

**MOLECULAR DETECTION OF HUMAN RHINOVIRUS IN RESPIRATORY SAMPLES OF SWINE FLU NEGATIVE NORTH INDIAN CHILDREN WITH FLU-LIKE ILLNESS**

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**ABSTRACT**

**Objectives:** Flu-like illness may also be caused by different respiratory viruses other than influenza. Human rhinovirus (HRV) shows almost flu-like symptoms. The purpose of this study is the molecular detection of HRV in throat swab of swine flu negative North Indian children during the years 2012 and 2013. Reverse transcriptase (RT) - polymerase chain reaction (PCR) amplification of 5' non-coding region (NCR) was used for HRV detection followed by cell culture isolation of HRV.

**Methods:** PCR confirmed swine flu negative throat swab samples were collected from the Department of Microbiology, Sanjay Gandhi Post Graduate Institute of Medical Sciences, Lucknow, Uttar Pradesh, India. The RNA isolation of samples was done using the QIAamp<sup>®</sup>Viral RNA Mini Kit (Qiagen), followed by single step RT-PCR amplification (AgPath-ID, Life Technologies). All PCR positive HRV samples were cell cultured in HeLa and HEp-2 cell lines for viral isolation.

**Results:** 135 swine flu negative throat swab samples were examined. Out of which 34 samples (25.2%) were found HRV positive by RT-PCR, while only four samples (11.8%) were culture positive on HeLa cell line. Younger children (0-4 year) were found more susceptible to HRV infection. This study indicated the highest prevalence of HRV (37.0%) during the months (September-October) of the Autumn season in 2012 and 57% in Winter-spring season (February-March) during 2013.

**Conclusion:** HRV may be a cause of flu-like symptoms in swine flu suspected North Indian children with a higher rate during Autumn and Spring season. Molecular detection of HRV using RT-PCR is more sensitive than cell culture assay.

**Keywords:** Human rhinovirus, Swine flu, Influenza-like illness, Lower respiratory tract infections.

**INTRODUCTION**

Human rhinoviruses (HRVs) association has been seen with severe respiratory illness in upper respiratory as well as lower respiratory tract infections (LRTIs) including influenza-like illness [1]. LRTIs have become the main cause of hospitalization and death of children under 5 years in developing countries [2]. These HRVs are also known as a major cause of common cold among children and immunocompromised patients. Pneumonic and asthmatic children are more susceptible to HRV infection with high mortality rates [3-5]. Symptoms may include a sore throat followed by sneezing, rhinorrhea, nasal obstruction, runny nose, headache, in infants. Sometimes HRV infections lead to muscle weakness, muscle aches, fatigue, malaise, and loss of appetite. Fever is usually common. The complication of HRV infection has been seen with otitis media, sinusitis, chronic bronchitis, and exacerbations of reactive airway disease [6,7]. These HRVs belonging to family Picornaviridae are small, non-enveloped virus with single-stranded, positive-sense RNA genome of 7.2-8.5 KB in length [8]. On the basis of cell-culture and antisera detection, two groups HRV-A, HRV-B, and 101 serotypes are identified. However, recently using polymerase chain reaction (PCR) technique, a different group HRV-C is newly identified concern to many new HRV strains [8,9]. These viruses mainly cause upper respiratory tract infection, but involvement is also found in LRTI [8]. Molecular detection of all prototypes of HRVs has not been completely applied with clinical samples [4,10]. Cell culture for HRV has several limitations as they are typical slow growers and the observation of the cytopathic

effect (CPE) of HRV identification takes several days, resulting in poor sensitivity of cell culture assay [11,12]. More than 100 serotypes are in existence that's why serological diagnosis is not possible [11]. According to several studies, reverse transcriptase (RT)-PCR is more sensitive than cell-culture [8,13-17]. Here, we used a 5' non-coding region (NCR) based RT-PCR method for rapid detection of HRV in clinical samples [5].

In previous studies, HRV is found a potent pathogen which causes up to 5% LRTI in children [18] and found associated with 42% of children with influenza-like illness [19] but the involvement of HRV in flu-like symptoms among swine flu negative infants and young children has not been studied previously in North India. The goal of this study is to analyze the prevalence of HRV in North Indian children found swine flu negative in 2012-2013.

**METHODS****Sample collection**

The PCR confirmed swine flu negative throat swab samples of North Indian children during the year 2012-2013 were collected from the Department of Microbiology, Sanjay Gandhi Post Graduate Institute of Medical Sciences, Lucknow, Uttar Pradesh, India.

**Viral RNA isolation**

The RNA Isolation of samples was done using the QIAamp<sup>®</sup>Viral RNA Mini Kit (Qiagen) as per protocol provided with the kit.

