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Review Article

LYNGBYA SP.: A SUITABLE CYANOBACTERIUM FOR HARVESTING ANTIMICROBIAL COMPOUNDS

MAHEEP KUMAR

Department of Botany, Guru Ghasidas Vishwavidyalaya, Koni, Bilaspur- 495009, India. Email: maheep.bot@gmail.com

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ABSTRACT

Lyngbya spp. is one of the most efficient groups of secondary metabolite producers and is very important from pharmaceutical point of view. Among its various species, *majuscula* is the major producers of commercially important biomolecules. Several species have been isolated and screened from the marine in the past decades. Although the chance of isolating a novel *Lyngbya* strain from a marine habitat, which would produce new biologically active metabolites, has increased, but rediscovery has also important aspect. The most relevant reason for discovering novel secondary metabolites is to circumvent the problem of resistant pathogens, which are no longer susceptible to the currently used drugs. Existence of *Lyngbya* has been reported in the hitherto untapped brackish ecosystem. Marine *Lyngbya* are efficient producers of new secondary metabolites that show a range of biological activities including antibacterial, antifungal, anticancer, insecticidal and enzyme inhibitor. Bioactive compounds from marine *Lyngbya* possess distinct chemical structures that may form the basis for synthesis of new drugs that could be used to combat resistant pathogens.

Keywords: Lyngbya · Bioactive compounds. Antibacterial. Antifungal. Secondary metabolites.

INTRODUCTION

Microbial natural products are an important source of both existing and new drugs. Among the producers of commercially important metabolites, cyanobacteria have proven to be a prolific source with a surprisingly small group of taxa accounting for the vast majority of compounds discovered till date [1-3]. Cyanobacteria produce a large number of compounds with varying bioactivities. A number of cyanobacteria and very few other microalgae, have been screened for antiviral activity so far, but the limited results available are promising [4]. Among these, Lyngbya spp. are the most economically and biotechnologically priceless prokaryotes. Representative species of Lybngbya include L. aestuarii, L. bouillonii, L. confervoides, L. <u>hieronymusii, L. kuetzingii, L. polychroa, L. semiplena, L. hieronymusii</u>, L. <u>hieronymusii</u> and several others, L. majuscula species has proved to be most potent. Although other species of Lyngbya may have potency but they are even untapped. The immense diversity of this habitat along with its underexploitation is the fundamental reason for attracting researchers towards it for discovering novel metabolite producers. Lyngbya spp. have also been isolated from terrestrial as well as brackish water bodies [5]. Lyngbya spp. as usual form bloom in marine habitat and produce noxious environment [6]. It has been found to contain a variety of chemicals that exerts a range of biological effects, including skin, eye and respiratory irritation. The toxins lyngbyatoxin A and debromoaplysiatoxins appear to give the most widely witnessed biological effects in relation to humans, and experiments involving these two toxins show the formation of acute dermal lesions. Studies into the epidemiology of the dermatitic, respiratory and eye effects of the toxins of this organism are reviewed and show that Lyngbya induced dermatitis has occurred in a number of locations [7]. Secondary metabolites produced by Lyngbya possess a wide range of biological activities [8]. The genus Lyngbya alone produces a large number (>100) of bioactive molecules [9-15]. It has an enormous biosynthetic potential that remains unchallenged without a potential competitor among other microbial groups. A large number of Lyngbya spp. have been isolated and screened from marine in the past several decades [16]. An important reason for discovering novel secondary metabolites is to circumvent the problem of resistant pathogens, which are no longer susceptible to the currently used drugs [17]. Secondary metabolites from marine Lyngbya strain may form the basis for the synthesis of novel therapeutic drugs, which may be efficient to combat a range of resistant microbes [18]. Many Lyngbya species isolates from oceans contain non-ribosomal

polyketide synthetase (NRPS) and polyketide synthetase (PKS) pathways, the hallmarks of secondary metabolite production[19-21].

