

SIMULTANEOUS REVERSE-PHASE ULTRA PERFORMANCE LIQUID CHROMATOGRAPHY METHOD DEVELOPMENT AND VALIDATION FOR ESTIMATION OF GRAZOPRE VIR AND ELBASVIR

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ABSTRACT

Objective: The objective of this study was to develop and validate rapid, specific, sensitive, and precise reverse-phase ultra performance liquid chromatography (RP-UPLC) method for the quantitative determination of grazoprevir and elbasvir, as there are no official monograph and no analytical method by UPLC.

Methods: Chromatographic separation was achieved on a Waters Acquity UPLC HSS C18 (2.1 mm × 100 mm, 1.8 micron) column with a 45:55 (v/v) mixture of 0.1% orthophosphoric acid (pH 2.8) and acetonitrile as a mobile phase, thermostated at 30°C with a short run time of 3.0 min.

Results: The retention times were 0.73 and 1.29 min for grazoprevir and elbasvir, respectively. Quantification is achieved with TUV detection at 254 nm over the concentration range of 25–150 µg/ml for grazoprevir and for elbasvir 12.5–75 µg/ml, with a correlation coefficient of 0.999 and 0.999, respectively. The developed method was validated according to the International Conference on Harmonization (ICH) guidelines with respect to linearity, accuracy, precision, specificity, and robustness. Forced degradation study was extended out under acidic, alkaline, oxidative, photolytic, and thermal conditions to demonstrate the stability-indicating capability of the developed UPLC method. The degradation products were well resolved from the main peak, thus proved the stability-indicating power of the method. The results of the analysis were validated statistically.

Conclusion: The method is precise, accurate, linear, robust, and fast. The short retention time allows the analysis of a large number of samples in a short period of time and, therefore, should be cost-effective for routine analysis in the pharmaceutical industry.

Keywords: Grazoprevir, Elbasvir, Ultra performance liquid chromatography, Development, Validation, Forced degradation.

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INTRODUCTION

Hepatitis C virus (HCV) is a single-stranded RNA virus that is estimated to affect 130–150 million people worldwide [1]. HCV is a single-stranded RNA virus that is categorized into nine distinct genotypes. HCV infection has been associated with both hepatic and extrahepatic manifestations that increase graft loss and mortality after transplantation [2]. The combination of antiviral drugs grazoprevir (Fig. 1), and elbasvir (Fig. 2), is more effective for the treatment of patients chronically infected with HCV. Elbasvir/grazoprevir (Zepatier) is a combination product with an FDA-approved indication for the treatment of chronic HCV genotypes (GTs) 1 and 4 in adults [3].

Grazoprevir chemically known as (1*R*,18*R*,20*R*,24*S*,27*S*) -*N*-{[(1*R*,2*S*)-1-[(cyclopropylsulfonyl) carbamoyl]-2-vinylcyclopropyl]-7-methoxy-24-(2-methyl-2-propanyl)-2,2,2,5-dioxo-2,2,1-dioxo-4,11,23,26-tetraazapentacyclo [24.2.1.0³.12.0⁵.10.0¹⁸.20] nonacosa-3,5,7,9,11-Benton-27-carboxamide is a direct acting antiviral medication used as a part of combination therapy to treat chronic hepatitis C, an infectious liver disease caused by infection with HCV. It is a protease inhibitor of HCV NS3/4A that prevents cleavage of the poly protein necessary for replication [4,5].

Elbasvir chemically known as dimethyl *N,N'*-{[(6*S*)-6*H*-indolo[1,2-*c*][1,3]benzoxazine-3,10-diyl]bis{1*H*-imidazole-5,2-diyl-(2*S*)-pyrrolidine-2,1-diyl}[(2*S*)-1-oxo-3-methylbutane-1,2-diyl]}] biscarbamate is a NSSA inhibitor, preventing hepatitis C viral RNA replication and virion assembly [6]. Elbasvir is a drug approved by the FDA for the treatment of Hepatitis C.

