INTRODUCTION

Rosaceae fruits (Fructus) are widely used in Russian medical practice crude herbal drugs (CHDs). Rosaceae fruits are delivered to the pharmacy in a packaged form for OTC distribution and used for the preparation of water extracts. Rosaceae fruits are part of medicinal tea composition, serve as raw materials for obtaining of tinctures and extracts, and are used in homeopathy [1,2]. Fresh fruits have the richest biologically active substances (BAS) composition [3,4]. Flavonoids (flavonols and anthocyanins), simple sugars, polysaccharides, tannins, and organic and hydroxycinnamic acids are among them [5-9]. Different complex drugs with wide range of pharmacological activity are produced from them [10,11].

Fresh fruits contain BAS complex presented in the plant in natural state. However, due to the high moisture content, fresh juicy fruits are exposed to enzymatic breakup and microbial contamination, these factors limit its use [12,13].

Conventionally, CHDs are subjected to drying [14,15]. An alternative method of conservation is freezing [16,17]. Due to the absence of a liquid phase, the activity of enzymes ceases, and as a result, biochemical processes are suspended. Both drying and freezing have their advantages and disadvantages, because during these processes, the content of BAS in the raw materials varies, which can lead to a decrease in the quality of CHD.

The drying method, temperature and velocity of drying influence the temperature and quality of BAS presented in CHD. Research data indicate that drying air temperatures between 50 and 60°C appear to be suitable for drying different CHD types [18]. In Aronia melanocarpa fruits, the highest content of total flavonoids, polyphenols, and total anthocyanins was detected in freeze-dried CHD, then sun and oven drying CHDs follow in descending order [19].

Freeze-drying (lyophilization) is another modern procedure that very effective preserves BAS in CHD, and much better than other preservation methods but requires special equipment. Review study shows that freeze-drying is imperfect method to preserve important BAS classes (such as carotenoids, volatiles, and phenolics) and should be used carefully [20].

It is important to study the variability of the chemical composition of BAS in fruits depending on the conservation method, to identify the most effective way of preserving the quality of CHD. Another important aspect is providing new types of CHD by introduction of new sources of raw materials (fresh, frozen CHD), along with the traditional dried ones.

METHODS

Plant materials

The subjects of the study were test samples of fresh, frozen, and dried fruits of Siberian hawthorn (Crataegus sanguinea Pall.).
mountain-ash (Sorbus aucuparia L.), black chokeberry (A. melanocarpa [Michx.] Elliott), European raspberry (Rubus idaeus L.), and cinnamon rose (Rosa cineraria L.), collected in the Botanical Garden of Sechenov First Moscow State Medical University and in the Moscow region. Plants and fruits have been identified by the head of Pharmaceutical Natural Sciences Department, Lavrov Alexander Nikolaevich and professor of Pharmaceutical Natural Sciences Department, Sergunova Ekaterina Vyacheslavovna; voucher specimens were deposited at the Pharmaceutical Natural Sciences Department Herbarium. CHD test samples were dried in an oven (laboratory drying cabinet) at a temperature of 60–80°C and stored at room temperature (20–25°C). CHD test samples were frozen according to GOST R 5.956-2010 “Quick-frozen fruits. General specifications.” Fruits were stored in a freezing chamber at a temperature of –18°C.

**Chemicals**

Commercially available individual substances were used as reference standards. Organic acids reference standards - L-ascorbic acid (CAS No. 50-81-7, 99.5%), citric acid monohydrate (CAS No. 598-29-1, 99, 5%), and DL-malic acid (CAS No. 6915-15-7, 99, 9%) - were received from Fusco Chemical Co., Ltd. (Japan) and rutoside (rutin trihydrate CAS No. 250249-75-3, 94.0%) was received from Sigma-Aldrich (USA).

**Determination of BAS content**

Galvanostatic coulometry method was applied to quantify the content of ascorbic acid and total free organic acids using coulometer Expert-006 (Ekoniks-Expert, Russia) with 5 mA current. The content of total free organic acids was determined by titration with electrically generated hydroxide ions with pH-metric fixation of the end point of titration. Ascorbic acid content was determined by titration with electrically generated iodine. The titration end point was determined by amperometrically [21].

