ASIAN JOURNAL OF PHARMACEUTICAL AND CLINICAL RESEARCH



# A COMPREHENSIVE REVIEW OF PHYTOCHEMICAL AND PHARMACOLOGICAL OVERVIEW ON CELOSIA CRISTATA FOR FUTURE PROSPECTIVE RESEARCH

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Received: 08 September 2020, Revised and Accepted: 20 October 2020

# ABSTRACT

*Celosia cristata* (CC) is used in traditional medicine to cure several disorders. It is a member of the genus Celosia and is commonly known as cockscomb, since the flower looks like the head on a rooster. Many sensitive ingredients were isolated from different parts of the plant. The recent studies showed that the plant exerted a wide range of pharmacological activities. The chemical constituents and pharmacological activities of CC were presented in this review.

### Keywords: Celosia cristata, cockscomb, chemical constituent.

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# INTRODUCTION

Medicinal plants have a wide range of pharmacological effects. *Celosia cristata* (CC) is an annual plant [1-6] of tropical origin and lacking a woody stem. It grows well in both humid and arid conditions and their flowers can last for up to 8 weeks. A high number of seeds can be produced by each flower up to 1500 per g. The plant often grows up to 30 cm in height, though many are smaller. The leaves are either green or bronze/maroon, depending on the cultivar. The flower can be broken into three parts: Their spikes, plumes, and crests vary from one another but have standard commonalities they are usually brightly colored, usually red, yellow, pink, or orange, though other colors can be present in hybrids [7-9]. The novelty of current review is to know phytochemical and pharmacological overview of CC for future prospective research. By knowing its value it can be utilized for the future discovery.

# NOMENCLATURE AND CLASSIFICATION

The *Celosia* species [6,8-20] is a small genus of edible and ornamental plants belonging to Amaranthacea. It belongs to kingdom Plantae, order Caryophyllales, family Amaranthaceae, genus *Celosia*, and species *Cristata*. The binomial name is *Celosia cristata* L.

## Common name

It is commonly known as Cockscomb, Crested celosia, Yellow toreador, Red cockscomb, Foxtail amaranth, Fire-flame bush, Shinaji tea, and Woodfordia.

#### DESCRIPTION

CC is an annual herb, hairless entirely. The stem [21-28] is erect, thick, little branched, green or tinged with red, ridged, and flat near the upper part. The simple leaves are alternate, petiolate; the blade is long-elliptical to ovally lanceolate, 5–13 cm long, 2–6 cm wide, acuminate or attenuate at the apex, gradually narrow and decurrent at the base, and entire marginally. The spikes are flat, succulent, and crest-like. Numerous flowers are present in down middle. The perianthial segments are light red to purplish red, yellowish white, or yellow, elliptically ovate, pointed at the tip, 5 in number. The bract, bractlet, and perianthial segments are scarious. Each flower has five stamens whose filaments are joined together to form a cup at the base. The fruit has an egg-shaped utricle. The seed is kidney-shaped, black, and lustrous.

# CULTIVATION

CC can grow of tropic origin. They can be grown in summer months in the colder climate. The plants [29-32] being annual plants grow for only about one-fourth of a year. The ideal temperature of soil is  $16^{\circ}$ C for growth.

### **PROPAGATION OF HERB**

Seed sow early to mid-spring in a warm greenhouse. Germination takes place within 2 weeks. When large enough to handle, prick the seedlings out into individual pots and plant them out after the last expected frosts.

# TRADITIONAL USES

Seeds were used as demulcent; for painful micturition and for dysentery. It is used medicinally in menorrhagia and as an astringent which are used to treat bloody stool, hemorrhoid bleeding, and diarrhea. The seed decoction is used to treat dysentery. The flowers [33-41] can be used as astringent, styptic, depurative, uterine sedative, constipating, antibacterial, and corrective of urinary pigments, febrifuge, and alexeteric. They can be used in the conditions of kapha and pitta, leprosy, burning sensation, skin diseases, diarrhea, dysentery, fever, headache, hemorrhoids, herpes, internal hemorrhage, leukorrhea, liver disorders, menorrhagia, ulcers, and wounds. Juice of leaves is beneficial for bilious sickness, a stimulant in pregnancy, blood shot eyes, blurring of vision, cataracts, and hypertension.

