

FABRICATION AND EFFECT OF PROCESS VARIABLES OF SITAGLIPTIN MICROSPHERES

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ABSTRACT

Objective: The study is to formulate and assess the effects of different variables on the release profile of sitagliptin microspheres.

Methods: The microspheres were prepared by emulsion-solvent diffusion method and ionotropic gelation method using ethyl cellulose and sodium alginate as the polymers, respectively. The formulations are optimized by applying 2³ factorial design based on the drug-polymer ratio, stirring speed, and method of preparation.

Results: The drug-polymer interaction was checked by the Fourier-transform infrared spectroscopy and differential scanning calorimetry the results of which indicated no incompatibility. The formulated sitagliptin microspheres were evaluated for shape, morphology, particle size, the degree of swelling, encapsulation efficiency, *in vitro* drug release studies for 12 h, and kinetics of drug release.

Conclusion: The results showed that the drug-polymer ratio and stirring speed affected the particle size and drug release. The release of the drug was found to be sustained, and diffusion path is following cube root law of Hixson-Crowell kinetics. The batch F3 was found to be desirable and was further characterized by scanning electron microscope for morphology.

Keywords: Sitagliptin, Factorial, Ethyl cellulose, Solvent diffusion.

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INTRODUCTION

Microspheres are impregnable particles ranging from 1 µm to 1 mm containing dispersed medication in either solution or microcrystalline contour. Microcapsules are belittled particles that contain a dynamic agent as a gist material and coating agent as a shell. At present, there is no generally acknowledged size range that particle must have to be named as microcapsules. Commercial microcapsules ordinarily have a width between 3 and 80 µm and contain 10–90 weight % cores. The microsphere is a quickly extending innovation. It is the way of applying moderately thin coatings to little particles of solids or droplets of fluids and dispersions [1]. The microsphere is accepting impressive consideration generally, formative and industrially. The microspheres comprise proteins or biodegradable polymers in nature which are usually free streaming powders. Strong biodegradable microcapsules consolidating a medication dispelled or dethawed all through the molecule framework have the potential for the controlled arrival of medication [2].

The World Health Organization stated that more than 180 million persons are suffering from abnormal high glucose level globally. The predominance of diabetes is anticipated to two-fold in next 15 years, goaded by untoward way of life changes. Sitagliptin is the new and foremost drug in this new class of medications to be sanctioned by Food and Drug Administration. For the patients who are not able to maintain the control over blood glucose, sitagliptin helps in keeping them in control. Sitagliptin has been affirmed as a monotherapy and as an extra treatment to two different sorts of oral diabetes meds, metformin, and thiazolidinediones. perhaps, sitagliptin is useful in averting diabetes in those patients with prediabetes [3].

MATERIALS

Sitagliptin is obtained as gift sample from Richer Pharmaceuticals, Hyderabad. Ethyl cellulose and sodium alginate are obtained as gift samples from Maan Pharmaceuticals, Ahmedabad. All other chemicals used were of analytical grade.

Compatibility study of drug and the polymer

Fourier-transform infrared absorption spectra: 2 mg of the substance being examined was triturated with 300–400 mg of finely powdered and dried potassium bromide. This quantity was usually sufficient to give a disc of 13 mm diameter and a spectrum of suitable intensity. The mixture was ground carefully, spread it uniformly in a suitable die, and submit in a vacuum to a pressure of about 800 MPa (8t.cm⁻²) [4].

Differential scanning calorimetry (DSC)

Thermal analyses of sitagliptin, ethyl cellulose, sodium alginate, calcium chloride, and physical mixture were performed using a DSC to study the thermal behavior of samples. All samples were heated in hermetically sealed aluminum pans at a constant scanning rate of 10°C/min from 40 to 260°C applying the minimum possible pressure under a nitrogen atmosphere. An empty aluminum pan was used as reference [5].

Formulation of sitagliptin microspheres

The sitagliptin microspheres are prepared by emulsion solvent diffusion method (ESD) and ionotropic gelation (IG) method.

ESD method

The drug sitagliptin was dissolved in 15 ml of acetone. Ethyl cellulose is dissolved in the solvent mixture of ethanol and dichloromethane (1:1). The drug is then dispersed in the polymer solution and stirred well for uniform dispersion. The polymer solution containing drug was then emulsified in an aqueous phase containing 100 ml of 0.1% Tween 80 and stirred well with a mechanical stirrer for 2 h at room temperature to allow the volatile solvent to evaporate. The prepared microspheres were then collected on Whatman filter paper, dried and stored in desiccator [6,7].

