INTRODUCTION

In pathological term, pain may be defined as "sensory consequence of neuronal activity triggered by noxious stimuli, inflammation or damage to specific nociceptive pathways in the nervous system [1]. It is quite evident for any clinician who treats patients in pain knows that one size does not fit all. There is no one standard dose or specific medication that will provide adequate analgesia to all patients. The optimal current approach is to titrate analgesics to produce optimal therapeutic benefit.

Gender-related influences on the experience of pain have been received considerable empirical attention in recent years [2-7]. It is generally accepted that males and females respond differently to painful conditions. Women report more pain than men and are at greater risk for developing many forms of chronic pain [2]. Sex differences are not limited to pain perception but may extend to the response to analgesics. There is little agreement about a differential response of men and women to opioid analgesics. In animals, male rats exhibit greater analgesia than female rats to equal doses of opioids [8,9]. In case of morphine, a gender difference is reported in its anti-nociceptive effect both in animals and human in spite of no gender-linked difference in serum levels of morphine [10-12]. In humans, sex differences in response to opioids have been described, but the findings are difficult to consolidate.

Leptin is secreted by adipocytes in proportion to the amount of body fat and exerts a potent inhibitory action on food intake. In humans, serum leptin concentrations correlate positively with percent body fat. Several studies have positively correlated the experience of pain with an increase in body mass index (BMI) [13,14]. It is not known whether obesity causes chronic pain, chronic pain causes obesity, or some other factor causes both concurrently. Obesity is hypothesized to lead to pain because of excess mechanical stresses and its pro-inflammatory state.

Differences in pain thresholds may have implications for pain management, as they may account in part for the variability in analgesic requirements between individuals. There is a need for further studies to investigate the correlation of gender and leptin with pain threshold and analgesic effect of opioids. Tramadol is a synthetic, centrally acting analgesic agent with two distinct, synergistic mechanisms of action, acting as both a weak opioid agonist and an inhibitor of monoamine neurotransmitter reuptake [15]. Earlier, we had reported the analgesic modulation of some analgesics in male and female Wistar rats [16]. Consequently, the aim of the present study was to investigate the correlation of gender and leptin with analgesic effect of tramadol in Wistar rats.

METHODS

Drugs and reagents

Active pharmaceutical ingredient of tramadol and rat leptin ELISA kit was procured from Sigma-Aldrich, Bengaluru, India. Acetic acid and other chemicals were purchased from Merck Life Sciences Pvt., Ltd., Mumbai, India. Normal saline (0.9% sodium chloride) was purchased from the pharmacy of Kasturba Hospital, Manipal, Karnataka, India.

Animals

A total of 48 (24 male and 24 female) Wistar rats weighing 100–150 g were housed in separate polycarbonate cages, maintained under standard conditions with temperature (22-24°C), 12-h light/12-h dark cycle and relative air humidity 40–60%. The animals were housed in separate polycarbonate cages, maintained under standard conditions with temperature (22-24°C), 12-h light/12-h dark cycle and relative air humidity 40–60%. The animals were housed in separate polycarbonate cages, maintained under standard conditions with temperature (22-24°C), 12-h light/12-h dark cycle and relative air humidity 40–60%. The animals were housed in separate polycarbonate cages, maintained under standard conditions with temperature (22-24°C), 12-h light/12-h dark cycle and relative air humidity 40–60%.
acclimatized to the laboratory conditions for 1 week before the start of the experiment. The animals were provided with a normal pellet diet (Amrit Feeds Ltd., Pune, India) and water *ad libitum*. The experimental protocol was approved by the Institutional Animal Ethics Committee (IAEC/KMC/41/2014) and experiments were conducted according to the ethical norms approved by the Ministry of Social Justice and Empowerment, Government of India, and Committee for the Purpose of Control and Supervision of Experiments on Animals guidelines.

**Experimental design**

A total of 48 Wistar rats (body weight 100–150 g). 24 each male and female Wistar rats were randomly divided into two groups (n=6/group) (Group I - Control- 0.9% NaCl; 1 ml/kg/day i.p. and Group II - Tramadol 10 mg/kg/day i.p.) for each nociception model - plantar test and acetic acid induced writhing test (Fig. 1). The treatment duration was of 5 days. On the last day of treatment (i.e. on the 5th day); 15 min after tramadol/normal saline treatment, animals were tested for the respective pain model. Paw withdrawal latency (PWL) was assessed using plantar test, and writhing movements were observed following administration of 0.8% acetic acid; 10 ml/kg i.p [16-18].

