INTRODUCTION

Diabetes mellitus is a chronic disease that affects 415 million people worldwide, and 5 million people died from diabetic complications [1]. Type 2 diabetes mellitus (T2DM) is manifested by hyperglycemia that results from lack of insulin or resistance to the action of insulin in the muscle, fat, and liver in addition to an inadequate response by the pancreatic beta cells [2].

Uncontrolled diabetes for the first few years can cause acute cardiac and renal complications which are life-threatening, whereas poorly controlled diabetes can cause long-term cardiac and renal complications [3]. This is due to a complex group of risk factors associated with T2DM including insulin resistance, hyperglycemia, diabetic dyslipidemia, hypertension, hyperinsulinemia, systemic inflammation, and adipose tissue-derived factors [4-6]. Worth mentioning, the changes in the mass and metabolism of adipose tissue may be paralleled by insulin resistance and visceral obesity commonly associated with T2DM [7].

Inflammatory mediators play a critical role in the development of cardiovascular disease (CVD). In particular, acute coronary syndrome (ACS) is an inflammatory disease and the serum levels of inflammatory factors, such as interleukin (IL)-6, IL-18 and C-reactive protein (CRP), are used to detect patients with CVD, especially coronary artery disease (CAD) [8]. Moreover, CRP has been found to be significantly elevated in patients with acute myocardial infarction [9].

CRP is a well-known acute inflammatory marker that has been used as a mediator of infection [10]. These two factors are easy in assessment, reliable, and inexpensive, and they are used for the diagnosis and follow-up of several diseases [10,11]. Procalcitonin (PCT) is produced during bacterial infections, sepsis, cardiogenic shock, major surgery, burns, multiple trauma, and after cardiac surgery [12,13]. It is a hormone of 116 amino acids and it is implicated in calcium metabolism. It is produced by the medullary C-cells of the thyroid gland [14-16]. Nonetheless, even thyroidectomized patients showed PCT response during acute inflammation [17] indicating that there are other probable origins of PCT production.

Some investigators have suggested that PCT may be produced by the liver and inflammatory cells [18,19]. The inflammatory response is an essential feature of ACS and myocardial infarction (MI). In acute MI (AMI), signs of inflammation are well identified and the amplified levels of acute phase reactants have been found to be linked to a worse short- and long-term prognosis [20]. Signs of the systemic inflammatory response, such as fever, leukocytosis, and the enhanced acute-phase reactants, are frequently shown in patients with ACS [21]. PCT has been manifested as a novel cardiac marker in AMI [22]. Circumstantial evidence showed that bacterial endotoxins and tumor necrosis factor-α (TNF-α) both induced PCT in vitro [13,23]. Based on this document, PCT may be considered as an alternative new valuable prognostic marker in ACS. Many research studies have cited higher PCT levels in patients versus healthy controls following severe sepsis, cardiac surgery, or trauma [24-26].

The focus of our interest was to detect the kinetics of procalcitonin production, compared with CRP, as a novel prognostic marker for cardiovascular complications in type 2 diabetic patients.

CLINICAL SIGNIFICANCE OF PROCALCITONIN AND C-REACTIVE PROTEIN IN THE PREDICTION OF CARDIOVASCULAR COMPLICATIONS IN PATIENTS WITH TYPE 2 DIABETES MELLITUS

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ABSTRACT

Objective: This work was delineated to assess procalcitonin (PCT) and C-reactive protein (CRP) as prognostic markers for cardiovascular complication in type 2 diabetic patients.

Methods: Forty diabetic patients without cardiovascular disease (CVD), 40 diabetic patients with CVD, and 20 healthy control counterparts were participated in this study. Serum PCT and CRP levels were assayed and correlated with metabolic parameters. Receiver operating characteristic (ROC) curve analysis was done for each biochemical marker.

Results: The mean level of PCT was 707.17±99.19 ng/l in diabetic patients versus 881.30±23.56 ng/l for the cardio-diabetic patients (p<0.0001). The mean value of CRP was 34.43±17.27 mg/l in diabetic patients versus 50.32±20.19 mg/l for the cardio-diabetic patients (p<0.0003). PCT levels were significantly amplified in the cardio-diabetic patients with increasing CRP, triglycerides (TG), fasting blood glucose (FBG), and cholesterol (p<0.004, 0.0005, 0.002, and 0.01, respectively). CRP levels were significantly enhanced in the cardio-diabetic patients with increasing TG, FBG, cholesterol, and microalbumin (p<0.002, 0.047, 0.003, and 0.001 respectively). ROC curve analysis for PCT and CRP revealed that the area under curve (AUC) was 0.878 and 0.727, respectively. These findings indicate the good validity of the above biomarkers especially PCT as a prognostic marker for cardiovascular complication in type 2 diabetic patients.

