

FORMULATION AND EVALUATION OF IMPLANTABLE DRUG DELIVERY SYSTEM OF TEMOZOLOMIDE BY USING HYDROPHILIC POLYMER

SINDHU V, BHAVYA S, SURESH KUMAR P, JEYABASKARAN M, PRAVEENKUMAR T, SD. YASMIN SULTHANA

Department of Pharmaceutics, Browns College of Pharmacy, Khammam, Telangana, India. Email: sindhu95.vemula@gmail.com

Received: 18 May 2017, Revised and Accepted: 31 May 2017

ABSTRACT

Objective: The present research study was carried out to formulate and evaluate the implants of temozolomide using hydrophilic polymer.

Methods: Temozolomide implants were formulated using extrusion method with different grades of carbopol. The powdered blend was evaluated for micromeritic properties such as angle of repose, bulk density, tapped density, Carr's index, and Hausner's ratio. The formulated implants were analyzed for drug content uniformity, thickness, weight variation, and short-term stability study. *In vitro* release study of implants was performed using 0.1N hydrochloric acid, and it is maintained at 37°C±0.5°C.

Results: *In vitro* release study demonstrated that the release rate of temozolomide from the implant matrix was a function of concentration of the polymer. As the concentration of polymer was increased, drug release from the matrix was extended. The release of drug from all implant formulations was found to be uniform and was extended over a period of 12 hrs. The implant formulations were found sterile, uniform in weight and size. The drug content was found to be in the range of 97.2-101.33%.

Conclusion: Drug interaction studies revealed that there were no chemical interactions between temozolomide and polymers used in the study. Short-term stability studies of implants revealed that implants were stable, and there were no significant changes in the physical appearance and drug content of the implant formulations. The results of the study demonstrated that implantable drug delivery system of temozolomide can be formulated using hydrophilic polymer.

Keywords: Carbopol, Cross linking agent, Hydrophilic polymer, Implants, Temozolomide.

© 2017 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>) DOI: <http://dx.doi.org/10.22159/ajpcr.2017.v10i11.20070>

INTRODUCTION

The objective of the present study was to formulate and evaluate implants of temozolomide using hydrophilic polymers. Nowadays, cancer is the major cause of mortality. Although scientists have done great research to know the causes of cancer, and for the diagnosis and the treatment, still mortality rate is high because the exact cure was not found. Cancer treatment is one of the major challenges in modern science as the drug delivery to solid tumors is a challenge to develop more effective cancer therapies. Drugs administered orally must be protected from denaturation in the gastrointestinal tract and should be capable of absorption across the wall of the intestine. After absorption and entering into hepatic circulation, it must be resistant to hepatic enzymes. The rate of drug absorption and elimination should be within the therapeutic range [1].

Nowadays, controlled drug delivery has achieved the sustained zero-order release of a drug substance over prolonged period of time. With the advancement in development and technology, various techniques such as osmotically driven pumps [2], matrices with controllable swelling [3] diffusion [4,5] or erosion rates [6], non-uniform drug loading profiles [7-9], multilayered matrices [10], and therapeutic molecule or protein in a schematic of a pulsatile or staggered fashion are used for formulating sustained release dosage forms. In the 1930s, a new system of sustained release implantable drug delivery system was administered by subcutaneous route was introduced [11]. Matrix systems are used as non-degradable implants. These systems consist of uniformly distributed drug throughout a solid non-biodegradable polymer [12]. Matrix systems rely on the diffusion of drug particles through non-degradable fibrous network of the polymer to achieve sustained release of the drug. The higher concentration of disintegrating agent within the matrix, greater would be the release of drug from the system [13].

Temozolomide is an oral alkylating agent used in the treatment of brain cancers and as first-line treatment for glioblastoma multiform and as a second-line treatment for astrocytoma. Temozolomide belongs to the class imidazotetrazines. These are organic polycyclic compounds containing an imidazole ring fused to tetrazine ring. Temozolomide is an imidazotetrazine derivative and an antineoplastic agent. It is a prodrug that has little to no pharmacological activity until it is hydrolyzed *in vivo* to 5-(3-methyltriazene-1-yl) imidazole-4-carboxamide (MTIC). After administration, temozolomide undergoes rapid, non-enzymatic hydrolysis at physiological pH to MTIC, which is the active form of the drug. About 38% of the administered temozolomide, total radioactive dose is recovered over 7 days. Elimination is by renal mechanism, 37.7% of drug is eliminated in urine and 0.8% in feces [14-17].