**Nociception models**

**Plantar test (Hargreaves’ method)**

Thermal pain threshold to radiant heat (IR-90) was quantified using the paw withdrawal test (Fig. 2). Rats were placed in a perspex enclosure, without restraint and a movable infrared radiant heat source placed directly under the plantar surface of the hind paw (Ugo Basile, Como, Italy). The PWL to radiant heat was defined as the time from onset of the radiant heat to the withdrawal of the rat hind paw. The cutoff time for PWL was 30 s. Testing was alternated between hind paws and carried out at 3 min intervals. The average of three estimations was taken to yield mean PWL. Before any testing was carried out, rats could adjust to their environments for at least 10 min.

**Acetic acid-induced writhing test**

Writhing movement was induced by administering an intraperitoneal injection of 0.8% acetic acid (10 ml/kg). 15 min after the tramadol/normal saline administration. After 10 min of acetic acid administration, the number of writhing movements such as abdominal constriction/elongation of body/arching of back/bind limb extension/forelimb extension/trunk twisting (Fig. 3) was cumulatively counted over 20 min for nociceptive evaluation [16-18].

**Collection of blood sample**

Following anesthesia with ketamine 60 mg/kg and xylazine 5 mg/kg; i.p., blood was withdrawn from retro-orbital plexus of rats through capillary tube [19]. Following collection of blood in micro-centrifuge tubes and its clot formation, serum was obtained by centrifugation of blood at 3,000 rpm for 20 min at 4°C using a refrigerated centrifuge (MIKRO 22R, Andreas Hettich GmbH & Co. KG, Germany). The
Estimation of serum leptin concentration

As per the assay protocol given along with rat leptin ELISA kit, all reagents and samples were kept at room temperature (22-25°C) before use. 100 µl of each standard and sample was added to appropriate wells. Wells were covered and incubated for 2.5 h at room temperature or overnight at 4°C with gentle shaking. The solution was discarded and washed 4 times with 1x wash solution. Washing by filling each well with wash buffer (300 µl) was done using a multichannel pipette. After the last wash, any remaining wash buffer was removed by aspirating or decanting. The plate was inverted and blotted against clean paper towels. 100 µl of 1x prepared Biotinylated Detection Antibody was added to each well followed by incubation for 1 h at room temperature with gentle shaking. The solution was discarded. Washing was repeated 4 times. 100 µl of prepared HRP-streptavidin solution was added to each well followed by incubation for 45 min at room temperature with gentle shaking. Solution was discarded and washed again 4 times. 100 µl of ELISA Colorimetric TMB Reagent was added to each well followed by incubation for 30 min at room temperature in the dark with gentle shaking. 50 µl of stop solution was added to each well and absorbance was read at 420 nm immediately. The mean absorbance for each set of duplicate standards, controls, and samples was entered in the Microsoft Excel sheet, and average zero standard optical density was subtracted from each mean absorbance. A standard curve with standard concentration on the X-axis and absorbance on the Y-axis was plotted and following that leptin concentration (pg/ml) was calculated.

Statistical analysis

Using Statistical Package for the Social Sciences version 16.0, data were expressed as mean±standard deviation and analyzed by one-way analysis of variance followed by post hoc Tukey test. The correlation of pain and serum leptin concentration was done using bivariate analysis followed by Pearson correlation coefficient and two-tailed significance. A level for p≤0.05 was considered as statistically significant.

RESULTS

PWL was significantly decreased (p<0.001) and both number of writhing movements and serum leptin concentrations were significantly increased (p<0.001) in female control group compared to male control group. Tramadol treated both male and female rats had significantly increased PWL (p<0.001) and decreased both number of writhing movements and serum leptin concentration (p<0.001) in comparison with both male and female control groups, respectively. In tramadol treated female rats, PWL was significantly decreased (p=0.005) and both number of writhing movements and serum leptin concentration were significantly increased (p<0.001) in comparison with the tramadol treated male rats (Fig. 4). PWL was negatively correlated with serum leptin concentration (Pearson correlation coefficient= −0.826 and two-tailed significance= 0.000), which means PWL was significantly decreasing (means more pain) with increased levels of serum leptin (Fig. 5). A number of writhing movements were positively correlated with serum leptin concentration (Pearson correlation coefficient= 0.505 and two-tailed significance=0.012), which means writhing movements were significantly increasing (means more pain) with increased levels of serum leptin levels (Fig. 6).

