

QUANTITATIVE ANALYSIS OF BIOACTIVE COMPOUNDS FROM *TARGETES ERECTA* (LINN.)DEVIKA R<sup>1,2\*</sup>, JUSTIN KOILPILLAI<sup>3</sup>

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## ABSTRACT

**Objectives:** This study investigates the quantitative analysis of the major phytoconstituents of *Tagetes erecta* (Linn.) and proves their therapeutic value as medicines in both leaf and flower extracts.

**Methods:** The various bioactive compounds were quantitatively analyzed as per standard methods.

**Results:** The present study registered carbohydrates, coumarine, alkaloids, and quinone which are of medicinal importance. During the study, quinone recorded the highest amount of phytocomponents in both the leaf and flower samples.

**Conclusion:** The significant result of the present study revealed that the flower extract exhibited more amounts of phytochemicals than the others, and this could be explored for potential drug development in future.

**Keywords:** Phytoconstituents, Coumarine, Alkaloids, Quinone, Extraction, Solvents.

## INTRODUCTION

Ever since Rig-Veda, various parts of plant sources were used as nutraceuticals, food supplements, traditional medicines, the major constituent in modern medicines and Ayurveda [1,2]. Attention on the use and research of medicinal plant constituents has increased all over the world from traditional to modern and even in the industrial sectors [3-6]. India is known for its vast diversity of plants species and currently about 2500 medicinal plants are used in various sectors of manufacturing industries [7,8] and there are many natural sources yet to be explored [9,10]. It is also evident that over 6000 plant species of India are said to have high therapeutic value, and they represent about 75% of third world countries [11,12]. Recently, because of the reduced risk, the bioactive compounds of plant sources such as *Embelia ribes*, *Phyllanthus emblica*, *Terminalia chebula*, *Eugenia iniflora*, and *Morinda citrifolia*, got focused attention as non-nutrient potentially bioactive compounds [13]. Among the plant secondary metabolites flavonoid constitute the major component, and they are found in all parts [14]. They are classified into six structural categories and contain 5000 different flavonoids [15-18]. *Hibiscus sabdariffa* extract revealed two classes of flavonoids such as flavonols (gossypetin) and anthocyanin [19,20]. The parts of *P. emblica* (Phyllanthaceae) are an important source with high amount of calcium, vitamin A, carotene, thiamine, niacin, minerals, amino acids, etc. [21-23]. *In vitro* evidences with leaf extract of *P. emblica* proved to be anti-neutrophilic and antiplatelet properties [24,25].

The extract of *Albizia lebbbeck* (Shirish) has been proved to be very effective against inflammatory pathologies such as asthma, arthritis, burns [26], antihistaminic [27], and saponin is claimed to be very effective against Alzheimer's and Parkinson's diseases [28]. *E. ribes* dried fruits contains 2,5-dihydroxy-3-undecyl-2-5-cyclohexadiene-1,4-benzo-quinone (Embelin) which forms the major constituents of the commercialized "Vidanga" drug which has wide clinical applications over cancer, diabetes, fertility, etc. from the ancient times [29,30]. *Alternanthera brasiliensis* (Amaranthaceae) contains flavonoids as 3-O-robinobioside derivatives of kaempferol and quercetin which is widely used in Brazilian medicines [31,32]. With this background of investigations, an attempt has been made to investigate the quantitative analysis of flower and leaf extract of *Tagetes erecta* (Linn.). Qualitatively,

the various parts of the target plant were investigated [33,34] and the various constituents were separated by column chromatography [35] and proved that the target plant is of high therapeutic value.

## METHODS

## Estimation of coumarin

1 g of each sample (leaf and flower) was mixed 5 ml of methanol, and the samples were spotted on the thin layer chromatographic plate as per standard method. The spots were observed at 254 nm and the zone were cut and dissolved in 2 ml methanol, and the solution was read at 365 nm and compared with standard solutions [36].

## Estimation of quinone

## Chromatographic system

Liquid chromatographic system is equipped with a 275 nm detector with 5 mm × 15 cm column at the flow rate of 11 minutes retention time at 35°C [37].

