

EFFECT OF GROWTH REGULATORS ON CALLUS INDUCTION FROM *CYCLEA PELTATA* (LAM.) HOOK. F. THOMS.

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Received: 14 August 2013, Revised and Accepted: 11 September 2013

ABSTRACT

Objective: To evaluate the effect of nutrient media and growth regulators on the stem explants of *Cyclea peltata* followed by the histological observations for the induction of callus.

Methods: The stem explants were inoculated on to MS, B5 and Nitsch media to check the suitable nutrient medium for the *in vitro* culture. Growth regulators such as 2, 4 - D, NAA, IAA, BAP and Kn were added to MS medium prior to the inoculation of stem explants and observed for the induction of callus, nature and color/pigmentation of the callus and initiation of roots and shoots. Callus induction was confirmed by histological observations.

Results: The young stem explants of *Cyclea peltata* cultured on MS medium supplemented with different concentrations of growth regulators showed the induction of callus. The callus obtained on the medium supplemented with either 2, 4 -D alone or in combination with BAP/Kn was yellowish cream and friable whereas, the callus obtained in the medium fortified with NAA/IAA alone or with BAP/Kn was compact and light to dark green in color. NAA (2mg/l), 2, 4 - D + BAP (1 + 3mg/l), 2, 4 - D + Kn (1 + 2, 2 + 0.1mg/l), NAA + BAP and IAA + BAP (3 + 2mg/l and 0.1 + 0.5mg/l) of concentrations induced significantly a higher number of calli. The histological study revealed the induction of callus from the cambial region of the explants.

Conclusion: The study revealed the induction of callus from the cambial cells of stem explants of *Cyclea peltata* when cultured on MS medium supplemented with combinations of auxin and cytokinins.

Keywords: *Cyclea peltata*, auxin, cytokinin, callus

INTRODUCTION

Cyclea peltata (Lam.) Hook. f. Thoms. (Menispermaceae) is a medicinally important plant used in the treatment of various health problems. *C. peltata* is reported to have anti-toxin function besides playing a role in diabetic disorders of skin, like boils and carbuncle and antidiabetic, anti-inflammatory activities, protection against nephrotoxicity and oxidative damage [1-5]. Jain [6] reported the use of *Cyclea peltata* for small pox and stomach ache.

The *in vitro* culturing of cells, tissues or organs depends on various physico-chemical factors such as light, temperature, humidity, nutrient supply, pH conditions, growth regulators, season for explant collection, type/age of explant and genotype. Nurazah et al. [7] studied the role of explant age on *in vitro* culturing in cotton at weekly intervals. The best result for the multiple shoot induction from the nodal explants of *Madhuca latifolia* Macb. was observed when the explants were collected in the month of May [8].

The role of different growth regulators on the induction of callus with varying colors were reported in *Ruta graveolens* with the induction of green to yellow colored callus in the medium with NAA [9], *Phyllanthus amarus* with the induction of dark green, nodular callus in the medium supplemented with BAP/TDZ [10] and *Pulsatilla koreana* with the induction of soft, light green callus in the medium with Kn [11]. In the present study an attempt was made to check the type of nutrient medium, age of the explant and type of suitable growth regulators to induce the optimum callus from different explants of *C. peltata*.

MATERIALS AND METHODS

Plant material and surface sterilization

The stem of *C. peltata* (Menispermaceae) were collected from natural forests of Dakshina Kannada District, Karnataka, India, and identified following the Flora of Udipi and Dakshin Kannada [12] and the voucher specimen (MU/AB/BN-02) were deposited at the herbarium of Department of Applied Botany.

The mature, young and youngest explants were collected to find out the suitable explant for *in vitro* culture. The collected plant materials were washed thoroughly under running tap water for 20-30min to remove the surface debris [13]. The explants were treated with Bavistine (30-45min), 70% alcohol (30sec) and 0.1% HgCl₂ (10min). After each step, the explants were washed in sterile distilled water. Later, the explants were cut into ~0.5cm and inoculated on to nutrient medium.