# CHEMICAL CONSTITUENTS

The extracts of CC contain flavonoids, mucilages, phenolic compounds and tannins, saponins, triterpenoids, alkaloids, carbohydrates, proteins, amino acids, gums, and steroids [41-51]. The plant contained choline esters of hyaluronic acid, betanin, and several sterols. The inflorescence contained amarantin, isoamarantin, celosianin, and isocelosianin. The seeds contain 10.1–12.8% of protein and yield 7.2–7.9% fatty oil. Six compounds were isolated from the ethanolic extract of CC, and identified as 4-hydroxyphenethyl alcohol, kaempferol, quercetin,  $\beta$ -sitosterol, 2-hydrox octadecanoic acid, and stigmasterol [52-56]. Isoflavone, cristatein (5-hydroxy-6-hy-droxymethyl -7,2 0 -dimethoxyisoflavone, 2), and five known flavonoids were also identified. Five saponin, cristatain , celosin A, celosin B, celosin C, and celosin D were obtained from the seeds of CC. Triterpenoid saponin and semenoside A are isolated from semen of CC. Two glycoproteins, CCP-25 and CCP-27, were purified from the leaves of CC. The compounds isolated from CC were p-hydroxyphenylethanol, kaempferol, quercetin, cristatain, celosin A, celosin B, celosin, celosin, sphingosine,  $\beta$ -sitosterol, stearic acid, stigmasterol, daucosterol, palmitinic acid, and n-hexacosanoic acid [57-62].

# PHARMACOLOGICAL EFFECTS

#### Hemostatic effect

The mice were given decoction of Flos CC with the dosage of 17 g/kg after 5 days, and compared with a control group. It showed that the bleeding time was shortened. After 7 days rabbits were given the same decoction [63-66] with the dosage of 1.7g/kg. It was observed that the coagulation time, prothrombin time, and plasma recovery were shortened, and the euglobulin lysis time was markedly shortened in comparison with control.

#### Hepatoprotective effects

The hepatoprotective activity of semenoside A with an oral dose of 1.0, 2.0, and 4.0 mg/kg, respectively, was observed by CCl<sub>4</sub>-induced hepatotoxicity in mice. The results showed that it had significant hepatoprotective effects. Cristatain saponin showed significant hepatoprotective effect on CCl4 and N, N-dimethylformamide-induced hepatotoxicity in mice, which were observed by significant decreases in the values of aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (ALP) of serum and histopathological examinations compared to controls [67-72].

### Cytotoxic effects

The cytotoxicity [73-76] of water and organic solvent extracts was determined in the fibroblast cells Cos7 and in four cancer cell lines: HeLa, HepG2, SK-Hep1, and LS 174T. IC50 of the water extracts against cancer cell lines is compared.

# Antioxidant effects

CC ethanol extract had antioxidant activity [77-80] in a dose-dependent manner in 1-diphenyl-2-picryl-hydrazyl (DPPH) radical scavenging. Ethanol extract had antioxidant activity in a dose-dependent manner. Silica dose-dependently increased the intracellular ROS generation in RAW 264.7 cells. CC ethanol extract showed anti-aging effects, the hyaluronidase inhibitory effects and elastase activity inhibitory effects were relatively strong which suggests the CC ethanol extract has hydration and anti-wrinkle property.

# Adipogenic effect

The *in vitro* the capacity of a CC extracts to impact the adipogenic potential of native human adipose tissue progenitor cells. Native adipose tissue [81-84] progenitor cells were isolated by depletion approaches from human subcutaneous adipose tissues. Cell culture conditions were used to assess the effect of CC extract on commitment and differentiation of progenitor cells. Results showed that CC extract reduces lipid content of progenitor cells undergoing differentiation.

### Antimicrobial and anthelmintic effects

The antimicrobial properties [85-88] of ethanolic, methanolic, and other solvent extracts of CC were evaluated against microorganisms, *Staphylococcus aureus, Bacillus subtilis, Salmonella typhimurium, Escherichia coli, Pseudomonas aeruginosa,* and *Candida albicans.* The minimal inhibitory concentration values of the extracts against animal pathogenic bacteria and yeast were assessed using the broth microdilution methods. Results obtained that the different extracts differed clearly in their antimicrobial activities.

