

HYPOGLYCEMIC ACTIVITY OF A NOVEL POLYHERBAL FORMULATION IN STREPTOZOTOCIN-INDUCED DIABETIC RATS: A THERAPEUTIC STUDY

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ABSTRACT

Objective: The concept of the synergistic effect of poly-herbalism was as old as medicine history. Present novel polyherbal formulation (PHF) composed of five different herbs. The present investigation aimed to evaluate the synergistic therapeutic hypoglycemic potential of PHF against streptozotocin (STZ) (60 mg/kg b.w, ip)-induced diabetic rats.

Methods: For this therapeutic study, the dose was framed orally once a day to the test objects after STZ dosing at 500 mg/kg/5 ml dosage levels for 21 days. The transformation of body weight and blood glucose level was examined, and the histopathological changes of beta cells of the pancreas, cellular architectures of liver and kidney were also perceived after scarification of the objects.

Results: The outcomes were compared to that of glibenclamide (5 mg/kg) treated group. Declines of body weight and blood glucose levels were perceived in STZ-induced diabetic animals very significantly ($p < 0.01$ or $p < 0.05$). However, these diabetic changes were significantly ($p < 0.01$ or $p < 0.05$) decreased in PHF-dosing groups revealed more encouraging effects compared to that of glibenclamide. In the other hand, various liver function and enzymes test (creatinine, urea, total bilirubin, total albumin, alkaline phosphatase, gamma-glutamyl transferase, aspartate transaminases, and alanine transaminases) and lipid profile (total cholesterol, triglycerides, high-density lipoprotein cholesterol, total protein, low-density lipoprotein [LDL], and very LDL) studies strongly indicate the potential action of this novel formulation.

Conclusions: It is deliberated that PHF has the favorable effect to normalize the blood glucose levels, and also rejuvenation and reproduction of beta cells lead a better futuristic ant diabetic therapy for diabetic management.

Key words: Polyherbal formulation, Streptozotocin, Glucose, Pancreas, Histopathology, Lipid profile.

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INTRODUCTION

High blood sugar generally termed as hyperglycemia. It is a condition of abnormally high blood glucose level in the blood characterized by the sign of diabetes and pre-diabetes conditions. Diabetes is a chronic metabolic disorder characterized by hyperglycemia due to inadequate insulin-making or non-appropriate response by cell or both. It obstructs directly in the metabolism of macronutrients. Patients with high blood sugar will characteristically experience polyuria, polydipsia, and polyphagia. Categories of diabetes are referred to as type 1 and type 2 which is non-insulin-dependent diabetes, or juvenile onset and adult-onset diabetes. Hyperglycemia indications to various diseases such as cardiovascular, hepatic, vision, dental, nerve, and foot ulcer. Around 6% world population currently suffering from this disorder and it also likely to be changing its dynamics frequently as especially in developing nations [1]. There is a rough estimate say, near about 36 million world populations are undiagnosed of this rapidly growing miserable problem. As per the IDF report, China, India, and the USA will go to host the world 80% total diabetic population on 2030 [2]. Treatment of various metabolic disorders by poly-herbalism has been recognized in ancient documents like "Sarangdhar Samhita" dated in 1300 A.D. The presence of various phytoconstituents leads to the significant therapeutic efficacy of herbal medicines further potentiated when formulated together with compatible herbals in formulations. It is evident that a better therapeutic effect can be reached with multi-constituent formulations. Combination of herbals may act on multiple targets at the same time to provide a thorough relief [3]. Multi-herbal combination therapy can achieve greater hypoglycemia than monotherapy and can also enhance the safety and tolerability of pharmacotherapy. The plant parts, namely, *Cassia auriculata* L. [4,5], *Mangifera indica* [6,7], *Ficus*

banghalensis [8,9], *Cinnamomum tamala* [10,11], and *Trichosanthes dioica* [12,13], are abundantly found throughout the Indian subcontinent and scientifically proven for their moderate hypoglycemic potentials and other pharmacological actions. However, the synergistic potential of those herbs on hyperglycemia in the form of the poly-herbal formulation (PHF) has not been investigated till date. Hence, the present investigation aimed to evaluate the synergistic therapeutic hypoglycemic potential of PHF against experimentally induced diabetic rat models *in vivo*.

