FORMULATION AND IN-VITRO-IN-VIVO EVALUATION OF ALGINATE-CHITOSAN MICROSPHERES OF GLIPIZIDE BY IONIC GELATION METHOD

MAYA SHARMA*, CHOUDHURY PK, SURESH KUMAR DEV
Department of Pharmaceutical Sciences, Mohanlal Sukhadia University, Udaipur - 313 001, Rajasthan, India.
Email: maya.sharma1703@gmail.com

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INTRODUCTION
The drug delivery systems that can precisely control the release rates or target drugs to specific body site have an enormous impact on the health-care system. The past two decades have witnessed an advancement in the field of polymer and material science, resulting in the development of novel drug delivery systems [1]. Microspheres acquire important features among the particulate drug delivery systems by virtue of their small size and efficient carrier characteristics. However, the success of microspheres delivery system is less due to their short residence time at the site of absorption [2]. The physicochemical characteristics of the active chosen vary considerably, so microspheres are often developed according to specific clinical needs [3]. Microspheres were prepared by the emulsion-gelation method for their ease and cost-effectiveness [4].

Diabetes mellitus (DM) is a chronic metabolic disorder affecting people worldwide, with significant morbidity and mortality caused by its microvascular and macrovascular complications, affecting various vital organs and structures in humans [5]. It has been estimated that by the year 2030, the diabetic population will rapidly increase. However, prevalence is much more, as many patients are asymptomatic and go undiagnosed. Glipizide is an antidiabetic drug used to treat Type II DM. Glipizide with a half life of 2-3 hrs was converted in sustained release (SR) formulation to reduce dosing frequency. A model dosage form is one, which attains the desired therapeutic concentration of drug in plasma and maintains it for the entire duration of treatment. The absorption of glipizide is reduced, when taken after meals hence the oral SR was developed as this formulation released the drug slowly into the GIT and maintained a stable drug concentration in the serum for a longer duration of time.

Conclusion: Drugs with short half life that are absorbed from the gastrointestinal tract (GIT) are eliminated rapidly from the blood flow. To avoid this, the oral SR was developed as this formulation released the drug slowly into the GIT and maintained a stable drug concentration in the serum for a longer duration of time.

Keywords: Glipizide, Sodium alginate, Chitosan, Sustained release microspheres, Scanning electron microscopy, Differential scanning calorimetry, In-vitro and in vivo activity.

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CONCLUSION
Drugs with short half life that are absorbed from the gastrointestinal tract (GIT) are eliminated rapidly from the blood flow. To avoid this, the oral SR was developed as this formulation released the drug slowly into the GIT and maintained a stable drug concentration in the serum for a longer duration of time.

Keywords: Glipizide, Sodium alginate, Chitosan, Sustained release microspheres, Scanning electron microscopy, Differential scanning calorimetry, In-vitro and in vivo activity.
up exclusively of one unit or the other, referred to as M-blocks or G-blocks, or as regions in which the monomers forms an alternating sequence. The calcium reactivity of alginates is a consequence of the particular molecular geometries of each of these regions [10]. Sodium alginate is capable of forming rigid gels by the action of calcium ion or multivalent cations. It is relatively easy to describe alginates in terms of M and G units, but the detailed molecular compositions of alginates in terms of block lengths and block distributions are quite difficult to determine [11].

The chitosan entrapped calcium pectinate microspheres have been used for SR of drugs or for targeting drugs to the colon [12].

**METHODS**

Glipizide was obtained as a gift sample from by Kreative Organics, Hydendbad. Chitosan from Kerala State chitosan plant, Kerela, and sodium alginate was purchased from CDH, New Delhi. All other chemicals used were of analytical grade.

