

for close contacts, can be used to model a slight “give” in the receptor and/or ligand. Docking studies of designed compounds were carried out using grid-based ligand docking with energetics (GLIDE) module version 5.9. Schrödinger, LLC, New York, NY, 2013. The software package running on multiprocessor Linux PC. GLIDE has previously been validated and applied successfully to predict the binding orientation of many ligands.

Protein structure preparation

The X-ray crystal structures of VP24 (PDB: 4U2X) protein of EBOV retrieved from the RCSB Protein Data Bank. Water molecules of crystallization were removed from the complex, and the protein was optimized for docking using the protein grounding and refinement utility provided by Schrödinger LLC.

Ligand structure preparation

The structures of energetic constituents of *A. indica* (Fig. 1) were constructed by means of the splinter dictionary of Maestro 9.3 (Schrodinger, LLC) using the optimized potentials for liquid simulations—all atom force field with the steepest descent followed by curtailed Newton conjugate gradient protocol.

Docking protocol

All docking calculations were performed using the “extra precision” mode of GLIDE program. For the binding site, an assortment of energy grids was premeditated and stored, is distinct in terms of two concentric cubes: The bounding box, which must contain the center of any satisfactory ligand pose, and the enclosing box, which must contain all ligand atoms of an satisfactory pose, with a root mean square deviation of <0.5 Å and a maximum atomic displacement of <1.3 Å were eliminated as unneeded to increase assortment in the retained ligand poses. The scale factor for van der Waals radii was applied to those atoms with absolute partial charges ≤ 0.15 (scale factor of 0.8) and 0.25 (scale factor of 1.0) electrons for ligand and protein, respectively. Energy minimization protocol includes dielectric constant of 4.0 and 1000 steps of conjugate gradient. Upon end of each docking calculation, for the most part, 100 poses per ligand were generated. The most excellent docked structure be preferred using a GLIDE score (G-score) function [17,18].

$$\text{Glide Score} = 0.065 * \text{vdW} + 0.130 * \text{Coul} + \text{Lipo} + \text{Hbond} + \text{Metal} + \text{BuryP} + \text{RotB} + \text{Site}$$

RESULTS AND DISCUSSION

The docking investigation was completed for the ligands with the target protein VP24 by means of the docking software GLIDE, and the docked images are given away in Figs. 2-7. The structures docked by GLIDE are usually ranked according to the GLIDE scoring function (more negative). The scoring function of GLIDE docking program is offered in the G-score form. The most clear-cut method of evaluating the precision of a docking procedure is to determine how intimately the lowest energy pose (binding conformation) predicted by the object scoring function. To study the molecular basis of interaction and likeness of binding of ligands to VP24 protein, all the ligands were docked into the active site of VP24. The docking result of these ligands is prearranged in Table 1.

The interaction energy includes van der Waals energy, electrostatic energy, as well as intermolecular hydrogen bonding were calculated for each minimized complex. The docking score by means of GLIDE varied from -1.54 to -7.95 against VP24. The GLIDE score for a standard brincidofovir docked with was -6.06. Naturally occurring products catechin, epicatechin, gallic acid, and nimbolide are potential than the standard brincidofovir and favipiravir, but azadirachtin, margolonone, mahmoodin, isomargolonone, gedunin, margolone, nimbidin, and nimbin have low binding likeness toward the target when compared with the standard. This proves that analogs of catechin, epicatechin, gallic acid, and nimbolide could be impending drugs for the target VP24 in antiviral drug development for EBOV. In generally, for low