Conclusion: This study evidences the usefulness of measuring serum levels of PCT and CRP in diagnosis of cardiovascular complication in type 2 diabetic patients.

Keywords: Procalcitonin, Diabetes mellitus, C-reactive protein, Cardiovascular complications.

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The statistical analysis revealed significant differences between the groups in several parameters. Venous blood and urine samples were collected from all participants and each blood sample was divided into two portions. The small portion was collected on EDTA-coated tube and the large portion was collected on EDTA-free coated tube. Serum samples were obtained by centrifugation and the biochemical variables were measured on the same day of the blood collection. Remaining serum specimens were stored at -20°C until analysis of PCT and CRP levels.

Quantitative determination of serum glucose was carried out colorimetrically using commercial kits purchased from Randox Laboratories (Crumlin, County Antrim, United Kingdom) using method of Thomas [27]. Quantitative estimation of serum cholesterol was done colorimetrically using commercial kits purchased from Randox Laboratories using method of Richmond [28]. Serum high density lipoprotein (HDL)-cholesterol was assayed colorimetrically using commercial kits purchased from Randox Laboratories using method of Assmann [29]. Low density lipoprotein (LDL)-cholesterol was quantified in serum using a kit provided by Spinreact (Girona, Spain) for the quantitative determination of serum LDL-cholesterol following the method of Okada et al. [30]. Triglycerides (TG) in serum were measured colorimetrically using commercial kit purchased from Randox Laboratories according the method of Jacobs and Van Denmark [31]. Glycated hemoglobin was determined using a kit provided by Spinreact (Girona, Spain) for the quantitative determination of serum glycated hemoglobin according to the method described by Trivelli et al. [32]. Serum CRP was measured by ELISA using commercial kit purchased from Immunospec Corporation (Canoga Park, USA) following the method of Hedlund [33]. Quantitative estimation of microalbumin in urine was done by immunoturbidimetric assay using commercial kit purchased from Pointe Scientific, INC (Canton, USA) according to the method of Mogens and Schmitz [34]. Serum PCT was evaluated by solid phase enzyme-linked immunosorbent assay (ELISA kit) using 96-well microplates supplied by Glory Science Co., Ltd (Del Rio, USA) in accordance with the method described by Arkader et al. [35].

**RESULTS**

Laboratory measurements of the parameters of the different groups are presented in Table 1. Cholesterol, CRP, FBG, glycated hemoglobin A1c (HbA1c), LDL, TG, microalbumin, and PCT levels were significantly higher in diabetic patients than in healthy individuals (p=0.022, p<0.0001, p<0.0001, p=0.009, p=0.007, p<0.0001, p<0.0001, respectively). Likewise, CRP, FBG, HbA1c, LDL, TG, cholesterol, PCT and microalbumin levels were significantly higher in cardio-diabetic patients than in healthy individuals (p<0.0001, p<0.0001, p<0.0001, p=0.009, p=0.007, p=0.001, and p<0.0001, respectively). In addition, CRP, LDL, PCT, and microalbumin levels were significantly higher in cardio-diabetic patients as compared to diabetic patients (p<0.0003, p<0.0001, p<0.0001, and p<0.0001, respectively). Whereas, HDL level showed significant drop in cardio-diabetic patients versus diabetic patients and controls (p=0.0002 and p<0.0001, respectively). Furthermore, it revealed significant decline in diabetic patients relative to healthy participants (p=0.038) (Table 1).

