



## MATERIALS AND METHODS

### Materials

Lafutidine powder was purchased from Hangzhou hyper chemicals limited, Zhejiang, China. Tween 80 and tween 20, was purchased from Scharlau S. L Spain. (PVP K-90) was provided by Hangzhou hyper chemicals limited, Zhejiang, China. Dialysis membrane70 provided by HIMEDIA (Mumbai, India). All other chemicals and solvents were of analytical reagent grade, and deionized water also was used in this study.

### Methods

#### Preparation of lafutidine nanosuspension by precipitation method

Nanosuspension precipitation method is used to prepare oral nanosuspension of lafutidine using a different concentration of polymer and surfactant. In brief, 10 mg of lafutidine was dissolved in an organic solvent (1 ml methanol). Deionized water containing the stabilizer (PVP K-90 or tween 80) alone at the drug: stabilizer ratio 1:8 or in the combination of PVP K-90 with tween 80 at the different concentration or in combination with co-surfactant (tween 20), which acts as the antisolvent system. This was followed by the addition of the

organic solution into the antisolvent solution by the help of a syringe pump, under mechanical agitation of different speeds using homo disperser for 60 min at 25±1°C to allow organic solvent to evaporate and get the desired nanosuspension [16]. The batches were prepared according to the formulation design (table 1).

#### Evaluation of the prepared nanosuspension

##### Particle size and size distribution

Particle size determination was done by using Angstrom Advanced Inc. ABT-9000 USA particle size analyzer which is a dynamic light scattering works by measuring the intensity of light scattered by the molecules in the sample as a function of time, at scattering angle 90° and a constant temperature of 25 °C. From the analysis, the average particle size which is also called volume moment mean, (mean diameter in nm) reflects the size of those particles which constitute the bulk of the sample volume and it was measured for all the prepared formulas. The polydispersity index (PDI) which is a measure of the width of the size distribution of each formula of LAF nanosuspension also determined, it is a measure of the distribution of particle size of nanoparticles obtained from a particle analyzer, PDI is an index of spread or variation or width within the particle size distribution. Also, the analyzer determines the specific surface area of each sample [17].

**Table 1: Composition of lafutidine nanosuspension formulation**

Substance	F1	F2	F3	F4	F5	F6	F7	F8	F9	F10
LAF (mg)	10	10	10	10	10	10	10	10	10	10
PVP-K90 (mg)	80	-	50	40	30	50	40	30	50	50
Tween 80 (mg)	-	74.8	28	37.4	46.7	14	18.7	23.4	28	28
Tween 20 (ml)	-	-	-	-	-	13.6	18.2	22.7	-	-
Methanol (ml)	1	1	1	1	1	1	1	1	1	1
Water (ml)	30	30	30	30	30	30	30	30	30	30
Stirring speed (rpm)	1500	1500	1500	1500	1500	1500	1500	1500	500	3000

F: formula, LAF: Lafutidine, PVP K-90: Polyvinylpyrrolidone k-90, rpm: round per min

#### Determination of entrapment efficiency (EE) of nanosuspension

10 ml of nanosuspension was centrifuged at 6000 rpm for 20 min. The supernatant solution was filtered and separated. 1 ml of this filtrate was diluted with water and the absorbance at maximum  $\lambda$  max was measured by UV spectrophotometer using water as blank [18]. The amount of free drug in the formulations was measured and the entrapment efficiency is then calculated from Eq.1

$$E. E\% = \frac{(\text{Total drug in formula} - \text{free drug}) \times 100}{\text{Total drug in formula}} \dots\dots (1)$$

The results were analyzed in triplicate and standard deviations are reported.

#### In vitro dissolution profile of nanosuspension

In vitro drug release for nanosuspension was done by using Himedia dialysis membrane (MWCO 12 KD). Volume containing 10 mg of lafutidine of (F3-F10) nanosuspension was placed in the pretreated dialysis bag and drug release was done using USP dissolution apparatus II containing 900 ml of dissolution medium at 37 ± 0.5 °C. The speed of the paddle was 100 rpm. The formulation of lafutidine nanosuspension was subjected to the drug release studies in media of 0.1N HCl (pH 1.2) in comparison with a pure drug. Samples (5 ml) were withdrawn at intervals of 5, 10, 15, 20, 30, 40, 50, 60, 70, 80, 100, 110 and 120 min respectively), and replenished with the equivalent volume of fresh dissolution medium to maintain the constant volume. Then, samples were filtered and assayed spectrophotometrically on UV spectrophotometer at 286 nm wavelength. The experiment repeated in triplicate for each formulation [19].

#### Zeta potential

Zeta potential of the selected formulation of lafutidine nanosuspension was measured by using (NanoSeries, Nano-ZS, Malvern Instruments, UK). A zeta potential value of ±30mV is

required as a minimum for physically stable nanosuspension stabilized by electrostatic repulsion only. While zeta potential of about ±20 mV is sufficient to of high-intensity the nanosuspension system stabilized by a combination of steric and electrostatic stabilization [20].

#### The freeze-drying of selected formula

Freeze drying or lyophilization was carried out using christ (ALPHA 1-4 LD plus) to retrieve nanoparticles in a dried-powder state from the nanosuspensions, to complete characterization of nanosuspension and show the effect of freeze drying on nanoparticles size and solubility so water-removal was conducted through freeze-drying. The selected formula was used containing 2% w/w mannitol as the cryoprotectant, to prevent agglomeration was frozen in a refrigerator at -70 °C for 24 h. Then the sample was lyophilized using vacuum freeze dryer at a controlled temperature of (-44) °C and the pump operating at the pressure of 2.5 × 10 pascals over the range of 48–72 hr. The obtained powder was used for further studies [21].

#### Fourier transforms infrared spectroscopy (FTIR)

The fourier transforms infrared spectroscopy spectra of pure lafutidine and lyophilized powder of the selected formula were obtained using FTIR spectrophotometer (FTIR-8300 Shimadzu, Japan) by potassium bromide (KBr) pellet method. This study was achieved to identify any sign of interaction between the drug and stabilizer used. The spectrum obtained was in between the wave number of 4000-400 cm<sup>-1</sup>[22].