Antibacterial compounds

The filamentous Cyanobacterial genus Lyngbya is being found to be a rich source of toxic and otherwise bioactive metabolites. Only bioassay of active fraction were done to prove that it has some antimicrobial activity Many of the compounds isolated from marine cyanobacteria have been shown to have specific targets in higher eukaryotic organisms (e.g., tubulin, actin) and have minimal or no antibacterial activity. Many novel structures have been elucidated as a result of systematic screenings for anticancer, antibacterial, antifungal, and protease inhibitory effects [22, 23]. Few are as follows. Four novel cyclic undecapeptides, lyngbyazothrins A, B, C , and D, were isolated from the cultured Lyngbya sp. The mixture of lyngbyazothrins A and B shows only low antimicrobial activity against Micrococcus flavus, whereas the mixture of lyngbyazothrins C and D was active against Bacillus subtilis, Escherichia coli, Pseudomonas aeruginosa, and Serratia marcescens [24]. Malyngolide is natural δ -lactones obtained from different strains of the marine cyanobacterium Lyngbya and characterized by a hydroxymethyl group and a long aliphatic chain attached to the δ -position of a sixmembered lactone ring. Malyngolide was isolated from L. majuscula collected in Hawaii [25]. Malyngolide displayed antibacterial activity against Mycobacterium smegmatis and Streptococcus pyogenes. Pahayokolide A showed specific inhibition of the two representatives of Gram-positive Bacillus sp. [26].

Antifungal compounds

Methanol extracts obtained by mechanical agitation of solvent with lyophilized coral sand sediment and *L. confervoides* filaments showed significant in vitro antifungal activity against *Candida albicans*. It was Lobocyclamides A-C which exhibited modest antifungal activity [27]. Lyngbyabellin B was isolated from a marine cyanobacterium, *L. majuscula,*. This new cyclic depsipeptide displayed potent toxicity toward brine shrimp and the fungus *Candida albicans* [28].

Other valuable compounds

Cytotoxic/Anticancer

A new stereoisomer of malyngamide C, 8-epi-malyngamide C, and the known compound lyngbic acid [(4E,7S)-7-methoxytetradec-4-

enoic acid] were isolated from a sample of L. majuscula. Lyngbic acid and malyngamide C were found cytotoxic against HT29 colon cancer and to inhibit bacterial quorum sensing [29]. Apratoxin A, a potent cytotoxin with a novel skeleton, has been isolated from the marine cyanobacterium L. majuscula. Apratoxin A has cytotoxicity against human tumor cell lines [30]. Cancer cell toxicity-guided fractionation of extracts of the marine cyanobacteria L. majuscula and L. sordida led to the isolation of apratoxin D [31]. Marine cyanobacterium L. bouillonii produces apratoxin E, a new peptidepolyketide hybrid of the apratoxin class of cytotoxins[30]. A new 36membered macrolactone, (25S,27S,29S,33S)-caylobolide A, was isolated from L. majuscula. Caylobolide A exhibited in vitro cytotoxicity against human colon tumor cells [32]. Marine cyanobacterium L. bouillonii afforded three hitherto undescribed analogues of the glycosidic macrolide lyngbyaloside, namely, 2-epilyngbyaloside and the regioisomeric 18E- and 18Z lyngbyalosides C. Concurrently we discovered two new analogues of the cytoskeletal actin-disrupting lyngbyabellins, 27-deoxylyngbyabellin A and lyngbyabellin J, a novel macrolide of the laingolide family, laingolide B, and a linear modified peptide, lyngbyapeptin D, along with known lyngbyabellins A and B, lyngbyapeptin A, and lyngbyaloside. The structures contains either brominated or chlorinated consistent with halogenation being a hallmark of many marine natural products. All extracts derived from these L. bouillonii collections were highly cytotoxic due to the presence of apratoxin A or apratoxin C [33]. In continuing investigations of this organism for minor metabolites of related structure, we have isolated and characterized two new natural products, curacins B and C. Both of these are toxic to brine shrimp, demonstrate strong cytotoxicity against murine L-1210 leukemia and human CA46 Burkitt lymphoma cell lines [34]. Two new linear lipopeptides, 1 and 2, and a known compound, curacin D, have been isolated from a marine cyanobacterium, brown L. polychroa. The new compounds were assigned the trivial names dragonamide C and dragonamide D, as their peptide moiety is related to previously reported dragonamides A and B [36]. Bioassay-guided fractionation of the organic extract of a Curaqao collection of L. majuscula led to the isolation of a new lipid, curacin A, with exceptional brine shrimp toxic and antiproliferative activities [37]. A lipophilic extract of an eastern Caribbean collection of L. majuscula yielded two new halogenated fatty acid amides, grenadamides B and C, and two new depsipeptides, itralamides A and B, along with the known compounds hectochlorin and deacetylhectochlorin. The recently reported depsipeptide carriebowmide was also present in the extract and isolated as its sulfone artifact, grenadamides B and C displayed marginal activity against the beet armyworm (Spodoptera exigua), rest others were assessed for general cell toxicity in human embryonic kidney (HEK293) cells, only itralamide B displayed significant cytotoxicity[36]. Investigation of the marine cyanobacterium L. majuscula has led to the isolation of a cyclodepsipeptide, hantupeptin A. Hantupeptin A showed cytotoxicity to MOLT-4 leukemia cells and MCF-7 breast cancer cells [39]. Hantupeptins B and C were isolated, along with the previously reported hantupeptin A, from the marine cyanobacterium, L. majuscula. Hantupeptins B and C showed moderate in vitro cytotoxicity when tested against MOLT-4 (leukemic) and MCF-7 (breast cancer) cell lines [40]. An organic extract of the cyanobacterium L. majuscula, led to the isolation of the known cyclic depsipeptide antanapeptin A. a new bioactive cyclic depsipeptide, homodolastatin 16. Homodolastatin 16, a higher homologue of the potential anticancer agent dolastatin 16, exhibited moderate activity against oesophageal and cervical cancer cell lines [41]. Lyngbyabellin A, a significantly cytotoxic compound with unusual structural features, was isolated from a Guamanian strain of the marine cyanobacterium L. majuscula. This novel peptolide is structurally related to dolabellin. Lyngbyabellin A was found to be a potent disrupter of the cellular microfilament network [42]. Kalkipyrone, a novel R-methoxy- \hat{a}, \hat{a} ¢-dimethyl-c-pyrone possessing an alkyl side chain, was isolated from an assemblage of the marine cyanobacteria L. majuscula and Tolypothrix Kalkipyrone, a novel Rmethoxy-â,â¢-dimethyl-ç-pyrone possessing an alkyl side chain, was isolated from an assemblage of the marine cyanobacteria L. majuscula and Tolypothrix sp. Kalkipyrone is toxic to brine shrimp and gold fish and is structurally related to the actinopyrones that were previously isolated from *Streptomyces* spp. Kalkipyrone is toxic to brine shrimp and gold fish and is structurally related to the actinopyrones that were previously isolated from *Streptomyces* spp[43].