The results of correlation between serum PCT concentration and metabolic parameters in the different studied groups are depicted in Table 2. Significant positive correlation between serum PCT and FBG, HbA1c, LDL, TG, microalbumin, and PCT levels were observed in the current study. In addition, twenty healthy participants who with no history of T2DM, other endocrine dysfunctions, hyperlipidemia, hypertension, or coronary heart diseases (CHDs) were enrolled in the study and served as controls. Clinical evidence of CVD included MI or coronary artery bypass surgery, stroke, and peripheral arterial disease. Patients in the group without vascular disease were T2DM patients who had no history of vascular disease and those with normal electrocardiogram findings at exercise and normal peripheral artery Doppler ultrasonography. Exclusion criteria involved the presence of sustained Type 1 DM, acute and chronic infections, malignancy, hepatic or renal disease, diabetic retinopathy and nephropathy, and other endocrine dysfunctions. This study was approved by Ethical Committee of Ethics Commission and Scientific Research of the General Authority for Hospitals and Educational Institutes.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control subject (C)</th>
<th>Diabetic patients (D)</th>
<th>Cardi-diabetic patients (CD)</th>
<th>P1</th>
<th>P2</th>
<th>P3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cholesterol (mg/dl)</td>
<td>194.00±36.70</td>
<td>225.70±54.40</td>
<td>233.02±57.47</td>
<td>0.022</td>
<td>0.007</td>
<td>0.56</td>
</tr>
<tr>
<td>CRP (mg/l)</td>
<td>14.00±252.00</td>
<td>135.00±383.00</td>
<td>69.00±378.00</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
<td>0.003</td>
</tr>
<tr>
<td>TG (mg/dl)</td>
<td>20.00±55.00</td>
<td>10.00±47.00</td>
<td>15.00±45.00</td>
<td>0.022</td>
<td>&lt;0.0001</td>
<td>0.0001</td>
</tr>
<tr>
<td>LDL (mg/dl)</td>
<td>92.00±95.00</td>
<td>146.25±64.94</td>
<td>202.47±55.94</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
<td>0.0001</td>
</tr>
<tr>
<td>HDL (mg/dl)</td>
<td>42.00±63.00</td>
<td>35.35±8.88</td>
<td>27.57±8.83</td>
<td>0.038</td>
<td>&lt;0.0001</td>
<td>0.0002</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>9.74±1.67</td>
<td>6.60±14.70</td>
<td>6.80±15.90</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
<td>0.325</td>
</tr>
<tr>
<td>Microalbumin (mg/ml)</td>
<td>7.40±2.30</td>
<td>11.60±7.07</td>
<td>18.70±15.90</td>
<td>0.016</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

P1: Diabetic group compared to control group. P2: Cardio-diabetic group compared to control group. P3: Cardio-diabetic group compared to diabetic group.
concentration and cholesterol, TG, CRP, and FBG has been determined in cardio-diabetic patients (p=0.011, p=0.0005, p=0.004, and p=0.002, respectively). As well, significant positive correlation between serum PCT concentration and LDL, cholesterol, TG, CRP, FBG, and HbA1C has been recorded in diabetic patients (p=0.052, p=0.013, p=0.003, p=0.0001, p=0.0001, and p=0.009, respectively). However, significant negative correlation has been observed between serum PCT and microalbumin in diabetic patients (p=0.016) (Figs. 1 and 2).

The data of correlation between serum CRP level and other measured biochemical markers in the different studied groups are illustrated in Table 3. Significant positive correlation between serum CRP level and cholesterol, TG, FBG, PCT, and microalbumin in cardio-diabetic patients has been demonstrated (p=0.003, p=0.002, p=0.047, p=0.004, and p=0.001, respectively). Similarly, significant positive correlation between serum CRP level and LDL, cholesterol, TG, FBG, HbA1C, and PCT has been detected in diabetic patients (p=0.004, p=0.001, p=0.004, p=0.0001, p=0.047, and p<0.0001, respectively). Meanwhile, significant negative correlation has been recorded between serum CRP level and HDL, and microalbumin in diabetic patients (p=0.047 and p=0.018, respectively) (Figs. 3 and 4).

The receiving operating characteristic (ROC) curve is designed for PCT and CRP (Figs. 5 and 6). The cut-off values for PCT and CRP were 750 ng/l, and 42.1 mg/l, respectively. AUC for PCT and CRP was 0.878 and 0.727, respectively. This result indicated the good validity of the above biochemical markers particularly PCT to discriminate diabetic patients from cardio-diabetic patients.

DISCUSSION

Type 2 diabetic patients have a high risk for CVD. This risk is accompanied by many factors such as hypertension, dyslipidemia, and obesity in these patients. However, the onset of CVD in type 2 diabetic patients is not linked with the high prevalence of traditional risk factors only, but other non-traditional risk factors may be implicated. Thus, CVD is increased in type 2 diabetic patients due to a complex combination of various traditional and non-traditional risk factors. This has a critical role to play in the evolution of atherosclerosis over its long natural history from endothelial function to clinical events [36]. The objective of this study was to assess PCT and CRP as prognostic markers for cardiovascular complications in patients with T2DM.