## Sample preparation

5 g of samples were mixed with dehydrated alcohol, and the samples were subjected to run in the methanol and dehydrated alcohol mobile phase.

## Estimation of alkaloids

## Chromatographic system

The liquid chromatographic system is equipped with a 275 nm detector with 5 mm × 15 cm column at the flow rate of 1.8 ml/minutes retention time at 35°C [37].

## Sample preparation

5 g of samples were mixed with 150 ml methanol and the samples were subjected to run in potassium phosphate, distilled water and suitable mobile phase.

## Estimation of carbohydrates (Antrone method [38])

The flower and leaf samples were ground with distilled water and the particulate matter were removed by centrifugation followed by filtering

through nylon mesh. A series of standard carbohydrate solutions were prepared, and a standard graph was obtained for calculation. Reading of absorbance was taken at 630 nm. The unknown samples (flower and leaf) were prepared, and the concentration of carbohydrates was determined through standard graph.

## RESULTS AND DISCUSSION

The dried and powered flower and leaf samples of *T. erecta* L. were subjected to quantitative analysis for alkaloids, quinones, coumarins, and carbohydrates as per standard procedures [36-38]. In the present investigation, alkaloid concentration in the flower extract was 3.11 mg/g and 12.3 mg/g in leaf extract. A reddish brown color formation with potassium bismuth iodide indicates the presence of alkaloids in *T. chebula* [1] and these alkaloids serve as antioxidants, antibacterial, antifungal, and antiviral [39]. Various parts (leaf, flower, stem, and seed) of *Mimosa hamate* revealed the presence of alkaloids in both ethanolic and methanolic extracts [9]. Alkaloids are effective against chronic diseases, and they are reducing headaches associated with hypertension [40] and their antimicrobial activity was due to the inhibition of alkaloid was 2.05 mg/g in *E. iniflora* (L.) leaves [11]. During the present investigation with *T. erecta* (L.), the quinone was quantitatively estimated and was found 36.6 mg/g in flower extract and 33.4 mg/g in the leaf extract and also reported that they are vital for good health and has high therapeutic value [41].

The other secondary metabolite which was extracted during the study was coumarine which are used as traces in medicines. The flower extract did not registered but about 2.55 mg/g was recorded in the case of leaf extract of *T. erecta* (L.) They are proved to be useful in pharmaceutical industries as effective nutraceuticals [42]. The presence of carbohydrate was proved quantitatively, and it was found 16.3 µg/g in the flower extract and 11.8 µg/g in the leaf extract. The presence of carbohydrate was proved qualitatively in *M. hamate* [9] and the total carbohydrate recorded in *Eugenia unifera* was 1.14 mg/g [11] and these plants were used as artificial sweetener and also proved to support the body in the rebuilding [43]. The qualitative and quantitative analysis of phytoconstituents of *M. citrifolia* fruit revealed different phytochemicals such as alkaloids, carbohydrates, proteins, phenols, flavonoids, glycosides, saponins, anthraquinones, and tannins and quantitatively the amount of carbohydrate was 11.32 g/100 g which was the highest recorded constituents than other phytochemicals [2]. The plant constituents of various family groups have proved to be highly beneficial and possess significant antimicrobial and are effective antioxidant properties [44-46]. However, further studies are needed to isolate, purify for authentication to be utilized as an industrial drug formulation.

## CONCLUSION

Further research and the accumulation of knowledge on the extraction of individual phytoconstituents and further research validation and characterization with the help of advanced technologies will pave a way for an effective plant derived drugs.