METHODS

Temozolomide was obtained as a gift sample from Alkem Laboratories Ltd., Mumbai, carbopol 931 was procured from Signet Chemical Corp., Mumbai, Maharashtra, India, acetic acid was procured from Loba Chemie, Mumbai, and glutaraldehyde solution was procured from S.D. Fine Chem. Ltd, Mumbai, Maharashtra, India.

Preparation of implants using extrusion method

Implants of temozolomide were prepared with different grades of carbopol as per the formula. The drug was dissolved in 5% acetic acid solution. Carbopol powder was added slowly to the drug solution, and it was allowed to soak for 10-15 minutes. The swollen mass so formed was mixed uniformly in a glass mortar and mixed thoroughly until it becomes a sticky dough mass. The dough mass was fed into the cylinder of the extruder and was extruded in the form of long rods through the nozzle. The rods were kept for drying overnight on a glass plate, and the rods were cut into 27 mm-sized implants. The implants were then dried at 40°C.

Cross linking of implants

About 25 ml of 25% of glutaraldehyde solution was taken in 100 ml beaker and was placed in an empty desiccator. A wire mesh containing implants was kept in a desiccator and was immediately closed. The implants were made to react with glutaraldehyde vapors for different time intervals (6 hrs, 12 hrs, and 24 hrs). Then, they were removed and air-dried for 72 hrs. Hence, that complete reaction between the carbopol and glutaraldehyde should take place. Afterwards, the implants were kept in an open atmosphere for a week to make the residual glutaraldehyde gets evaporated. The cross linking was carried out at low temperature. The residual glutaraldehyde can also be removed using an aqueous (2%) sodium metabisulfite solution and then immediately removed from it and placed in absolute alcohol bath (Table 1).

Evaluation of pre-compression parameters of the powder blend

The prepared powder formulation was subjected to measurement of angle of repose, bulk density, tapped density, Carr's compressibility index, and Hausner's ratio as per the standard procedure suggested [18,19].

Evaluation parameters for implants

Uniformity of weight

This test is performed to maintain the uniformity of weight of each implant. This is done by weighing 20 implants at random, and average weight is calculated. Not more than two of the individual weights deviate from the average weight by more than the percentage, and none deviate by more than twice the percentage. The mean and standard deviation were determined and reported [20].

Diameter of implants

The length and diameter of implants from every batch were measured with the help of Vernier calipers. Three samples were taken for the study from each batch, and mean value was reported [21,22].

Procedure for drug content uniformity test

Drug content of implants from every batch was estimated. The implants were cut into small pieces and were taken into 50 ml volumetric flask, 45 ml of glacial acetic acid was added and shaken thoroughly to dissolve the drug, and the volume was made up to 50 ml with glacial acetic acid. This solution was suitably diluted with glacial acetic acid and assayed for temozolomide content by measuring the absorbance at 330 nm. Temozolomide contents were calculated, using the standard calibration curve [20].

% swelling index

To study swelling index, the implant formulations were immersed into swelling solution phosphate buffer pH 7. The implants were placed in swelling solution and weight of implant was measured after 1 hr, and the excess of solution was removed gently by tapping the surface with a dry piece of filter paper [23]. The degree of swelling for each implant formulation at given time was calculated using the following equation:

$$H = \frac{W_t - W_0}{W_0} \times 100$$

Where, W_t and W_0 are the sample's weight at any given time and in the dry state, respectively.

In vitro dissolution studies

Dissolution test was carried out using USP XXIV (model DISSO, M/s. Lab India, Hyderabad) rotating paddle method (apparatus 2). 0.1N hydrochloric acid was used as dissolution medium (900 ml), and the stirring rate was maintained at 50 rpm and temperature at $37^\circ\text{C} \pm 0.5^\circ\text{C}$. Samples of 5 ml were withdrawn at predetermined time intervals, filtered, and replaced with 5 ml of fresh dissolution medium. The collected samples were suitably diluted with dissolution fluid wherever necessary and were analyzed for the temozolomide at 330 nm using a double-beam ultraviolet spectrophotometer (Shimadzu-2000). Each dissolution study was performed for three times, and the mean values were taken [24].

Stability study

The purpose of stability testing (the International Conference on Harmonization [ICH], 2004) is to provide evidence on how the quality of a drug substance or drug product varies with time under the influence of a variety of environmental factors such as temperature, humidity, and light, enabling recommended storage conditions, retest periods, and shelf lives. The ICH guidelines stability studies were carried out at $25^\circ\text{C}/75\% \text{RH}$ for the selected formulation for 3 months. The selected formulations were wrapped in butter paper, were then stored at $37^\circ\text{C}/75\% \text{RH}$ for 3 months, and evaluated for their physical appearance and drug content at specified intervals of time.