Effect of nutrient media and growth regulators on the stem explants of *C. peltata*

Three different types of culture media *viz:* MS, B5 and Nitsch were used in the present study to find out the suitable medium for callus induction. The stem explants with ~5cm size were inoculated on to MS medium with varying concentrations of growth regulators either singly or in combinations. The selected growth regulators include 2, 4 - D, NAA, IAA, BAP and Kn (0.1- 3mg/l). All the cultures were incubated at 25 ± 2°C with 16hr of photoperiod and 40.0 ± 3.0 μmol m⁻² s⁻¹ light intensity unless specified. The explants were observed at the end of 25-30d of incubation for various details such as development of callus from mature or immature regions of the explants, nature of callus, color/pigmentation of the callus, initiation of roots and shoots. The histological studies of the Carnoy's B fluid fixed cultures were carried out to observe the induction of callus from the explants.

Statistical analysis of the data

The data presented are the average of three replicates and expressed as Mean ± SD. The statistical analysis of the data was carried out using SAS package (version 9.0) and the treatment means were compared using Duncun's Multiple Range Test (DMRT) at a level of 5% significance.

RESULTS AND DISCUSSION

Effect of nutrient media on the stem explants of *C. peltata*

There was no significant variation between the callus induced in three types of media by the explants of *C. peltata*. Therefore, the most commonly used MS medium was selected for further experiments. However, there was a significant variation in callus induction with respect to the concentrations of growth regulators used in the study and age of the explants taken. MS medium is the most common and suitable nutrient medium used for the *in vitro* culturing of many plant species. The culturing of four genotypes of rice in MS, N6 and R media showed a better callus induction on MS medium compared to N6 and Regeneration (R) media [14]. Similarly, a fairly good callus was observed in the present study irrespective of the media used. Thus, MS medium was used in all further experiments as this is the versatile medium used in culture studies.

The young explants of *C. peltata* responded well with the initial swelling followed by the emergence of callus from the explants after 10d of incubation. In three different tomato cultivars, Hana, Premium and Money maker, the *in vitro* regeneration of shoots was observed better when 8-10d old cotyledon and hypocotyl explants were used for the culturing [15]. The *in vitro* culturing of lettuce for callus induction showed better result with 3d old explants indicating the age as the critical factor for callus induction [16].

Effect of growth regulators on the stem explants of *C. peltata*

The stem explants of *C. peltata* cultured on medium supplemented with auxin (2, 4 -D, NAA, IAA) or cytokinin (BAP, Kn) or combinations of auxin with cytokinin showed morphologically different types of callus irrespective of the concentrations of growth regulators used in the study (Table 1). The explants showed a twist and swelling (Fig. 1a) soon after a week of incubation followed by the emergence of callus all over the explants. The twisting and swelling of the explant observed in *C. peltata* may be due to the internal pressure developed during the incubation. Similar to this result, the swelling of explants during the induction of callus was reported in the hypocotyl and leaf explants of *Meconopsis simplicifolia* [17], nodal explants of *Dendrocalamus hamiltonii* Nees et Arn. Ex Munro [18] and leaf explants of *Mentha piperita* (L.) [19].

Table 1: Effect of growth regulators on callus, root and shoot induction from *C. peltata* stem explants

Growth regulator s	Callus texture	Callus color	Root induction	Shoot induction
2, 4 - D, 2, 4 - D + BAP/ Kn	Friable	Yellowish cream	Nil	Nil
NAA, NAA+ BAP/Kn	Compact	Light to dark green	Nil	Nil
IAA, IAA + BAP/Kn	Compact	Cream to green	Nil	Nil
BAP, Kn	Hard	Light green	Nil	Yes

The medium supplemented with either 2, 4 -D alone or in combination with BAP/Kn induced yellowish cream, friable callus (Fig. 1b). The callus obtained in the medium fortified with NAA/IAA or NAA/IAA + BAP/Kn was compact and light to dark green in color (Fig. 1c). This result is in agreement with the report of Shirin et al. [20], where in, they have observed the formation of green colored calli of *Solanum tuberosum* in the medium supplemented with NAA and BAP and yellow colored calli in 2, 4 - D and green colored calli in the medium supplemented with NAA alone or in combination with BAP. The induction of callus from other members of Menispermaceae viz. *Stephania cepharantha* Hayata [21], *Coscinium fenestratum* [22] and *Tinospora cordifolia* [23] were reported when cultured on MS medium supplemented with 2, 4 - D + Kn/BAP. Based on the results obtained in different plant species and from the present study, it is clear that the callus nature, texture and color are highly species specific.