### Molecular detection by RT-PCR

RT-PCR was done by using 5 µl of extracting viral nucleic acid and one-step RT-PCR Kit (AgPath-ID, Life Technologies), as per the manufacturer's instructions. The forward primer 5'-GGGACCAACTACTTTGGGTGTCCG-3' and reverse primer 5'-CACGGACACCCAAAGTAGT-3' were used to amplify a region within the 5'NCR as previously described [6]. The conditions during the single step PCR were as follows- reverse transcription for cDNA synthesis at 50°C (30 minutes), Initial denaturation 95°C (10 minutes), followed by 40 cycles of denaturation at 95°C (30 seconds), annealing at 55°C (30 seconds), and elongation at 72°C (45 seconds). Resulting in amplified fragment, approximately 400 bp in length confirmed in agarose gel electrophoresis.

### Cell culture

Monolayer of HeLa and HEp-2 cells were grown separately in cell culture flasks (Corning), using the minimal essential medium (Sigma-Aldrich) supplemented with Earle's salts, L-glutamine, Penicillin-Streptomycin (Sigma), and 10% Fetal Bovine Serum (Gibco-Life Technologies). Subconfluent HeLa and HEp-2 cell monolayers were washed with Dulbecco's phosphate buffered saline (PBS) (Sigma) three times, and then inoculated with filtered (0.22 µm filter by Millipore), PCR confirmed HRV positive throat swab samples. After 45 minutes at room temperature, the cells were washed with PBS, overlaid with 2% MEM, then moved to an incubator (35°C, 5% CO<sub>2</sub>), under observation for 5 days [20]. The passage was done for confirmation of CPE.

### Statistical analysis

All statistical analysis was done using GraphPad Prism software, Version 5.01. (GraphPad Software, Inc., USA)

### RESULTS

In this study, molecular detection of HRV in swine flu negative samples was done by RT-PCR amplification of 5'NCR region. Among 135 samples, 34 samples (25.2%) were HRV positive (Fig. 1). 55.9% of total HRV positive children were between 0 and 4 year age group. 35.3% children were 5-9 years old while 8.8% HRV positive children were 10-14 years old (Fig. 2). All PCR positive samples were found cell culture negative in HeLa and HEp-2 cell lines in primary inoculation, while only four samples (11.8%) showed CPE in HeLa cell line in the first passage (Table 1 and Fig. 3). The positive rate of HRV was found highest (37%) during Autumn season (September-October) in 2012 and (57%) in Winter-Spring season (February-March) during 2013 (Fig. 4).

### DISCUSSION

In clinical samples, detection of HRV has increased with the improvement of molecular techniques [7,16], providing the evidence of HRV association with LRTI including influenza-like illness [21]. The molecular method based on the 5'NCR has been used previously for rapid detection and typing of all serotypes of HRV [6]. HRVs have been successfully cultured on HeLa cell lines [22]. However, in this study, the cell culture rate of HRV was found very low, and only four samples

(11.8%) were cultured successfully. A recent study showed 26% prevalence of HRV among children under 5 years of age, hospitalized with flu-like symptoms [14,23].

The HRV infection occurs around the year, but during Autumn (September-November) and Spring (March, April) seasons, the incidence of HRV associated infections increased up to 80% with flu-like symptoms [24]. In a previous study, the prevalence of HRV found 15.4% by RT-PCR method during 2000-2006 in Beijing with highest positive results 32.61% in September 2004 and 35.3% in February 2005. Among these HRV positive patients, 44.8% were under the age of 1 year [25]. However, the association of HRV in flu-like symptoms among swine flu negative infants and young children has not been studied previously in North India. In a flu-like illness, clinical specimens are tested for common respiratory viruses such as Influenza A and B, parainfluenza 1, 2, 3, and RSV, while HRV may also play a role in flu-like illness in influenza suspected children.

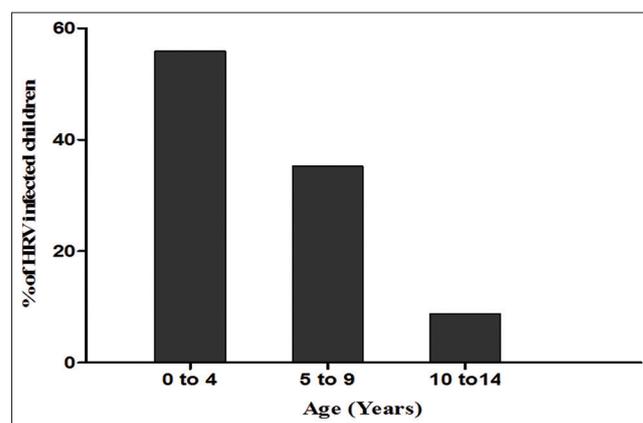


Fig. 2: Percentage of human rhinovirus infected children with respect to their age groups