Lagunamides A and B are new cyclic depsipeptides isolated from the marine cyanobacterium *L. majuscula*. Lagunamides A and B displayed significant antimalarial properties, against *Plasmodium falciparum*. Lagunamides A and B also possessed potent cytotoxic activity against P388 murine leukemia cell lines, respectively [44].

A new cyclic depsipeptide Lyngbyabellin B was also isolated from L. majuscula which show toxicity against brine shrimp and fungus Candida albicans [45]. This cyanobacterium also produce series of Lynbyabellin E-I, analogues of Lyngbyabellin B which has same toxicity activity towards human lung tunor and neurobalstoma cell line [46]. L. majuscula also produces Lyngbyastatin 1, a new cytotoxic analogue of dolastatins 12 and 11. These compounds proved toxic with only marginal or no antitumor activity when tested against colon adenocarcinoma or mammary adenocarcinoma [47]. There are lot of cytotoxic compounds isolated from L. *majuscula*, Lyngbyastatin and norlyngbyastatin, are new cytotoxic analogues of dolastatin G and nordolastatin G, respectively[48]. Another but least cytotoxic compound Malyngamide, cocosamide A and B were isolated from lipophilic extract of L. majuscula. These compound show weak cytotoxicity against MCF 7 breast cancer and HT-29 colon cancer cells [49]. There are other compound which show resemblance with malyngamide, malyngamide C, 8-O-acetyl-8-epi-malyngamide C and 8-epi-malyngamide C. these Compounds show moderate cytotoxicity to NCI-H460 human lung tumor and neuro-2a cancer cell lines [50].

CONCLUSION

Thus there are various species of *Lyngbya* found in various environment. Specially the marine strains of *Lyngbya* are more potent to produce multiple of structurally diverse compound/ secondary metabolites which act as diverse. This cyanobacterium not only produce bioactive antibacterial, antifungal but also a good source of anticancerous compounds.

As with the discovery of novel antimicrobial drugs from microorganism, the resistant against this also originated and not the situation become more difficult, those resistant microbes become multidrug resistant. In such situation screening of antimicrobial compounds from the cyanobacteria is become necessary. We have seen that various metabolites isolated from that single genus *Lyngbya* which are very effective. This give the information and excitation to search for novel drugs from that microorganism of similar to that microorganism.