#### Scanning electron microscopy (SEM)

Scanning electron microscope used for observing the morphology of samples, SEM of pure lafutidine, and lyophilized powder was confirmed by direct deposition of powder on double-sided carbon tape and coated with gold; the analysis of the particle size was

performed using the UTHSCSA Image Tool software. Five to six high resolution and high magnification pictures of SEM representative of the sample were used to find the particle size. The program was used to measure the average size of the particles seen in each of the SEM picture [23].

#### X-ray powder diffraction

Powder x-ray diffraction can be used to confirm the crystalline nature of materials. So, this information is used to verify whether the substances are crystalline or amorphous. The diffractograms of lafutidine and lyophilized powders of the selected formulation were obtained for analysis. The study was confirmed by using Shimadzu XRD-6000 powder X-ray diffractometer at continuous scan range of 10-80 degree. The operating voltage was 40 (kV) and current 30mA [24].

#### Statistical analysis

The experimental results are given as mean triplicate samples standard deviation (SD) and were analyzed according to one-way analysis of variance (ANOVA) using Sigma Plot 11 software at which the results would be significant if  $p < 0.05$ , and the results would be nonsignificant if  $p > 0.05$ .

#### RESULTS AND DISCUSSION

##### Particle size analysis and polydispersity index measurement

The effect of different parameters on the particle size and polydispersity index was studied using ten different formulations. The mean particle size (effective diameter) for formulations varied in the wide range from  $6.358 \pm 0.0$  nm to  $1188 \pm 1.7$  nm. The particle size and PDI for different formulations of different parameters is showing in table 2.

Table 2: The particle size, and PDI of different formulations

Formula no.	P. S. $\pm$ SD*	PDI $\pm$ SD*
F1	1188 $\pm$ 1.7	0.324 $\pm$ 0.0
F2	805.2 $\pm$ 1.7	0.489 $\pm$ 0.0
F3	22.9 $\pm$ 0.0	0.048 $\pm$ 0.0
F4	73.0 $\pm$ 11.9	0.893 $\pm$ 0.0
F5	63.55 $\pm$ 0.0	0.278 $\pm$ 0.0
F6	19.08 $\pm$ 0.0	0.405 $\pm$ 0.0
F7	15.89 $\pm$ 1.8	1.0 $\pm$ 0.0
F8	6.358 $\pm$ 0.0	0.196 $\pm$ 0.0
F9	36.14 $\pm$ 0.0	0.481 $\pm$ 0.0
F10	59.5 $\pm$ 0.0	0.703 $\pm$ 0.0

\*SD standard deviation, n=3, P. S.: Particle size, PDI: Polydispersity index

#### Effect of stabilizer type on the particle size and polydispersity index

Two formulations were used to show this effect F1, F2. The best stabilizer was in F2 (tween 80) which has particle size  $805.2 \pm 1.7$  nm. Also, this formulation show PDI in the value of 0.489 and this low value will indicate good stability of the nanosuspension. The effect of stabilizer type shown in the fig. 2. The choice of suitable stabilizers are the most important factors to control the size and stability of the nanosuspension during nanoprecipitation methods [25]. Although; the mechanism of action of PVP K-90 which was used as hydrophilic polymers to stabilize nanosuspensions physically by steric stabilization; but they gave larger particle in ratios 1:8 drug: stabilizer in F1 ( $p < 0.05$ ) than particle size of F2 which contain tween 80, this may be attributed to the insufficient affinity of stabilizers to drug molecule. However, if there is no affinity between the particle surface and the polymer, the attractive forces between two particles become dominant due to depletion of polymer from the gap of two particles (depletion force) [26].

#### Effect of combination of surfactant with polymer on particle size and polydispersity index

F3 was used to show the effect of combination of tween 80 (used as surfactant) with PVP K-90 (used as polymer) as compared with F1 and F2 that used these stabilizers alone (fig. 3). When PVP K-90 used as a primary stabilizer in F1, could not significantly decrease the particle size ( $p > 0.05$ ) to nanosized particles; therefore, it could not be used as the single stabilizer for the nanosuspension. The use of tween 80 combined with another stabilizer (PVP K-90) as in fig. 3 may disperse drug particle into the aqueous solution and decrease the particle size [27]. The combination of surfactant (tween80) with PVP K-90 provide the electrostatic-steric mechanism, surfactant found to influence the particle size which helps in preventing growth and stabilize the particles primarily by adsorption at the solid-liquid interface and reduction of the interfacial tension leading to an increased rate of nucleation [28].

The most significant effect ( $p < 0.05$ ) of surfactant combination with one polymer was shown in formulas that contain tween 80 or PVP-K90 as primary stabilizer, as in F1 and F2 respectively at ratio 1:8 (drug: primary stabilizer), the particle size was 1188 nm and 805.2

nm respectively, while in F3 in which tween 80 combined with PVP K-90 at drug: polymer: surfactant ratio 1:5:3 the particle sizes were 22.9 nm. That means the combination has the good surface affinity and could form a substantial mechanical and thermodynamic barrier at the interface of drug molecule [29]. Polydispersity index values of F3 was 0.048, indicate that these formulas are monodisperse standard, there are significant differences ( $p < 0.05$ ) from PDI values of F1 and F2 that were 0.324-0.489 indicate that these formulas are middispersed standard.

#### Effect of stabilizer concentration on the particle size and polydispersity index

Three formulations were used to show this effect F3, F4, and F5. In fig. 4 the particle size of F5 containing PVP K-90 and tween 80 at drug: polymer: surfactant ratio 1:5:3 was increased significantly ( $p < 0.05$ ) from 22.9 nm to 73 nm in F4 when the surfactant concentration raised to drug: polymer: surfactant ratio 1:4:4 and then the particle size decrease significantly ( $p < 0.05$ ) to 5.248 nm in F5.

It was observed that with an increase in surfactant concentration in the nanosuspension from the particle size of the nanosuspension decreases. This was due to the decrease in relative viscosity, which led to decrease in particle size. It means that hydrodynamic diameter of particle decreased with increase in the concentration of the surfactant. The concentration of surfactant affected on particle size because too little concentration of stabilizer induces agglomeration or aggregation and too much concentration promotes Ostwald ripening [30].