METHODS

Chemicals used

Various experimental chemicals were procured from multiple sources, viz. Streptozotocin (STZ), analytical grade laboratory reagents and chemicals from HiMedia Laboratories, Mumbai, India, SD Fine Chem, Bengaluru, respectively, and Commercial reagent kits obtained from Karnataka fine Chem Ltd., Bengaluru.

Collection of the plant

Parts of the plants, namely *C. auriculata*, *C. tamala*, *Ficus benghalensis*, *M. indica*, and *Trichosanthes dioica* were collected from in an around Kerala state and authenticated. Each plant specimen voucher was deposited in the Department of Pharmacognosy, Rajiv Gandhi Institute of Pharmacy, Trikaripur, India, for future reference. The respective voucher numbers were given for each herb as shown in Table 1.

Extraction of plant material and extract preparation

All the procured parts of plants (Table 1) were subjected to shade drying and after powdered and passed through 20# sieve. Those

Table 1: Authentication of herbs present in the PHF

Herbs	Family	Code	Part used	Voucher No
<i>Cassia auriculata</i> L.	Caesalpinaceae	CA	Flower	PG/CA/F-001
<i>Cinnamomum tamala</i>	Lauraceae	CT	Leaves	PG/CT/L-003
<i>Ficus benghalensis</i> L.	Moraceae	RB	Root bark	PG/FB/B-003
<i>Mangifera indica</i> L.	Anacardiaceae	MI	Leaves	PG/MI/L-002
<i>Trichosanthes dioica</i>	Cucurbitaceae	TD	Arial parts	PG/TD/AP-001

PHF: Polyherbal formulation

prepared plant powder materials were individually macerated with distilled water (1:10), and all the aqueous plant extracts were imperiled to lyophilization. Lyophilized extracts were store at airtight container and store in a dark, cool place for further use.

Experimental animals

The animal experimentation was carried out using either sex of adult Wistar albino rats (180±10 g). Before the experiment, all the animals were placed in-house in polyacrylic cages with dark and light 24 h cycle at ambient room temperature with free access of water and rodent pellets diet. The animal experimentation protocol was approved by IAEC of Rajiv Gandhi Institute of Pharmacy and experimentation was carried out as per the CPCSEA guidelines. Preliminary screening for phytochemical 1 g each of lyophilized plant extracts was dissolved in 100 ml of distilled water and subjected to qualitative analysis of phytochemicals such as alkaloids, tannins, glycosides, flavonoids, phenolic compounds, and saponins as per standard chemical methods [2].

Development of PHF

About 1 g each of lyophilized plant extracts were dissolved in 100 ml of distilled water and subjected to qualitative analysis of phytochemicals such as alkaloids, tannins, glycosides, flavonoids, phenolic compounds, and saponins as per standard chemical methods [2].

Total phenolic content (TPC) determination

Sample stock preparation was done with 50% methanol in 100 ml volumetric flask with triplicate maceration of 1 g PHF in methanol. 1 ml aliquot diluted 10 times with distilled water and later added with 1.5 ml Folin-Ciocalteu's reagent. The solution mixture was incubated for 5 min at room temperature. After incubation solution mixture was further added with 4 ml of 20% Na₂CO₃ (w/w) and make up the volume up to 25 ml with distilled water. The reaction mixture was agitated and keeps for 30 min at room temperature. The absorbance was measured at 765 nm in UV spectrophotometer against the blank. Similarly, standard gallic acid stock sample also prepared with methanol (100 µg/ml), and various dilutions concentrations were prepared such as 6.25, 12.5, 25, and 50 µg/ml and standard gallic acid curve also been plotted against the absorbance at 765 nm by UV spectrophotometer against the blank [14].

$$\% \text{ TPC (w/w)} = \text{GAE} \times \text{V} \times \text{D} \times 10 - 6 \times 100 / \text{W}$$

GAE - Gallic acid equivalent (µg/ml), V - total volume of sample (ml), D - Dilution factor, W - Sample weight (g).