REFERENCES

- Burja AM, Banaigs B, Abou-Mansour E, Burgess JC, Wright PC. Marine cyanobacteria- a prolific source of natural products. Tetrahedron 2001; 57: 9347-9377.
- Kumar M, Tripathi MK, Srivastava A, Gour JK, Singh RK, Tilak R, Asthana RK. Cyanobacteria, *Lyngbya aestuarii* and *Aphanothece bullosa* as antifungal and antileishmanial drug resources. Asian Pacific Journal of Tropical Biomedicine 2013; 3(6): 458-463.
- Kumar M, Tripathi MK, Srivastava A, Nath G, Asthana RK. A Comparative Study of Antibacterial Activity of Brackish and Fresh Water Cyanobacterial Strains. Asian Journal of Experimental Biological Sciences 2012; 3(3):548-552.
- 4. Borowitzka MA. Microalgae as sources of pharmaceuticals and other biologically active Compounds. Journal of Applied Phycology1995; 7: 3-15.
- Edwards DJ, Marquez BL, Nogle LM, McPhail K, Goeger DE, Roberts MA, Gerwick WH. Structure and Biosynthesis of the Jamaicamides, New Mixed Polyketide-Peptide Neurotoxins from the Marine Cyanobacterium Lyngbya majuscule, Chemistry & Biology2004; 11: 817–833.
- 6. Dennison WC, O'Neil JM, Duffy EJ, Oliver PE, Shaw GR. Blooms of the cyanobacterium *Lyngbya majuscule* in

coastal waters of Queensland. Australian Bulletin Institute of Oceanography 1999; 19: 501–506.

- Berry JP, Gantar M, Gawley RE, Rein KS. Isolation of Bioactive Metabolites from a *Lyngbya* Species Isolated from Periphyton of the Florida Everglades, Florida Fish and Wildlife Conservation Commission, Florida Institute of Oceanography, and Intergovernmental Oceanographic Commission of UNESCO 2004; 192-194.
- 8. Gerwick WH, Tan LT, Siachitta N. Nitrogen-containing metabolites from marinecyanobacteria. In The Alkaloids; Academic Press: San Diego, CA, USA 2001; pp. 75–184.
- Ainslie RD, Barchi JJ, Kuniyoshi M, Moore RE. Mynderse, Structure of malyngamide C. Journal of Organic Chemistry 1985; 50: 2859–2862.
- Cardellina J H, Marner F-J, Moore RE. Structure and absolute configuration of malyngolide, an antibiotic from the marine blue-green alga *Lyngbya majuscula* Gomont. Journal of American Chemical Society. 1979; 101: 240– 242.
- 11. Moore RE. In Marine Natural Products: Chemical and Biological Perspectives; Scheuer, P. J., Ed.; Academic Press: New York, 1981; 4: pp 1-52.
- Mynderse JS, Moore RE. Malyngamides D and E, two trans-7-methoxy-9-methylhexadec-4-enamides from a deep water variety of the marine cyanophyte *Lyngbya* majuscula. Journal of Organic Chemistry 1978; 43: 4359– 4363.
- Luesch H, Yoshida WY, Moore RE, Paul VJ, Corbett TH. Total Structure Determination of Apratoxin A, a Potent Novel Cytotoxin from the Marine Cyanobacterium Lyngbya majuscula, Journal of American Chemical Society2001;123: 5418-5423.
- Marner F-J, Moore RE, Hirotsu K, Clardy J. Majusculamides A and B, two epimeric lipodipeptides from *Lyngbya* majuscula Gomont. Journal of Organic Chemistry 1977; 42: 2815–2819.
- Blunt J, Munro M, Upjohn M. The Role of Databases in Marine Natural Products Research. Handbook of Marine Natural Products 2012; pp 389-421
- 16. Bebout BM, Fitzpatrick MW, Paerl HW. Identification of the sources of energy for nitrogen fixation and physiological characterization of nitrogen- ¢ xing members of a marine microbial mat community. Applied Environmental Microbiology 1993; 59: 1495-1503.
- 17. Ekwenye UN, Kazi E. Investigation of plasmid DNA and antibiotic resistance in some pathogenic organism. African Journal of Biotechnology 2007; 6: 877–880.
- Lewis K. Multidrug resistance pumps in bacteria: variations on a theme. Trends in Biochemical Science 1994; 19: 119-123.
- Soria-Mercado IE, Pereira A, Cao Z, Murray TF, Gerwick WH. Alotamide A, a Novel Neuropharmacological Agent from the Marine Cyanobacterium *Lyngbya bouillonii*. Organic letter 2009; 11: 4704-4707.
- Sharp K, Arthur KE, Gu L, Ross C, Harrison G, Gunasekera SP, Meickle T, Matthew S, Luesch H, Thacker RW, Sherman DH, Paul VJ. Phylogenetic and Chemical Diversity of Three Chemotypes of Bloom-Forming Lyngbya Species (Cyanobacteria: Oscillatoriales) from Reefs of Southeastern Florida, Applied and Environmental Microbiology 2009; 2879–2888.
- Moore RE. Cyclic peptides and depsipeptides from cyanobacteria: a review. Journal of Industrial Microbiology1996; 16:134–143.
- Namikoshi M, Rinehart KL. Bioactive compounds produced by cyanobacteria. Journal of Industrial Microbiology 1996; 17:373–384.
- Zainuddin EN, Jansen R, Nimtz M, Wray V, Preisitsch M, Lalk M, Mundt S. *Lyngbya*zothrins A-D, Antimicrobial Cyclic Undecapeptides from the Cultured Cyanobacterium *Lyngbya* sp. Journal of Natural Product 2009; 72: 1373– 1378.
- 24. Cardllina JH, Moore RE. Structure and absolute configuration of Malyngolide, an antibiotic from the