# Antidiabetic effect

The effect of the methanolic extract of CC leaves on blood glucose level, superoxide dismutase (SOD), catalase (CAT), ALT, AST, ALP activities, and malondialdehyde (MDA) level was examined in diabetic rats. The results obtained a significant increase in serum AST, ALP, and ALT activities and reduction in SOD and CAT activities compared with normal control groups.

### Antinociceptive effect

Methanol extract of the whole plant of CC was used to evaluate the antinociceptive activity. The antinociceptive effect of CC was carried out in thermal (hot plate and tail immersion test) and chemical (acetic acid, formalin, and glutamate-induced nociception test) pain models in mice at various doses. Central and peripheral mechanisms are associated with CC showing significant antinociceptive effect [89,90].

#### Other pharmacological effects

CC was considered as one of the herbal therapy [91-97] acting as antitussive. Choline esters of hyaluronic acid from the plant, when fed to rats showed antiulcer and gastroprotective effect. The plant prevented fluoride toxicity, the food supplemented with calcium can reduce the effect of high fluorine, and the food supplemented with both calcium and CC extracts is better.

## CONCLUSION

CC possessed a wide range of therapeutic activities which were proved that this plant have a potential regenerator capacity of various cells, antiproliferative activity, antimicrobial potentiality, adipogenic potentiality, and cytotoxic potential. The wide range of therapeutic potentialities of CC is mainly due to the presence of various bioactive molecules in flowers, roots, stems, leaves, and herbs.

#### **AUTHORS' CONTRIBUTIONS**

All authors have equally contributed for making this case report to be successful.

# **CONFLICTS OF INTEREST**

None.

## REFERENCES

- 1. Rubini D, Sudhahar D, Anandarajagopal K. Phytochemical investigation and anthelmintic activity of *Celosia cristata*. IRJP 2012;3:335-7.
- Surse SN, Shrivastava B, Sharma P, Sharma J, Gide PS. Pharmacognostic standardisation of whole plant of *Celosia argentea*, var. *Cristata* (L). Int J Pharm Res Sch 2014;3:2277-7873.
- 3. De Bao W. Da G, Jie XY, Qin W. The nutritional components of *Celosia* cristata L cv plumosa. J Plant Resour Environ 1994;3:32-5.
- Woo K, Ko J, Song S, Lee J, Kang J, Seo M, et al. Antioxidant compounds and antioxidant activities of the methanolic extracts from cockscome (*Celosia cristata* L.) flowers. Planta Med 2011;77:77-8.
- Yaolin W, Tofazzal I, Satoshi T. Phenolic constituents of *Celosia* cristata L. susceptibleto Spinach root rot pathogen *Aphanomyces* cochlioides. Biotechnol Biochem 2010;70:2567-70.
- Wang Y, Lou Z, Wu QB, Guo ML. A novel hepatoprotective saponin from *Celosia cristata* L. Fitoterapia 2010;81:1246-52.
- Sun ZL, Gao GL, Xia YF, Feng J, Qiao ZY. A new hepoprotective saponin from semen *Celosia cristata*. Fitoterapia 2011;82:591-4.
- Gholizadeh A, Kapoor HC. Modifications in the purification protocol of *Celosia cristata* antiviral proteins lead to protein that can be N-terminally sequenced. Protein Pept Lett 2004;11:551-61.
- Wang Y. Chemical Constituents and Bioactivities of *Celosia Cristata* L. and *Penthorum Chinense* Pursh, Ph D Thesis. China: Second Military Medical University; 2012.
- Kim YS, Hwang JW, Sung SH, Jeon YJ, Jeong JH, Jeon BT, et al. Antioxidant activity and protective effect of extract of *Celosia* cristata L. flower on tert-butyl hydroperoxide-induced oxidative hepatotoxicity. Food Chem 2015;68:572-9.
- Herrmann F, Romero MR, Blazquez AG, Kaufmann D, Ashour ML, Kahl S, *et al.* Diversity of pharmacological properties in Chinese and European medicinal plants: Cytotoxicity, antiviral and antitrypanosomal screening of 82 herbal drugs. Diversity 2011;3:547-80.
- Pyo YH, Yoon MY, Son JH, Cho TB. The effect of *Celosia cristata* L. Ethanol extract on anti-oxidant and anti-aging activity. Korean J Biotechnol Bioeng 2008;23:431-8.
- Wen Y, Islam MT, Tahara S. Phenolic constituents of *Celosia cristata* L. Susceptible to Spinach root rot pathogen *Aphanomyces cochlioides*. Biosci Biotechnol Biochem 2006;70:2567-70.
- Fitoussi R, Estève D, Delassus A, Vié K. Impact of *Celosia cristata* extract on adipogenesis of native human CD34+/CD31-cells. JCDSA

2013;3:55-63.