Drug polymer interaction study

The IR spectra of temozolomide and its formulations were obtained by KBr Pellet method using Perkin Elmer Fourier transform infrared (FTIR) series model 1615 spectrometer. The subdermal implants of temozolomide prepared with carbopol were tested for compatibility of the drug with the excipients by IR study.

RESULTS AND DISCUSSION

Pre-compression evaluation parameters of temozolomide formulation blend

The powder blends were prepared by mixing of various ingredients mentioned and used for characterization of various flow properties of powder. The bulk density of all the formulations was found to be in the range of 0.48 ± 0.05 to 0.58 ± 0.06 (g/cm^3) showing that the powder has good flow properties. The tapped density of all the formulations was found to be in the range of 0.57 ± 0.01 to 0.69 ± 0.04 . The compressibility index of all the formulations was found to be ranging between 16.21 ± 0.06 and 17.97 ± 0.02 . All the formulations have shown the Hausner ratio ranging between 0.64 ± 0.03 and 1.17 ± 0.02 , indicating the powder has good flow properties (Table 2).

Evaluation parameters of temozolomide implants

Physical characteristics

The physical characteristics of temozolomide implants (F1-F9) such as weight variation and drug content were determined, and results of the formulations (F1-F9) found to be within the limits specified in official books.

Drug content

All the implant formulations shown good uniformity in drug content and they contain 97.2-101.33% of temozolomide which is within the specified limit.

Table 1: Formulation composition for implants

Formulation code	F1	F2	F3	F4	F5	F6	F7	F8	F9
Temozolomide (mg)	20	20	20	20	20	20	20	20	20
Carbopol 931 (mg)	200	400	600	-	-	-	-	-	-
Carbopol 934 (mg)	-	-	-	200	400	600	-	-	-
Carbopol 971 (mg)	-	-	-	-	-	-	200	400	600
5% acetic acid (ml)	5	5	5	5	5	5	5	5	5
25% glutaraldehyde solution	Qs								

Diameters of implants

The diameter determined for formulated implants is tabulated in Table 3. Implants mean diameters were almost uniform in all the batches of implants formulations and were found to be in the range of 1.05-1.70 mm.

Uniformity of weight

The weight variations for all formulations are shown in Table 3. All the implants passed weight variation test as the % weight variation was within the pharmacopoeial limits. The weights of all the implants formulations were found to be in the range of 50±5 mg.

% swelling index

The % swelling index of the prepared implants ranged from 87-146 %.

In vitro drug release

Dissolution test was carried out using USP XXIV (model DISSO, M/s. Lab India, Hyderabad) rotating paddle method (apparatus 2) at 50 rpm using 0.1N hydrochloric acid as dissolution medium. Each dissolution study was performed for three times, and the mean values were taken. The *in vitro* dissolution studies of implants of temozolomide were conducted in simulated gastric fluid 0.1N HCl for 12 hrs. Formulations F1-F3 were prepared with carbopol 931. Formulation F1 showed complete drug release within 2 hrs, whereas F2 and F3 showed complete drug release in 3 and 4 hrs, respectively. The implants were unable to retain their shape and integrity for not more than 4 hrs. Hence, they were not considered. The formulations prepared with carbopol 934 retarded drug release. Formulations F4 and F5 showed complete drug release within 5 and 6 hrs. Formulations F4 and F5 were unable to retard drug release up to desired time period. F6 formulation retarded the drug release up to 12 hrs, and it showed a maximum of 89.87 in 12 hrs.

Formulations F7-F9 were prepared with carbopol 971. Formulations F7, F8, and F9 were retarded the drug release for more than 12 hrs. The formulation F7 was shown 98.78% in 12 hrs, whereas the formulation F8 and F9 showed only 84% and 78% of drug release in 12 hrs, respectively. It was observed that as the concentration of polymer increases, the drug release was also retarded. Initially, the formulations

containing low concentration and low viscosity showed 50-100% drug release within 4-6 hrs. Formulations containing high viscosity and high concentration were able to retard drug release for more than 12 hrs. Hence, based on dissolution study, formulation F7 is considered as the best formulation (Table 4 and Figs. 1-3).