The literature survey revealed that there are a very few analytical methods [7-10] reported for the determination of elbasvir in single and in combination with grazoprevir. Referable to the deficiency of reported ultra performance liquid chromatography (UPLC) methods describing determination of the mixtures under investigation, it was deemed useful to produce simple, sensitive, and selective reverse-phase UPLC (RP-UPLC) method that could be useful for the simultaneous determination of grazoprevir and elbasvir. The suggested method was designed to be suitable for the quality assessment of these mixes in a tablet dosage form. The proposed method was validated as per the guidelines suggested by ICH [11].

METHODS

Instruments and apparatus

The Acquity waters UPLC system equipped with TUV detector and autosampler integrated with Empower Software was used to perform development and validation of the method. A Acquity HSS C18, 100 mm × 2.1 mm, a column with particle size of 1.8 µm was used as a stationary phase for chromatographic separation and determination of grazoprevir and elbasvir. Sartorius analytical balance was used for all weighings, Metsar pH meter was used for the pH measurement of buffer solution, and Labman Sonicator was used to dissolve the standard and samples.

Materials

Working standards of grazoprevir and elbasvir were gifted by spectrum laboratories (Hyderabad, India). Dosage form ZEPATIER tablets purchased from the market. HPLC-grade acetonitrile, methanol, and

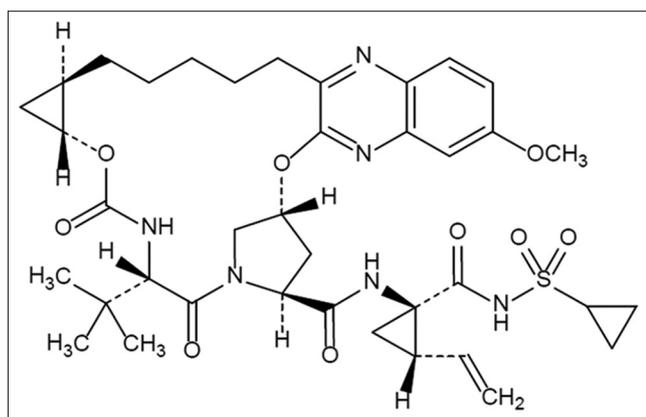


Fig. 1: Grazoprevir structure

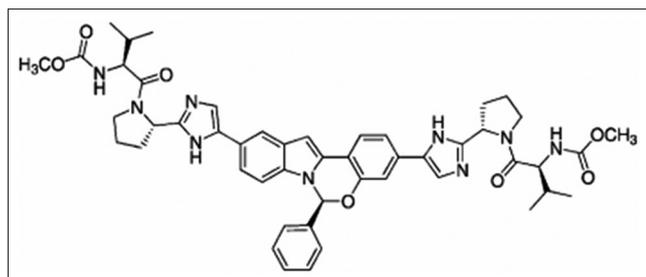


Fig. 2: Elbasvir structure

orthophosphoric acid (OPA) purchased from Merck Ltd, Mumbai, India, were used in the study.

Preparation of solutions

Preparation of standard stock solutions

Accurately weighed 10 mg of grazoprevir and 5 mg of elbasvir transferred to 10 ml of volumetric flasks separately. 8 ml of diluent was added to both of these flasks and sonicated for 10 min. Flasks were made up with diluents and labeled as standard stock solution 1 and 2 (1000 µg/ml of grazoprevir and 500 µg/ml of elbasvir).

Preparation of standard working solution (100% solutions)

About 1 ml of each stock solution was pipetted out and taken into a 10 ml volumetric flask and made up with diluents (100 µg/ml of grazoprevir and 50 µg/ml of elbasvir).

Preparation of marketing sample stock solution

A total of 10 tablets were weighed and the average weight of each tablet was calculated, then, the weight equivalent to one tablet was transferred into a 10 ml volumetric flask, 8 ml of diluent was added and sonicated for 25 min, further, the volume was made up with diluent and filtered by UPLC filters (1000 µg/ml of grazoprevir and 500 µg/ml of elbasvir).