The nodal explants showed the development of dark green colored callus (Fig. 1d). However, a few of the nodal explants showed the induction of single axillary shoot (Fig. 1e) and these shoots failed to induce roots upon transfer to medium with IBA or IAA. The direct organogenesis in *Tinospora cordifolia* from the nodal explants [24] and *Psoralea corylifolia* [25] from the shoot tips were reported when cultured on MS medium fortified with varying concentration of growth regulators. The *in vitro* regeneration of *C. peltata* using nodal explants was reported by Abraham et al. [26] where, the explants were cultured on MS medium supplemented with 3mg/l of BAP and 0.5mg/l of IAA to induce the highest regeneration. However, in the present work, the use of same growth regulators at the same concentration induced only 1-2 axillary shoots and these failed to induce roots. This may be due to various factors like environmental conditions, genotype of the explant, culture conditions, and concentration of endogenous growth regulators.

The initiation of callus in the stem explants of *C. peltata* was observed from the cambial region (Fig. 1f). The cells were arranged one above the other with a prominent nucleus. The induction of callus from the cambial cells, phloem and cortical cells was reported in tea [27] and black pepper [28]. In *Vigna radiata* W. the induction of callus started from the outer layer of cambial cells [29]. Similarly, in *C. peltata* the histological studies confirmed the induction of callus from the cambial region of the explants.

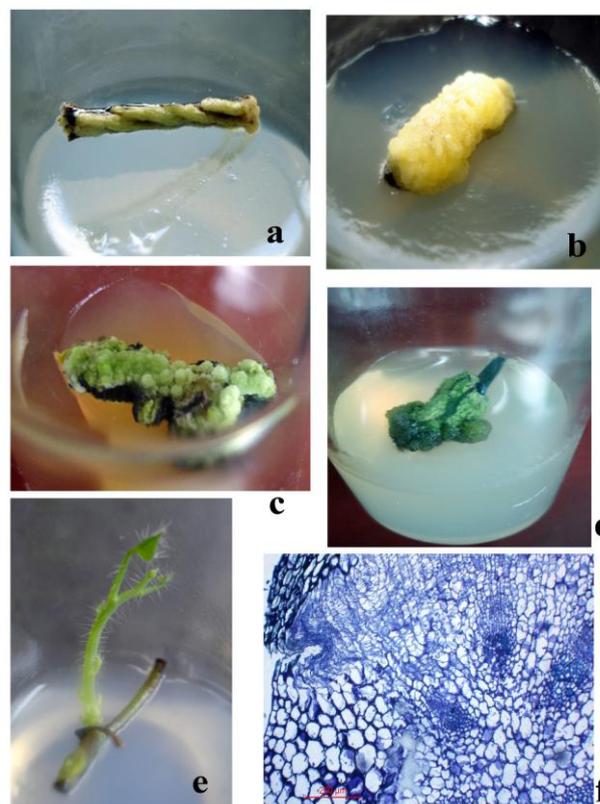


Fig. 1: a Twisted explant showing the induction of callus; **b** Callus induction on medium supplemented with 2, 4 - D; **c** Callus induction on medium supplemented with NAA; **d** Callus induction from nodal explant; **e** Emergence of single axillary shoot from nodal explant; **f** Stem explant showing callus induction from the cambial region (bar= 0.2mm)

The stem explants responded positively by inducing the callus when cultured on medium supplemented with either auxin or cytokinin with a highest mean number of 6.66 ± 0.57 explants in the medium with NAA at the concentration of 2mg/l (Table 2). The stem explants showed a highest callus induction (8.50 ± 0.71) when cultured on medium supplemented with 2, 4 - D + BAP at the concentrations of 1 + 3mg/l (Table 3).