IG method

Sodium alginate was dissolved in distilled water, and the drug was dispersed in the polymer solution with vigorous agitation. The

drug-polymer solution was then introduced gently into 5% w/v solution of calcium chloride through 21G stainless steel needle and stirred at a constant speed for 2 h to improve their mechanical strength. Then, microspheres are decanted, washed with water, allowed to dry at room temperature for 24 h and stored in a desiccator (Table 1) [6,7].

Statistical optimization technique

The optimization was designed statistically using 2³ factorial design using Minitab® Statistical Software (Version 17). A 2-level 3-factor full-factorial design consists of 8 full-factorial design points. According to the model, 8 experiments were conducted in total (Table 2). For this study, X₁ - method of preparation, X₂ - drug: Polymer concentration, and X₃ - stirring speed were selected. The dependent variables were Y₁ - particle size analysis, Y₂ - degree of swelling, Y₃ - encapsulation efficiency, and Y₄ - % drug release. The factors and levels of independent variables and independent variables are as shown in Tables 3 and 4, respectively. The results obtained from the experiment were statistically analyzed for response variables using Minitab® Statistical Software (Version 17). The statistical model incorporating interactive and polynomial terms was used to evaluate the response:

$$Y = b_0 + b_1X_1 + b_2X_2 + b_3X_3 + b_{12}X_1X_2 + b_{13}X_1X_3 + b_{23}X_2X_3 + b_{123}X_1X_2X_3$$

Physicochemical evaluation of sitagliptin microspheres

Shape and surface morphology

Shape and surface morphology of the microspheres were observed using scanning electron microscopy (SEM). Samples mounted on an aluminum stub were sputter coated with gold under the reduced pressure and a thick gold coating was applied using a sputter coater. The sample was placed under the microscope and vacuum was applied. The microspheres were observed under SEM [5].

Particle size analysis

The particle size of the microsphere was determined using optical microscopy method. The microspheres were counted approximately for particle size using a calibrated optical microscope fitted with an ocular micrometer and a stage micrometer [7,8].

Yield of microspheres

The prepared microspheres were collected and weighed. The actual weight of obtained microspheres divided by the total amount of all drug and polymer material that was used for the preparation gives the yield of microspheres [7].

$$\% \text{Yield} = \frac{\text{Actual weight of the microspheres}}{\text{Total weight of the drug and polymer}} * 100$$

Degree of swelling of microspheres

The degree of swelling was calculated using phosphate buffer pH 6.8 without enzyme. In all the formulations, the quantities of microspheres were accurately weighed and placed in the Petri dish which was completely immersed in the phosphate buffer pH 6.8. After 2 h, the microspheres were removed dried by filter paper and

weighed accurately again [9]. Then, the degree of swelling was calculated as,

$$\text{Degree of swelling} = \frac{W_2 - W_1}{W_1} * 100$$

Where,

W₁ = Initial weight of the dry microspheres

W₂ = Final weight of the swollen microspheres

Encapsulation/incorporation efficiency

An accurately weighed quantity of microspheres equivalent to 100 mg of the drug was crushed and dissolved in 100 ml of phosphate buffer pH 6.8 in a volumetric flask and stirred for 12 h. After stirring, the solution was filtered through Whatman filter paper, and the filtrate was diluted using phosphate buffer pH 6.8 and absorbance was measured for the determination of un-entrapped drug at 267 nm using UV spectrophotometer. Values are taken to calculate the drug loading efficiency [9].

$$\text{Encapsulation efficiency} = \frac{\text{Calculated drug content}}{\text{Theoretical drug content}} * 100$$

In vitro drug release

The drug release rate of the microspheres was determined in 900 ml of phosphate buffer pH 6.8 using USP XXII dissolution apparatus type 2 (paddle type). An equivalently weighted amount of microspheres equivalent to 100 mg was placed in a non-reacting muslin cloth having a smaller mesh size than the microspheres. The cloth was tied with a nylon thread to avoid the escape of any microspheres. The temperature of the medium was maintained at 37.0±0.5°C at 50 rpm. The sample aliquots were collected at specified time intervals, diluted with the same medium and analyzed at 267 nm for drug sitagliptin. Samples withdrawn were replaced with equal volume of the dissolution medium to maintain *in vitro* sink condition [9].