Total flavonoid content (TFC) determination

Standard rutin stock solution is prepared (100 µg/ml) using 80% methanol and various dilute concentrations such as 10, 20, 40, 80, and 100 µg/ml been prepared. Each 0.5 ml standard rutin dilution is added with 1.5 ml 95% methanol, 0.1 ml of 10% aluminum chloride, 0.1 ml 1M potassium acetate, and 2.8 ml of distilled water and incubated for 30 min at room temperature. Absorbance measured at 415 nm using UV spectrophotometer against the blank. Similarly, 1 g of PHF dissolved separately in 25 ml of 80% methanol for determination of flavonoids content as described above [14].

$$\% \text{ total flavonoids content (w/w)} = \text{RE} \times \text{V} \times \text{D} \times 10 - 6 \times 100 / \text{W}$$

RE - rutin equivalent (µg/ml), V - total volume of sample (ml), D - dilution factor, W - sample weight (g).

Study for acute toxicity

The study was performed as per the OECD guideline-425. Experimental Wistar rats were fasted overnight before the study. Fasted animals were divided into four groups of six animals each. Individual groups were administered orally with PHF dose from 250, 500, and 1000 and 2000, 4000, and 5000 mg/kg b.w, respectively. All the groups of animals were initially observed for 1 h and periodically for next 24 h up to 14 days for any gross behavioral changes such as drowsiness, restlessness, writhing, convulsions, symptoms of toxicity, and mortality.

Oral glucose tolerance test (OGTT) on normal and diabetic animals

Experimental animals were randomly selected and grouped into three of six each. The Groups I and II were treated with the vehicle carboxymethylcellulose solution (0.5% w/v in normal saline) and PHF (500 mg/kg b.w), respectively. Whereas Group III was treated with standard glibenclamide (1 mg/kg b.w), Groups II and III were administered with PHF and glibenclamide and after 30 min all the experimental animals including Group I was fed orally with 2 g/kg of glucose solution. After a particular time interval, namely 0, 30, 60, 90, and 120 min, blood was withdrawn from the tail vein. The estimation of blood glucose level was measured by Accu-Chek, Roche Diabetes Care India [15].

Hypoglycemic activity in STZ-induced diabetic rats

Experimental diabetes was induced by 0.5 ml/kg b.w IP injection of freshly prepared STZ (50 mg/kg b.w. in 0.1 M citrate buffer at pH 4.5) in overnight fasted animals [16]. After 24 h, the STZ treated animals were given with glucose solution (5% w/v) at the dose of 2 ml/kg bw to prevent the mortality due to hypoglycemic shock. The induced rats with fasting blood glucose >300 mg/dl were considered as diabetics and taken for hypoglycemic activity. Experimental animals (male Wistar rats) were divided into three groups with six animals each. Groups II-IV were designated as diabetic control. Glibenclamide (5 mg/kg bw) treated and PHF (500 mg/kg, bw) treated groups. Another six animals were taken as normal control without STZ as Group I. Except the Groups I and II; all other groups were treated with standard glibenclamide and PHF (suspended in 1% w/v carboxymethylcellulose) through oral gavage for 21 consecutive days. During this time period blood, samples were been taken through the retro-orbital route of the animal in a specific time interval such as 0 day, 7th day, 14th day, and 21st day for estimation of blood glucose level. The estimations were done by Chem Biochemistry Analyzer (AN Biotech, India). Body weight of experimental animals also measured in this specified time intervals.

Blood collection and harvestmen of organs

Blood collection and harvesting of organs were taken place after a 21st day of experimentation. The animals were sacrificed by ether anaesthetization, and an aliquot of blood (5 ml) was collected from a jugular vein in plain and ethylenediaminetetraacetic acid tubes tube and centrifuged at 1282 g × 5min. After centrifugation, the serum was separated with Pasteur Pipette in plain tubes and subjected for biochemical estimations [17]. Meanwhile, the rats were dissected, and the organs were isolated such as whole liver, pancreas, and two kidneys. After isolation, the specimens are freed from extracellular fats and deposited in formalin solution for further histopathological investigations.

Lipid profile and total protein estimations

The entire estimations of lipid profiles such as total cholesterol, triglycerides, high-density lipoprotein (HDL)-cholesterol, and low-

density lipoprotein (LDL)-cholesterol along with total protein were screened with the standard procedure using Chem Biochemistry Analyzer (AN Biotech, India) [22].

Liver and kidney function tests

Estimation of serum creatinine [19], urea [20], uric acid [21], total bilirubin [22], albumin [23], and alkaline phosphatase (ALP) [24], gamma-glutamyltransferase (GGT) [25], aspartate transaminases (AST), and alanine transaminases (ALT) [26] was carried out using standard procedure.