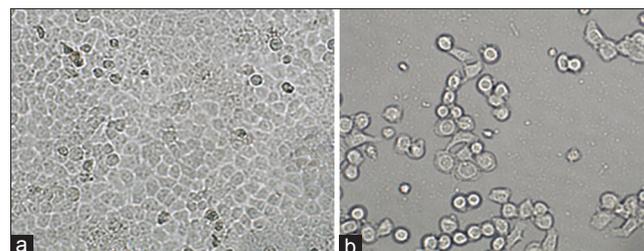


Fig. 3: (a and b) Cell line images (40× magnification). (a): HeLa cell line (Control), (b): Inoculated HeLa cells showing cytopathic effect in the first passage

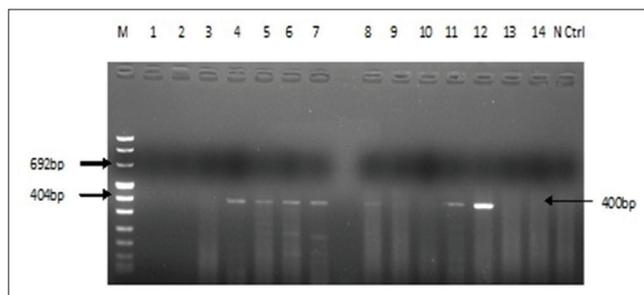


Fig. 1: Agarose gel image showing polymerase chain reaction amplification of the 5' non-coding region (NCR) region of viral RNA. Lane M: Marker 1000 bp. Lane 4 to 7 and Lane 11 to 12: Amplified 5' NCR region of 400 bp

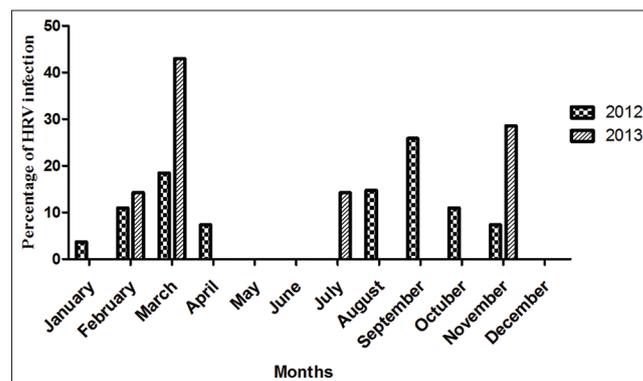


Fig. 4: Percentage of human rhinovirus infection in different months of years 2012-2013

Table 1: Cell culture result of PCR positive HRV samples inoculated in cell lines

Serial number	Cell lines	Total number of PCR positive HRV samples inoculated on cell lines	Cell culture results	
			Primary inoculation	First passage
1	HeLa	34	All negative	Four positive
2	HEp-2	34	All negative	All negative

PCR: Polymerase chain reaction, HRV: Human rhinovirus

## CONCLUSIONS

HRV may be a cause of flu-like symptoms in swine flu suspected North Indian children with a higher rate during Autumn and Spring season due to rapid fluctuation in climate conditions. RT-PCR is rapid and sensitive molecular method for diagnosis of HRV in clinical swab samples. Younger children under the age of 5 years are more susceptible to HRV infection. The cell culture of HRV is not very sensitive for isolation of all HRV strains.