## REFERENCES

- Tariq AL, Reyaz AL. Quantitative phytochemical analysis of traditionally used medicinal plant *Terminalia chebula*. Int Res J Biotechnol 2013;4(5):101-5.
- Sangeetha M, Preethi S. Qualitative and quantitative analysis of bioactive compounds in *Morinda citrifolia* fruit. Indian J Adv Plant Res 2014;16:5-8.
- Ncube NS, Afolayan AJ, Okah AL. Assessment technology of antimicrobial properties of natural compounds of plant origin: Current methods and future trends. Afr J Biotechnol 2008;7(12):1797-806.
- Abu-Rabia A. Urinary diseases and ethnobotany among pastoral nomads in the Middle East. J Ethnobiol Ethnomed 2005;1:4.
- Raveen N, Nayak S, Kar DM, Das P. Pharmacological evaluation of ethanolic extracts of the plant *Altermanthera sessilis* against temperature regulation. J Pharm Res 2010;3(6):1381-3.
- Harborne JB. Phytochemical Methods. London: Chapman and Hall. Ltd.; 2012. p. 49-133.
- Raja A, Gajalakshmi P, Raja MM. Drugs from the natural bio sources for human diseases. Int J Pharm 2010;463:360-3.
- Jasuja ND, Saxena R, Chandra S, Sharma R. Pharmacological characterization and beneficial uses of *Punica granatum*. Asian J Plant Sci 2012;11:251-7.
- Saxena R, Sharma R, Nandy BC, Jasuja ND. Qualitative and quantitative estimation of bioactive compounds in *Mimosa hamata*. Int J Pharm Pharm Sci 2014;6(6):72-5.
- Edeoga W, Taxeira EW, Negri G, Saltino A. Plant origin of green propolis: Plant anatomy and chemistry. Evidence based complementary alternative medicine. Int J Pharm 2005;2:85-92.
- Daniel G, Krishakumari S. Quantitative analysis of primary and secondary metabolites in aqueous hot extract of *Eugenia uniflora* (L) leaves. Asian J Clin Res 2015;8(1):334-8.
- Rajalekharan PE. Herbal medicine. In: World of Science. Employ News 2002;21(27):3.
- Humadi SS, Istudor V. Quantitative analysis of bioactive compound *Hibiscus sabdariffa* L. extracts Note 1: Quantitative analysis of flavonoids. Farmacia 2008;LVI, 6:699-707.
- Steinmetz KA, Potter JD. Vegetables, fruit, and cancer. II. Mechanisms. Cancer Causes Control 1991;2(6):427-42.
- Harborne JB, Williams CA. Advances in flavonoid research since 1992. Phytochemistry 2000;55(6):481-504.
- Bruneton J. Pharmacognosie: Phytochemie, Plantes Medicinal. Paris: TEC and DOC, Lavoisier; 1999. p. 310-7.
- Istudor V. Farmacognozie, Fitochimie, Fitoterpie. Vol. 1. Scientific Basis Boca Raton: Medicala Bucuresti; 1998. p. 148-50.
- Bodea C. Manuals for universities and higher education institutes, vol 2, Washington DC, Educational Academic Bureau, 1965, p.889-13.
- Brachadeo V. Physicians Disk Reference. Montvale: Thomson PDR; 2004. p. 435-6.
- Bisset N, Wichtl M. Herbal Drugs and Phytopharmaceuticals: A Hand Book for Practice on a Scientific Basis. Boca Raton, Ann Arbor, London, Tokyo: CRC Press; 1994. p. 266-7.
- Sukanya MK, Suku S, Aruna SR. Phytochemical analysis, antimicrobial screening and antihelminthic properties of *Phyllanthus emblica*. Int J Pharm Biol Sci 2013;4(4):55-64.
- Srinivasan M. Vitamin C in plants: Indian gooseberry *Phyllanthus emblica*. Nature 1994;153:684-8.
- Dhale DA. Pharmacognostic evaluation of *Phyllanthus emblica* Linn. (Euphorbiaceae). Int J Pharm Biol Sci 2012;3(3):210-7.
- Raghu HS, Ravindra P. Antimicrobial activity and phytochemical study of *Phyllanthus emblica* Linn. J Pharmacol 2010;14:119-28.
- Summanen JO. An anti-inflammatory activity of extracts from leaves of *Phyllanthus emblica*. Plant Med 1997;63:518-24.
- Faisal M, Sing PP, Ircchariya R. Review on *Albizia lebbek* - A potent herbal drug. Int Res J Pharm 2012;3(5):63-8.
- Babu NP, Pandikumar P, Ignacimuthu S. Anti-inflammatory activity of *Albizia lebbek* Benth. an ethnomedicinal plant, in acute and chronic animal models of inflammation. J Ethnopharmacol 2009;125(2):356-60.
- Sanjay K. Saponins of *Albizia lebbek* in Alzheimer's and Parkinson's disease. Int J Nat Prod 2003;19:42-8.
- Souravi K, Rajasekharan PE. A review on the pharmacology of *Embella ribes* Burm F.A threatened medicinal plant. Int J Pharm Biol Sci 2014;5(2):443-56.
- Mitra R. Vidanga (*Embelia ribes*) an Ayurvedic drug can help family planning. Appl Bot Abstr 1995;15(4):267-82.
- Durate MR, Debur MC. Character of the leaf and stem morpho-anatomy of *Altermanthera brasiliiana* (L.) O. Kuntze, Amaranthaceae. Braz J Pharm Sci 2004;40(1):85-92.
- Brochado CO, Almeida AP, Barreto BP, Costa LP, Ribeiro LS, Pereira RL. Flavonol robinobiosides and rutosides from *Altermanthera brasiliiana* (Amaranthaceae) and their effects on lymphocyte proliferation *in vitro*. J Braz Chem Soc 2003;14(3):449-51.
- Devika R, Koilpillai J. Phytochemical screening studies of bioactive compounds of *Tagetes erecta*. Int J Pharm Biol Sci 2012;3(4)(B):596-602.
- Devika R, Koilpillai J. Screening and evaluation of bioactive components of *Tagetes erecta* L. by GC-MS analysis. Asian J Pharm Clin Res 2014;7(2):58-60.
- Devika R, Koilpillai J. Column chromatographic separation of bioactive compounds from *Tagetes erecta* Linn. Int J Pharm Sci Res 2015;6(2):762-6.
- Suresh Kumar CA, Varadharajan R, Muthumani P, Meera R, Devi P, Kameswari B. Pharmacognostic and preliminary investigations on the