The results obtained in this study showed that cholesterol, LDL, and TG are significantly increased in diabetic patients relative to controls whereas, HDL is significantly decreased in diabetic patients versus the controls. These results are in conformity with those of Ali and Al Hadidi [37] who stated that all the above parameters are significantly higher in T2DM group, whereas HDL is significantly lower in T2DM group in respect with the control group. These results are explained by Ronald [38] who cited that insulin resistance may contribute in the development of dyslipidemia in diabetic patients. As type 2 diabetes, insulin resistance increases the flow of free fatty acids from adipose tissue and impairs insulin-mediated skeletal muscle uptake of free fatty acids resulting in increased fatty acid flow to the liver [39,40]. The increased free fatty acid levels in individuals with impaired glucose tolerance suggesting that insulin resistance is related to the elevated free fatty acid levels which occur before the onset of hyperglycemia [41]. Many investigators have demonstrated a relationship between plasma free fatty acid levels and insulin resistance [42]. Free fatty acids in the form of TG are deposited in the muscle, liver, heart, and pancreas in the presence of insulin resistance. Furthermore, insulin resistance increases the activity of hepatic lipase, which is responsible for hydrolysis of phospholipids into LDL and HDL particles with consequent formation of very small and dense LDL particles and a reduction in HDL particles [43,44]. This hypothesis is appreciated when some drugs that lowered the high level of free fatty acids (thiazolidinediones) could improve insulin sensitivity in the muscle, liver, and adipose tissues [45,46].

FBG and HbA1c levels are significantly enhanced in diabetic patients comparing with healthy individuals. These results are in harmony with

### Table 2: Correlation between serum procalcitonin concentration and metabolic parameters in the different studied groups

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Serum procalcitonin level in control groups</th>
<th>Serum procalcitonin level in diabetic group</th>
<th>Serum procalcitonin level in cardio-diabetic groups</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>r</td>
<td>p</td>
<td>r</td>
</tr>
<tr>
<td>Cholesterol (mg/dl)</td>
<td>0.231</td>
<td>0.325</td>
<td>0.385</td>
</tr>
<tr>
<td>TG (mg/dl)</td>
<td>0.688</td>
<td>0.0008**</td>
<td>0.448</td>
</tr>
<tr>
<td>HDL (mg/dl)</td>
<td>0.245</td>
<td>0.296</td>
<td>-0.281</td>
</tr>
<tr>
<td>LDL (mg/dl)</td>
<td>0.132</td>
<td>0.576</td>
<td>0.308</td>
</tr>
<tr>
<td>CRP (mg/l)</td>
<td>-0.133</td>
<td>0.575</td>
<td>0.760</td>
</tr>
<tr>
<td>FBG (mg/dl)</td>
<td>-0.416</td>
<td>0.068</td>
<td>0.718</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>0.337</td>
<td>0.146</td>
<td>0.4036</td>
</tr>
<tr>
<td>Microalbumin (mg/ml)</td>
<td>0.230</td>
<td>0.327</td>
<td>-0.377</td>
</tr>
</tbody>
</table>

r=Correlation coefficient, *p<0.05, **p<0.01, not significant (p>0.05). CRP: C-reactive protein, FBG: Fasting blood glucose, HbA1c: Hemoglobin A1c, HDL: High-density lipoprotein, LDL: Low-density lipoprotein, TG: Triglycerides

### Table 3: Correlation between serum CRP level and and metabolic parameters in the different studied groups

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Serum CRP level in control groups</th>
<th>Serum CRP level in diabetic group</th>
<th>Serum CRP level in cardio-diabetic groups</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>r</td>
<td>p</td>
<td>r</td>
</tr>
<tr>
<td>Cholesterol (mg/dl)</td>
<td>0.490</td>
<td>0.028*</td>
<td>0.476</td>
</tr>
<tr>
<td>TG (mg/dl)</td>
<td>0.327</td>
<td>0.158</td>
<td>0.442</td>
</tr>
<tr>
<td>HDL (mg/dl)</td>
<td>0.278</td>
<td>0.234</td>
<td>-0.315</td>
</tr>
<tr>
<td>LDL (mg/dl)</td>
<td>0.265</td>
<td>0.257</td>
<td>0.444</td>
</tr>
<tr>
<td>FBG (mg/dl)</td>
<td>-0.049</td>
<td>0.844</td>
<td>0.588</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>-0.265</td>
<td>0.258</td>
<td>0.315</td>
</tr>
<tr>
<td>Microalbumin (mg/ml)</td>
<td>0.530</td>
<td>0.016*</td>
<td>0.370</td>
</tr>
<tr>
<td>Procalcitonin (ng/l)</td>
<td>-0.133</td>
<td>0.575</td>
<td>0.760</td>
</tr>
</tbody>
</table>

