STABILITY-INDICATING HIGH-PERFORMANCE LIQUID CHROMATOGRAPHY METHOD FOR DETERMINATION OF ANTIHISTAMINE DRUG AZELASTINE

SHITAL PATEL*, PASHA TY
Department of Pharmaceutical Analysis, Parul Institute of Pharmacy, Ta.-Waghodia, Vadodara - 391 760, Gujarat, India.
Email: shital_3012@yahoo.com

Received: 27 February 2018, Revised and Accepted: 01 May 2018

ABSTRACT

Objective: The objective of this research was to develop a simple, precise, accurate, and stability-indicating reverse-phase high-performance liquid chromatographic method for estimation of azelastine hydrochloride (AZL) in nasal spray preparation.

Methods: Chromatography was performed on a 250 mm×4.6 mm, 5-μm particle size, Waters Spherisorb CN column using (50:50 v/v) mixture of potassium dihydrogen phosphate buffer and acetonitrile as mobile phase. The detection was carried out at 290 nm and flow rate employed was 1.0 ml/min. The degradation of AZL was studied under different ICH recommended stress conditions.

Results: The retention time was 4.34 min for AZL. Linearity was established in the concentration range of 5–120 µg/ml with a correlation coefficient of 0.9996. Limit of detection (LOD) and limit of quantitation (LOQ) were found to be 0.81 µg/ml and 2.44 µg/ml, respectively. Percentage recovery was found between 99 and 102%. The values of percentage relative standard deviation (<2%) proved the high precision of the proposed method. The method was found to be robust regarding any small variation in the column temperature, pH of mobile phase, and mobile phase ratio. AZL was found stable in 5 M HCl at 80°C for 5 h, 5 M NaOH at 80°C for 5 h, 30% H₂O₂ at 80°C for 5 h, and in oven at 70°C for 8 h.

Conclusion: The results obtained in this research work clearly proved that the proposed HPLC method for the assay of AZL in nasal spray preparation is simple, precise, specific, accurate, and stability indicating. It clearly indicates that the method is suitable for analysis of AZL in the raw material and the pharmaceutical product without interference from excipients.

Keywords: Azelastine hydrochloride, High-performance liquid chromatography, Degradation study, Stability-indicating method.

INTRODUCTION

Allergic reaction produces inflammation, and people, who suffer from nasal allergies such as hay fever, dust, mite, and pet allergies, tend to experience a variety of symptoms due to this inflammation. Inflammation of the nasal passage forces fluid out of the nasal tissues, resulting in a runny and blocked nose. The three main types of drugs available for anti-inflammatory and antiallergic effect are corticosteroids, antihistamines, and decongestants [1].

Azelastine hydrochloride (AZL), which is a second-generation H₁ receptor antagonist, chemically, it is known as 4-[(4-Chlorophenyl)methyl]-2-{hexahydro-1-methyl-1H-azepin-4-yl}-1-{2H}-phthalazinone, hydrochloride. The chemical structure of AZL is shown in Fig. 1. AZL occurs as a white, almost odorless, crystalline powder with a bitter taste. It has a molecular weight of 418.37. It also inhibits lipooxygenase and platelet aggregation. It is bronchodilator agent also. It is indicated as antiallergic agent [2,3].

AZL is an official drug in British Pharmacopoeia [2], which recommends a titrimetric method for the determination of AZL in pure form using 0.1 M perchloric acid as a titrant. Various methods such as liquid chromatography (LC) [4], LC-electrospray ionization tandem mass spectrometry [5], high-performance LC (HPLC) [6,7], capillary HPLC [8], and electrokinetic capillary HPLC [8] had been reported for the determination of azelastine and its metabolite in biofluids. Visible spectrophotometric method [9], HP thin-layer chromatography (HPTLC) [10], reversed-phase HPLC (RP-HPLC) [11], and Fourier transform infrared [12] have been developed for estimation of AZL in formulation. RP-HPLC [13] method has been reported for simultaneous