Total flavonoids and total anthocyanins content in the fruits was determined by spectrophotometry method [22,23] using Cary Varian 4000 spectrophotometer (Agilent Technologies, USA). Rutoside was chosen as the reference standard to determine total flavonoids content in hawthorn, cinnamon rose, and mountain-ash fruits. The complexes of the fruits flavonoids and rutoside with aluminum chloride have the same absorption maximum at a wavelength of 410 nm, the procedure described in pharmacopoeial monograph “St. John’s Wort herb - Herba Hyperici” [2] (Fig. 1). The total anthocyanins content was determined in terms of cyanidin-3,5-diglucoside. Cyanidin-3,5-diglucoside spectrum has identical characteristics with water-alcohol raspberry fruit extract (absorption maximum at 510–520 nm), the procedure described in monograph “Cornflower flowers - Flores Centaureae cyanus” [2]. Cyanidin-3-O-glucoside (absorption maximum at 534 nm) is relevant reference standard for black chokeberry fruits according to monograph “Bilberry fruit, fresh - Myrtilli fructus recens” (Fig. 2) [24].

The content of total polysaccharides was determined by gravimetry method according “Broadleaf plantain leaves - Folia Plantaginis majoris” [2]. Content of total tannins in terms of tannin was determined by titration with potassium permanganate according to general pharmacopoeial monograph “Determination of the content of tannins in medicinal plant raw materials” [25].

**Ascorbic acid assay**

An analytical sample weighing 5.0 g (exact weight sample) of dried fruit (crushed) and fresh-frozen fruit (homogenized suspension) was placed in a porcelain mortar where it was carefully grounded with glass powder (about 5 g), 1.50 ml of the purified water was added and remained for infusion for 10 min. The resulting extract was mixed and filtered through a paper fold filter.

About 0.5 ml of the extract from the fruit and the standard solution was placed into a coulometric cell filled with electrolyte - 0.1 M KI solution in hydrochloric buffer solution (pH=1.2), and a measurement was performed using coulometric titrator. Titration was performed by electrically generated iodine.

**Total free organic acids assay**

Approximately 5.0 g (exact weight sample) of dried fruit (crushed) and fresh-frozen fruits (homogenized suspension) was placed into 200 ml glass-stoppered flask, 150 ml of the purified water was added and held for 2 h in a boiling water bath. The resulting extract was cooled and filtered through a paper fold filter into a 200 ml volumetric flask. The volume of the extract was adjusted to the mark with the purified water and mixed.

About 0.5 ml of the extract was placed into the coulometric cell filled with a background electrolyte, an aqueous solution of potassium sulfate. The measurement was performed using coulometric titrator. The titration was carried out with hydroxide ions generated by the device.

**Total anthocyanins assay in raspberry fruits**

Approximately 5.0 g (exact weight sample) of fresh, frozen, and dried (crushed) fruits was placed into a 250 ml glass-stoppered flask; 100 ml of a 1% hydrochloric acid solution was added, and the flask was kept in a water bath at a temperature of 40–45°C for 15 min. The extract was filtered through cotton wool into 250 ml volumetric flask. The cotton wool with raw material was placed into a flask again, 100 ml of a 1% solution of hydrochloric acid was added, preliminary washing the particles of the raw material from the funnel into a flask, and the extraction process was repeated by the above method. Then, the contents of the flask were filtered through cotton wool into the same volumetric flask. The filter cake was washed with 40 ml of a 1% hydrochloric acid solution. After the filtrate was cooled, the volume of extract was adjusted to the mark with a 1% solution of hydrochloric acid. The resulting extract was filtered through a paper filter into a 250 ml flask, discarding the first 10 ml of the filtrate and measuring the optical density of the filtrate on a spectrophotometer at a wavelength of 510 nm in a cell with layer thickness of 10 mm. 1% solution of hydrochloric acid was used as the reference solution. The content was calculated using the specific absorption index of cyanidin-3,5-diglucoside (453).