Table 2: Effect of growth regulators on callus induction from *C. peltata* stem explants: Auxin and cytokinin

Growth regulators	Number of explants showing callus (Mean±SD)				
	0.1	0.5	1	2	3
2, 4- D	1.33±0.57 ^{bcd}	2.66±2.08 ^{bcd}	5.33±0.57 ^a	3.66±1.15 ^b	1.33±0.57 ^{bcd}
NAA	1.00±0.00 ^{bcd}	4.00±1.00 ^a	6.33±1.15 ^a	6.66±0.57 ^a	2.33±1.52 ^{bcd}
IAA	5.00±0.00 ^a	3.66±1.52 ^b	2.33±0.57 ^{bcd}	2.00±2.00 ^{bcd}	3.00±1.00 ^{bc}
BAP	2.33±1.15 ^{bcd}	4.33±0.57 ^a	4.33±0.57 ^a	4.00±2.64 ^a	0.33±0.57 ^{bcd}
Kn	2.66±2.30 ^{bcd}	5.33±0.57 ^a	5.66±1.52 ^a	1.66±1.52 ^{bcd}	0.66±0.57 ^{bcd}

SE/plot = 1.25, CV (%) = 38.24

Values with the different letters indicate significant difference at 5% level.

Table 3: Effect of growth regulators on callus induction from *C. peltata* stem explants: 2, 4 - D + BAP and 2, 4 - D + Kn

2, 4-D	Number of explants showing callus (Mean±SD)				
	0.1	0.5	1	2	3
BAP					
0.1	5.00±0.00 ^b	4.50±0.71 ^b	5.00±1.41 ^b	7.00±1.41 ^a	6.50±0.71 ^a
0.5	5.50±0.71 ^b	6.00±0.00 ^a	5.50±0.71 ^b	6.00±1.41 ^b	7.50±0.70 ^a
1	7.50±0.70 ^a	4.50±0.71 ^b	8.50±0.70 ^a	7.00±0.00 ^a	8.50±0.71 ^a
2	6.50±0.70 ^a	7.50±0.70 ^a	6.00±1.41 ^b	4.50±0.71 ^b	5.50±0.71 ^b
3	5.00±1.41 ^b	5.50±2.12 ^b	5.50±0.71 ^b	4.50±2.12 ^b	5.50±0.71 ^b
SE/plot=1.03, CV (%) = 17.10					
Kn					
0.1	3.50±0.71 ^{b-l}	6.00±1.41 ^{b-i}	4.00±1.41 ^{b-l}	5.50±0.71 ^{b-k}	5.00±0.00 ^{b-k}
0.5	3.50±0.70 ^{b-l}	2.50±0.70 ^{b-l}	2.50±0.70 ^{b-l}	3.50±0.70 ^{b-l}	2.50±0.71 ^{b-l}
1	3.00±1.41 ^{b-l}	6.50±0.71 ^{b-g}	8.00±1.41 ^a	9.50±0.71 ^a	8.50±0.71 ^a
2	9.50±0.71 ^a	8.00±0.00 ^a	9.00±0.00 ^a	7.50±0.71 ^{bc}	7.50±0.71 ^{bc}
3	5.50±0.70 ^{b-k}	8.50±0.71 ^a	6.50±0.70 ^{b-g}	6.00±0.00 ^{b-i}	8.50±0.70 ^a

SE/plot = 0.81, CV (%) = 13.49

Values with the different letters indicate significant difference at 5% level.

The stem explants showed a highest mean number of 9.50 ± 0.71 explants for callus induction when cultured on medium supplemented with at 2, 4 - D + Kn at the concentration of 1 + 2 and 2 + 0.1mg/l (Table 3). The stem explants showed a highest mean number of 9.50 ± 0.71 and 8.50 ± 2.12 explants for callus induction when cultured on medium supplemented with NAA + BAP and IAA + BAP at the concentrations of 3 + 2mg/l and 0.1 + 0.5mg/l respectively (Table 4 and 5).