r=Correlation coefficient, *p<0.05, **p<0.01, not significant (p>0.05). CRP: C-reactive protein, FBG: Fasting blood glucose, HbA1c: Hemoglobin A1c, HDL: High-density lipoprotein, LDL: Low-density lipoprotein, TG: Triglycerides
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the study of Ali and Al Hadidi [37] who revealed that FBG and HbA1C levels are significantly higher in diabetic patients than in healthy individuals. Furthermore, Makris et al. [47] recorded a significant relation between FBG and HbA1C in diabetic patients. The studies of Peterson et al. [48] and Miedema [49] have shown that the increased blood glucose leads to the increased attachment of glucose molecules to the hemoglobin in red blood cells. The long period of hyperglycemia in the blood, the more glucose binds to hemoglobin in the red blood cells and the higher in glycated hemoglobin. It is formed in a non-enzymatic glycation pathway of hemoglobin exposure to plasma glucose; then, reaction occurs between glucose and the N-end of the beta chain in hemoglobin. In diabetes mellitus, higher amounts of glycated hemoglobin indicating a poorer control of blood glucose levels with consequent complications such as CVD, nephropathy, neuropathy, and retinopathy.

CRP level is significantly elevated in diabetic patients relative to healthy individuals. These results are in keeping with those of Belfki et al. [50] who demonstrated that levels of CRP are significantly elevated in patients with T2DM versus controls. Morohoshi et al. [51] and Guha et al. [52] mentioned that hyperglycemia stimulates the liberation of the inflammatory cytokine such as IL-6 and TNF-α from different cell types and results in the secretion of acute phase reactants by adipocytes. Grunfeld et al. [53] and Hirschfield et al. [54] proved that CRP is an acute phase reactant that is produced primarily in the liver under the activation of adipocyte-derived proinflammatory cytokines.

Serum PCT level is significantly amplified in diabetic patients relative to controls. Our results are in harmony with those of Soylemez et al. [55] who reported that PCT levels are elevated in type 2 diabetic patients compared with healthy controls. In addition, Schiopu et al. [56] found that PCT is positively correlated with the presence of hyperglycemia and with systolic blood pressure. Moreover, hyperglycemia is accompanied by increased systemic inflammatory activation and thus, it seems that this inflammation may stimulate PCT production.

These investigators explained the elevated levels of PCT in type 2 diabetic patients by the fact that T2DM is related to oxidative stress and advanced glycation end-products (AGEs) elevation. AGEs interact with its receptor called RAGE. Potentiation of RAGE leads to regulation of the transcription factor, nuclear factor-kB, and its target genes and also of activator protein-1 (AP-1). These factors could ultimately lead to upregulation of PCT gene expression.

Fig. 1: Correlation between different metabolic parameters and serum procalcitonin concentration in diabetic group
There are significantly higher differences in microalbumin between diabetic patients and controls. These findings echo those of previous study of Chowta et al. [57] who found high prevalence of microalbuminuria (37%) in T2DM, and the incidence of microalbuminuria increases with the increased duration of diabetes mellitus. Mogensen et al. [58] proved a positive correlation between microalbuminuria and the duration of diabetes mellitus. Long duration of diabetes has significant contribution for the development of microalbuminuria. Prolonged exposure to hyperglycemia causes AGEs accumulation. Bucala et al. [59] and Tan et al. [60] stated that hyperglycemia may cause tissue damage by several mechanisms, one of which is non-enzymatic glycation of intra- and extra-cellular proteins. Glucose has a reactive aldehyde moiety which reacts non-enzymatically with the amino groups of proteins in the extracellular matrix, producing reversible Amadori products, and AGEs that can impair degradation of proteins, and induce cytotoxic pathways. Hence, serum levels of AGEs increased in type 2 diabetic patients, and this leads to increased level of microalbumin.

