). Similarly, EAE was found to be active with the zone of inhibition against *B. subtilis* (9.17±0.29 mm), *K. pneumoniae* (9.00±0.00 mm), *E. coli* (8.00±0.00 mm) and *C. albicans* (8.23±0.21 mm) and loss of activity against *P. aeruginosa*, *S. aureus* and *S. faecalis* (0.00±0.00).

**V. altissima**

Agar well diffusion method was used to assess the antimicrobial activity against the human affected pathogens by measuring the zone of inhibition (mm). Most of the extracts showed significant antimicrobial activity against the tested organisms at the same concentration of

4 mg/ml (Fig. 5). In general, AE demonstrated higher antimicrobial activity than the other solvent extracts. The ethyl acetate leaves extract was the most effective against the tested organisms. It recorded the highest zone of inhibition of 19.67±1.53 mm against *B. subtilis* and 19.00±1.00 mm against *S. aureus* at 4 mg/ml. This is significant activity compared with the activity of the standard gentamicin which showed zones of inhibition ranges from 17.33±0.58 to 29.00±1.00 mm. The results of antimicrobial activity are given in which clearly show that the EAE of *V. altissima* has both antibacterial and antifungal activity against the tested organisms.

**DISCUSSION**

The presence of phytochemicals in the plant extracts highly correlated to the biological activity [18]. From this revealed significant antimicrobial activity possess in the all the studied plant extracts. Nowadays, number researcher evaluated the antimicrobial compounds from plant material, they act as lesser side effect in the human body. The present study corroborates with the antimicrobial activity of various plant extracts against *S. aureus* and *E. coli* [18]. In earlier researcher reported the ME produce moderate antimicrobial activity against the microbial pathogens [19]. AE of *M. fragrans* seed extract showed antibacterial and antifungal activity against *S. aureus* (13.8±0.42 mm) and *Aspergillus niger* (14.4±0.37 mm) with zone of respectively for 10 µg/ml of extract solution [20]. In earlier record showed ethanol root extract of *A. boonei* exhibited inhibition against various pathogens such as *S. aureus* (8-10 mm), *B. subtilis* (6-8 mm), *P. aeruginosa* (5-8 mm), *E. coli* (5-8 mm) and fungus *C. albicans* (5-9 mm) with volume of extract solutions ranging from 2.5 to 10 mg/ml [21]. In previous study reported

contrary result on MEs of *H. isora* leaf showed antimicrobial activity against *E. coli* (15.28 mm) and fungus *A. niger* (11.30 mm) showed good zone of inhibition and concentration was (10 mg/ml) [22]. The present study coincides with the antimicrobial activity of aqueous extracts of *V. altissima* [23]. *Atalantia* species showed good antimicrobial activity against the tested pathogens are *Atalantia hydrophila* followed by *Proteus mirabilis*, *P. aeruginosa*, *Proteus vulgaris* and *E. coli* while our findings also exhibited the same results against *P. aeruginosa* and *E. coli* [24]. In this study, showed good zone of inhibition against all the tested pathogens and activities were compared with positive control gentamicin.

## CONCLUSION

The significant inhibitory activity of all the tested extracts *M. fatua*, *A. boonei*, *H. isora*, *V. altissima* and *A. racemosa* was noted against different pathogenic microorganisms. These five plant extracts could be studied further as future control the common pathogenic microbes. Phytochemical analysis could be carried out to isolate the bioactive compounds of these plant species, which act as antimicrobial agents.

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