#### Effect of addition co-surfactant on particle size and polydispersity index

The addition of **co-surfactant** is also preferred for long-term stabilization and decrease particle size significantly ( $p < 0.05$ ) as in fig. 5, F6-F8 studied the effect of the combination of surfactants (tween 80 and tween 20) on the particle size that range 6.358-19.08 nm which smaller than F3, F4 and F5 that contain one surfactant (tween 80), these findings are in accordance with another study whose particle sizes was lower and better stability when stabilized the formulation with surfactant mixtures compared with

formulations with only one surfactant [31]. PDI of F8 and F6 were 0.196, and 0.405 respectively this show that these formulas were dispersed while F7 was polydisperse because it had PDI equal to 1.0.

**Effect of stirring speed on the particle size and polydispersity index of prepared nanosuspension**

Three different speeds 500, 1500 and 3000 rpm were used to prepare three formulations F9, F3, and F10 to show this effect as in fig. 6. In this study the optimum speed at the drug: polymer: surfactant ratio 1:5:3 was found to be 1500 rpm that produce mean particle size 22.9±0.0 nm. PDI of these formulations was in the range of 0.048-0.703. The result suggested that lafutidine nanoparticles were nonsignificantly affected ( $p>0.05$ ) by stirring speed, the decreased particle size with increased stirring speed is due to an intensification of micromixing (i. e, mixing of molecular level) between multi-phases. High micromixing efficiency increased mass transfer and the rate of diffusion between the multi-phases which induced high homogeneous supersaturation in short time, thus rapid nucleation to produced smaller drug particle. In addition to that, a high stirring speed prevents the particle growth by preventing their aggregation. Hence the formation of smaller and more uniform drug particles with high stirring speed [32].

However, further increase in the stirring speed led to increasing in the particle size ( $p>0.05$ ) of nanosuspension during precipitation method that resulted in non-uniform formulations with visible settling due to the aggregation of particles, this could be due to mechanical agitation and non-uniform mixing producing local regions of supersaturation during anti-solvent precipitation [33].

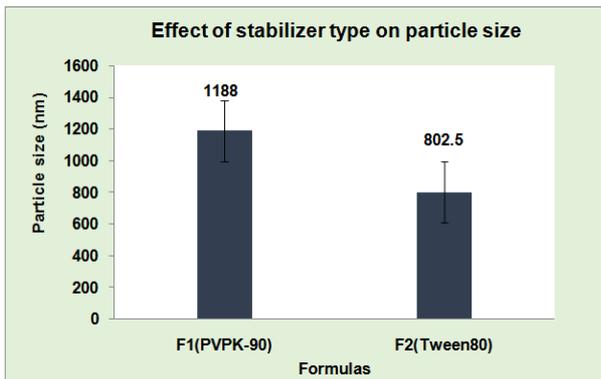


Fig. 2: Effect of stabilizer type on the particle size of nanosuspension (Results as an as mean, n=3)

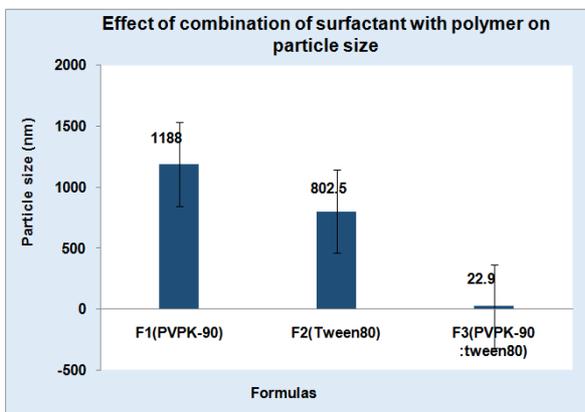


Fig. 3: Effect of a combination of the surfactant with polymer on the particle size of nanosuspension (n=3)

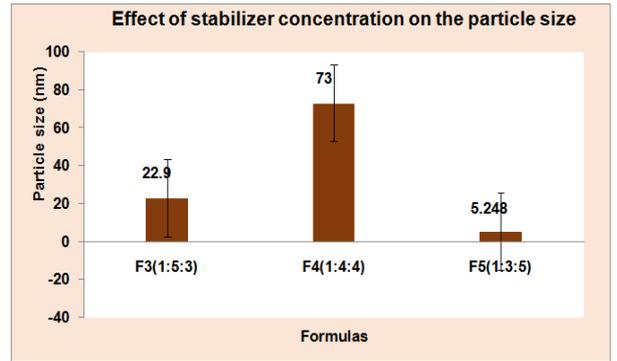


Fig. 4: Effect of stabilizer concentration on the particle size of nanosuspension (Results are expressed as mean, n=3)

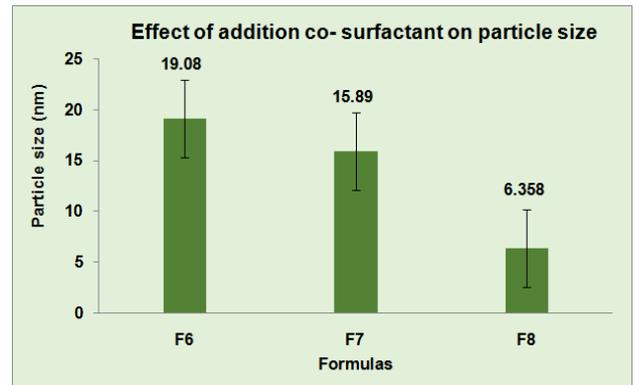


Fig. 5: Effect of addition of co-surfactant on particle size (n=3)