Fig. 1: Chemical structure of azelastine hydrochloride
METHODS

Instrumentation and chromatographic condition
The quantitative analysis was performed on Agilent HPLC 1260 equipped with quaternary solvent manager, autosampler, and photodiode array (PDA) detector integrated with Ez chrome software. Separation was achieved on Waters Spherisorb CN (250×4.6 mm, 5 µm) column. The standards and samples were separated using mobile phase consisting of acetonitrile:0.05 M potassium dihydrogen phosphate buffer pH 3.0 (50:50 %v/v). The flow rate was 1.0 ml/min and detection was set at 290 nm. The column temperature was set at 45°C, and the injection volume was 20 µl.

Materials and reagents
AZL (cerified purity of 99.9%, batch no. 0301425316) was kindly gifted by Cadila Health Care Limited, Moraiya (Gujarat). Arzep nasal solution was purchased from local pharmacy. Acetonitrile and water (HPLC grade) were purchased from Merck, India. Potassium dihydrogen phosphate, O-phosphoric acid (OPA), sodium hydroxide, hydrochloric acid, and hydrogen peroxide used were of analytical grade and purchased from SD Fine Chemicals, India.

Standard stock solution preparation
A standard solution of AZL containing 1000 µg/ml was prepared in acetonitrile and diluted as appropriate with the same solvent to obtain the working concentration range. Solutions were found to be stable for at least 5 days when kept at room temperature.

Buffer preparation (pH 3.0)
About 6.805 g of potassium dihydrogen phosphate was dissolved in 600 ml HPLC water and then final volume was made up to 1000 ml with HPLC water. Buffer pH was adjusted to 3.0 with ortho-OPA.

Validation of the method
The method was validated for system suitability, specificity, linearity, precision, accuracy, limit of detection (LOD), limit of quantification (LOQ), and robustness as per the ICH guideline.

System suitability
The system suitability was checked by seven replicate analyses of 100 µg/ml solution of AZL and chromatographic parameters were evaluated.

Linearity
Accurately measured volumes of the AZL standard solution were successively transferred into a series of 10 ml volumetric flasks to obtain final concentrations of 5–120 µg/ml and diluted to the mark with acetonitrile and solutions were mixed properly. 20 µl aliquots of each solution were injected 3 times and were chromatographed. The average peak area of AZL was plotted against the drug concentration, and the regression equation was derived.

Precision
Repeatability of the method was assessed by analysis of seven injections of 100 µg/ml AZL solution. Percentage relative standard deviation (%RSD) for retention time and peak area of drug was calculated. The intraday and interday precision for analysis of AZL was performed with three concentrations (20, 40, and 60 µg/ml) for 3 times on the same day and for three different days, respectively.

Specificity - forced degradation studies
The forced degradation study was executed to find whether the analytical method was stability indicating and could determine the AZL in the presence of impurities and degradation products. AZL standard solution was stressed under acid, base, oxidative, thermal, and photolytic stress conditions. 2 ml of the standard solution of AZL (1000 µg/ml) was transferred to three 20 ml volumetric flask followed by 2 ml of 5.0 M HCl, 2 ml of 5.0 M NaOH, and 2 ml of 30% H2O2 to different flask and then heated in a thermostatic water bath at 80°C for 5 h to induce degradation by acid, base, and oxidation, respectively. Then, the mixtures were diluted with acetonitrile up to the mark. 50 mg AZL powder was placed in convection oven and exposed to heat at 70°C for 8h and further preceded as per sample preparation for thermal degradation. To check photolytic degradation, AZL drug (50 mg) was taken in two Petri dishes. One Petri dish was wrapped with aluminum foil and other was not wrapped. Both the Petri dishes were kept in ultraviolet chamber (365 nm) for 24 h at room temperature and further proceeded as per sample preparation. The photostability of a drug substance depends on the wavelength, time, and intensity of exposure [17]. All stressed samples were analyzed by proposed method.