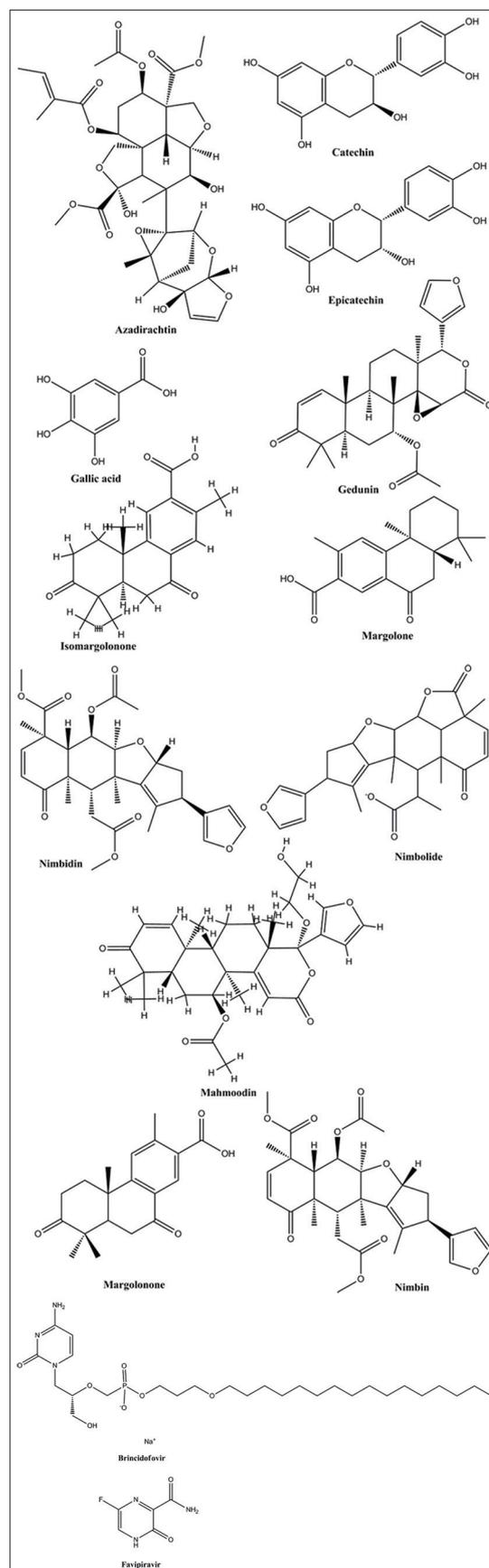


Fig. 1: Ligand structures from *Azadirachta indica*

GLIDE score, good ligand affinity to the receptor may perhaps expect. Catechin and epicatechin be evidence for the most excellent inhibition