Histopathological analysis

The preserved organs were further taken to the microscopic slide with egg albumin and allowed to dry. After dehydration, the tissues were subjected to xylene solution for 15–20 min for further cleaning. Clean tissues were put in paraffin infiltration in the automatic tissue processing unit, and tissue blocks were prepared. The thin sections (5 μ m) of tissue were obtained using microtome. The sections were further taken to the microscopic slide with egg albumin and allowed to dry. Finally, staining of the sections done with an acidic and basic dye, namely Eosin and hematoxylin, respectively [27,28].

Statistical analysis

The experimental data produced as mean \pm SEM. All statistical significance were found in groups were established by one-way analysis of variance (ANOVA) followed by Dunnett's *t*-test *post hoc* test. $p < 0.05$ was considered significant [29].

RESULTS

Preliminary screening of phytochemicals

Preliminary phytochemical screening of PHF indicates the presence of phytochemicals such as alkaloids, glycosides, tannins, flavonoids, and phenolics in the mixture.

TPC and TFC of PHF

In this PHF, TPC and TFC were found to be 81.01 ± 15 GE/g and 10.16 ± 11 RE/g (Table 2), respectively.

Table 2: TPC and TFC of PHF

TPC ^a	81.01 \pm 15.0
TFC ^b	10.16 \pm 1.13

Values are means \pm SEM of three replicate studies. ^aGE/g: Mg gallic acid equivalent, ^bQE/g: Mg rutin equivalent. TPC: Total phenolic content, TFC: Total flavonoid content, PHF: Poly-herbal formulation

Study of acute toxicity

PHF was found to be non-toxic at the optimum dose of 5000 mg/kg bw. There was no evidence of behavioral change in the animals observed even no mortality as well. Any other symptoms such as breathing, sensory nervous system, gastrointestinal effects, or toxicity also were found absent in all experimental animals during the observational period indicating the high margin of safety of PHF.

OGTT on normal and experimental rats

There was a significant ($p < 0.05$) reduction of blood plasma glucose level in PHF-treated group and glibenclamide (0.25 mg/kg)-treated group compared to normal control (Table 3) whereas no significant change observed in normal control animals during the period of study.

Hypoglycemic activity of PHF in diabetic-induced rats

PHF-treated group has been shown a significant reduction about 91.83 ± 2.96 mg/dl of blood sugar compared to the diabetic control group, whereas 123.83 ± 2.11 mg/dl found in glibenclamide-treated group. These results indicate the normalization of blood sugar level with PHF in diabetic rats (Tables 4 and 5) as compared to normal control group (90.83 ± 1.77 mg/dl) of animals.

Total protein and lipid profile test

The automatic analyzer has been employed for the screening of total cholesterol, triglycerides, HDL-cholesterol, total protein, LDL, and VLDL, respectively, in blood serum of experimental animals (Table 6) [18].

Liver and kidney function tests

The concentrations of creatinine [19], urea [20], total bilirubin [22], albumin [23] as well as the activities of ALP [24], GGT [25] and AST, ALT, and BUN [26] were determined in the serum (Table 7).

Histopathologic analysis

Kidney

The histological sections showed intact cellular architecture and normal cellularity in the glomerulus. The normal group showed the intact tubules (Fig. 1a) and blood vessels. Interstitium showed to be unremarkable. Most of the tubules in the diabetic group showed degenerative changes (Fig. 1b) with dispersed interstitial inflammatory mononuclear infiltration. The glibenclamide treated groups showed moderate degenerative changes in few tubules (Fig. 1c), whereas minimal degenerative tubules changes (Fig. 1d) found in the PHF treated group.