**Preparation of microspheres**

**Glipizide chitosan-alginate microspheres**

The required quantity of sodium alginate was dissolved in distilled water (30 ml). Calculated quantity of the drug was added and homogenized. Chitosan in required proportion was dissolved in 2% v/v acetic acid with pH (5.5) adjusted with 10% NaOH solution. 5% calcium chloride dehydrate solution was added to it. Drug alginate mixture was added to the chitosan-calcium chloride solution dropwise at a constant rate of 30 ml/h with gentle stirring over thermal controlled magnetic stirrer. The formed microspheres were filtered, washed with acetone and dried in an oven at 35°C. The dried formulations were stored in an amber colored bottle and kept in a desiccator until used (Table 1).

**Percentage yield**

The percentage yield of the microspheres was calculated for each batch by dividing the weight of microspheres by the total weight of drug and polymer.

\[
\text{Percentage yield} = \frac{\text{Practical yield}}{\text{theoretical yield}} \times 100
\]

**Size distribution and size analysis**

Glipizide chitosan microspheres were separated into different size fractions by sieving for 10 minutes using a mechanical shaker containing standard sieves as per Indian Pharmacopeia specifications. The particle size distribution was determined and means particle size of gel beads was calculated by the following formula and shown in Fig. 2.

\[
\text{Mean particle size} = \frac{\sum (\text{Mean particle size of the fraction} \times \text{weight fraction})}{\sum \text{Weight fraction}}
\]

Table 1: Formulation process codes and parameter for the formulation of the glipizide microspheres by ionic gelation method

<table>
<thead>
<tr>
<th>Code</th>
<th>Drug &amp; Sod. alginate</th>
<th>Chitosan</th>
<th>Distilled water</th>
<th>CaCl₂ (5%)</th>
<th>D:P</th>
</tr>
</thead>
<tbody>
<tr>
<td>GCS1</td>
<td>100 mg +</td>
<td>+</td>
<td>+</td>
<td>30 ml</td>
<td>100 ml</td>
</tr>
<tr>
<td>GCS2</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>GCS3</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>GCS4</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>GCS5</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>GCS6</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>GCS7</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

D:P: Drug to polymer ratio, +: Present

**Differential scanning calorimetry (DSC)**

Thermograms of drug, polymers, and microspheres were obtained using DSC 822e (Mettler Toledo) calorimeter. DSC measures the amount of heat energy absorbed or released by a sample, as it is heated, cooled or held at a constant temperature. Thermogram shows that no interaction seen between polymers and also between the polymer and drug. The drugs and polymers showed separate peaks in thermogram (Fig. 3).

**Scanning electron microscopy (SEM)**

The samples for the SEM analysis were prepared by sprinkling the microspheres on one side of the double adhesive stub. The stub was then coated with fine gold dust. The microspheres were then observed under the SEM (JEOL Model JSM-6390LV) at 15 kV. It showed discreet, more and less uniform and uniformly distributed microspheres (Fig. 4).

**Entrapment efficiency**

The prepared microspheres were dissolved, and the amount of drug present in weighed samples was determined by analyzing spectrophotometrically at 276 nm after filtration and suitable dilutions [13]. The method obeyed Beer’s law in the concentration range of 0-35 µg/ml. The entrapment efficiency was calculated from the theoretical amount and actual amount of the drug in dry microspheres.

\[
\text{Drug entrapment efficiency (\%)} = \frac{\text{Calculated drug content - theoretical drug content}}{\text{theoretical drug content}} \times 100
\]

**Swelling index**

Swelling index was determined by measuring the extent of swelling of microspheres in a particular buffer solution. To ensure complete equilibrium weighed, the amount of microspheres was allowed to swell in a buffer solution. The excess liquid adhering to the drops were removed by blotting, and the swollen microspheres were weighed using microbalance. The hydrogel microspheres were then dried in an oven at 60°C for 5 hrs until there was no change in the dried mass of the sample. The swelling index of the microsphere was calculated by the following formula [14].

\[
\frac{\text{Mass of swollen microspheres} - \text{Mass of dry microspheres}}{\text{Mass of dried microspheres}} \times 100
\]

![Fig. 1: Chemical structure of glipizide](image1)

![Fig. 2: Size distribution of sodium alginate microspheres of drug glipizide (Drug to polymer-1: 0.5-1:2)](image2)

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Mucoadhesive studies
Approximately 50 microspheres were taken and spread uniformly over a wet glass slide, which was held to the walls of the beaker with glue. This assembly was then introduced into the USP disintegration apparatus. A number of microspheres still adhering to the glass slide was counted at regular intervals and assessed for the mucoadhesive nature of the microspheres. All the readings were taken in triplicate and results depicted in ± standard deviation (SD) (Table 2).