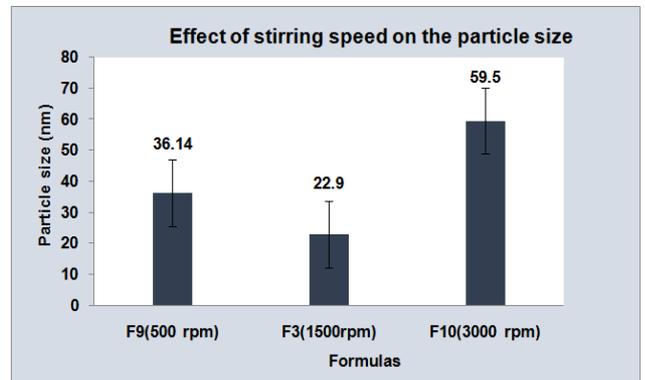


Fig. 6: Effect of addition of stirring speed on particle size (Results are expressed as mean, n=3)

**Drug entrapment efficiency**

The Percentage drug entrapment efficiency of the formulations was calculated and the results were 95.604%, 98.54%, 96.34%, 98.43%, 98.74%, 98.54, 94.56% and 97.5 for F3, F4, F5, F6 F7, F8, F9 and F10 respectively. The drug entrapment efficiency of F7 was high when compared to other formulations. Drug entrapment efficiency was significantly affected by the addition of non-ionic surfactants (combination of tween80 with tween 20) with ratio of drug: polymer: surfactant (1:4:4) with higher EE% (98.74%) in F7 (fig. 8) which could be due to decreased partitioning of LAF into the outer aqueous phase and better dispersion obtained by adding a hydrophilic surfactants [34].

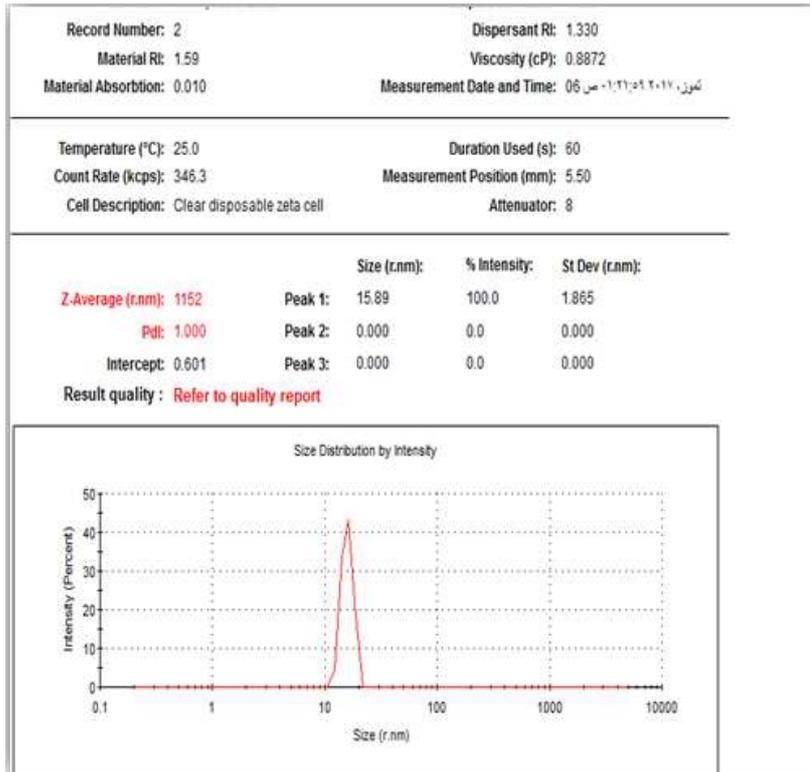


Fig. 7: Particle size distribution of the selected formula (F7)

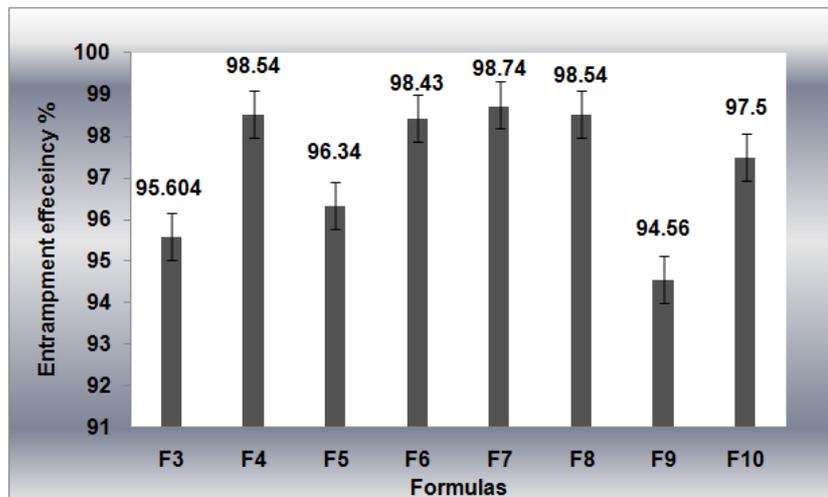


Fig. 8: Drug entrapment efficiency of the prepared formulation of lafutidine nanosuspension (Results are expressed as mean, n=3)

**In vitro dissolution study**

The dissolution profile of the formulas that in nanosize (F3-F10) were studied in 0.1N HCl of pH 1.2 to determine the best formula that gives the best release in first 10 min. The release of F3-F10 is presented 0.1N HCl of pH 1.2 in fig. (9). From the study, the results showed that the formula F7 that contain PVP K-90, tween 80 and tween 20 stabilizers gave the best release in 10 min in comparison with other formulas and the formula shows a maximum cumulative percentage drug release of 100 % within 10 min.

From the above result depending on the particle size and PDI will be the formula no. F7 which has the mean particle size (15.89±0.0 nm) and E. E% (98.74). F7 was considered as the selected formula because it had the highest dissolution rate, highest entrapment efficiency percentage and low particle size when compared with

another formula. This formula is selected for zeta potential measurement study, lyophilization, and further study.

**Zeta potential**

The zeta potential for the selected formulation of lafutidine nanosuspension was -23 mV as shown in fig. 11. The charge was negative due to adsorbed tween 80, tween 20 and PVP K-90 on the drug particles; however, this result of zeta potential proposes that the nanosuspension was adequately stabilized. It reflects the electrical potential of particles and is influenced by the composition of the particles and the medium in which it is dispersed. Zeta potential gives certain information about the surface charge properties and furthers the long-term physical stability of the nanosuspension. The obtained value for selected formulation indicates stable nanosuspension [35].