Table 3: OGTT

Groups	Blood glucose levels(mg/dl)					
	Treatments	0min	30min	60min	90min	120min
Group1	Normal control	92.28 \pm 2.57	143.33 \pm 2.08	134.33 \pm 2.99	120.19 \pm 2.99	95.66 \pm 2.06
Group2	PHF(500 mg/kg)	92.76 \pm 3.62	127.34 \pm 1.96	116.45 \pm 4.25	103.22 \pm 3.23*	96.51 \pm 3.59
Group4	Glibenclamide(0.25mg/kg)	91.58 \pm 2.96	119.21 \pm 2.11**	102.65 \pm 3.02**	96.35 \pm 2.96**	90.54 \pm 3.52

Values are mean \pm SEM, n=6, * $p < 0.05$, ** $p < 0.01$ compared to normal control group. One-way ANOVA followed by Dunnett's multiple comparison test, OGTT: Oral glucose tolerance test, PHF: Poly-herbal formulation

Table 4: Blood glucose level in normal and experimental animals

Biochemical Parameters	Blood sugar(mg/dl)			
	0Day	7 th Day	14 th Day	21 st Day
Normal control	90.83 \pm 3.93	95.33 \pm 3.09	92.66 \pm 4.06	90.83 \pm 1.77
Diabetic control	370.00 \pm 3.00	390.50 \pm 3.50	403.16 \pm 4.94	415.33 \pm 3.54
Diabetic+glibenclamide	371.66 \pm 2.68	277.66 \pm 4.42**	190.66 \pm 2.28*	123.83 \pm 2.11*
Diabetic+PHF	364.16 \pm 4.29*	272.00 \pm 4.19**	141.83 \pm 3.80**	91.83 \pm 2.96**

All values are expressed as mean \pm S.E.M(n=6). * $p < 0.05$, ** $p < 0.01$ as compared to diabetic control. One-way ANOVA followed by Dunnett's multiple comparison test, PHF: Poly-herbal formulation

Pancreas

The histological study reveals the septal separation of pancreatic lobules. Few of the lobules showed small areas of light-staining islets of Langerhans. Around 65% of small beta cells in central islet cells were been found in normal control groups (Fig. 1e [long-arrow]), and 30% large alpha-cells seen in the periphery (Fig. 1e [short-arrow]). In diabetic control section, most of the beta cells show degenerative changes with 40% beta-cell in center of islet cells (Fig. 1f [long-arrow]), while the periphery comprises 55% large alpha-cells (Fig. 1f [short-arrow]). The glibenclamide-treated animal pancreatic cells showed some degenerative changes of the beta cells. The center of islet cells consists of 50% small beta-cells (Fig. 1g [long-arrow]), while the periphery comprises 45% large alpha-cells (Fig. 1g [short-arrow]). In PHF-treated section studied shows the center of islet cells consist of 65% small beta-cells (Fig. 1h [long-arrow]), while the periphery comprises 30% large alpha-cells (Fig. 1h, [short arrow]). Intervening these cells is seen vascular spaces in all the sections.

Liver

All the sections showed liver parenchyma with intact architecture, the periportal, perivenular, and mid zonal hepatocytes appear unremarkable. In Normal control, the central veins and Sinusoids appear unremarkable (Fig. 1i). Sinusoids appear unremarkable. Whereas in diabetic control Section showed the periportal region had dense mononuclear inflammatory infiltration (Fig. 1j). Standard glibenclamide-treated group section revealed the unchanged central veins (Fig. 1k) and sinusoids. In the case of PHF-treated groups section

studied shows that the periportal region shows moderate mononuclear inflammatory infiltration (Fig. 1l). The central veins and sinusoids appear unremarkable as like all the sections.

DISCUSSION

The concept of poly-pharmacy has been practicing since from ancient time. The proven examples are listed in various Ayurveda literature. This trend also recognized by the WHO in some clinical ailment like diabetes which is a metabolic disorder associated with polydipsia and polyphagia.