Kinetics of drug release

To know the mechanism of the drug release from the microspheres, the results obtained from the *in vitro* dissolution process were fitted into different kinetic equations as follows and coefficient of correlation (r) values was calculated by regression analysis as follows [10,11]:

1. Zero-order drug release: Cumulative % drug release versus time.
2. First-order drug release: Log cumulative % drug retained versus time.
3. Higuchi's equation: Cumulative % drug release versus square root of time.
4. Peppas-Korsmeyer exponential equation: Cumulative % drug release versus log time.
5. Hixson-Crowell cube root plot: Cube root of % drug remaining versus time.

RESULTS AND DISCUSSION

Compatibility study

It was concluded that the drug along with the polymers showed no change in any characteristic peak of the drug, which confirms that

Table 1: Formulation of sitagliptin microspheres

Ingredients	F1	F2	F3	F4	F5	F6	F7	F8
Sitagliptin (parts)	1	1	1	1	1	1	1	1
Ethyl cellulose (parts)	2	2	3	3	-	-	-	-
Sodium alginate (parts)	-	-	-	-	2	2	3	3
Calcium chloride (% w/v)	-	-	-	-	5	5	5	5
Stirring speed (rpm)	600	1200	600	1200	600	1200	600	1200
Method of preparation	ESD	ESD	ESD	ESD	IG	IG	IG	IG

ESD: Emulsion solvent diffusion, IG: Ionotropic gelation

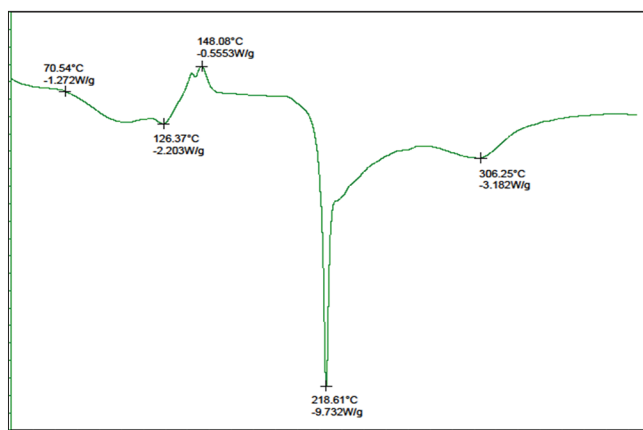


Fig. 3: Differential scanning calorimetry of sitagliptin

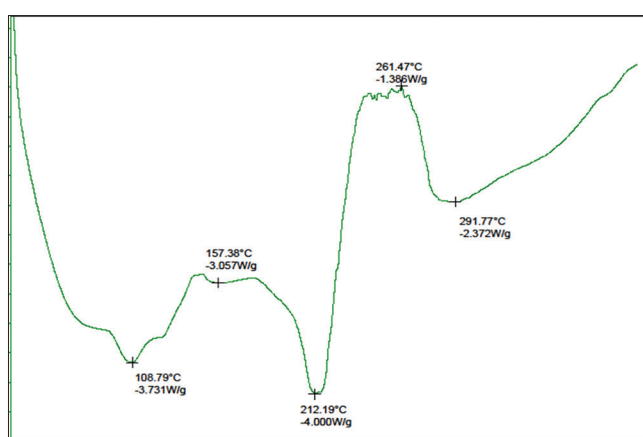


Fig. 4: Differential scanning calorimetry of sitagliptin + ethyl cellulose + sodium alginate