Table 4: Effect of growth regulators on callus induction from *C. peltata* stem explants: NAA+BAP and NAA + Kn

NAA	Number of explants showing callus (Mean±SD)				
	0.1	0.5	1	2	3
BAP					
0.1	8.50±0.71 ^a	6.00±0.00 ^{b-e}	6.50±0.71 ^{b-d}	7.00±1.41 ^b	7.00±0.00 ^b
0.5	6.00±0.00 ^{b-e}	5.50±0.71 ^{b-e}	5.00±1.41 ^{b-h}	7.00±1.41 ^b	6.00±1.41 ^{b-e}
1	7.50±0.70 ^a	6.00±0.00 ^{b-e}	5.00±1.41 ^{b-h}	7.00±1.41 ^b	3.50±2.12 ^{b-i}
2	4.00±0.00 ^{b-i}	4.50±0.71 ^{b-i}	9.00±1.41 ^a	5.00±1.41 ^{b-h}	3.00±1.41 ^{b-i}
3	7.50±0.71 ^a	7.50±0.70 ^a	4.00±1.41 ^{b-i}	9.50±0.71 ^a	9.00±1.41 ^a
SE/plot=1.10, CV (%) = 17.64					
Kn					
0.1	6.00±0.00	8.00±1.41	9.50±0.71	9.00±1.41	9.00±1.41
0.5	7.50±0.71	6.50±0.71	8.50±2.12	8.00±0.00	6.00±1.41
1	7.00±1.41	6.50±0.71	8.00±2.83	7.00±1.41	5.50±2.12
2	9.50±0.70	7.50±0.70	9.00±1.41	8.00±2.83	6.50±2.12
3	10.00±0.00	10.00±0.00	9.00±0.00	10.00±0.00	9.50±0.70

SE/plot=1.37, CV (%) = 17.05

Values with the different letters indicate significant difference at 5% level.

Table 5: Effect of growth regulators on callus induction from *C. peltata* stem explants: IAA+BAP and IAA + Kn

IAA	Number of explants showing callus (Mean±SD)				
	0.1	0.5	1	2	3
BAP					
0.1	1.00±1.41 ^{b-l}	8.50±2.12 ^a	6.50±0.71 ^{b-d}	8.00±1.41 ^a	7.00±2.83 ^b
0.5	3.50±0.71 ^{b-k}	7.00±1.41 ^b	8.00±1.41 ^a	5.50±0.71 ^{b-f}	5.00±1.41 ^{b-g}
1	5.00±0.00 ^{b-g}	4.50±0.71 ^{b-h}	2.50±0.70 ^{b-l}	3.50±0.70 ^{b-k}	1.00±0.00 ^{b-l}
2	1.50±2.12 ^{b-l}	2.00±1.41 ^{b-l}	1.00±1.41 ^{b-l}	6.00±1.41 ^{b-d}	4.00±0.00 ^{b-i}
3	4.00±1.41 ^{b-i}	2.50±0.71 ^{b-l}	2.00±0.00 ^{b-l}	2.50±0.71 ^{b-l}	1.50±2.12 ^{b-l}
SE/plot = 1.32, CV (%) = 31.86					
Kn					
0.1	3.00±1.41	3.00±0.00	2.50±0.71	1.00±1.41	1.00±1.41
0.5	5.50±0.71	3.00±1.41	3.50±0.70	1.00±1.41	2.50±3.55
1	3.50±2.12	4.50±0.71	2.50±0.71	2.50±0.71	4.00±0.00
2	5.50±0.71	2.00±0.00	3.00±1.41	5.00±0.00	2.50±0.71
3	4.00±1.41	4.50±0.71	4.00±1.41	4.00±1.41	3.00±1.41

SE/plot = 1.29, CV (%) = 40.01

Values with the different letters indicate significant difference at 5% level.

The influence of different concentrations and combinations of growth regulators on the induction of callus from the nodal explants of *Rauwolfia serpentina* L. Benth showed a better response in the medium supplemented with BAP + NAA at the concentration of 0.5 + 2mg/l [30]. Nurazah et al.[7] studied the effect of growth regulators on callus induction from the flower petal explants of *Cananga odorata* and observed a better response in the medium supplemented with NAA + BAP (3 + 0.5mg/l). The induction of callus using auxin alone or in combination with cytokinin in MS medium was reported in different explants of various plant species viz. cotyledon explants of *Mucuna pruriens* (L.) DC. in the medium with 6.7µM of 2, 4 - D [31] and leaf explants of *Phyllanthus amarus* in the medium with 2.26µM of 2, 4 - D and 2.32µM of Kn [10]. In the present study, the stem explants of *C. peltata* responded well when cultured on the medium supplemented with combinations of auxin and cytokinins selected in the study. This suggests that the callus induction and adventitious shoot regeneration from the selected explants depended on the genotype and combinations of growth regulators as reported by Zhang et al. [32]. The callus obtained may be maintained on suitable culture condition either to regenerate the plantlets or to produce pharmaceutically important bioactive compounds.