Lyophilized aqueous plant extracts were been used to formulate the PHF to minimize the residual solvent toxicity. All those used herbs were also proven for their antioxidant properties. The PHF consisting of various important phytoconstituents such as alkaloids, tannins, glycosides, flavonoids, and phenolic compounds, where later two are natural antioxidants, reported to significantly increase superoxide dismutase (SOD), glutathione and catalase activities eventually promotes the expression of SOD, catalase, and glutathione [27]. The endothelial dysfunctions due to hyperlipidemia, hyperinsulinemia, and hyperglycemia are the result of oxidative stress [28], which leads to diabetes mellitus. The reduction of antioxidant enzymes, inactivation of enzymes and lipid peroxidation cause cell death while free radicals get excess produces. The presence of a significant amount of phenolic and flavonoids contents makes this formulation potent scavenging and antioxidants in biological systems. As per existing studies on diabetes mellitus, it has been observed that the progression of this disorder leads to boosting the free radicals and potentially decrease the cellular antioxidant capacity leads to oxidative stress even both insulin dependent and independent diabetes mellitus [29]. The tolerability with high dose (5000 mg/kg b.w) in experimental animals reflects the effect of altered carbohydrate metabolism during the post-glucose administration. The prolonged hypoglycemic study evident significant decrease in blood glucose level may be due to the synergistic effect of all bioactive phytochemicals in various bio levels. The results also correlate with the study carried by El-Baz *et al.*, 2017, on *Haematococcus pluvialis* where the bioactive principles found were the flavonoids [27]. Rats orally treated with the PHF at a dose level of 500 mg/kg of b. w were found to have significantly reduced activities of these enzymes, thus indicating less damage to hepatocytes. Similar action was also

Table 5: Change of body weight in normal and experimental rats

Groups	Body weight(g)			
	0Day	7 th Day	14 th Day	21 st Day
Normal control	181±2.8	182±3.8	184±2.0	186±3.1
Diabetic control	178±3.7	174±3.2	170±2.9	168±3.4
Diabetic+glibenclamide	190±2.1	190±2.2**	194±3.1**	196±1.6**
Diabetic+PHF	180±1.9	182±3.1*	185±2.6*	189±3.2*

All values are expressed as mean±S.E.M(n=6). *p<0.05, **p<0.01 as compared to diabetic control. One-way ANOVA followed by Dunnett's multiple comparison test. PHF: Poly-herbal formulation

Table 6: Analysis of serum lipid profile and total protein

Biochemical Parameters	Normal control	Diabetic control	Diabetic+Glibenclamide	Diabetic+PHF
Cholesterol(mg/dl)	53.57±2.95	63.01±2.46	55.82±4.59	50.25±2.85
Triglyceride(mg/dl)	30.98±2.81	48.79±2.89	32.70±1.90*	29.06±3.66*
HDLc(mg/dl)	17.75±0.35	23.11±0.44	20.25±1.47	19.25±0.14*
Total protein	07.21±0.12	07.27±0.30	07.22±0.26	07.23±0.21
LDL(mg/dl)	07.03±1.85	15.91±1.50	05.91±2.63*	05.17±1.55*
VLDL(mg/dl)	06.19±0.56	09.75±0.58	06.54±0.38*	05.85±0.73*

All values are expressed as mean±S.E.M(n=6). HDLc: High-density lipoprotein-cholesterol, LDL: Low-density lipoprotein, VLDL: Very low-density lipoprotein, *p<0.05, **p<0.01 as compared to diabetic control. One-way ANOVA followed by Dunnett's multiple comparison test

Table 7: Liver and kidney function tests

Biochemical Parameters	Normal control	Diabetic control	Diabetic+glibenclamide	Diabetic+PHF
Creatinine(mg/dl)	0.536±0.01	0.606±0.03	0.556±0.01	0.540±0.02*
Urea(mg/dl)	42.61±2.80	59.44±1.79	46.53±3.15*	45.48±6.02*
Total bilirubin(μmol/l)	0.24±0.08	0.228±0.08	0.216±0.084	0.235±0.09
Albumin(g/dl)	3.32±0.13	3.15±0.34	3.22±0.10	2.93±0.13**
ALP(U/L)	261.83±4.23	568.83±4.27	280.86±3.99*	288.45±5.19*
GGT(U/L)	2.01±1.56	8.04±5.85	2.95±2.71*	3.05±2.10
AST(U/L)	155.8±4.14	263.50±3.91	159.93±4.63*	155.11±7.22*
ALT(U/L)	66.05±2.90	131.96±3.86	86.4±4.31	90.06±3.18*
BUN(mg/dl)	17.57±0.35	27.77±0.25	24.23±0.51	25.15±0.34

All values are expressed as mean±S.E.M(n=6). ALP: Alkaline phosphatase, GGT: Gamma-glutamyl transferase, AST: Aspartate transaminases, ALT: Alanine transaminases, *p<0.05, **p<0.01 as compared to diabetic control. One-way ANOVA followed by Dunnett's multiple comparison test