Accuracy
Accurately measured 2 ml of nasal spray equivalent to 20 mg of AZL was taken in three different 10 ml volumetric flasks. 10 mg, 20 mg, and 30 mg of AZL standard was added to three different volumetric flasks containing nasal spray preparation and diluted up to mark with acetonitrile. Further, they were diluted to have 40 µg/ml of sample in each flask. The experiment was performed in triplicate. Percentage recovery was calculated for each level.

Robustness
To determine the robustness of the method, the experimental conditions were deliberately changed. The flow rate of the mobile phase (1.0±0.1 ml/min), pH of mobile phase (3.0±0.2), mobile phase composition (±2%), and column temperature (45±1°C) was varied. In each case, the %RSD values were calculated for the obtained retention time and peak area. The number of theoretical plates and tailing factor was compared with that obtained under the optimized conditions.

LOD and LOQ
The LOD and LOQ were estimated mathematically. The mathematical formulas used were as follows:

- LOD = 3.3 (SD of y-intercept/slope of the calibration plot)
- LOQ = 10 (SD of y-intercept/slope of the calibration plot)

Analysis of the marketed formulation
To determine the AZL content of a nasal spray (label claim 1% w/w, 10 ml), solution of the container was taken out in 100 ml volumetric flask and mixed with acetonitrile (10 ml) and sonicated for 20 min, then it was diluted up to the mark with acetonitrile which made 1 mg/ml AZL concentration. 1 ml of this solution was transferred to a 10 ml volumetric flask and diluted to volume with acetonitrile which made 100 µg/ml. The analysis was repeated 3 times. The possibility of interference of excipients with the analysis was studied.

RESULTS

Development of method
To develop stability-indicating HPLC method for antihistamine drug AZL, mobile phases of various compositions were investigated and system suitability test was performed for method optimization. The mobile phase consisting of acetonitrile:0.05 M potassium dihydrogen phosphate buffer pH 3.0 (50:50 %v/v) was found optimum. The effect of flow rate was also optimized and 1.0 ml/min was considered optimum for the separation method. The column temperature was set at 45°C.

Patel and Pasha

249
Validation of method
The proposed method was validated according to the ICH guideline to prove its suitability for the intended purpose. Method validation data are summarized in Table 1.

Linearity
Peak area and concentration were subjected to linear regression analysis to calculate the calibration equation and correlation coefficient (Fig. 3). The regression data revealed a good linear relationship over the concentration range of 5–120 µg/ml.

Precision
The precision of an analytical method gives information on random error. The precision of the method was evaluated by repeatability, interday and intraday precision. The values of %RSD for all found <2% proved the high precision of the proposed method.

Accuracy
When the method was used for subsequent analysis of AZL from the nasal spray solution after spiking of AZL standard at three different levels for 3 times, the recovery was found between 99 and 102% (Table 2). The percentage recovery was calculated from the amount recovered and actual amount added. Amount recovered was calculated from comparison of area before and after spiking the standard drug.

LOD and LOQ
LOD and LOQ were found to be 0.81 µg/ml and 2.44 µg/ml, respectively, from calculation.

Robustness
The method was found to be robust regarding any small variation in the column temperature (45±1°C), pH of mobile phase (3.0±0.2), and mobile phase composition (±2%) as revealed by the constancy of the retention time and peak area. In case of flow rate of mobile phase (1±0.1 ml/min), where minor variation resulted in a significant change in the retention and peak area of the AZL.

Stability-indicating aspects
Any significant change in peak area of AZL and any additional peak were not observed when chromatographed after refluxing with 5.0 M HCl, 5.0 M NaOH, and 30% H₂O₂ for 5 h in separate flasks. No additional peak was observed and no significant change in initial concentration of AZL when solid AZL drug kept at 365 nm for 24 h. AZL samples kept under dry heat conditions furnished no additional peaks. Peak purity value of AZL was found 1.00 after exposure to forced degradation condition. Hence, AZL is stable under acid, alkali, oxidative, dry heat, and 365 nm for 24 h photolytic condition.

Assay of the pharmaceutical formulation
The proposed validated method was successfully applied to determine AZL in Arzep nasal spray solution. The recovery obtained was around 96.14%. In chromatograms of drug sample from nasal spray preparation, no interference was observed from excipients.