In-vitro release studies
Preparation of standard plot in methanol
Glipizide (50 mg) was dissolved in 100 ml of methanol. It was then suitably diluted for graded solutions in a range of 0-35 µg/ml. The absorbance was read using an ultraviolet (UV) spectrophotometer (Shimadzu 1800) at λ\text{max} 276 nm (Table 3 and Fig. 5).

In-vitro release of microspheres
The in-vitro release studies were conducted at 37±0.5°C and at 100 rpm by buffer change method using 0.1 N HCl (1 hrs), 4 pH (1 hrs), 6 pH (3 hrs), 6.8 pH (3 hrs), and 7.4 pH (2 hrs) phosphate buffers (200 ml) in sink conditions using a diffusion cell. Accurately weighed sample of prepared microspheres was added to the donor cell. At preset time intervals; 5 ml of aliquots were withdrawn and replaced by an equal volume of fresh dissolution medium. The aliquots were analyzed UV spectrophotometrically at λ\text{max} 276 nm after proper dilution (Table 4 and Fig. 6).

In-vivo hypoglycemic activity
The in-vivo hypoglycemic activity was conducted in healthy and hyperglycemia-induced male albino Wistar rats of 175±25 g body weight by measuring the hyperglycemic effect produced after oral administration of a microspheres dose equivalent to 5 mg/kg body weight of glipizide in comparison to the pure drug at the same dose.

All animal experiments were approved by Institutional Animal Ethics Committee (IAEC) of Pinnacle Biomedical Research Institute (PBRI) Bhopal (Reg. No. 1824/PO/ERs/S/15/PCBRA). Protocol Approval Reference No. PBRI/IAEC/PN-16042. Animals were housed in separate cages under controlled conditions of temperature (22±2°C). All animals were given standard diet (golden feed, New Delhi) and water regularly [15].

Animals were kept at 12 hrs in alternating light and dark cycles. Animals were further divided into eight groups with six animals in each group (Table 5) [16].
The treatment was started on the 4th day after the STZ injection, considering it as the 1st day of treatment. The treatment was continued till 7 days.

### Induction of diabetes
1. Diabetes was induced in rats by intraperitoneal injection of streptozotocin (STZ) at a dose of 60 mg/kg body weight. STZ dissolved in ice cold 0.1 M citrate buffer at 4°C.
2. The animals were allowed to drink 5% glucose solution overnight to overcome STZ induced hyperglycemia.
3. The animals were considered diabetic, if their blood glucose value was above 200 mg/dl on the 3rd day of STZ injection.
4. The treatment was started on the 4th day after the STZ injection, considering it as the 1st day of treatment. The treatment was continued till 7 days.
5. Blood glucose level and body weight were observed on 0, 3, 5, and 7th day of post-treatment (Table 6).

### Experimentation
All animals were divided randomly into six groups with six animals in each group. Group I, received normal saline orally only and served as vehicle control, Groups II, III, IV, V, VI, and VII were made diabetic with STZ. Group II received saline, Groups III and IV received glipizide (5 mg/kg) per day oral, Groups V, VI, and VII received formulation GCS3 (5 mg/kg) per day oral. All dosing of test samples was done orally throughout the experimentation (Table 7 and Fig. 7).

### Stability studies of formulations
Stability testing is an integral part of the formulation development. It generates information about the shelf life of drug substances and their formulations and recommends appropriate guidelines for storage [17,18]. In the formulations GCS1-GCS7, there was no significant change in stability. Loss of drug content was seen in formulations with low polymer concentrations.