- Jayanthy V, Shafna A. Use of flowers as antimelanocyte agent against UV radiation effects. Am J Biopharm Biochem Life Sci 2012;1:A068.
- Yun SM, Choi BH, Ku HO, Lee MH, Nam HM, Lee KJ, et al. Antimicrobial activities of the flower extract of *Celosia cristata* L. Planta Med 2008;74:PA31.
- Baranwal VK, Verma HN. Localized resistance against virus infection by leaf extract of *Celosia cristata*. Plant Pathol 1998;41:633-8.
- Balasubrahmanyam A, Baranwal VK, Lodha ML, Varma A, Kapoor HC. Purification and properties of growth stage-dependent antiviral proteins from the leaves of *Celosia cristata*. Plant Sci 2000;154:13-21.
- Hamzah RU, Lawal AR, Madaki FM, Erukainure OL. Methanolic extract of *Celosia argentea* var. Crista leaves modulates glucose homeostasis and abates oxidative hepatic injury in diabetic rats. Comp Clin Pathol 2018;27:1065-71.
- Shanta I, Shafiullah SM, Ahmed T. Antinociceptive effect of methanol extract of *Celosia cristata* Linn. In mice. BMC Complement Altern Med 2016;16:400.
- Gholizadeh A, Kumar M, Balasubrahmanyam A, Sharma S, Narwal S, Lodha ML. Antioxidant activity of antiviral proteins from *Celosia cristata*. J Plant Biochem Biotechnol 2004;13:13-8.
- Rajendran R. Preliminary phytochemical analysis and anti-bacterial activity of *Mimosa pudica* Linn leaves. JGPT 2009;1:76.
- Balick JM, Cox PA. Plants, People and Culture: The Science of Ethnobotany. New York: Scientific American Library; 1996. p. 228.
- Jagtap NS, Khadabadi SS, Ghorpade DS, Banarase NB, Naphade SS. Antimicrobial and antifungal activity of *Centella asiatica* (L.) Urban. Umbeliferae Res J Pharm Tech 2009;2:328-30.
- Wijesekera RB. Plant derived medicines and their role in global health. In: The Medicinal Plant Industry. United States: CRC Press; 1991.
- Chitravadivu C, Bhoopathi M, Balakrishnan V, Elavazhagan T, Jayakumar S. Antimicrobial activity of laehiums prepared by herbal venders, South India. Am Eur J Sci Res 2009;4:142-7.
- Mohammed R, Das AK, Ariful M, Mollik H, Jahan R, Khan M, et al. An ethnomedicinal survey of Dhamrai Sub-district in Dhaka district, Bangladesh. Am Eur J Sust Agric 2009;3:881-8.
- Dana E, Sanz-Elorza M, Sobrino E. Plant Invaders in Spain (Checklist); 2003. Available from: http://www.med-alienplants.org/checklist.