- stem of *Saccharum spontaneum*. J Pharm Sci Res 2009;1(3):129-6.
37. Ayoolal GA, Cokerl HA, Adesegun AA, Bellol A, Obaweyal K, Ezennal TO, *et al*. Phytochemical screening and antioxidant activities of some selected medicinal plants used for malaria therapy in Southwestern Nigeria. Trop J Pharm Res 2008;7(3):1019-24.
  38. Hedge JE, Hofreiter BT. In: Carbohydrate Chemistry. 17<sup>th</sup> ed. New York: Academic Press; 1962.
  39. Singh R, Jasrai YT. Mimosa hamate (Wild) aplant with antipathogenic properties. Int J Med Aromat Plants 2012;2:677-83.
  40. Agitey Smith E, Addae Mensah I. Phytochemical, nutritional and medicinal properties of some leafy vegetables consumed by Edo people of Nigeria. West Afr J Pharm Drug Res 1977;4:7-8.
  41. Bonjean K, De Pauw Gillet MC, Defresne MP, Colson P, Houssier C, Dassonneville L. Physicochemical analysis of leaf extract of some tropical medicinal plants. J Ethnopharmacol 1998;69:241-6.
  42. Di Carlo G, Mascolo N, Izzo AA, Capasso F. Flavonoids: Old and new aspects of a class of natural therapeutic drugs. Life Sci 1999;65(4):337-53.
  43. Obdoni BO, Ochuko PO. Phytochemical studies and comparative efficacy of the crude extracts of some homostatic plants in Edo and Delta states of Nigeria. Glob J Pure Appl Sci 2001;8:203-8.
  44. Freeze HH. Disorders in protein glycosylation and potential therapy: Tip of an iceberg? J Pediatr 1998;133(5):593-600.
  45. Prashiith KT, Vihiyaka KS, Soumya KV, Ashwini SK, Kiran R. Antibacterial and antifungal activity of methanolic extract of *Abrus pulchellus wall and Abrus precatorius* Linn. A comparative study. Int J Toxicol Pharmacol Res 2010;2(1):26-9.
  46. Luziatelli G, Sorensen M, Thellade L, Molgaard L, Bhattacharya S, Haldar PK, *et al*. Evaluation of anti-inflammatory activity of *Vernonia cinera* Less extracts in rats. Phytomedicines 2010;6:21-7.