DISCUSSION
The present study was aimed at developing a precise, specific, robust, and accurate HPLC method for the analysis of AZL in bulk drug and in pharmaceutical dosage form. The author developed stability-indicating HPLC method and validated for the determination of AZL in bulk and nasal spray solution using Waters Spherisorb CN column (250 mm×4.6 mm, 5 µm). The mobile phase consists of potassium dihydrogen phosphate buffer (pH 3.0):acetonitrile (40:50 v/v) was used throughout the analysis. The flow rate was 1.0 ml/min, the injection volume was 20 µl, column temperature was 45°C, run time 6 min, and detection was performed at 290 nm using a PDA detector. The retention time of AZL was found to be 4.34 min. The linearity was found satisfactory in the range of 5–120 µg/ml and showed good correlation coefficient values 0.9996. Less %RSD values showed good precision of the method. The results obtained by the forced degradation studies were enough to say that the drug is stable and method is stability indicating one. The comparison of the developed method with published methods shows the developed method is simple, robust, and economical one.

### Table 1: Validation data

<table>
<thead>
<tr>
<th>Data</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Linear range</td>
<td>5–120 µg/ml</td>
</tr>
<tr>
<td>Correlation coefficient</td>
<td>0.9996±0.0002</td>
</tr>
<tr>
<td>LOD</td>
<td>0.81 µg/ml</td>
</tr>
<tr>
<td>LOQ</td>
<td>2.44 µg/ml</td>
</tr>
<tr>
<td>Accuracy</td>
<td>100.05±0.95</td>
</tr>
<tr>
<td>Precision (%RSD)</td>
<td></td>
</tr>
<tr>
<td>Repeatability (n=7)</td>
<td>1.381</td>
</tr>
<tr>
<td>Interday (n=3)</td>
<td>0.800</td>
</tr>
<tr>
<td>Intraday (n=3)</td>
<td>0.372</td>
</tr>
<tr>
<td>Robustness</td>
<td>Robust in column temperature, pH of mobile phase (3.0±0.1), and mobile phase composition</td>
</tr>
<tr>
<td>Specificity</td>
<td>Specific</td>
</tr>
</tbody>
</table>

Note: *a* = standard deviation, *b* = relative standard deviation, LOD: Limit of detection, LOQ: Limit of quantitation, %RSD: Percentage relative standard deviation

### Table 2: Results of accuracy for AZL

<table>
<thead>
<tr>
<th>% Addition</th>
<th>µg/ml added</th>
<th>µg/ml recovered</th>
<th>% recovery±SD (n=3)</th>
</tr>
</thead>
<tbody>
<tr>
<td>50</td>
<td>20</td>
<td>19.91</td>
<td>99.54±1.29</td>
</tr>
<tr>
<td>100</td>
<td>40</td>
<td>40.46</td>
<td>101.14±0.70</td>
</tr>
<tr>
<td>150</td>
<td>60</td>
<td>59.67</td>
<td>99.46±0.72</td>
</tr>
</tbody>
</table>

AZL: Azelastine hydrochloride
CONCLUSION
The proposed HPLC method for the assay of AZL in nasal spray preparation is simple, precise, specific, accurate, and stability indicating. It proves that the method is suitable for analysis of AZL in the raw material and the pharmaceutical product without interference from excipients. A stress degradation study was conducted to investigate the degradation behavior of AZL under ICH recommended condition; it shows that AZL is stable drug.

ACKNOWLEDGMENT
The authors would like to thank Cadila Health Care Limited, Moraiya, Gujarat, for providing AZL as gift sample for this research work.

AUTHOR’S CONTRIBUTION
Shital Patel has performed the work presented here. Dr. T. Y. Pasha has guided this project.

CONFLICTS OF INTEREST
All authors have none to declare.

REFERENCES
16. ICH Harmonised Tripartite Guideline Validation of Analytical Procedure: Methodology (Q2B); 1996.