Preparation of sample working solution (100 % solution)

About 1 ml of filtered sample stock solution was transferred to 10 ml volumetric flask and made up with diluent (100 µg/ml of grazoprevir and 50 µg/ml of elbasvir).

Buffer preparation (0.1%OPA)

About 1 ml of OPA solution taken in a 1000 ml of volumetric flask, add about 100 ml of Milli-Q water, and final volume makes up to 1000 ml with mill-Q water. Buffer pH adjusted to 2.8 with triethylamine.

Diluent

Based on the solubility of the drugs, diluent was selected, acetonitrile and water taken in the ratio of 50:50.

Method development

Chromatography

Chromatographic analysis was performed on Acquity UPLC HSS C18 (2.1 mm × 100 mm, 1.8 micron). The mobile phase consists of 0.1% OPA (pH 2.8) buffer:acetonitrile (45:55 v/v) was used throughout the analysis. The flow rate was 0.3 ml/min, the injection volume was 1.0 µl, column temperature was 30°C, run time 3 min, and detection was performed at 254 nm using a TUV detector.

Calibration curve of grazoprevir

Aliquots of working standard solution (100 µg/ml) of grazoprevir (0.25, 0.5, 0.75, 1, 1.25, and 1.5 ml) were transferred into a series of 10 ml volumetric flasks and volume was adjusted to the mark with diluents to get concentrations 25, 50, 75, 100, 125, and 150 µg/ml. Solutions were injected into the system with stated chromatographic conditions. The graph of area of peak obtained versus respective concentration was plotted. The mean area and its standard deviation were calculated.

Calibration curve of elbasvir

Aliquots of working standard solution (50 µg/ml) of elbasvir (0.25, 0.5, 0.75, 1, 1.25, and 1.5 ml) were transferred into a series of 10 ml volumetric flasks and volume was adjusted to the mark with diluent to get concentrations 12.5, 25, 37.5, 50, 62.5, and 75 µg/ml. Solutions were injected into the system with stated chromatographic conditions. The graph of area of peak obtained versus respective concentration was plotted. The mean area and its standard deviation were calculated.

Method validation

System suitability parameters

The system suitability parameters were determined by preparing standard solutions of grazoprevir (100 µg/ml) and elbasvir (50 µg/ml), and the solutions were injected 6 times and the parameters such as peak tailing, resolution, and USP plate count were determined.

Linearity

Linearity was demonstrated from 25% to 150% of standard concentration using minimum six calibration levels (25%, 50%, 75%, 100%, 125%, and 150%) for both the title drugs. The method of linear regression was used for data evaluation.

Accuracy

The accuracy of an analytical method expresses the nearness between the reference value and found value. The accuracy of the method was measured in triplicate at three concentration levels, i.e., 50%, 100%, and 150% of standard solutions of grazoprevir and elbasvir [12-14].

Limit of detection (LOD) and limit of quantitation (LOQ)

Increasingly, dilute solution of each drug was injected into the chromatograph and signal-to-noise (S/N) ratio was calculated at each concentration. The LOD and LOQ were calculated on the basis of signal-to-noise ratio of 3:1 and 10:1, respectively.

Robustness

Robustness of the method was determined to ensure its capacity to remain unaffected by small deliberate variation in the method parameters such as a mobile phase ratio, temperature of the column, and flow rate of the mobile phase [15,16].

Forced degradation studies

A thorough verification of method selectivity was carried out by forcing degradation studies, also known as stress testing. They are performed to determine possible degradation products and confirm the ability of the developed method to detect and separate impurities, which can possibly arise during the lifetime of an API or drug product. Stress tests are conducted in conditions exceeding those used in accelerated stability testing. Stress studies were performed under conditions of dry heat (thermal studies), hydrolysis (in the presence of acidic, alkaline, and water), oxidation, and photolysis. A minimum of four samples was generated for every stress condition, the blank solution

stored under normal conditions. 100% sample solution subjected to stress treatment. Hydrolytic decomposition of grazoprevir and elbasvir was conducted at 30°C in 2 N HCl, water, and 2 N NaOH. For oxidative stress studies, sample was dissolved in 20% H₂O₂ and kept for 1 day at room temperature. A photolytic study, drug solution was exposed to ultraviolet light for 1 day [17].