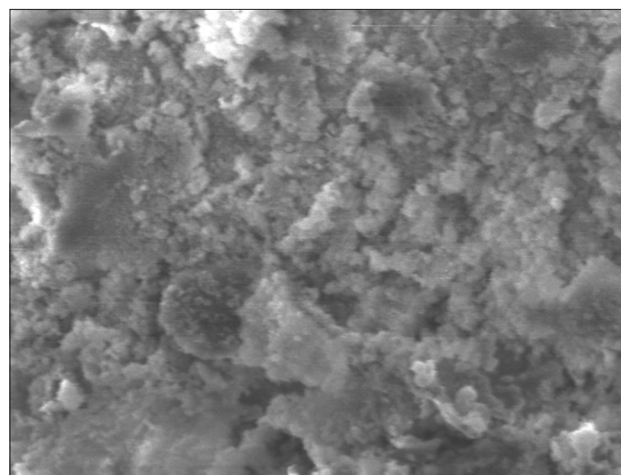


Fig. 5: Scanning electron microscopy of batch F3 microparticles ×600

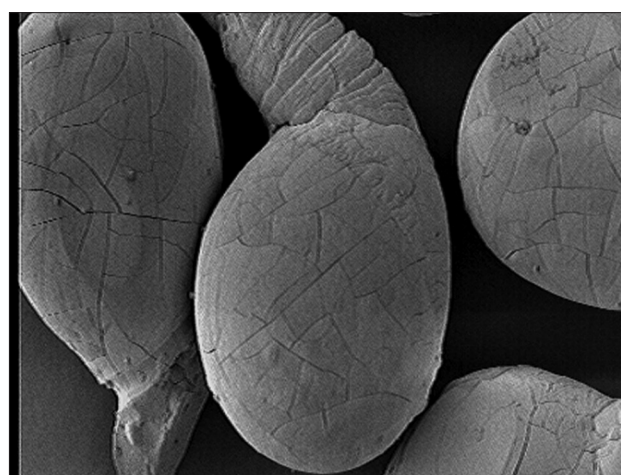


Fig. 6: Scanning electron microscopy of batch F6 microspheres at ×40

The ESD formulations were best expressed by zero-order drug release and Hixson-Crowell as the plots showed the highest linearity. The IG formulations were best expressed by zero-order drug release and Higuchi. The drug release pattern of the formulations shows the best fit with the highest correlation coefficients for Hixson-Crowell indicating that the release of sitagliptin is controlled by diffusion. This indicates that the change in surface area, the diameter of the dissolved particles and the change in diffusion path length during the dissolution process follows the cube root law [15,16].

DISCUSSION [15,17-19]

F1-F4: ESD method

The microparticles of formulations from F1 to F4 which are prepared by ESD method were irregularly spherical in shape. The percentage yield increased with increase in polymer concentration from 88.3% to 93.6%. Stirring speed does not show any valid effect on the percentage of yield. Particle size increased with increase in the polymer concentration due to the viscosity character of the polymer which comes in contact with the drug to form microparticles. The particle size decreased with increase in stirring speed. Encapsulation efficiency was increased with the increase in the polymer to drug ratio. Since the drug is well soluble in aqueous media, the encapsulation efficiency is good overall in this method. Stirring speed had a decreased effect on the encapsulation efficiency. The degree of swelling is less when compared to that of the other method because ethyl cellulose is not much swellable than sodium alginate. Encapsulation efficiency increases with increase in polymer concentration. The *in vitro* drug release rate decreased significantly with increasing the amount of

polymer because of the thickness of polymer around the drug particle which takes more time to diffuse out throughout the polymer matrix. The drug release in having a positive effect with an increase in the stirring speed which may be because the particle size is reduced and surface area is more [15,17-19].

F5-F8: IG method

The microparticles of formulations from F5 to F8 which are prepared by IG method where almost spherical in shape with some tail like formation in it due to the concentration of sodium alginate. There is no much effect on percentage yield due to polymer concentration or stirring speed. The particle size of the microparticles has been significantly increased when compared to that of the other method because of the swelling and viscous character of sodium alginate, and there was a slight decrease in particle size with respond to increase in stirring speed. Thus, stirring speed has a negative effect on particle size in the microparticles prepared by IG method using sodium alginate. The degree of swelling is considerable high because sodium alginate is aqueous soluble and forms a gel with maximum swelling when comes into contact with the aqueous solvent. The polymer concentration shows positive effect with a degree of swelling. Here also, the encapsulation efficiency increases with increase in polymer concentration. The *in vitro* drug release rate decreased with increase in polymer concentration which makes the drug difficult to diffuse out of the polymer matrix. The decrease in particle size increased the dissolution rate due to the increase in the effective surface area of the particle to that of the aqueous medium [15,17-19].

Factorial equations

All the polynomial equations were found to be statistically significant determined using Minitab® statistical software. The equation can draw a conclusion after considering the magnitude of the coefficient and mathematical sign carried (Table 5).