Stability studies

Stability studies were carried out at 25°C/75% RH and 37°C/75% RH for the selected formulation for 3 months. The selected formulations

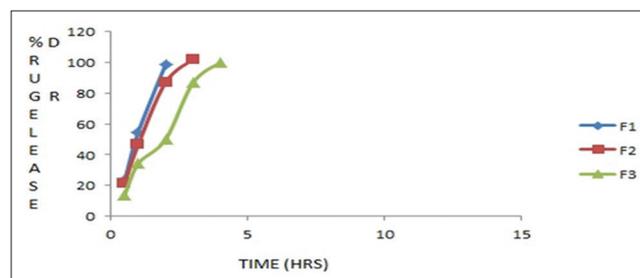


Fig. 1: *In vitro* drug release of implants with carbopol 931 (F1, F2, and F3)

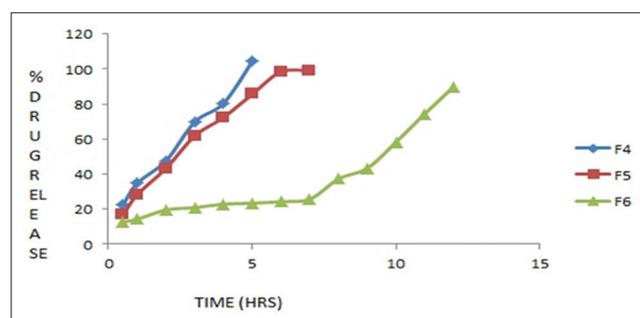


Fig. 2: *In vitro* drug release of implants with carbopol 934 (F4, F5, and F6)

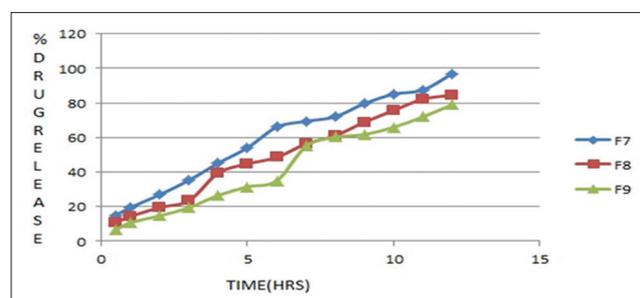


Fig. 3: *In vitro* drug release of implants with carbopol 971 (F7, F8, and F9)

Table 2: Micromeritic properties of powder blend

Formulation code	Bulk density	Tapped density	Compressibility index	Hausner's ratio
F1	0.49±0.07	0.57±0.01	16.21±0.06	0.86±0.06
F2	0.56±0.06	0.62±0.05	16.87±0.05	0.98±0.05
F3	0.52±0.03	0.68±0.07	17.11±0.01	0.64±0.03
F4	0.54±0.04	0.64±0.08	17.67±0.08	1.12±0.04
F5	0.53±0.06	0.67±0.03	16.92±0.04	1.2±0.08
F6	0.56±0.05	0.66±0.06	17.65±0.09	1.06±0.09
F7	0.58±0.06	0.69±0.04	16.43±0.05	0.76±0.03
F8	0.48±0.05	0.57±0.02	17.97±0.02	1.15±0.09
F9	0.54±0.08	0.62±0.03	17.54±0.09	1.17±0.02

Table 3: Evaluations of physical parameters of implants

Formulation code	Weight variation (mg) (±SD)	Drug content (%) (±SD)	% swelling index	Diameter of implants
F1	55±0.04	100.8±0.01	89	1.02
F2	51±0.01	97.8±0.02	112	1.54
F3	49±0.02	99.9±0.09	134	1.36
F4	46±0.05	101.33±0.03	104	1.28
F5	51±0.08	100.07±0.08	114	1.07
F6	53±0.09	95.6±0.09	145	1.67
F7	46±0.01	98.9±0.07	87	1.74
F8	55±0.08	100.2±0.04	134	1.48
F9	51±0.07	99.8±0.08	176	1.59

SD: Standard deviation

Table 5: Stability studies for optimized formulation (F7)

S.No.	Optimized formulation (F3) duration (months)	25°C (75% RH)	37°C (75% RH)
1	1	97.85	97.92
2	2	97.35	97.80
3	3	97.10	97.75

Compatibility studies by FTIR

The drug and excipient compatibility studies were carried out by FTIR study. The study showed peaks for the corresponding functional groups in temozolomide. When the study was carried out with temozolomide and polymers, there was no major changes in the peaks. By observing the above FTIR spectrums, there is no difference between internal structures and confirmation of these samples at the molecular level. It was shown that there is no interaction between the drug and polymers used (Figs. 4 and 5).

CONCLUSION

Temozolomide implants were prepared using Hydrophilic polymer. Nine formulations were prepared using carbopol 931, carbopol 934, and carbopol 971. The pre-formulation parameters were carried out for the powder blend. All the formulations results were within the limits. Implants were prepared using extrusion method. The physical parameters of all the formulations were found to be within the limit. The *in vitro* dissolution test was conducted to all the nine formulations, among them, the formulation F7 was shown 98.78% drug release in 12 hrs, whereas the others shown less drug release. FTIR studies revealed that there were no chemical interactions between temozolomide and the polymers used in the study. Short-term stability studies of promising formulations indicated that there were no significant changes in appearance and drug content of implants.