Cholesterol, TG, and LDL are significantly increased in cardio-diabetic group relative to healthy control group. Meanwhile, HDL is significantly decreased in cardio-diabetic patients versus healthy individuals. This finding is in respect with the study of Haddad et al. [61] who found that cholesterol, LDL-C, and TG levels are increased but HDL-C level is decreased in diabetic patients with CAD compared with those in control group. These data are explained by Celermajer [62] who mentioned that dyslipidemia is a critical mechanism by which atherosclerosis and endothelial dysfunction can occur in diabetic patients. Healthy endothelium regulates activation of platelet, tone of the blood vessel, leukocyte adhesion, inflammation, and thrombogenesis. Thus, healthy endothelium is antiatherogenic, vasodilatory, and anti-inflammatory [62]. Affection of these mechanisms results in the generation of atherosclerosis. Therefore, both insulin resistance and insulin deficiency lead to dyslipidemia accompanied by increased glycosylation, oxidation, and TG enrichment of lipoproteins.
Furthermore, Dokken [63] has shown that oxidized LDL is proatherogenic because when the particles of LDL are oxidized, they showed new properties that are recognized by the immune system as “foreign.” Furthermore, oxidized LDL induces many abnormal biological responses, such as promoting the capability of leukocytes to ingest lipids and differentiate into foam cells, attracting leukocytes to the intima of the vessel, and stimulating leukocytes, endothelial cells (ECs), and smooth muscle cell (SMC) proliferation [64]. All of these results in the generation of atherosclerotic plaque. Furthermore, in diabetic patients, LDL particles can glycated, in a process similar to the glycation of hemoglobin. Glycation of LDL prolongs its half-life [65] and therefore increased the capability of LDL to induce atherogenesis.

FBG and HbA1C levels are significantly elevated in cardio-diabetic patients relative to the controls. These results are in harmony with those of Anping et al. [66] who stated that levels of HbA1C are gradually increased in unstable angina (UA) and AMI patients versus healthy individuals. Biologically, glycated hemoglobin is an advanced glycosylation end-product, and the increased level of HbA1C causes the formation of advanced glycosylation end-product, which attaches to the vessel wall and leads to dysfunction of endothelium and oxidative stress progression [67,68]. Furthermore, the binding of advanced glycosylation end-product is associated with overproduction of inflammatory mediators like CRP [69]. Increased CRP level has been shown to be significantly linked with the instability of plaque [70,71]. This explains why that after adjusting CRP, there is no significant association between HbA1c and the severity of CAD. Finally, increased level of advanced glycosylation end-product interferes with the endogenous fibrinolytic system which may lead to high risk of coronary artery stenosis [72].

CRP is significantly elevated in cardio-diabetic patients in respect to healthy individuals. Our results come in line with those of Ridker [73] who proved that the increased level of CRP is related to an 8-fold increase.
in cardiovascular mortality. Furthermore, Liang et al. [74] stated that the level of CRP is significantly enhanced in AMI and UA patients than in stable angina patients and healthy control. These observations are interpreted by Shrivastava et al. [75] who reported that atherosclerotic process is characterized by low-grade inflammation and increased level of the inflammatory modulators. In addition, CRP is also generated locally the in atherosclerotic lesions by inflamed SMCs lymphocytes and monocytic cells.

Paffen and DeMaat [76] and Pfützner et al. [77] found that CRP plays a critical role in many aspects of atherogenesis including activation of the classical pathway of the complement system. Through this action, CRP directly facilitates the innate immunity, a process that has already been related to the development and progression of CHD [77]. CRP increases LDL uptake into macrophages and enhances the capability of macrophages to form foam cells. Moreover, CRP upregulates adhesion molecules expression in ECs that can attract monocytes to the site of injury. Therefore, CRP is a high sensitive biomarker that can be used for diagnosis, management, and prognosis of CHD [78].