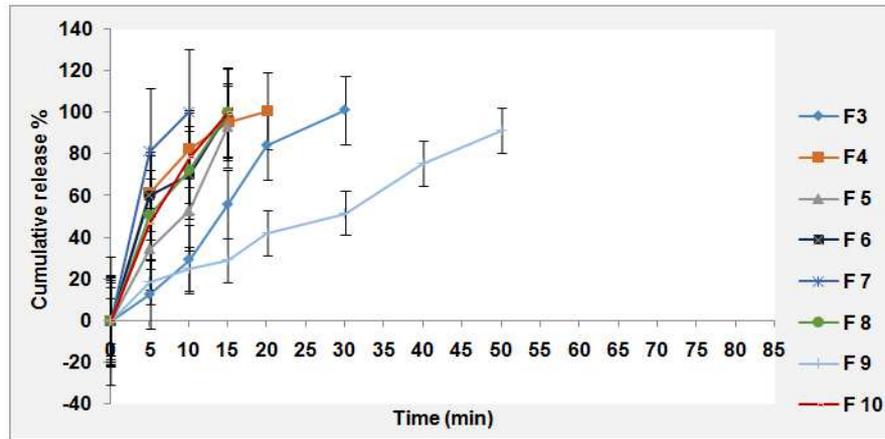


Fig. 9: *In vitro* drug release profile of lafutidine formulation nanosuspension in 0.1N HCl at 37±0.5 °C (n=3)

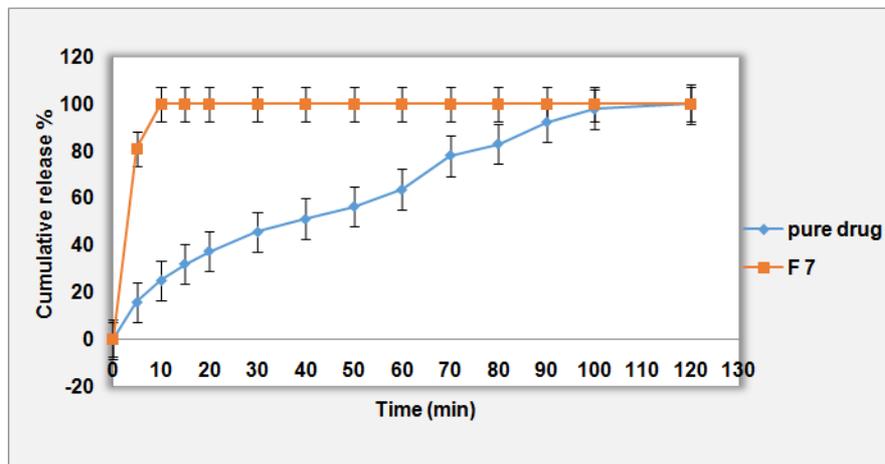


Fig. 10: *In vitro* drug release profile of selected formula (F7) compared with pure powder lafutidine in 0.1N HCl at 37±0.5 °C (Results are expressed as mean, n=3)

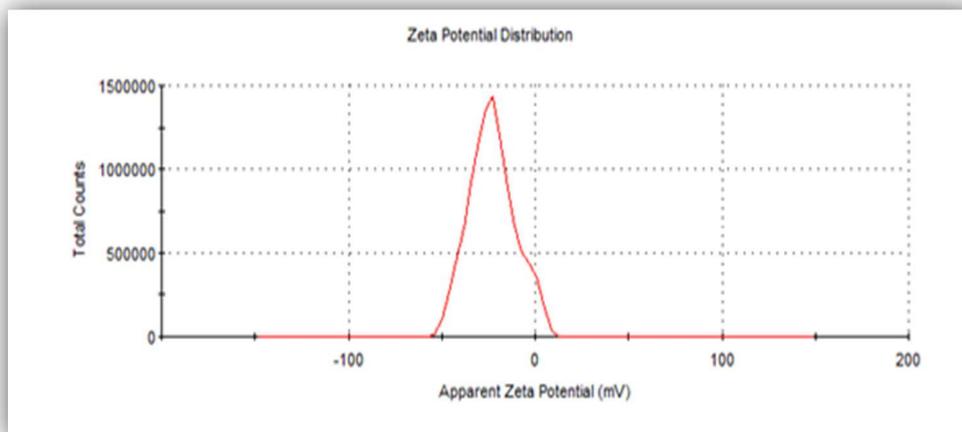


Fig. 11: Zeta potential of the selected formula (F7)

**Saturation solubility of freeze-drying nanosuspension**

The batch F7 (lafutidine: PVP K-90: tween 80: tween 20 1:4:2:2) was selected for freeze drying. In this study, 2% of mannitol was added as the cryoprotectant for avoiding nanocrystals aggregation/agglomeration. Mannitol containing samples showed good re-dispersibility upon manual shaking. The results of the solubility