![Fig. 1: Absorption spectra of rutoside complexes (1) and flavonoids from cinnamon rose fruits (2) with aluminum chloride](image1)

![Fig. 2: The absorption spectra of cyanidin-3-O-glucoside (1) and anthocyanins from fruits of black chokeberry (2)](image2)
Total anthocyanins assay in black chokeberry fruits
Approximately 5.0 g (exact weight sample) of fresh, frozen, and dried (crushed) fruits was placed into a 100 ml glass-stoppered flask, 50 ml of 60% ethanol containing 1% of hydrochloric acid was added. The flask was closed with a grounded glass stopper. The flask was attached to a reflux condenser and heated in a boiling water bath for 90 min. The extract was filtered through a paper filter. 1 ml of the obtained extract was placed in a 25 ml volumetric flask and adjusted to the mark with 1% solution of hydrochloric acid in 95% ethanol. The optical density was measured in a cell with a layer thickness of 1 cm at a wavelength of 534 nm. 95% ethanol was used as the reference solution. The total anthocyanins content was calculated using the specific absorption index of cyanidin-3-O-glucoside (100).

RESULTS AND DISCUSSION
Research data presented in Table 1 show that the fruits freezing results in a slight (by 5–7%) decrease in the amount of total polysaccharides and ascorbic acid. The content of total flavonoids and total tannins in frozen fruits decreased on average by 10–20%, total free organic acids content was preserved up to 90% of their initial value in fresh CHD, and the content of total anthocyanins has dropped by 30%.

The results of total anthocyanins determination are presented for raspberry and black chokeberry fruits as total flavonoid content. Under the influence of high temperatures, the content of total free organic acids decreased by 45–60%; total tannins, ascorbic acid, and
Fig. 4: Dynamics of biologically active substances content during the storage of dried Rosaceae fruits. The results of total anthocyanins determination are presented for raspberry and black chokeberry fruits as total flavonoid content.

Total free organic acids

Ascorbic acid

Total flavonoids

Total tannins

Y-axis - part of BAS from the initial content, %
X-axis - time, months

Hawthorn fruits
Mountain-ash fruits
Raspberry fruits
Cinnamon rose fruits
Black chokeberry fruits

It has been established that during frozen fruits storage, the content of total flavonoids and total polysaccharides did not change after 12 months, and the content of total free organic acids even increased. The tendency to a constant decrease in the amount was observed in ascorbic acid and total tannins: In the first 6 months, an average of 15% and 12%, respectively, during the period from 9 to 12 months, losses of ascorbic acid in the fruits were 28%, total tannins - 24%.

During the dried fruits storage (from 1 to 12 months), the content of total free organic acids, total polysaccharides, total flavonoids, and total tannins did not change significantly. A decrease from the initial content to 40–50% is typical for the ascorbic acid in dried fruits after 12 months of storage. Furthermore, the content of total anthocyanins in the dried raspberry fruits has been reduced by 17% by the end of the storage.
Table 1: The content of biologically active substances in different types of Rosaceae fruits (n=5, f=4, P=95%, T (f, P)=2.78)