### Morphological characteristics and release profile
The formulated microspheres were stable throughout the course of study of about (90 days) at 5°C in the freezer; at room temperature (32°C) and at 45°C in oven [19,20]. There was no significant morphological change or changes in release characteristics. Slight color changes were observed at elevated temperature because of degradation of polymers.

### RESULT AND DISCUSSION
The effects of process variables on various physicochemical, morphological and in-vitro release characteristics were studied. The low coefficient of variations showed that the method is highly reproducible.

Microspheres of glipizide were formulated using sodium alginate and chitosan by orifice ionic gelation method. Different concentrations of the polymer were used to entrap the drug, and various process variable parameters were analyzed.

The microspheres were discrete, free flowing and monolithic matrix type. A coat of chitosan was also produced on the surface of the alginate microparticles.

The mucoadhesive property of the microspheres increased with increase in the concentration of polymer. The size distribution was in the range of 200-500 µm. It was observed that increase in the polymer ratio, increased the size of the microspheres. Incorporation of more polymers resulted in increased size due to the distribution of the insoluble drugs in the interstitial spaces of the matrix.

It was observed that higher proportion of polymer in the formulation resulted in the microspheres being smoother. The drug release rate decreased with increase in chitosan concentration. Microspheres showed the inherent brown color of the polymers. When the proportion of polymer was increased, the percentage yield and the drug entrapment also improved.

Drug release characteristics were studied in simulated gastric and intestinal fluid without pepsin to simulate the gastrointestinal environment. Release was slow and uniformly spread over a period of time. This could be attributed to the matrix nature of the formulation. Poor water soluble drugs seem to rush initially into the interstitial spaces of the polymer, from where the water phase forces the drug into the gel portion of the polymer. The swelling hydrophilic membrane then acts as a flux controlling barrier [21,22]. Thus, alginate microspheres with chitosan, prepared by simple ionic gelation method, could be used for sustain release of the drug [23]. The drug release was diffusion as well as erosion controlled.

### Table 2: Physicochemical properties of the glipizide chitosan-alginate microspheres (D:P=1:0.5-1:2), n=3

<table>
<thead>
<tr>
<th>Code</th>
<th>% yield</th>
<th>% entrapment±SD</th>
<th>Shape</th>
<th>Color</th>
<th>% Swelling index</th>
<th>Mucoadhe. property (%±SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>GCS1</td>
<td>90.6</td>
<td>87.1±0.10</td>
<td>Spherical</td>
<td>Light brown</td>
<td>11.5</td>
<td>49.1±0.60</td>
</tr>
<tr>
<td>GCS2</td>
<td>85.2</td>
<td>84.4±0.15</td>
<td>Spherical</td>
<td>Light brown</td>
<td>15.7</td>
<td>63.0±1.50</td>
</tr>
<tr>
<td>GCS3</td>
<td>84.3</td>
<td>89.6±0.10</td>
<td>Spherical</td>
<td>Light brown</td>
<td>17.2</td>
<td>65.2±1.14</td>
</tr>
<tr>
<td>GCS4</td>
<td>92.0</td>
<td>88.4±0.10</td>
<td>Spherical</td>
<td>Light brown</td>
<td>19.0</td>
<td>69.5±1.52</td>
</tr>
<tr>
<td>GCS5</td>
<td>91.1</td>
<td>72.1±0.07</td>
<td>Spherical</td>
<td>Light brown</td>
<td>20.5</td>
<td>72.5±0.45</td>
</tr>
<tr>
<td>GCS6</td>
<td>85.1</td>
<td>89.1±0.28</td>
<td>Spherical</td>
<td>Light brown</td>
<td>24.5</td>
<td>81.3±0.43</td>
</tr>
<tr>
<td>GCS7</td>
<td>83.3</td>
<td>82.0±0.10</td>
<td>Spherical</td>
<td>Light brown</td>
<td>27.0</td>
<td>88.2±0.50</td>
</tr>
</tbody>
</table>