marine blue-green alga *Lyngbya majuscula* Gomont. Journal of Organic Chemistry 1979; 44: 4039-4042

- 25. Berry JP, Gantar M, Gawley RE, Wang M, Rein KS. Pharmacology and toxicology of pahayokolide A, a bioactive metabolite from a freshwater species of *Lyngbya* isolated from the Florida Everglades. Comparative Biochemistry and Physiology 2004; Part C 139: 231–238.
- MacMillan, J. B., Ernst-Russell, M. A., de Ropp, J.S., and Molinski, T. F., Lobocyclamides A-C, Lipopeptides from a Cryptic Cyanobacterial Mat Containing Lyngbya confervoides, J. Org. Chem. 2002, 67, 8210-8215.
- Milligan KE, Marquez BL, Williamson R T, Gerwick WH. Lyngbyabellin B, a Toxic and Antifungal Secondary Metabolite from the Marine Cyanobacterium Lyngbya majuscule. Journal of Natural Product 2000; 63: 1440-1443.
- Kwan JC, Teplitski M, Gunasekera SP, Paul VJ, Luesch H. Isolation and Biological Evaluation of 8-epi-Malyngamide C from the Floridian Marine Cyanobacterium Lyngbya majuscule. Jouranl of Natural Product 2010; 73: 463–466.
- 29. Luesch H, Yoshida WY, Moore RE, Paul VJ, Corbett TH. Total Structure Determination of Apratoxin A, a Potent Novel Cytotoxin from the Marine Cyanobacterium *Lyngbya majuscule*. Joural of Americal Chemical Society 2001; 123: 5418-5423.
- Gutie'rrez M, Suyama TL, Engene N, Wingerd JS, Matainaho T, Gerwick WH. Apratoxin D, a Potent Cytotoxic Cyclodepsipeptide from Papua New Guinea Collections of the Marine Cyanobacteria Lyngbya majuscula and Lyngbya sordid. Jouranl of Natural Product 2008; 71: 1099–1103.
- Matthew S, Schupp PJ, Luesch H. Apratoxin E, a Cytotoxic Peptolide from a Guamanian Collection of the Marine Cyanobacterium *Lyngbya bouillonii*. Jouranl of Natural Product 2008; 71: 1113–1116.
- 32. MacMillan JB, Molinski TF. Caylobolide A, a Unique 36-Membered Macrolactone from a Bahamian *Lyngbya majuscule*. Organic letter 2002; 4: 1535-1538.
- 33. Pereira AR, McCue CF, Gerwick WH. Cyanolide A, a Glycosidic Macrolide with Potent Molluscicidal Activity from the Papua New Guinea Cyanobacterium *Lyngbya bouillonii*. Journal of Natural Product. 2010, *73*, 217–220.
- Matthew S, Salvador LA, Schupp PJ, Paul VJ, Luesch H. Cytotoxic Halogenated Macrolides and Modified Peptides from the Apratoxin-Producing Marine Cyanobacterium Lyngbya bouillonii from Guam, Journal of Natural Product 2010; 73: 1544–1552.
- Yoo H-D, Gerwick WH. Curacin B and C, new antimitotic natural product from the marine cyanobacterium *Lyngbya majuscule*. Journal of Natural Products 1995; 58(12): 1961-1965.
- Gunasekera SP, Ross C, Paul VJ, Matthew S, Luesch H. Dragonamides C and D, Linear Lipopeptides from the Marine Cyanobacterium Brown Lyngbya polychroa. Journal of Natural Product 2008; 71: 887–890.
- 37. Gerwick WH, Proteau PJ, Nagle DG, Hamel E, Blokhin A, Slate DL. Structure of Curacin A, a Novel Antimitotic, Antiproliferative, and Brine Shrimp Toxic Natural Product from the Marine Cyanobacterium Lyngbya majuscula, Journal of Organic Chemistry 1994; 59: 1243-1245.
- Jime'nez JI, Vansach T, Yoshida WY, Sakamoto B, Po'rzgen P, Horgen D. Halogenated Fatty Acid Amides and Cyclic Depsipeptides from an Eastern Caribbean Collection of the Cyanobacterium Lyngbya majuscula, Journal of Natural Product 2009; 72: 1573–1578.
- Tripathi A, Puddick J, Prinsep MR, Lee PPF, Tan LT. Hantupeptin A, a Cytotoxic Cyclic Depsipeptide from a Singapore Collection of *Lyngbya majuscule*. Journal of Natural Product 2009; 72: 29–32.
- Tripathi A, Puddick J, Prinsep MR, Lee PPF, Tan LT. Hantupeptins B and C, cytotoxic cyclodepsipeptides from the marine cyanobacterium *Lyngbya majuscule*. Phytochemistry 2010; 71: 307–311.