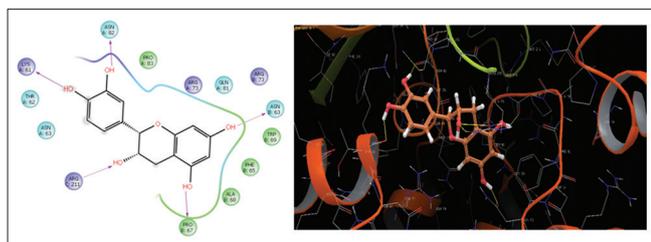


Fig. 2: Ligand interaction of catechin with VP24 (PDB: 4U2X) protein

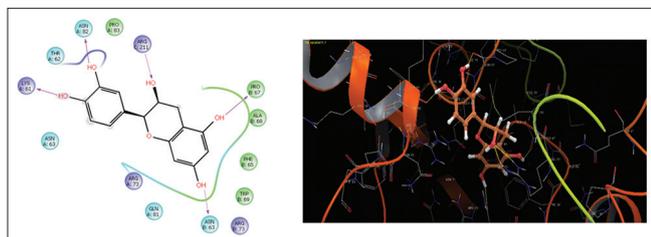


Fig. 3: Ligand interaction of epicatechin with VP24 (PDB: 4U2X) protein

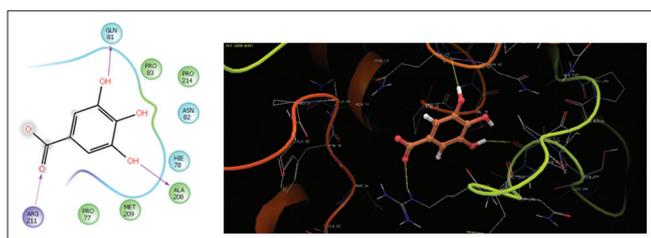


Fig. 4: Ligand interaction of gallic acid with VP24 (PDB: 4U2X) protein

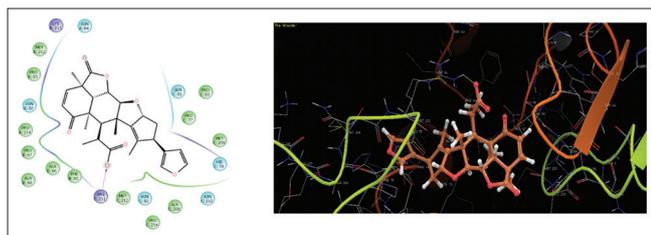


Fig. 5: Ligand interaction of nimbolide with VP24 (PDB: 4U2X) protein

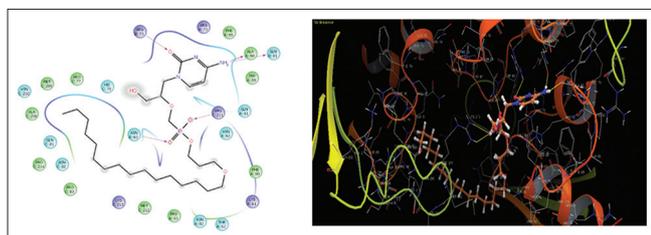


Fig. 6: Ligand interaction of brincidofovir with VP24 (PDB: 4U2X) protein

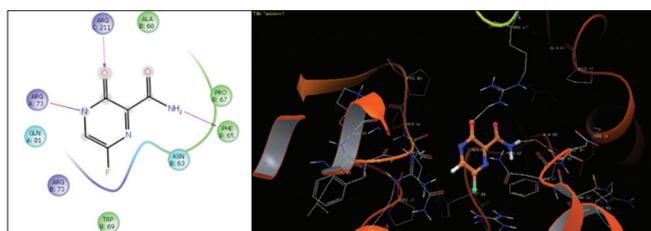


Fig. 7: Ligand interaction of favipiravir with VP24 (PDB: 4U2X) protein

Table 1: Glide score for the docked ligands

S. No.	Compound	Glide score
1	Catechin	-7.95
2	Epicatechin	-7.95
3	Gallic acid	-6.37
4	Nimbolide	-6.09
5	Brincidofovir (standard)	-6.06
6	Azadirachtin	-5.05
7	Margolonone	-4.96
8	Mahmoodin	-4.31
9	Isomargolonone	-3.74
10	Favipiravir (Standard)	-3.52
11	Gedunin	-3.36
12	Margolone	-3.26
13	Nimbidin	-1.54
14	Nimbin	-1.54

for the VP24 with -7.95 glide score against VP24 protein receptor. We found a fantastic agreement between the localization of the inhibitor on docking and from the crystal structure of the protein. Conformational analysis of different docked complexes also shows that residues LYS 61, ASN 63, ARG211, ASN 82, ARG 73, ALA 68, and GLN 81 for VP24 protein play imperative role in this receptor's activity. Docking studies performed by GLIDE have inveterate that above inhibitors fit into the binding pocket of the VP24 receptor. From the results, we may monitor that for successful docking, intermolecular hydrogen bonding and lipophilic interactions between the ligand and the receptor are very significant. The main reason for the increase in GLIDE score is due to the penalties for close intraligand contacts.

CONCLUSION

In conclusion, we have a notorious molecules catechin, epicatechin, gallic acid, and nimbolide, an inventive drug candidate that was docked against VP24 protein in a premeditated attempt to ascertain a new drug candidate, which is able to obstruct with diverse key target points of VP24 in treating EBOV. This compound catechin, epicatechin, gallic acid, and nimbolide well calculated as a best (lead) molecule and we necessitate design analogs, synthesis and evaluate its effectiveness against viral disease caused by EBOV through the molecular level and *in vivo* studies.