- Norazaidah AY, Hainida EK. Antioxidant activity and phenolic content of raw and blanched *Amaranthus* species. Food Chem 2006;94:47-52.
- Roriz CL, Barros L, Carvalho AM, Santos-Buelga C, Ferreira IC. *Pterospartum tridentatum, Gomphrena globosa* and *Cymbopogon citratus*: A phytochemical study focused on antioxidant compounds. Food Res Int 2014;62:684-93.
- Salvador MJ, Ferreira EO, Mertens-Talcott SU, De Castro WV, Butterweck V. Isolation and HPLC quantitative analysis of antioxidant flavonoids from *Alternanthera tenella* Colla. Z Naturforsch C J Biosci 2006;61:19-25.
- 32. Salvador MJ, Pereira PS, França SC, Candido RC, Ito IY. Bioactive chemical constituents and comparative antimicrobial activity of callus culture and adult plant extracts from *Alternanthera tenella*. Z Naturforsch C J Biosci 2009;64:373-81.
- 33. Biella CA, Salvador MJ, Dias DA, Dias-Baruffi M, Crotti LS. Evaluation of immunomodulatory and antiinflammatory effects and phytochemical screening of *Alternanthera tenella* Colla (*Amaranthaceae*) aqueous extracts. Memorias Inst Oswaldo Cruz 2008;103:569-77.
- 34. Teixeira CG, Piccoli A, Costa P, Soares L, Da Silva-Santos JE. Involvement of the nitric oxide/soluble guanylate cyclase pathway in the antioedematogenic action of *Pfaffia glomerata* (Spreng) Pedersen in mice. J Pharm Pharmacol 2006;58:667-75.
- 35. Freitas CS, Baggio CH, Da Silva-Santos JE, Rieck L, de Moraes Santos CA. Involvement of nitric oxide in the gastroprotective effects of an aqueous extract of *Pfaffia glomerata* (Spreng) Pedersen, *Amaranthaceae*, in rats. Life Sci 2004;74:1167-79.
- Sanoko R, Esperanza G, Pizza C, De Tommasi N. Triterpene saponins from *Alternanthera repens*. Phytochemistry 1999;51:1043-47.
- Wang P, Li S, Ownby S, Zhang Z, Yuan W. Ecdysteroids and a sucrose phenylpropanoid ester from *Froelichia floridana*. Phytochemistry 2009;70:430-6.
- Cai Y, Sun M, Corke H. Antioxidant activity of betalains from plants of the *Amaranthaceae*. J Agric Food Chem 2003;51:2288-94.
- Liu H, Cao J, Jiang W. Evaluation of physiochemical and antioxidant activity changes during fruit on-tree ripening for the potential values of unripe peaches. Sci Horticult 2015;193:32-9.
- Souza JG, Tomei RR, Kanashiro A, Kabeya LM, Azzolini AE. Ethanolic crude extract and flavonoids isolated from *Alternanthera maritima*: Neutrophil chemiluminescence inhibition and free radical scavenging