ACKNOWLEDGMENTS

Senior author (BN) is grateful to UGC, New Delhi for financial assistance under RFSMS scheme. Authors are thankful to VGST, Department of IT, BT and S & T, Govt. of Karnataka, India for financial assistance.

REFERENCES

- Begum and Nath. Ethnobotanical review of medicinal plants used for skin diseases and related problems in Northeastern India, Journal of Herbs, Spices and Medicinal Plants 2000; 7: 55-93
- Bhandary MJ and Chandrashekar KR. Herbal therapy for herpes in the ethno-medicine of Coastal Karnataka. Ind. J. Trad. Knowledge 2011; 10(3): 528-532
- Christina AJM, Lakshmi PM, Nagarajan M and Kurian S. Modulatory effect of *Cyclea peltata* Lam. on stone formation induced by ethylene glycol treatment in rats. Methods Find Exp Clin Pharmacol 2002; 24: 77
- Hullatti KK and Sharada MS. Comparative antipyretic activity of Patha: An Ayurvedic drug, Phcog Mag. 2007; 3: 173
- Vijayan FP, Rani VK, Vineesh VR, Sudha KS, Michael MM and Padikkala J. Protective effect of *Cyclea peltata* Lam on cisplatin-induced nephrotoxicity and oxidative damage. J Basic Clin Physiol Pharmacol. 2007; 18: 101-114
- Jain SK. Dictionary of Indian folk medicine and ethnobotany, A reference manual of man-plant relationships and ethnobotanists, Deep publications, New Delhi, India. 1991; 110
- Nurazah Z, Radzali M, Syahida A and Maziah M. Effect of plant growth regulators on callus induction from *Cananga odorata* flower petal explants. Afr J Biotechnol. 2009; 8: 2740-2743
- Bansal YK and Chibbar T. Micropropagation of *Madhuca latifolia* Macb. through nodal culture, Plant Biotechnol. 2000; 17: 17-20
- Tejavathi DH, Devaraj VR, Murthy SM, Anitha P and Nijagunaiah R. Regeneration of multiple shoots from the callus cultures of *Macrotyloma uniflorum* (Lam.) Verdc. Indian J Biotechnol. 2010; 9: 101-105
- Nitnaware KM, Naik DG and Nikam TD. Thidiazuron-induced shoot organogenesis and production of hepatoprotective lignan phyllanthin and hypophyllanthin in *Phyllanthus amarus*, Plant Cell Tiss Organ Cult. 2011; 104: 101-110
- Lin G-Z, Zhao X-M, Hong S-K and LianY-J. Somatic embryogenesis and shoot organogenesis in the medicinal plant *Pulsatilla koreana* Nakai. Plant Cell Tiss Organ Cult. 2011; 106: 93-103
- Bhat KG. Flora of Udupi, Indian Naturalist (Regd.), Udupi, Karnataka, India. 2003.
- Bhagya N, Chandrashekar KR, Muralidharan K, Amarnath CH. Phytochemical analysis and antioxidant activity of *in vitro* cultured stem callus of *Cyclea peltata* (Lam.) Hook. f. & Thoms. J. Trop. Med. Plants. 2012; 13(2): 117-123
- Khatun MM, Ali MH and Desamero NV. Effect of genotype and culture media on callus formation and plant regeneration from mature seed scutella culture in rice. Plant Tissue Cult. 2003; 13: 99-107
- Gubis J, Lajchova Z, Farago J and Jurekova Z. Effect of growth regulators on shoot induction and plant regeneration in tomato (*Lycopersicon esculentum* Mill.), Biologia (Bratislava). 2004; 59: 405-408.
- Mohebodini M, Javaran MJ, Mahboudi F and Alizadeh H. Effects of genotype, explant age and growth regulators on callus induction and direct shoot regeneration of Lettuce (*Lactuca sativa* L. AJCS. 2011; 5(1):92-95