REFERENCES

1. Yea W, Chie W. Novel Drug Delivery System. 2nd ed. New York, NY: Marcel Dekker, Inc.; 1992. p. 269.
2. Sefton MV. Implantable pumps. CRC Crit Rev Biomed Eng 1987;14:201-40.
3. Conte U, Maggi L. A flexible technology for the linear, pulsatile and delayed release of drugs, allowing for easy accommodation of difficult *in vitro* targets. J Control Release 2000;64(1-3):263-8.
4. Lee ES, Kim SW, Kim SH, Cardinal JR, Jacobs H. Drug release from hydrogel devices with rate-controlling barriers. J Memb Sci 1980;7:293-303.
5. Korsmeyer RW, Peppas NA. Macromolecular and modelling aspects of

- swelling- controlled systems. In: Roseman TJ, Mansdorf SZ, editors. Controlled Release Delivery Systems. New York: Marcel Dekker; 1983. p. 77-90.
6. Yang L, Fassihi R. Modulation of diclofenac release from a totally soluble controlled release. Drug delivery system. J Control Release 1997;44:135-40.
7. Hildgen P, McMullen JN. A new gradient matrix: Formulation and characterization. J Control Release 1995;34:263-71.
8. Lu S, Anseth KS. Photopolymerization of multilaminated poly(HEMA) hydrogels for controlled release. J Control Release 1999;57(3):291-300.
9. Lu S, Ramirez F, Anseth K. Photopolymerized, multilaminated matrix devices with optimized non-uniform initial concentration profiles to control drug release. J Pharm Sci 2000;89:45-51.
10. Qiu Y, Chidambaram N, Flood K. Design and evaluation of layered diffusional matrices for zero-order sustained-release. J Control Release 1998;51(2-3):123-30.
11. Danckwerts M, Fassihi A. Implantable controlled release drug delivery systems: A review. Drug Dev Ind Pharm 1991;17:1465-502.
12. Dash AK, Cudworth GC 2nd. Therapeutic applications of implantable drug delivery systems. J Pharmacol Toxicol Methods 1998;40(1):1-12.
13. Higuchi T. Rate of release of medicaments from ointment bases containing drugs in suspension. J Pharm Sci 1961;50:874-5.
14. Overington JP, Al-Lazikani B, Hopkins AL. How many drug targets are there? Nat Rev Drug Discov 2006;5(12):993-6.
15. Imming P, Sinning C, Meyer A. Drugs, their targets and the nature and number of drug targets. Nat Rev Drug Discov 2006;5(10):821-34.
16. Zaremba T, Curtin NJ. PARP inhibitor development for systemic cancer targeting anticancer agents. Med Chem 2007;7(5):515-23.
17. Dinca EB, Sarkaria JN, Schroeder MA, Carlson BL, Voicu R, Gupta N, et al. Bioluminescence monitoring of intracranial glioblastoma xenograft: Response to primary and salvage temozolomide therapy. J Neurosurg 2007;107(3):610-6.
18. Nalnees B. Formulation and evaluation of sustained- release matrix tablets of nitrofurantoin. Int J Chem Technol Res 2013;5(1):491-501.
19. Lachman L, Lieberman HA, Kanig JL. The Theory and Practice of Industrial Pharmacy. 3rd ed. Vargheese Publishing House; 1991.
20. Jameela SR, Kumary TV, Lal AV, Jayakrishnan A. Progesterone-loaded chitosan microspheres: A long acting biodegradable controlled delivery system. J Control Release 1998;52(1-2):17-24.
21. Islam S, Islam S, Urmi AB. Observation of the release of aspirin from gelatin-sodium alginate polymeric implant. J Chem Pharm Res 2012;4(12):5149-56.
22. Salaria B, Murthy RS, Solanki A. Preparation and evaluation of chloroquine phosphate microspheres using cross linked gelatin for long term drug delivery. Indian J Pharm Sci 2002;64:48-52.
23. Karina CR, Riesta P, Esti H. Preparation and evaluation of ciprofloxacin implants using bovine hydroxyapatite-chitosan composite and glutaraldehyde for osteomyelitis. Int J Pharm Pharm Sci 2016;8(1):45-51.
24. Brahmankar DM, Jaiswal SB. Biopharmaceutics and Pharmacokinetics: A Treatise. 1st ed. New Delhi: Vallabh Prakashan; 1995.