Effect of formulation variables on particle size (Y₁)

The mean particle size of the microspheres ranged from 300 μm to 2 mm. The mean size increased with increasing X₂ (1:2-1:3) which produces a significant increase in the viscosity, leading to the formation of larger size emulsion droplets and finally a higher microsphere size, particularly in IG method. The mean size was also influenced by X₃. It is observed that microspheres prepared using ESD method does not show a significant variation in their mean size value. Notably, in IG method, when X₂ ratio was increased from 1:2 to 1:3 there was the formation of microspheres with larger sizes due to an increase in solution viscosity of the polymer sodium alginate (Fig. 11).

Effect of formulation variables on degree of swelling (Y₂)

The degree of swelling of the microspheres ranged from 2 to 5. The contour plot clearly shows that X₂ influences the Y₂ and X₃ have no influence on the swelling capacity of the polymer. This shows that the swelling capacity is decided by the nature of the polymer, not by the X₃. Furthermore, with increase X₂ (1:2-1:3) viscosity also increases which ultimately results in more swelling. Sodium alginate is a hydrophilic polymer, and ethyl cellulose is a hydrophobic polymer which actually influences the swelling of microspheres and viscosity of the solution.

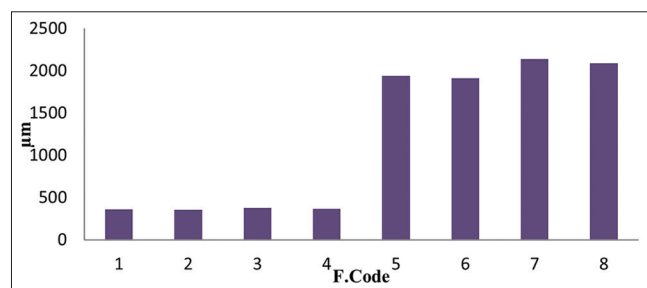


Fig. 7: Schematic representation of particle size analysis of F1-F8

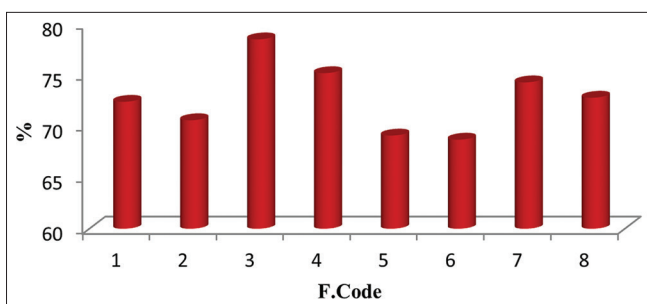


Fig. 8: Schematic representation of encapsulation efficiency of F1-F8

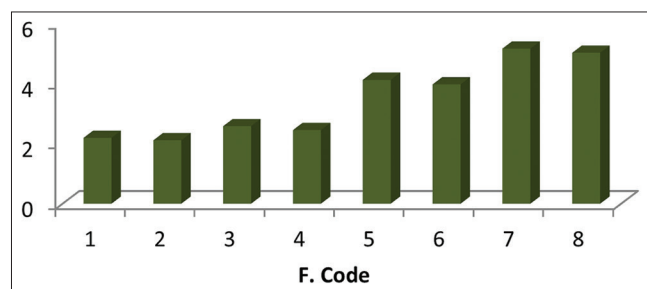


Fig. 9: Schematic representation of degree of swelling of F1-F8

It is observed that microspheres prepared using ESD method does not show a significant Y₂ and IG method, Y₂ is greatly charmed with an increase in X₂ (Fig. 12).

Effect of process variables on encapsulation efficiency (Y₃)

The entrapment efficiencies Y₃ ranged from 68% to 78%. The entrapment efficiency of sitagliptin is dependent on its solubility. Since sitagliptin is soluble in both water and ethanol, the Y₃ in both methods is nearly equal in both the method of preparations. The increase in X₂ has well influenced Y₃. The polymer concentration X₂ is directly proportioned to the entrapment efficiency Y₃. Hence, lower the polymer concentration lesser the entrapment efficiency and vice versa. However, the highest entrapment efficiency is observed in ESD method, may be due to the less viscosity of the ethyl cellulose when compared to the sodium alginate. The X₃ stirring speed has opposite effect on the entrapment efficiency Y₃, i.e., the stirring speed X₃ is inversely proportional to the entrapment efficiency Y₃. Thus, as the speed is increased the entrapment efficiency has been decreased and vice versa (Fig. 13).