- Sulaiman IM, Rangaswamy NS and Babu CR. Formation of plantlets through somatic embryogeny in Himalayan blue poppy, *Meconopsis simplicifolia* (Papaveraceae). Plant Cell reports. 1991; 9: 582-585
- Godbole S, Sood A, Thakur R, Sharma M and Ahuja PS. Somatic embryogenesis and its conversion into plantlets in a multipurpose bamboo, *Dendrocalamus hamiltonii* Nees et Arn. Ex Munro. Cur Sci. 2002; 83: 885-889
- Sujana P and Naidu CV. Indirect Plant Regeneration from Leaf Explants of *Mentha piperita* (L.) – An Important Multipurpose Medicinal Plant. Journal of Phytology. 2011; 3: 19-22
- Shirin F, Hossain M, Kabir MF, Roy M and Sarker SR. Callus induction and plant regeneration from internodal and leaf explants of four potato (*Solanum tuberosum* L.) cultivars. World J Agricultural Sci. 2007; 3: 01-06
- Suzuki S, Fujino H, Tatsuo Y, Yamazaki N and Yoshizaki M. Japan J Breed. Rapid propagation of *Stephania cephalantha* Hayata by tissue culture. 1992; 42: 769-777
- Nair AJ, Sudhakaran PR, Rao JM and Ramakrishna SV. Berberine synthesis by callus and cell suspension cultures of *Coscinium fenestratum*. Plant Cell Tissue Organ Cult. 1992; 29: 7-10
- Chintalwar GJ, Gupta S, Roja G and Bapat VA. Protoberberine alkaloids from callus and cell suspension cultures of *Tinospora cordifolia*. Pharmaceut Biol. 2003; 41: 81-86
- Gururaj HB, Giridhar P and Ravishankar GA. Micropropagation of *Tinospora cordifolia* (Willd.) Miers ex Hook. F and Thoms: a multipurpose medicinal plant. Cur Sci. 2007; 92:23-26.
- Pandey P, Mehta R and Upadhyay R. In vitro propagation of an endangered medicinal plant *Psoralea corylifolia* Linn. Asian Journal of Pharmaceutical and Clinical Research. 2013; 6(3): 115-118.
- Abraham J, Cheruvathur MK, Mani B and Thomas TD. A rapid *in vitro* multiplication system for commercial propagation of pharmaceutically important *Cyclea peltata* (Lam) Hook and Thoms. based on enhanced axillary branching. Ind Crop Prod. 2010; 31: 92-98.
- Frisch CH and Camper ND. Effect of synthetic auxins on callus induction from tea stem tissue, Plant Cell Tissue Organ Cult. 1987; 8: 207-213.
- Sujatha R, Babu CL, Nazeem PA. Histology of organogenesis from callus cultures of black pepper (*Piper nigrum* L.). J Trop Agr. 2003; 41:16-19.
- Park J-B, Lee K-B, Lee S. Histological study of callus formation and root regeneration from mung bean (*Vigna radiata* W.). J Plant Biol. 2002; 45:170-176.
- Salma U, Rahman MSM, Islam S, Haque N, Jubair TA, Haque AKMF and Mukti IJ. The influence of different hormone concentration and combination on callus induction and regeneration of *Rauwolfia serpentina* L. Benth. Pakistan J Biol Sci. 2008; 11: 1638-1641
- Vibha JB, Choudhary K, Singh M, Rathore MS and Shekhawat NS. An efficient somatic embryogenesis system for velvet bean [*Mucuna pruriens* (L.) DC.]: a source of anti parkinson's drug. Plant Cell Tissue Organ Cult. 2009; 99: 319-325
- Zhang C-L, Chen D-F, Elliott MC and Slater A. Efficient procedures for callus induction and adventitious shoot organogenesis in sugar beet (*Beta vulgaris* L.) breeding lines. In Vitro Cell Dev Biol-Plant. 2004; 40: 475-481