<table>
<thead>
<tr>
<th>CHD</th>
<th>BAS content, %</th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Total free organic acids</td>
<td>Ascorbic acid</td>
<td>Total flavonoids</td>
<td>Total tannins</td>
<td>Total polysaccharides</td>
</tr>
<tr>
<td>Hawthorn fruits</td>
<td>2.61±0.02</td>
<td>0.90±0.002</td>
<td>0.19±0.01</td>
<td>2.13±0.22</td>
<td>5.69±0.12</td>
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<tr>
<td>Fresh</td>
<td>2.57±0.03</td>
<td>0.076±0.004</td>
<td>0.16±0.02</td>
<td>1.78±0.14</td>
<td>5.07±0.08</td>
</tr>
<tr>
<td>Frozen</td>
<td>0.90±0.02</td>
<td>0.030±0.002</td>
<td>0.15±0.01</td>
<td>1.36±0.07</td>
<td>4.53±0.05</td>
</tr>
<tr>
<td>Black chokeberry fruits*</td>
<td>4.72±0.01</td>
<td>1.31±0.03</td>
<td>0.21±0.01</td>
<td>7.12±0.11</td>
<td>4.32±0.12</td>
</tr>
<tr>
<td>Fresh</td>
<td>4.67±0.06</td>
<td>1.17±0.06</td>
<td>0.20±0.01</td>
<td>6.41±0.07</td>
<td>4.22±0.06</td>
</tr>
<tr>
<td>Frozen</td>
<td>2.15±0.04</td>
<td>0.32±0.02</td>
<td>0.18±0.01</td>
<td>3.24±0.08</td>
<td>4.17±0.08</td>
</tr>
<tr>
<td>Mountain-ash fruits</td>
<td>5.90±0.03</td>
<td>0.31±0.07</td>
<td>0.24±0.01</td>
<td>4.27±0.02</td>
<td>4.43±0.07</td>
</tr>
<tr>
<td>Fresh</td>
<td>5.78±0.02</td>
<td>0.26±0.07</td>
<td>0.22±0.03</td>
<td>3.66±0.04</td>
<td>4.52±0.06</td>
</tr>
<tr>
<td>Frozen</td>
<td>3.35±0.02</td>
<td>0.12±0.03</td>
<td>0.19±0.02</td>
<td>2.35±0.07</td>
<td>4.03±0.08</td>
</tr>
<tr>
<td>Dried</td>
<td>7.40±0.02</td>
<td>0.18±0.01</td>
<td>0.90±0.01</td>
<td>5.06±0.14</td>
<td>6.33±0.13</td>
</tr>
<tr>
<td>Raspberry fruits*</td>
<td>6.53±0.04</td>
<td>0.17±0.01</td>
<td>0.71±0.02</td>
<td>4.03±0.09</td>
<td>5.85±0.09</td>
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<tr>
<td>Fresh</td>
<td>5.30±0.00</td>
<td>0.02±0.01</td>
<td>0.69±0.02</td>
<td>2.41±0.12</td>
<td>5.28±0.01</td>
</tr>
<tr>
<td>Dried</td>
<td>5.19±0.04</td>
<td>0.19±0.07</td>
<td>5.90±0.07</td>
<td>9.41±0.14</td>
<td>5.22±0.08</td>
</tr>
<tr>
<td>Frozen</td>
<td>4.92±0.01</td>
<td>0.17±0.08</td>
<td>4.08±0.06</td>
<td>8.50±0.09</td>
<td>4.88±0.12</td>
</tr>
<tr>
<td>Dried</td>
<td>3.24±0.02</td>
<td>0.08±0.02</td>
<td>2.54±0.05</td>
<td>4.43±0.07</td>
<td>4.20±0.09</td>
</tr>
</tbody>
</table>

n: Number of repeat tests, f: Number of degrees of freedom, p %: Confidence figure, T (f, P): Student’s coefficient.

CONCLUSION

For the 1st time, new data were obtained showing the effect of negative temperatures on the content of certain BAS groups (total free organic acids, total polysaccharides, total flavonoids, total anthocyanins, and total tannins) in five kinds of Rosaceae fruits (Siberian hawthorn, mountain-ash, black chokeberry, European raspberry, and cinnamon rose). Results show that freezing as a conservation method allows to preserve more BAS in the Rosaceae fruits. Frozen CHD can be proposed as an alternative to fresh and dried CPB for the preparation of folkpathic and homeopathic medicines. CHD developed using oven drying was found to have the smallest BAS content compared to other conservation procedures that also confirm the research data [18,20,26].

Dynamics of BAS content studies during the storage of frozen Rosaceae fruits indicates that shelf life should be 12 months in a freezer at the temperature no higher than −18°C in polyethylene bags (Grade H - food grade).

ACKNOWLEDGMENTS

Supported by the “Russian Academic Excellence Project 5-100”.

AUTHORS’ CONTRIBUTIONS

Sergunova E.V. was engaged in scientific development and carrying out experiment. Bokov D.O. compiled literary review and wrote the draft of the manuscript.

CONFLICTS OF INTEREST

There are no conflicts of interest.

REFERENCES