SD: Standard deviation

### Table 3: Glipizide pure drug absorbance in UV spectrophotometer

<table>
<thead>
<tr>
<th>S.No.</th>
<th>Concentration (µg/ml)</th>
<th>Absorbance at (276 nm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>2</td>
<td>5</td>
<td>0.115</td>
</tr>
<tr>
<td>3</td>
<td>10</td>
<td>0.238</td>
</tr>
<tr>
<td>4</td>
<td>15</td>
<td>0.368</td>
</tr>
<tr>
<td>5</td>
<td>20</td>
<td>0.482</td>
</tr>
<tr>
<td>6</td>
<td>25</td>
<td>0.599</td>
</tr>
<tr>
<td>7</td>
<td>30</td>
<td>0.714</td>
</tr>
<tr>
<td>8</td>
<td>35</td>
<td>0.824</td>
</tr>
</tbody>
</table>

UV: Ultraviolet

---

**Fig. 7:** % reduction in plasma glucose level in hyperglycemic rats after administration of pure drug and formulation GCS3

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**Table 3: Glipizide pure drug absorbance in UV spectrophotometer**
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Statistical analysis of in-vivo data

All data were analyzed by one-way ANOVA followed by Bonferroni test. p<0.05 was considered as the level of significance. All data are presented in mean±SD.

Glipizide showed a significant decrease in plasma glucose levels in both normal and hyperglycemic rats.

A 25% reduction in glucose is considered a significant hypoglycemic effect. The sustained hypoglycemic effect with microspheres is due to the slow release and absorption of glipizide over extended periods of time.

The plasma glucose level of diabetic controlled rats increased significantly from day 3 to day 7 of STZ injection. Hyperglycemic rats were selected for the study as discussed above. The plasma glucose data obtained indicates that the drug is entrapped in microspheres and produced the consistent antihyperglycemic effect. This effect was pronounced in the case of hyperglycemic rats whereas normal rats showed comparatively lesser alterations in plasma glucose level after formulation administration.

CONCLUSION

A new SR system of glipizide microspheres of alginate-chitosan was designed and formulated by an ionic gelation method. It’s morphological, and release characteristics were studied. The microspheres were easy to prepare, and the mean diameter of microspheres increased with increase in the amount of the polymers increase. The pore size of microspheres was affected by the concentration of the alginate and chitosan. Stirring at high speed above 200 rpm causes the destruction of microspheres. The microspheres showed excellent in-vivo activity and SR characteristics as compared to the conventional oral dosage forms. Thus, drug entrapment technique is a useful tool for the development of multiparticulate system even for a water-insoluble drug.

ACKNOWLEDGMENT

Authors are grateful to Sophisticated Test and Instrumentation Centre, Cochin University, Kochi, Kerala, for providing DSC and SEM facility and also PBRI, Bhopal, for providing in-vivo facility.

REFERENCES


Table 4: In-vitro release kinetic equation data of glipizide chitosan-alginate microspheres (D:P-1: 0.5-1:2), n=3 for values

<table>
<thead>
<tr>
<th>Code</th>
<th>R0</th>
<th>R1</th>
<th>R2</th>
<th>R3</th>
<th>R4</th>
<th>R5</th>
</tr>
</thead>
<tbody>
<tr>
<td>GCS1</td>
<td>0.987</td>
<td>0.955</td>
<td>0.985</td>
<td>0.744</td>
<td>0.982</td>
<td></td>
</tr>
<tr>
<td>GCS2</td>
<td>0.991</td>
<td>0.958</td>
<td>0.991</td>
<td>0.754</td>
<td>0.971</td>
<td></td>
</tr>
<tr>
<td>GCS3</td>
<td>0.979</td>
<td>0.958</td>
<td>0.976</td>
<td>0.774</td>
<td>0.962</td>
<td></td>
</tr>
<tr>
<td>GCS4</td>
<td>0.980</td>
<td>0.968</td>
<td>0.981</td>
<td>0.814</td>
<td>0.978</td>
<td></td>
</tr>
<tr>
<td>GCS5</td>
<td>0.976</td>
<td>0.966</td>
<td>0.976</td>
<td>0.842</td>
<td>0.974</td>
<td></td>
</tr>
<tr>
<td>GCS6</td>
<td>0.981</td>
<td>0.983</td>
<td>0.980</td>
<td>0.873</td>
<td>0.986</td>
<td></td>
</tr>
<tr>
<td>GCS7</td>
<td>0.982</td>
<td>0.974</td>
<td>0.980</td>
<td>0.901</td>
<td>0.955</td>
<td></td>
</tr>
</tbody>
</table>