DISCUSSION

The present study has demonstrated the correlation of gender and leptin with analgesic modulation of tramadol in Wistar rats. We found the significant increase in serum leptin levels and decrease in pain threshold (more pain sensitivity to noxious stimuli) in female rats than male rats. The significant increase in serum leptin concentration in female rats in the present study is in agreement with Mendonca et al, who found a positive and statistically significant correlation between estrogen and leptin independent of body mass index [20]. According to Shimizu et al. and Messinis et al., estrogen can be an important regulator of leptin production in women [21,22]. The gender difference in serum leptin concentration is well established and in vitro results suggest that gonadal hormones, such as testosterone, may be important regulators of leptin secretion [23-25]. In both humans and rodents, males have lower plasma leptin concentrations than their female counterparts at any level of adiposity [26,27]. A strong inverse association between serum levels of leptin and testosterone was recently reported in untreated and testosterone-treated hypogonadal men [28,29]. In the present study also, we found significantly lower levels of leptin in male rats compared to female rats which might be due to downregulation of serum leptin by testosterone. The mechanism underlying this effect of testosterone remains to be elucidated.

Fig. 4: Comparison of paw withdrawal latency (in seconds), number of writhing movements, and serum leptin concentration (pg/ml) among experimental groups
In the present study, tramadol treatment at the dose of 10 mg/kg for 5 days has decreased serum leptin level in both male and female rats. Some of the studies also suggest the decrease in serum leptin concentration with tramadol (opioid agonist) treatment at the dose of 0.5 mg/kg/day orally when administered for 4 weeks [30,31]. If we calculate the total amount of tramadol for 5 days in our study which comes to 50 mg/kg and in the above-mentioned studies it is 14 mg/kg in the total 4 weeks duration then the dose and duration of tramadol treatment in our study is even more than these studies where serum leptin level was decreased. Decrease in serum leptin level by tramadol at the dose of 10 mg/kg for 5 days treatment duration could be one of the additional mechanisms of tramadol to reduce pain.

There was a significant positive correlation found between pain and serum leptin levels in our study. One of the effects of increased serum leptin levels may be greater sensitivity to pain. The relationship between leptin and pain may be modulated by other factors, with estrogen being one possibility [32]. In rat and mouse models, leptin has been shown to reduce the threshold for pain [33]. Leptin has also been shown to increase levels of interleukin-1, a cytokine known to cause hyperalgesia [34]. Recent studies have shown that leptin, an adipocytokine, played a significant role in nociceptive behavior induced by nerve injury in rats [35,36]. Maeda et al. suggests that peripheral effect of leptin on neuropathic pain is mediated via macrophage stimulation [36]. Following more leptin levels in female rats compared to males, these could be the possible reasons for more pain in female rats than male rats.

CONCLUSIONS
The present study revealed that female rats have more serum leptin concentration than male rats which could be one of the possible reasons for having more pain sensitivity to noxious stimuli in female rats compared to male rats. Further information regarding the link between leptin and pain may help drive novel treatments for pain. Tramadol treatment at the dose of 10 mg/kg for 5 days has decreased serum leptin level in rats which might be one of the additional mechanisms of tramadol to reduce pain. Additional research to elucidate the mechanisms driving sex hormones and leptin in pain responses is needed to foster future interventions to reduce these disparities in pain and analgesic modulation of tramadol. Gender and body mass index-specific tailoring of pain treatments may become a conceivable outcome in the foreseeable future.

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AUTHOR'S CONTRIBUTION
The first author has designed the study, carried out the experimental part of the research study, data compilation, statistical analysis, interpretation of results, and manuscript writing. The second author has guided the first author and monitored the experiment, statistical analysis, and interpretation of result. The third author has coguided the first author.

CONFLICTS OF INTEREST
All the authors declare that they do not have any conflicts of interest.

REFERENCES