Effect of process variables on drug release characteristics

The drug release ranged from 78% to 98%. The contour plot clearly shows that drug-polymer concentration X₂ influences more in drug release Y₄. The drug release Y₄ is directly proportional to the polymer concentration X₂. As the polymer concentration increases in a microsphere, the drug takes a long time to diffuse through the polymer, so that drug release is getting slower and vice versa. The stirring speed X₃ has an indirect effect on drug release Y₄ because of small particle size in ESD method. Since the particle size Y₁ is slightly affected by stirring speed X₃, the drug release Y₄ is also slightly disturbed because of the same (Fig. 14).

CONCLUSION

In the present endeavor, the microspheres were prepared in two different methods of preparations for their ease of preparation. The reason for using two different polymers is to observe the release-modifying characteristics of the polymers. The shape of the

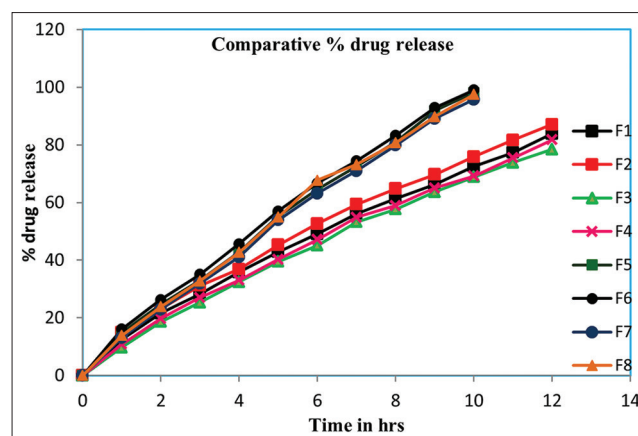


Fig. 10: Comparative % drug release

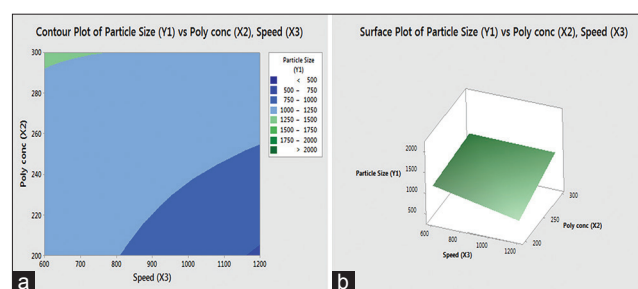


Fig. 11: (a and b) Contour and surface plots of particle size Y₁

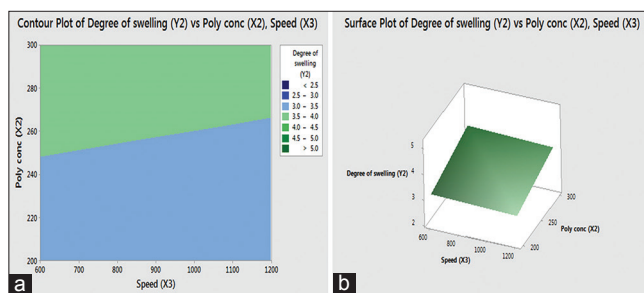


Fig. 12: (a and b) Contour and surface plots of degree of swelling Y_2

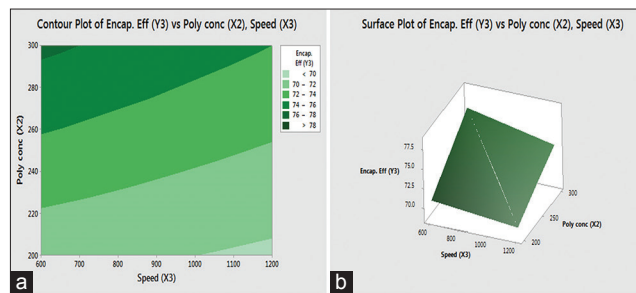


Fig. 13: (a and b) Contour and surface plots of encapsulation efficiency Y_3

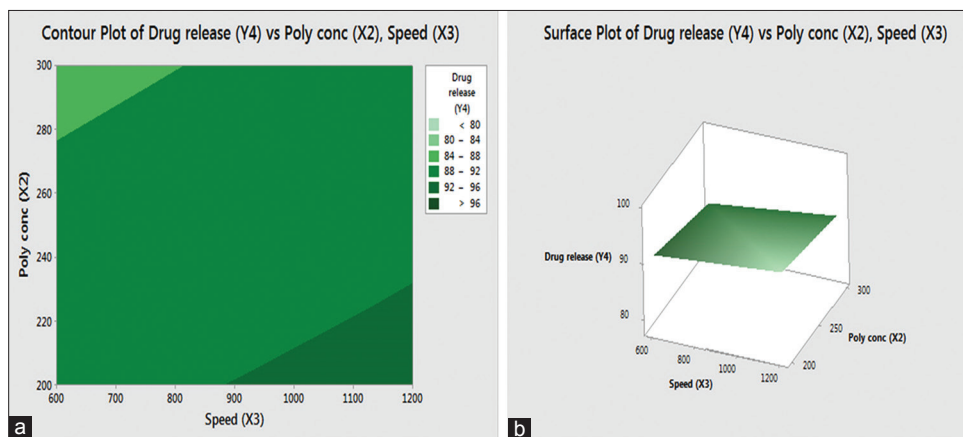


Fig. 14: (a and b) Contour and surface plots of drug release Y_4

Table 4: Kinetic values of formulations F1 to F8

Formulation code	Zero-order		First-order		Higuchi		Peppa's		Hixson-Crowell	
	R^2	R^2	R^2	R^2	R^2	n	R^2	R^2		
F1	0.988	0.972	0.968	1.201	0.752	0.992				
F2	0.985	0.966	0.971	1.185	0.728	0.991				
F3	0.990	0.986	0.960	1.238	0.794	0.997				
F4	0.990	0.974	0.962	1.223	0.777	0.993				
F5	0.996	0.814	0.943	1.367	0.743	0.935				