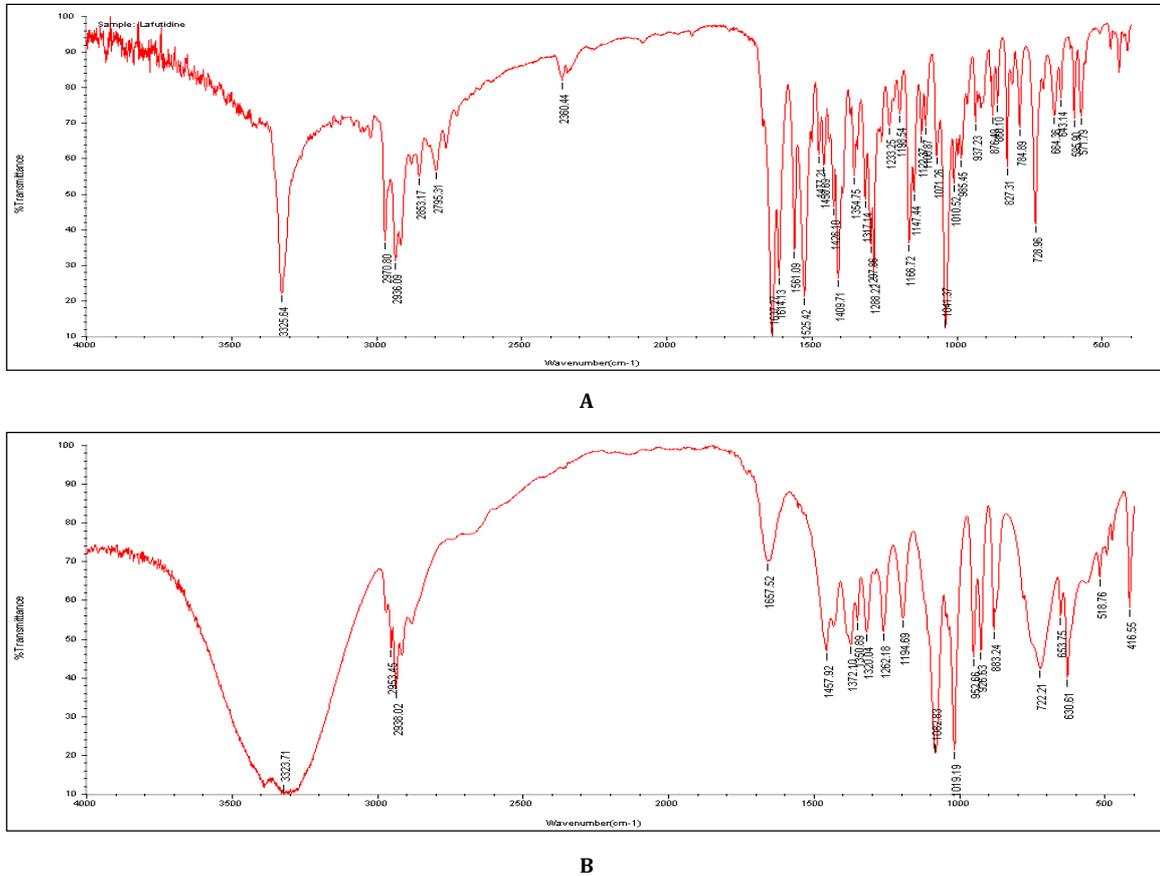
study indicated that pure lafutidine had a very low solubility in water (0.078 mg/ml) which considered as practically insoluble in water, while the saturation solubility of lafutidine nanoparticle in lyophilized powder was (125.5 mg/ml) greater than that of pure drug which considered as freely soluble in water. This may be due to the reduction in particle size of lafutidine to nano size which results in exposure of more surface area and enhanced hydrophilicity; these

were responsible for the significantly enhanced ( $p < 0.05$ ) saturation solubility [36].

**Fourier transforms infrared spectroscopy**

The Fourier transforms infrared spectra of pure lafutidine shown in fig. 12 (A) and that of the lyophilized powder of the selected formula

(F7) intensity in fig. 12 (B). FTIR spectra of lafutidine nanosuspension show no change in shifting the position of the lafutidine nanosuspension functional groupspeak and this will indicate there was no an interaction between the drug and the stabilizer PVP K-90 and other excipients (tween80 and tween 20) used in the formulation [37].



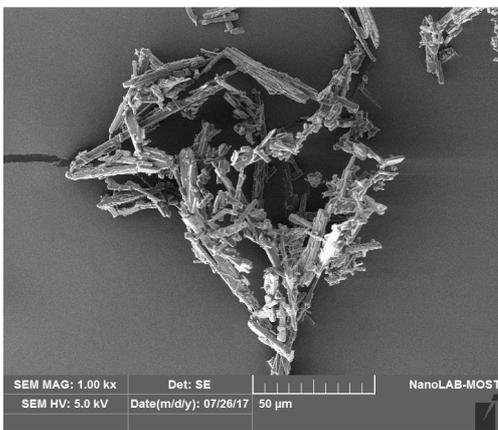
**Fig. 12: Fourier transforms infrared spectroscopy of A) lafutidine and B) lyophilized powder**

**Scanning electron microscope**

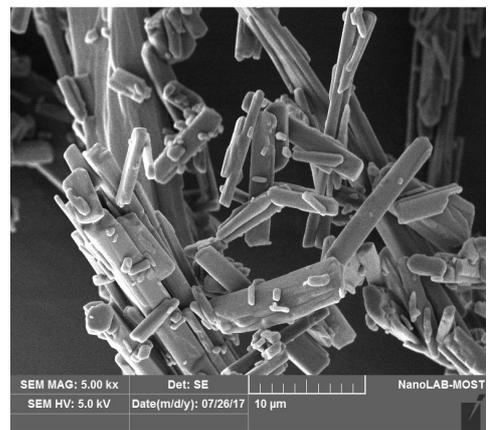
Scanning electron microscope of pure lafutidine, PVP K-90 and lyophilized powder are presented in fig. (13, 14, and 15) at 1K, 5K, 10K, 50K and 500x (this only for PVP K-90) magnification. The scanning electron microscope study for pure lafutidine powder showed large particles with crystalline needle shape with the smooth surface, while scanning electron microscope of PVP K-90

elucidated the irregular shape with the rough surface of PVP K-90 particle because it was the amorphous polymer [38].