R: Regression coefficient value, D:P: Drug to polymer ratio

Table 5: For in-vivo grouping of animals

<table>
<thead>
<tr>
<th>Group No.</th>
<th>Treatment</th>
<th>Dose</th>
<th>Number of animals</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Normal saline</td>
<td>5 ml/kg p.o.</td>
<td>06</td>
</tr>
<tr>
<td>2</td>
<td>STZ (STZ control)</td>
<td>60 mg/kg i.p.</td>
<td>06</td>
</tr>
<tr>
<td>3</td>
<td>Glipizide</td>
<td>5 mg/kg p.o.</td>
<td>06</td>
</tr>
<tr>
<td>4</td>
<td>GCS3</td>
<td>5 mg/kg p.o.</td>
<td>06</td>
</tr>
</tbody>
</table>

STZ: Streptozotocin

Table 6: Blood glucose level (mg/dl)

<table>
<thead>
<tr>
<th>S.No.</th>
<th>Treatment</th>
<th>0 Day</th>
<th>3 Day</th>
<th>5 Day</th>
<th>7 Day</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Normal saline (5 ml/kg)</td>
<td>87.90±5.626</td>
<td>89.60±5.873</td>
<td>90.48±6.485</td>
<td>89.20±6.429</td>
</tr>
<tr>
<td>2</td>
<td>STZ control</td>
<td>284.35±11.734</td>
<td>289.43±12.438</td>
<td>293.92±12.020</td>
<td>296.73±11.853</td>
</tr>
<tr>
<td>3</td>
<td>Glipizide (5 mg/kg)</td>
<td>282.93±8.574</td>
<td>222.65±8.025*</td>
<td>166.90±8.773*</td>
<td>119.45±9.211*</td>
</tr>
<tr>
<td>4</td>
<td>GCS3 (5 mg/kg)</td>
<td>287.37±6.423</td>
<td>240.72±5.101*</td>
<td>192.23±5.737*</td>
<td>145.70±5.757*</td>
</tr>
</tbody>
</table>

Values are expressed as mean±SD at n=6. One-way ANOVA followed by Bonferroni test, *p<0.05 significant compared to the diabetic control group.

Table 7: Reduction in plasma glucose levels after administration of pure drug and glipizide formulation in rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>1 day</th>
<th>3 day</th>
<th>5 day</th>
<th>7 day</th>
</tr>
</thead>
<tbody>
<tr>
<td>STZ control group</td>
<td>glipizide</td>
<td>18.101</td>
<td>21.013</td>
<td>42.011</td>
</tr>
<tr>
<td>Glipizide (5 mg/kg)</td>
<td>18.213</td>
<td>23.072</td>
<td>43.215</td>
<td>59.751</td>
</tr>
<tr>
<td>GCS3 (5 mg/kg)</td>
<td>10.351</td>
<td>16.829</td>
<td>34.597</td>
<td>50.898</td>
</tr>
</tbody>
</table>

STZ: Streptozotocin

Statistical analysis of in-vivo data

All data were analyzed by one-way ANOVA followed by Bonferroni test. p<0.05 was considered as the level of significance. All data are presented in mean±SD.

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