RESULTS

System suitability

The column efficiency, resolution, and peak symmetry were calculated for the standard solutions. The retention time of grazoprevir and elbasvir was found to be 0.72 and 1.28 min, respectively. The peaks obtained for grazoprevir and elbasvir were sharp and have clear baseline separation (Fig. 3). It was observed from the results that the system suitability parameters meet the requirement of method validation. System suitability and method validation results are summarized in Table 1.

Linearity

The calibration curves plotted for grazoprevir and elbasvir were linear over the concentration range of 25–150 µg/ml for grazoprevir, 12.5–75 µg/ml for elbasvir. Peak areas were plotted against concentrations and linear regression analysis performed for the resultant curve (Figs. 4 and 5). The correlation coefficient values of grazoprevir and elbasvir are 0.999 and 0.999, thus indicating excellent correlation between peak areas and concentrations of the analyte. The results are summarized in Table 2.

Table 1: Results from system suitability studies and validation

Parameter	Grazoprevir	Elbasvir
Theoretical plates	4589	9546
K prime (retention factor)	6.28	11.93
Tailing factor	1.86	1.55
Linearity (r ²)	0.999	0.999
% RSD for accuracy	0.88**	0.92**
% RSD for repeatability and intraday precision	0.9* and 0.8*	0.9* and 0.7*
LOD	0.20 µg/ml	0.13 µg/ml
LOQ	0.62 g/ml	0.39 µg/ml

*Results are mean of six injections. **Results are mean of three injections, LOD: Limit of detection, LOQ: Limit of quantitation

Precision

The precision of an analytical method gives information on the random error. It expresses of agreement between a series of measurements obtained from multiple sampling of the same homogeneous sample under prescribed conditions. The proposed method was evaluated by studying the precision as percentage relative standard deviation (% RSD). The % RSD was found to be less than 2 for all the drugs, which indicate that the method is precise. Data on the relative retention times obtained in a series of four consecutive injections also showed acceptable repeatability when analyzed not only on the same day but also on the 2nd day. This is confirming good precision of the method.

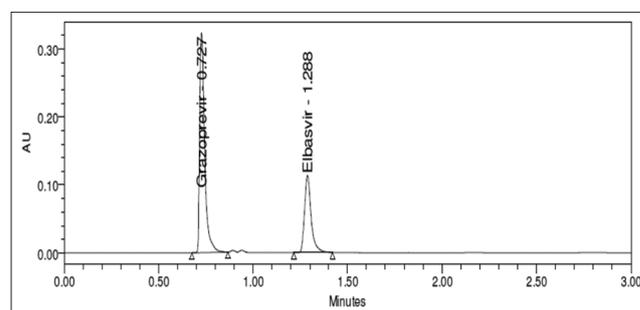


Fig. 3: Ultra performance liquid chromatography chromatogram of standard

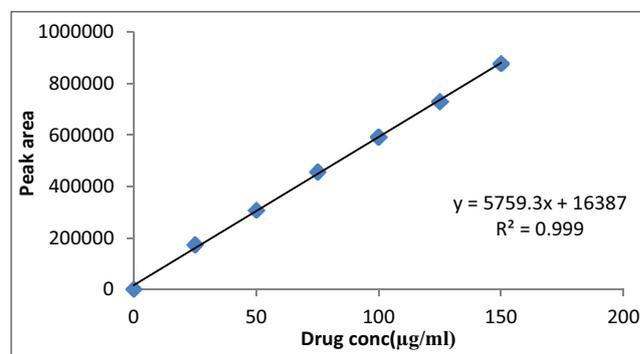


Fig. 4: Linearity curve for grazoprevir

Table 2: Linearity data for grazoprevir and elbasvir

% Level (approx)	Grazoprevir		Elbasvir	
	Concentration (µg/ml)	Peak area	Concentration (µg/ml)	Peak area
25	25	172958	12.5	69748
50	50	308953	25	140597
75	75	456108	37.5	208627
100	100	592958	50	274082
125	125	728638	62.5	348669
150	150	878708	75	411259
Slope	5759.3		5500.3	
Intercept	16387		1308	