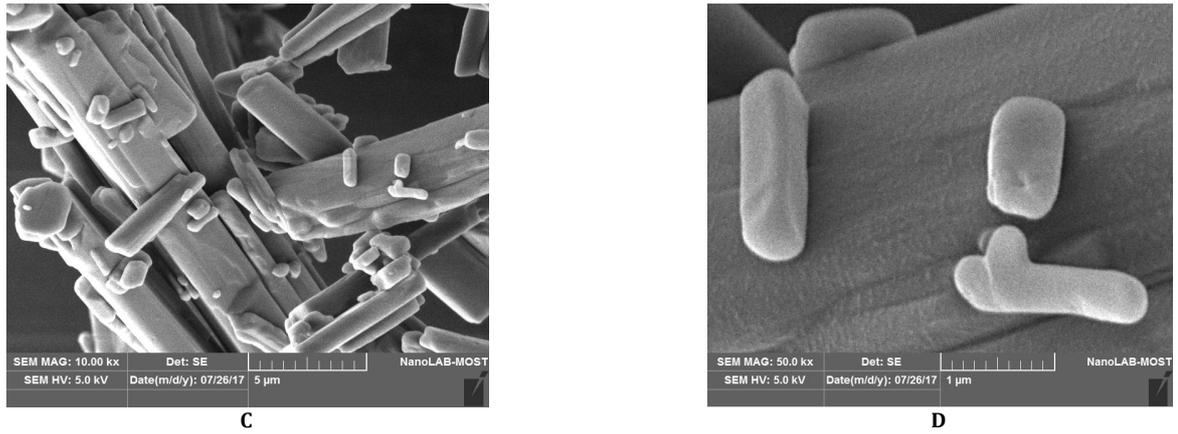
Scanning electron microscope of lyophilized powder at 1K, 5K, 10K and 50K of magnification revealed that the decreased the particle size with some of the crystalline needle structure of drug this problem may mean that by the addition more stabilizer and more cryoprotectant.



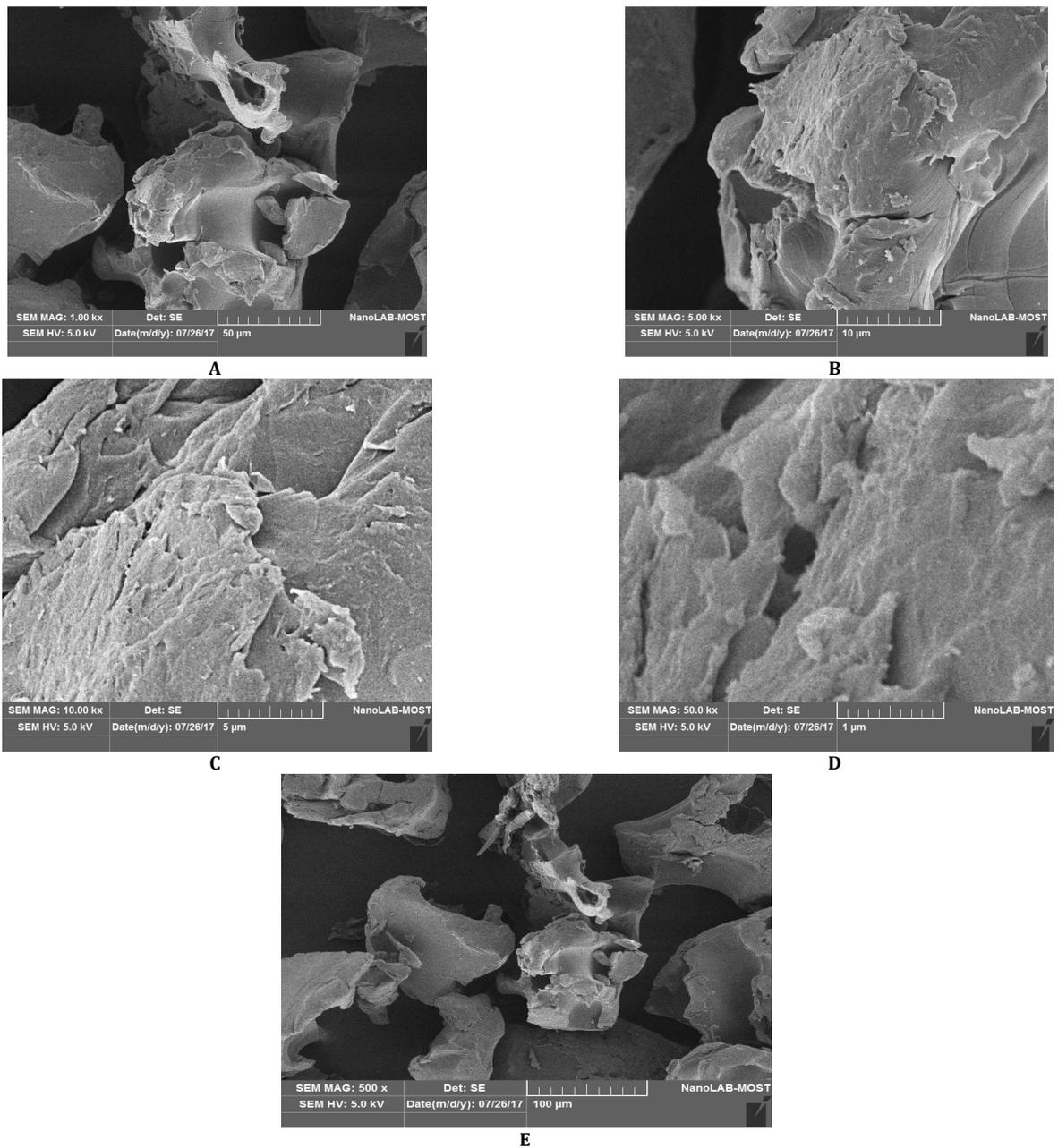
**A**



**B**



**Fig. 13: Scanning electron microscope of pure lafutidine (A, B, C, and D) at 1K, 5K, 10K and 50K magnification respectively**



**Fig. 14: Scanning electron microscope of PVP K-90 (A, B, C, D, and E) at 1K, 5K, 10K, 50K and 500X magnification respectively**