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## REFERENCES

- Hayden FG. Rhinovirus and the lower respiratory tract. *Rev Med Virol* 2004;14(1):17-31.
- Bryce J, Boschi-Pinto C, Shibuya K, Black RE; WHO Child Health Epidemiology Reference Group. WHO estimates of the causes of death in children. *Lancet* 2005;365(9465):1147-52.
- Rawlinson WD, Waliuzzaman Z, Carter IW, Belessis YC, Gilbert KM, Morton JR. Asthma exacerbations in children associated with rhinovirus but not human metapneumovirus infection. *J Infect Dis* 2003;187(8):1314-8.
- Savolainen-Kopra C, Blomqvist S, Kaijalainen S, Jounio U, Juvonen R, Peitso A, et al. All known human rhinovirus species are present in sputum specimens of military recruits during respiratory infection. *Viruses* 2009;1(3):1178-89.
- Lau SK, Yip CC, Tsoi HW, Lee RA, So LY, Lau YL, et al. Clinical features and complete genome characterization of a distinct human rhinovirus (HRV) genetic cluster, probably representing a previously undetected HRV species, HRV-C, associated with acute respiratory illness in children. *J Clin Microbiol* 2007;45(11):3655-64.
- Kiang D, Kalra I, Yagi S, Louie JK, Boushey H, Boothby J, et al. Assay for 5' noncoding region analysis of all human rhinovirus prototype strains. *J Clin Microbiol* 2008;46(11):3736-45.
- Loens K, Ieven M, Ursi D, De Laat C, Sillekens P, Oudshoorn P, et al. Improved detection of rhinoviruses by nucleic acid sequence-based amplification after nucleotide sequence determination of the 5' noncoding regions of additional rhinovirus strains. *J Clin Microbiol* 2003;41(5):1971-6.
- Blomqvist S, Skyttä A, Roivainen M, Hovi T. Rapid detection of human rhinoviruses in nasopharyngeal aspirates by a microwell reverse transcription-PCR-hybridization assay. *J Clin Microbiol* 1999;37(9):2813-6.
- Steininger C, Aberle SW, Popow-Kraupp T. Early detection of acute rhinovirus infections by a rapid reverse transcription-PCR assay. *J Clin Microbiol* 2001;39(1):129-33.
- Lee WM, Kiesner C, Pappas T, Lee I, Grindle K, Jartti T, et al. A diverse group of previously unrecognized human rhinoviruses are common causes of respiratory illnesses in infants. *PLoS One* 2007;2(10):e966.
- Bizzintino J, Lee WM, Laing IA, Vang F, Pappas T, Zhang G, et al. Association between human rhinovirus C and severity of acute asthma in children. *Eur Respir J* 2011;37(5):1037-42.
- Khetsuriani N, Lu X, Teague WG, Kazerouni N, Anderson LJ, Erdman DD. Novel human rhinoviruses and exacerbation of asthma in children. *Emerg Infect Dis* 2008;14(11):1793-6.
- Brownlee JW, Turner RB. New developments in the epidemiology and clinical spectrum of rhinovirus infections. *Curr Opin Pediatr* 2008;20(1):67-71.
- Miller EK, Lu X, Erdman DD, Poehling KA, Zhu Y, Griffin MR, et al. Rhinovirus-associated hospitalizations in young children. *J Infect Dis* 2007;195(6):773-81.
- Savolainen C, Blomqvist S, Hovi T. Human rhinoviruses. *Paediatr Respir Rev* 2003;4(2):91-8.
- Lu X, Holloway B, Dare RK, Kuypers J, Yagi S, Williams JV, et al. Real-time reverse transcription-PCR assay for comprehensive detection of human rhinoviruses. *J Clin Microbiol* 2008;46(2):533-9.
- Loens K, Goossens H, de Laat C, Foolen H, Oudshoorn P, Pattyn S, et al. Detection of rhinoviruses by tissue culture and two independent amplification techniques, nucleic acid sequence-based amplification and reverse transcription-PCR, in children with acute respiratory infections during a winter season. *J Clin Microbiol* 2006;44(1):166-71.
- Huang T, Wang W, Bessaud M, Ren P, Sheng J, Yan H, et al. Evidence of recombination and genetic diversity in human rhinoviruses in children with acute respiratory infection. *PLoS One* 2009;4(7):e6355.
- Renwick N, Schweiger B, Kapoor V, Liu Z, Villari J, Bullmann R, et al. A recently identified rhinovirus genotype is associated with severe respiratory-tract infection in children in Germany. *J Infect Dis* 2007;196(12):1754-60.
- Amineva SP, Aminev AG, Gern JE, Palmenberg AC. Comparison of rhinovirus A infection in human primary epithelial and HeLa cells. *J Gen Virol* 2011;92:2549-57.
- Mackay IM. Human rhinoviruses: The cold wars resume. *J Clin Virol* 2008;42(4):297-320.
- Arruda E, Crump CE, Rollins BS, Ohlin A, Hayden FG. Comparative susceptibilities of human embryonic fibroblasts and HeLa cells for isolation of human rhinoviruses. *J Clin Microbiol* 1996;34(5):1277-9.
- Smuts HE, Workman LJ, Zar HJ. Human rhinovirus infection in young African children with acute wheezing. *BMC Infect Dis* 2011;11:65.
- Arruda E, Pitkäranta A, Witek TJ Jr, Doyle CA, Hayden FG. Frequency and natural history of rhinovirus infections in adults during autumn. *J Clin Microbiol* 1997;35(11):2864-8.
- Zhao LQ, Qian Y, Zhu RN, Deng J, Wang F. Study on the status of human rhinovirus infections in infants and young children with acute respiratory infections in Beijing, from 2002 to 2006. *Zhonghua Liu Xing Bing Xue Za Zhi* 2007;28(7):683-5.