- Davies-Coleman MT, Dzeha TM, Gray CA, Hess S, Pannell LK, Hendricks DT, Arendse C E. Isolation of Homodolastatin 16, a New Cyclic Depsipeptide from a Kenyan Collection of *Lyngbya majuscule*. Journal of Natural Product 2003; 66: 712-715.
- 42. Luesch H, Yoshida WY, Moore RE, Paul VJ, Mooberry SL. Isolation, Structure Determination, and Biological Activity of *Lyngbyabellin A* from the Marine Cyanobacterium *Lyngbya majuscula*, Journal of Natural Product 2000; 63: 611-615.
- Graber MA, Gerwick WH. Kalkipyrone, a Toxic *ç*-Pyrone from an Assemblage of the Marine Cyanobacteria *Lyngbya majuscula* and *Tolypothrix* sp. Journal of Natural Product1998; 61: 677-680.
- 44. Tripathi A, Puddick J, Prinsep MR, Rottmann M, Tong L. Lagunamides A and B: Cytotoxic and Antimalarial Cyclodepsipeptides from the Marine Cyanobacterium *Lyngbya majuscule*. Journal of Natural Product 2010; *73:* 1810–1814.
- 45. Milligan KE, Marquez BL, Williamson RT, Gerwick WH. Lyngbyabellin B, a Toxic and Antifungal Secondary Metabolite from the Marine Cyanobacterium Lyngbya majuscule. Journal of Natural Product 2000; 63: 1440-1443.

- Luesch H, Yoshida WY, Harrigan GG, Doom JP, Moore RE, Paul VJ. *Lyngbya*loside B, a New Glycoside Macrolide from a Palauan Marine Cyanobacterium, *Lyngbya* sp. Journal of Natural Product 2002; 65: 1945-1948.
- 47. Harrigan GG, Yoshida WY, Moore RE, Nagle DG, Park PU, Biggs J, Paul VJ, Mooberry S L, Corbett TH, Valeriote FA. Isolation, Structure Determination, and Biological Activity of Dolastatin 12 and Lyngbyastatin 1 from Lyngbya majuscula/Schizothrix calcicola Cyanobacterial Assemblages. Journal of Natural Product1998; 61: 1221-1225.
- Luesch H, Yoshida WY, Moore RE, Paul VJ. Lyngbyastatin 2 and NorLyngbyastatin 2, Analogues of Dolastatin G and Nordolastatin G from the Marine Cyanobacterium Lyngbya majuscule. Journal of Natural Product1999; 62:1702-1706.
- Gunasekera SP, Owle CS, Montaser R, Luesch H, Paul VJ. Malyngamide 3 and Cocosamides A and B from the Marine Cyanobacterium *Lyngbya* majuscula from Cocos Lagoon, Guam. Journal of Natural Product 2011; 74: 871–876.
- 50. Gross H, McPhail KL, Goeger DE, Valeriote FA, Gerwick WH. Two cytotoxic stereoisomers of malyngamide C, 8-epi-malyngamide C and 8-O-acetyl-8-epi-malyngamide C, from the marine cyanobacterium Lyngbya majuscule. Phytochemistry 2010; 71: 1729–1735.