Table 3: Robustness data for grazoprevir and elbasvir

Condition	Grazoprevir			Elbasvir		
	RT	Peak area	% RSD	RT	Peak area	% RSD
Flow rate (-) 0.2 ml/min	0.81	701762	0.8	1.49	328367	1.3
Flow rate (+) 0.4 ml/min	0.65	225665	0.8	1.14	225665	0.8
Mobile phase (-) 40B: 60A	0.71	524490	0.6	1.20	234585	0.6
Mobile phase (+) 50B: 60A	0.71	522215	0.5	1.31	233907	0.4
Temp (-) 25°C	0.71	520713	0.5	1.31	231380	0.9
Temp (+) 35°C	0.72	553682	0.7	1.30	246118	1.1

Table 4: Degradation data of grazoprevir and elbasvir

Type of degradation	Grazoprevir			Elbasvir		
	Area	% Recovered	% Degraded	Area	% Recovered	% Degraded
Acid	566244	95.00	5.00	262482	95.19	4.81
Base	580458	97.38	2.62	267861	97.14	2.86
Peroxide	586306	98.36	1.64	271028	98.29	1.71
Thermal	590308	99.03	0.97	274131	99.42	0.58
UV	591869	99.29	0.71	273416	99.16	0.84
Water	592662	99.43	0.57	273398	99.15	0.85

Accuracy

The accuracy of the proposed method was determined by analyzing grazoprevir and elbasvir samples spiked at three different concentration levels in triplicate. To find out the accuracy, a known amount of standard drug was added to the fixed amount of preanalyzed sample solution at three different concentration levels in triplicate. Percent recovery of the drugs was calculated by comparing the area before and after the addition of the standard drug. Percent recovery of grazoprevir ranged from 98.15% to 100.88% and for elbasvir 98.36–101.03% showing better accuracy of the method.

LOD and LOQ

The LOD for grazoprevir and elbasvir was found to be 0.20 µg/ml and 0.13 µg/ml, respectively (Fig. 6). The LOQ for grazoprevir and elbasvir was found to be 0.62 µg/ml and 0.39 µg/ml, respectively (Fig. 7).

Robustness

No significant effect was observed on system suitability parameters such as theoretical plates, purity angle, and purity threshold when small but deliberate changes were made for chromatography conditions such as change in flow rate (±5%), temperature (±5%), and organic content (±2%). The results are summarized in Table 3.

Forced degradation studies

The specificity of the method was also evaluated by the forced degradation study. The peak purity angle is smaller than that of peak threshold angle means there was no interface with the analyte peak from degradation products. Major degradation occurred for grazoprevir and elbasvir under acid hydrolysis condition up to 5% and 4.81%, respectively. The results are summarized in Table.4.

Assay

The validated method was applied for the determination of grazoprevir and elbasvir in commercially available ZEPATIER tablets. The results of the assay (n=3) undertaken yielded 99.69% (% RSD = 0.47) of grazoprevir and 98.85% (% RSD = 0.61) of elbasvir. The data are given in Table 5.

DISCUSSION

The present study was aimed at developing a precise, sensitive, rapid, and accurate UPLC method for the analysis of grazoprevir and elbasvir in bulk drug and in pharmaceutical dosage form. Developing a UPLC method was to reduce the run time of the method and solvent consumption for routine analysis such as assay, dissolution, and content uniformity during quality assurance. The author developed stability indicating RP-UPLC method and validated for simultaneous estimation of grazoprevir and elbasvir in bulk and tablet dosage form using Acquity UPLC HSS C18 (100 × 2.1 mm, 1.8 mm). The mobile phase consists of 0.1% OPA buffer: Acetonitrile (45:55 v/v) was used throughout the analysis. The flow rate was 0.3 ml/min, the injection volume was 1.0 µl, column temperature was 30°C, run time 3 min, and detection was performed at 254 nm using a TUV detector. The retention time of grazoprevir and elbasvir was found to be 0.73 and 1.29 min, respectively. The linearity was found satisfactory in the range of 25–150 µg/ml and 12.5–75 µg/ml for grazoprevir and elbasvir showed good correlation coefficient values 0.999 and 0.999, respectively. Less % RSD values showed the good precision and accuracy of the method.