F6	0.993	0.773	0.952	1.352	0.727	0.928				
F7	0.996	0.880	0.940	1.387	0.763	0.959				
F8	0.995	0.824	0.939	1.374	0.752	0.936				

Table 5: The regression coefficients for each term in the regression model are summarized as follows

$$\begin{aligned}
 Y_1 &= 1090 - 725.1 X_1 - 153.8 X_2 + 114.3 X_3 + 145.9 X_1 X_2 - 110.4 X_1 X_3 + 99.73 X_2 X_3 - 100 X_1 X_2 X_3 \\
 Y_2 &= 3.450 - 1.118 X_1 - 0.3525 X_2 + 0.06250 X_3 + 0.1700 X_1 X_2 - 0.01000 X_1 X_3 - 0.005000 X_2 X_3 - 0.007500 X_1 X_2 X_3 \\
 Y_3 &= 72.70 + 1.475 X_1 - 2.500 X_2 + 0.8750 X_3 - 0.1750 X_1 X_2 + 0.4000 X_1 X_3 - 0.3250 X_2 X_3 - 0.05000 X_1 X_2 X_3 \\
 Y_4 &= 90.21 - 7.406 X_1 + 1.844 X_2 - 1.171 X_3 + 0.8637 X_1 X_2 - 0.4763 X_1 X_3 + 0.1337 X_2 X_3 - 0.08125 X_1 X_2 X_3
 \end{aligned}$$

microparticles is observed to be irregularly spherical in shape. The microspheres prepared by the method of ESD had a sustained effect of polymers more than 12 h may be because of the polymer ethyl cellulose which is not aqueous soluble, and diffusivity of the drug through it is difficult when compared to another polymer. The microparticles prepared by the IG method showed complete drug release in 10 h may be due to aqueous solubility of the polymer sodium alginate even though the size of the particles is larger than that of the other. Based on the percentage of drug release and encapsulation efficiency, the

formulation F3 behaves the best formulation among these optimized formulations showing the sustained diffusion drug release of about 78.5% following Hixson-Crowell kinetics.

AUTHORS CONTRIBUTION

S. Revathi has carried out the work and investigated the study. Dr. Dhanaraju is the principal advisor who supported with ideas and provided the mechanical resources of research work to do the overall study.

CONFLICTS OF INTEREST

The authors declare no competing interests.

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