activity. Z Naturforsch C J Biosci 2007;62:339-47.

- Simpson MG. Plant Systematics. 2<sup>nd</sup> ed. United States: Academic Press, Elsevier; 2010. p. 301-2.
- Uusiku NP, Oelofse A, Duodu KG, Bester MJ, Faber M. Nutritional value of leafy vegetables of Sub-Saharan Africa and their potential contribution to human health: A review. J Food Compos Anal 2010;23:499-509.
- Koh HL, Chua TK, Tan CH. A Guide to Medicinal Plant, an Illustrated, Scientific and Medicinal Approach. Singapore: World Scientific; 2009. p. 292.
- Wee YC. A Guide to Medicinal Plants. Singapore: Singapore Science Centre Publication; 1992.
- The National Academies Press. Lost Crops of Africa. Vol. 2. Washington, DC: The National Academies Press; 2006.
- Haya F, Nirit B, Moshe B, Ilona R, Zeev BN, Atara Z. Application of secondary-treated effluents for cultivation of sunflower (*Helianthus annuus* L.) and celosia (*Celosia argentea* L.) as cut flowers. Sci Horticult 2007;115:62-9.
- Etkin NL. Medicinal cuisines: Diet and ethnopharmacology. Int J Pharmacogn 1996;34:313-26.
- Pieroni A. Medicinal plants and food medicines in the folk traditional of the upper Lucca Province, Italy. J Ethnopharmacol 2000;70:235-7.
- 49. Wiart C. Medicinal Plants of Southeast Asia. Malaysia: Pelanduk Publications; 2000.
- Nidavani RB, Mahalakshmi AM, Seema M, Krishna KL. Pharmacology of *Celosia argentea* L. J Atoms Mol 2014;4:635-44.
- Jagtap SD, Deokule SS, Pawar PK, Harsulkar AM. Traditional ethnomedicinal knowledge confined to the Pawra tribe of Satpura hills, Maharashtra, India. Ethnob Leaf 2009;13:98-115.
- Vetrichelvan T, Jegadeesan M, Devi B. Anti-diabetic activity of alcoholic extract of *Celosia argentea* LINN seeds in rats. Biol Pharm Bull 2002;25:526-8.
- Li S, Long C, Liu F, Lee S, Guo Q, Li R, et al. Herbs for medicinal baths among the traditional Yao communities of China. J Ethnopharmacol 2006;108:59-67.
- Ticktin T, Dalle SP. Medicinal plant use in the practice of midwifery in rural Honduras. J Ethnopharmacol 2005;96:233-48.
- Grosvenor PW, Supriono A, Gray DO. Medicinal plants from Riau Province, Sumatra, Indonesia. Part 2: Antibacterial and antifungal activity. J Ethnopharmacol 1995;45:97-111.
- Dung N, Loi DT. Selection of traditional medicines for study. J Ethnopharmacol 1991;32:57-70.
- Chen CP, Lin CC, Tsuneo N. Screening of Taiwanese crude drugs for antibacterial activity against *Streptococcus mutans*. J Ethnopharmacol 1989;27:285-95.
- Houghton PJ, Osibogun IM. Flowering plants used against snakebite. J Ethnopharmacol 1993;39:1-29.
- Grubben GJ, Denton OA. Plant Resources of Tropical Africa 2. Vegetables. Wageningen, Leiden: PROTA Foundation, Backhuys, CTA; 2004.
- Omonike OO, Adebayo AG, Edith OA. Ethnobotanical survey of plants used in treatment of inflammatory diseases in Ogun state of Nigeria. Eur J Sci Res 2010;43:183-91.
- Padal SB, Murty PP, Rao DS, Venkaiah M. Ethnomedicinal plants from Paderu division of Visakhapatnam district, A.P, India. J Phytol 2010;2:70-91.
- Markandeya AG, Firke NP, Pingale SS, Gawale SS. Quantitative elemental analysis of *Celosia argentea* leaves by ICP-OES techniques using different digestion methods. Int J Chem Anal Sci 2013;4:175-81.
- Patil HM, Bhaskar VV. Medicinal uses of plants by tribal medicine men of Nandurbar district in Maharashtra. Natl Prod Radiance 2006;5:125-30.
- 64. Anonymous. The Wealth of India. New Delhi: CSIR; 1992. p. 414-5.
- Hase K, Kadota S, Basnet P. Immunostimulating activity of celosian, an antihepatotoxic polysaccharide isolated from *Celosia argentea*. Planta Med 1997;63:216-9.
- Vetrichelvan T, Jegadeesan M. Hepatoprotective effects of traditional medicines. Isolation of the active constituent from seeds of *Celosia* argentea. Indian Drugs 2000;37:286-8.
- Hase K, Kadota S, Basnet P, Takahashi T, Namba T. Research article hepatoprotective effects of traditional medicines. Phytother Res 1996a;10:387-92.
- Haribabu S, Adupa SK. Phytochemical screening and hepatoprotective activity of *Celosia argentea* Linn. J Pharm Res 2014;8:405-9.
- Jain A, Katewa SS, Galav PK, Sharma P. Medicinal plant diversity of Sitamata wildlife sanctuary, Rajasthan, India. J Ethnopharmacol 2005;102:143-57.
- 70. Katewa SS, Chaudhary BL, Jain A. Folk herbal medicines from tribal

area of Rajasthan, India. J Ethnopharmacol 2004;94:41-6.