Table 5: Assay of formulation

Drug	Label claim (mg)	Quantity found (mg)	% RSD	% Assay
Grazoprevir	100	99.69	0.47	99.69
Elbasvir	50	49.42	0.61	98.85

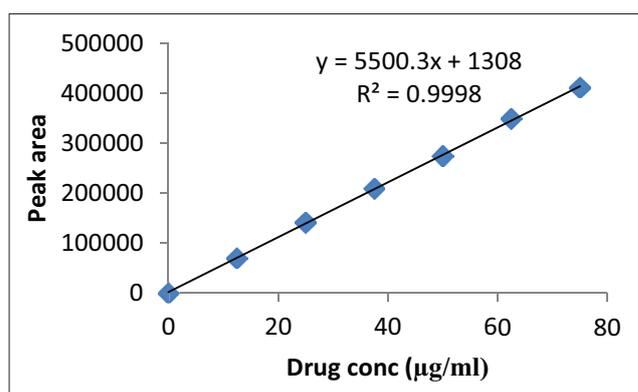


Fig. 5: Linearity curve for elbasvir

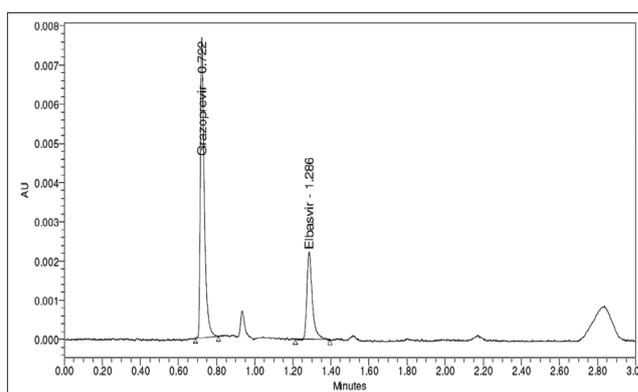


Fig. 6: Ultra performance liquid chromatography chromatogram for limit of detection of grazoprevir and elbasvir

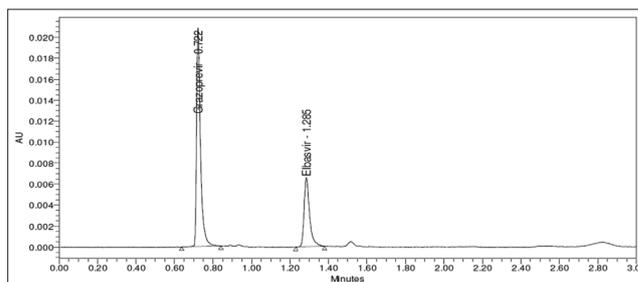


Fig. 7: Ultra performance liquid chromatography chromatogram for limit of quantitation of grazoprevir and elbasvir

The results obtained by the forced degradation studies were good enough to say that developed method was stable. The comparison of the developed method with published methods shows the developed method is new simple, rapid, robust, and economical method.

CONCLUSION

A rapid and robust method for the analysis of grazoprevir and elbasvir was developed and its applicability as a method for analyzing stability was checked. The method was found to be specific, accurate, precise, and reproducible. Force degradation studies confirmed its ability to determine stability because no interference from degradation products was observed. Moreover, no influence from excipients was found, allowing it to be used in the final drug product analysis. The method was also validated in accordance with ICH requirements.

CONFLICTS OF INTERESTS

All authors declare no conflicts of interests.

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Nil.

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