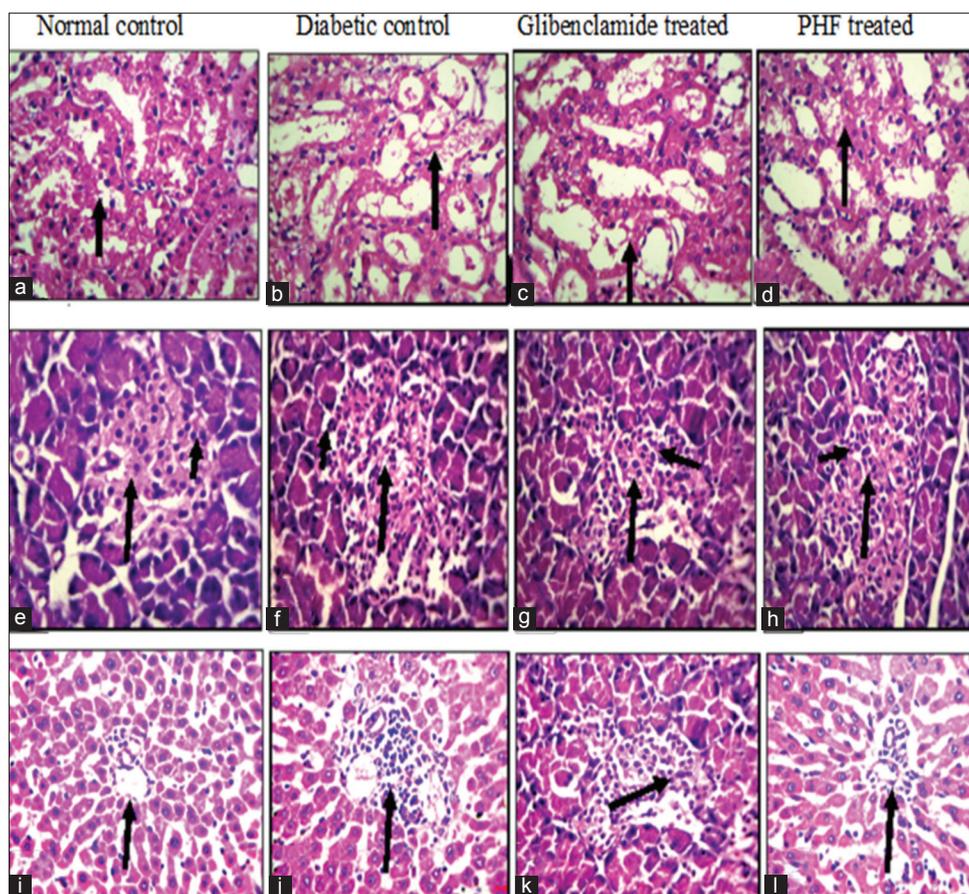


Fig. 1: Histopathological analysis of organs (kidney [a-d], pancreas [e-h], and liver [i-l]) H and E, ×400

been reported by Talha et al., 2015, in their study on the effects of the ethanolic extract of *Alocasia indica rhizomes* [30]. STZ-induced diabetic rats are associated with hyperlipidemia and increased levels of serum creatinine which all significantly controlled by PHF. The lowering of these lipid substances and serum creatinine in the blood of treated rats is presumed mainly to be a manifestation of lowering of blood glucose level. Low protein degradation found in PHF treated animals which leads to high serum protein concentration. Due to the reduction in lowering of glucose levels thus sparing the body fat and muscle protein which otherwise are utilized in diabetic rats probably indicates the elevation of body weight among the rats administered with PHF.

CONCLUSION

The common symptoms of diabetes, that includes polyphagia, polydipsia, and weight loss, were found to be lessened by the PHF dose level of 500mg/kg of b. w in diabetic rats. Along with the significant reduction of fasting glucose level and lipid profile of diabetic rats were the outcome of this study. The PHF was found significantly decreasing the activities of liver enzymes and ALP in diabetic rats. The histopathological investigation along with the biochemical evaluations strongly suggests the hypoglycemic potential of PHF. The results also indicate the effect of PHF on the regeneration of pancreatic β -cells leading to blood glucose regulation in the body.

AUTHORS' CONTRIBUTIONS

All the authors contribute the same contributions to this research.

CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest regarding the publication of this paper.

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