- Rajendra MK, Nitin BG, Nanasaheb PD, Mangesh MK, Sudhir MV, Sneha AK. Use of *Celosia argentea* Linn aqueous flower extract as a natural indicator in acid base titration. Int J PharmTech Res 2014;6:80-3.
- Hayakawa Y, Fujii H, Hase K, Ohnishi Y, Sakukawa R, Kadota S, *et al.* Anti-metastatic and immunomodulating properties of the water extract from *Celosia argentea* seeds. Biol Pharm Bull 1998;21:1154-59.
- Devhare SV, Nirmal SA, Rub RA, Dhasade VV, Zaware BB, Mandal SC. Immunomodulating activity of *Celosia argentea* Linn aerial parts. Lat Am J Pharm 2011;30:168-71.
- Gbadamosi IT, Alia AE, Okolosi O. *In-vitro* antimicrobial activities and nutritional assessment of roots of ten Nigerian vegetables. N Y Sci J 2012;5:234-40.
- Diéméléou CA, Zoué LT, Niamké SL. Antioxidant and antifungal properties of seed oils extracted from three leafy vegetables plants consumed in Côte d'Ivoire. J Nat Prod Plant Resour 2013;3:7-13.
- Eseoghene O, Kola' KA. Antimicrobial activity of *Celosia argentea* L. *Amaranthaceae*. Am J Res Commun 2015;3:123-33.
- Bhujbal SS, Chitlange SS, Suralkar AA, Shinde DB, Patil MJ. Antiinflammatory activity of an isolated flavonoid fraction from *Celosia* argentea Linn. J Med Plants Res 2008;2:52-4.
- Qingbin W, Yan W, Meili G. Triterpenoid, saponins from the seeds of *Celosia argentea* and their anti-inflammatory and antitumor activities. Chem Pharm Bull 2011;59:666-71.
- Joshi PC, Patil SA, Sambrekar SN. Evalution of antiurolithiatic activity of ethanolic extract of *Celosia argentea* (seed) in rats. Univ J Pharm 2012;1:52-60.
- Malomo SO, Ore A, Yakubu MT. *In vitro* and *in vivo* antioxidant activities of the aqueous extract of *Celosia argentea* leaves. Indian J Pharmacol 2011;43:278-85.
- Dalimartha S. Atlas Tumbuthan Obat Indonesia. Jakarta: Niaga Swadaya; 2007. p. 2-4.
- Wu JN. An Illustrated Chinese Materia Midica. Oxford: Oxford University Press; 2005. p. 168.
- Ashok KC, Divya Sree MS, Joshna A, Mohana LS, Satheesh KD. A review on South Indian edible leafy vegetables. J Glob Trends Pharm Sci 2013;4:1248-56.

- Ying T, Hai-Liang X, Mei-Li G. Review on research of the phytochemistry and pharmacological activities of *Celosia argentea*. Rev Bras Farmacogn 2016;26:787-96.
- Jun'ichi K, Hayato S, Kazutaka S, Koichi T, Hiroshi M. Celogentins AC. New antimitotic bicyclic peptides from the seeds of *Celosia argentea*. J Org Chem 2001;66:6626-33.
- Cai Y, Sun M, Schliemann W, Corke H. Chemical stability and colorant properties of betaxanthin pigments from *Celosia argentea*. J Agric Food Chem 2001;49:4429-35.
- Schliemann W, Cai Y, Degenkolb T, Schmidt J, Corke H. Betalains of Celosia argentea. Phytochemistry 2001;58:159-65.
- Morita H, Suzuki H, Kobayashi J. Celogenamide A, a new cyclic peptide from the seeds of *Celosia argentea*. J Nat Prod 2004;67:1628-30.
- Xue Q, Suna ZL, Guo ML, Ying W, Zhang G, Wang XL. Two new compounds from semen *Celosiae* and their protective effects against CCl<sub>4</sub>-induced hepatotoxicity. Natl Prod Res 2011;25:772-80.
- Satyanarayana V. Preliminary phytochemical screening and TLC profile of selected four plants of Tirupati hills in Chitoor district, Andhra Pradesh. J Pharmacogn Phytochem 2016;5:259-64.
- Satyavathi K, Lakshmi KK, Prasenjit M, De A, Jana S, Aneela S. Evaluation of anthelmintic activity of *Carica Papaya Latex* using *pheritima posthuma* anthelmintic activity. Pharm Sci 2012;2:10-2.
- Neha S. Anthelmintic activity of extracts of some medicinal plants. Int J Comput Sci Mathe 2011;3:183-7.
- Ojieh AE, Adegor EC, Lawrence EO. Preliminary phytochemical screening, analgesic and anti-inflammatory properties of *Celosia Isertii*. Eur J Med Plants 2013;3:369-80.
- Harborne JB. A Guide to Modern Technique of Plant Analysis. London: Chapman and Hill; 1973. p. 279.
- Brain KR, Turner TD. Practical Evaluation of Phytopharmaceuticals. 1<sup>st</sup> ed. Bristol: Wright-Scientechnica; 2013. p. 69-380.
- Mythili I T, Ravindhran R. Phytochemical screening and antimicrobial activity of Sesbania sesban (L.) Merr. Asian J Pharm Clin Res 2012;5:179-82.
- Hacene E. Effect of prodigiosin from *Serratia marcescens* br1 strain as an antioxidative, antimicrobial, and *in vivo* wound healing. Asian J Pharm Clin Res 2020;13:175-9.