Serum PCT level is significantly enhanced in cardio-diabetic patients versus healthy controls. Similar results have been obtained by Sinning et al. [78] who cited that patients with ACS have increased concentration of PCT. Likewise, Sponholz et al. [79] found that PCT level is increased in patients with cardiovascular events. Erren et al. [80] reported that the increased PCT level is linked to the extent of atherosclerosis in CAD patients and peripheral arterial disease. In atherosclerotic patients, ischemia and inflammatory processes cause PCT production. In addition, increased levels of PCT in the setting of CAD are more as a result of non-specific liberation of cytokine in the context of local tissue damage to myocardium due to ischemia and necrosis. This explains the association between PCT and low-grade inflammatory activity within the vascular wall caused by atherosclerosis. Schlitt et al. [81] found that PCT mRNA expression by peripheral blood mononuclear cells is

**Fig. 4: Correlation between different metabolic parameters and serum C-reactive protein level in cardio-diabetic group**
found that permeability to macromolecules, leukocyte adhesion, and vascular impairment could be existed depending on which function is affected and maintenance of organ function. Therefore, many kinds of endothelial impairment can be identified by any change and CVD [86,87]. Endothelial impairment can be explained by the association of microalbuminuria with generalized endothelial impairment. Indeed, microalbuminuria in type 1 and type 2 diabetes is usually paralleled by endothelial impairment regarding the regulation of hemostasis, fibrinolysis, leukocyte adhesion, and no synthesis and/or availability [86]. Jager et al. [88] and Stehouwer et al. [89] mentioned that chronic, low-grade inflammation is related to the occurrence and progression of microalbuminuria and with risk for atherothrombotic disease. From the above considerations, endothelial impairment and chronic low-grade inflammation are essential candidates to explain the association between microalbuminuria and CVD.

In view of our data, significant positive correlation between serum PCT and cholesterol, TG, CRP, LDL, Hba1c, and FBG in diabetic patients has been found. Likewise, significant positive correlation has been detected between PCT and cholesterol, TG, CRP, and FBG in cardio-diabetic patients. These findings are in concert with the reports of Schiopu et al. [56] who found that PCT is associated with many of the already established cardiovascular risk factors (CRP, hypertension, diabetes, and renal function). Furthermore, Sponholz et al. [79] stated that PCT level is associated with CRP and TG concentrations in patient with CAD.

A significant positive correlation between serum CRP level and cholesterol, TG, FBG, Hba1c, and LDL in diabetic patients, and also between CRP and cholesterol, TG, FBG, and microalbumin in cardio-diabetic patients has been demonstrated. These results converge with the previous report of Seo et al. [90] which showed that levels of hs-CRP are significantly correlated with the level of glycosylated hemoglobin, LDL/HDL ratio, LDL/total cholesterol (TC) ratio, and TC/TG ratio in diabetic patients or patients with CVD. Furthermore, Bahceci et al. [91] observed positive correlation between serum hs-CRP and glycated hemoglobin in T2DM with or without CHD. Safullah et al. [92] found that fasting plasma glucose, TG, LDL-cholesterol, and Hba1c are higher in diabetic patients with abnormal hs-CRP than the diabetic patients with normal hs-CRP.

ROC curve was done to detect the best cut-off value of serum PCT in diabetic and cardio-diabetic patients. It has proved that PCT at concentration 750 ng/l has 87.5% sensitivity and 72.5% specificity. Sponholz et al. [79] and Farzaad et al. [93] revealed that procalcitonin level is high in patients with cardiovascular disease. In addition, the studies of Sponholz et al. [79] and Erren et al. [80] reported that the elevated PCT level is related to the extent of atherosclerosis in patients with CAD and peripheral arterial disease. In addition, these findings indicate that PCT is a biomarker of CAD in type 2 diabetic patients.

The best cut-off value of serum CRP in diabetic and cardio-diabetic patients was 42.1 mg/l with 62.5% sensitivity and 82.5% specificity. These results are comparable to the findings of Ridker [73] who mentioned that increased level of CRP is related to 8-fold increase in cardiovascular mortality. Furthermore, Bahceci et al. [91] found that hs-CRP levels are increased in patients with CHD, whether they are diabetic or non-diabetic than in controls, and the levels of hs-CRP in diabetic patient with CHD are also higher than patients with CHD only. These observations suggest that CRP is a biomarker of CAD in type 2 diabetic patients.

CONCLUSION

The present findings provide a clear evidence favoring the clinical value of measuring serum level of PCT and CRP as diagnostic candidates for cardiovascular complication in patients with T2DM.

REFERENCES


Fibrinogen, viscosity, and white blood cell count are major risk factors for ischemic heart disease. The Caerphilly and Speedwell collaborative heart disease studies. Circulation 1991;83(3):386-44.