REFERENCES

- De Clercq E. Antiviral drugs in current clinical use. *J Clin Virol* 2004;30(2):115-33.
- Hupfeld J, Efferth T. Review. Drug resistance of human immunodeficiency virus and overcoming it by natural products. *In vivo* 2009;23(1):1-6.
- Ghosh T, Chattopadhyay K, Marshall M, Karmakar P, Mandal P, Ray B. Focus on antivirally active sulfated polysaccharides: From structure-activity analysis to clinical evaluation. *Glycobiology* 2009;19(1):2-15.
- Ginsberg HS, Goebel WF, Horsfall FL Jr. Inhibition of mumps virus multiplication by a polysaccharide. *Proc Soc Exp Biol Med* 1947;66(1):99.
- Branswell H. Ebola war. *Sci Am* 2015;312:42-9.
- WHO Ebola Response Team, Aylward B, Barboza P, Bawo L, Bertherat E, Bilivogui P, et al. Ebola virus disease in West Africa - The first 9 months of the epidemic and forward projections. *N Engl J Med* 2014;371(16):1481-95.
- Murin CD, Fusco ML, Bornholdt ZA, Qiu X, Olinger GG, Zeitlin L, et al. Structures of protective antibodies reveal sites of vulnerability on Ebola virus. *Proc Natl Acad Sci U S A* 2014;111(48):17182-7.
- Kugelman JR, Sanchez-Lockhart M, Andersen KG, Gire S, Park DJ, Sealfon R, et al. Evaluation of the potential impact of Ebola virus genomic drift on the efficacy of sequence-based candidate therapeutics. *MBio* 2015;6(1):pii:E02227-14.
- Mateo M, Carbonnelle C, Reynard O, Kolesnikova L, Nemirov K, Page A, et al. VP24 is a molecular determinant of Ebola virus virulence in guinea pigs. *J Infect Dis* 2011;204 Suppl 3:S1011-20.
- Reid SP, Leung LW, Hartman AL, Martinez O, Shaw ML, Carbonnelle C,

- et al. Ebola virus VP24 binds karyopherin alpha1 and blocks STAT1 nuclear accumulation. *J Virol* 2006;80(11):5156-67.
11. Subapriya R, Nagini S. Medicinal properties of neem leaves: A review. *Curr Med Chem Anticancer Agents* 2005;5(2):149-6.
 12. Biswas K, Chattopadhyay I, Banerjee RK, Bandyopadhyay U. Biological activities and medicinal properties of neem (*Azadirachta indica*). *Curr Sci* 2002;82:1336-45.
 13. Badam L, Joshi SP, Bedekar SS. 'In vitro' antiviral activity of neem (*Azadirachta indica*. A. Juss) leaf extract against group B coxsackieviruses. *J Commun Dis* 1999;31(2):79-90.
 14. Saha S, Galhardi LC, Yamamoto KA, Linhares RE, Bandyopadhyay SS, Sinha S, et al. Water-extracted polysaccharides from *Azadirachta indica* leaves: Structural features, chemical modification and anti-bovine herpesvirus Type 1 (BoHV-1) activity. *Int J Biol Macromol* 2010;47(5):640-5.
 15. Tiwari V, Darmani NA, Yue BY, Shukla D. *In vitro* antiviral activity of neem (*Azadirachta indica* L.) bark extract against herpes simplex virus Type-1 infection. *Phytother Res* 2010;24(8):1132-40.
 16. SaiRam M, Ilavazhagan G, Sharma SK, Dhanraj SA, Suresh B, Parida MM, et al. Anti-microbial activity of a new vaginal contraceptive NIM-76 from neem oil (*Azadirachta indica*). *J Ethnopharmacol* 2000;71:377-82.
 17. Parasuraman P, Suresh R, Premnath D. Balancing anti-amyloid and anti-cholinesterase capacity in a single chemical entity: *In silico* drug design. *Int J Pharm Pharm Sci* 2014;6(2):571-4.
 18. Amudha M, Rani S. *In silico* molecular docking studies on the phytoconstituents of *Cadaba fruticosa* (L.) Druce for its fertility activity. *Asian J Pharm Clin Res* 2016;9(2):48-50.