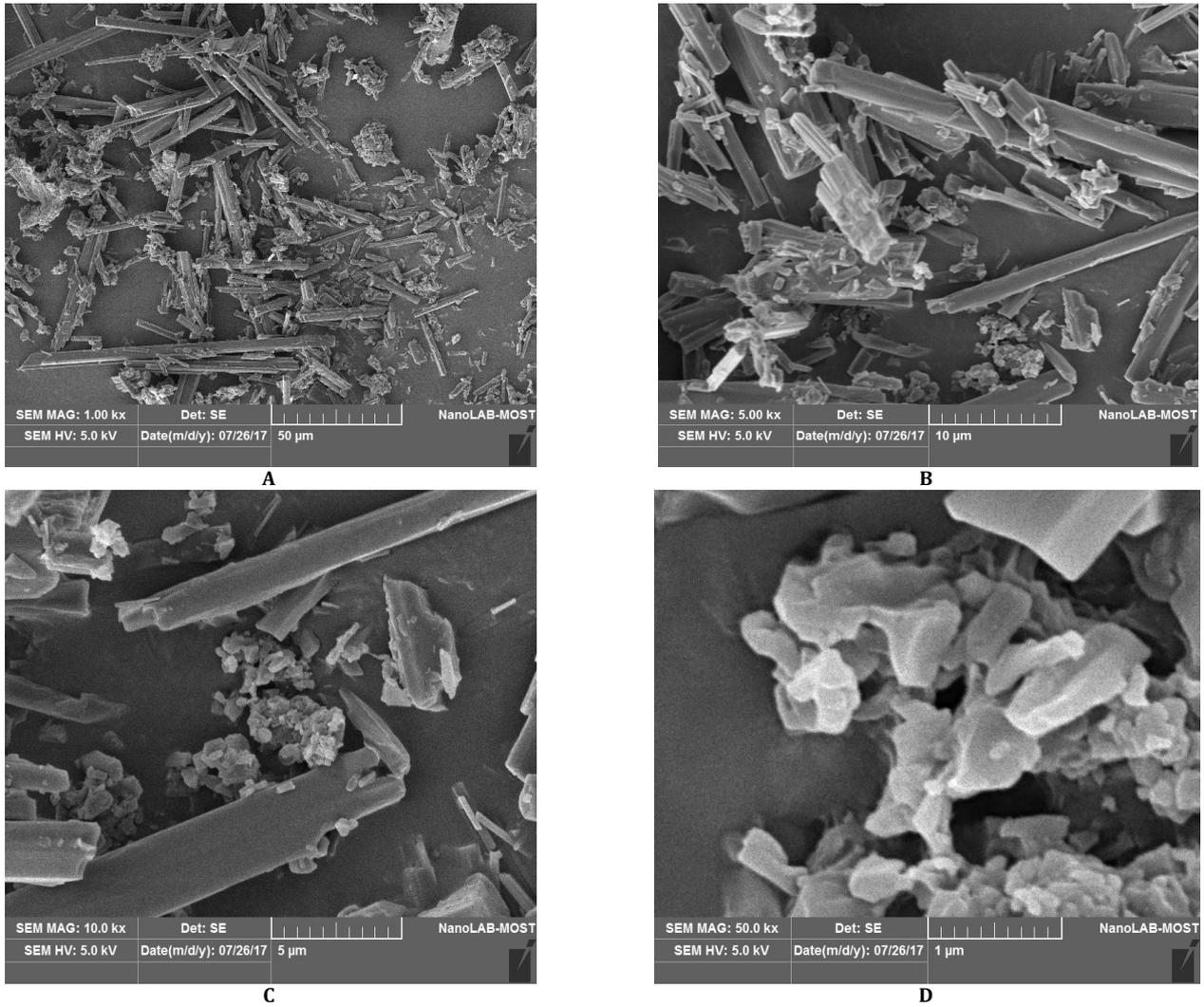


Fig. 15: Scanning electron microscope of lyophilized powder (A, B, C, and D) at 1K, 5K, 10K and 50K magnification respectively

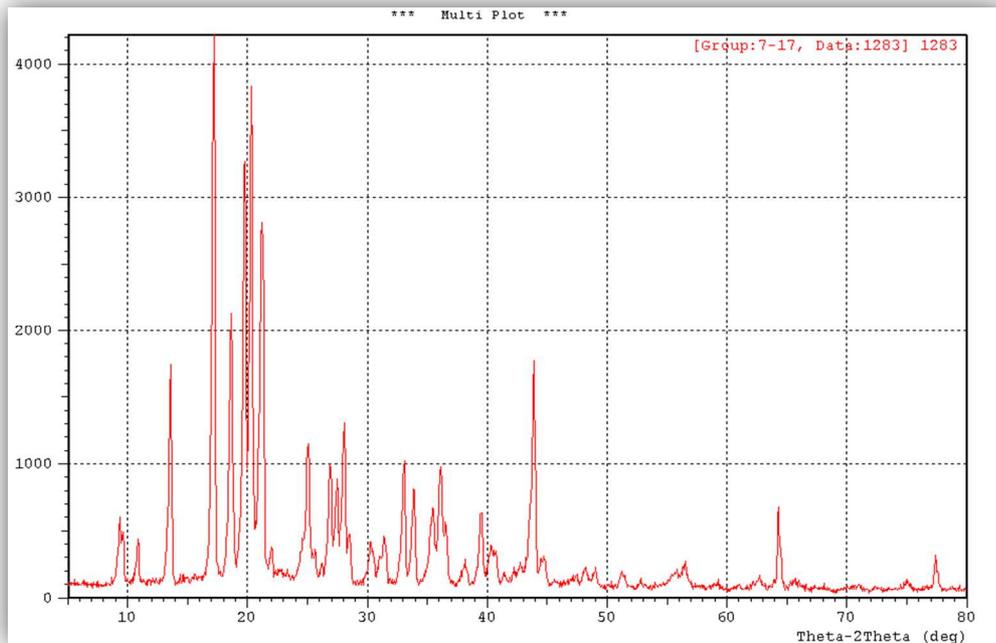


Fig. 16: Powder X-ray diffraction analysis of pure lafutidine

### Powder x-ray diffraction analysis (PXRD)

To confirm the physical state of lafutidine, PVP K-90 and lyophilized powder x-ray diffraction were performed. The Powder X-ray diffraction analysis patterns of lafutidine as a pure drug exhibited intense crystalline peaks at the  $2\theta$  angle as in fig. 16, the strongest three peaks were  $17.2^\circ$ ,  $19.8^\circ$ , and  $20.3^\circ$ . The diffraction patterns of PVP K-90 demonstrated in fig. 17. PVPK-90 indicated the halo pattern was revealing amorphous nature of polymer as in it's diffractogram and characterized by low intense peak. However, the strongest peaks of PVP K-90 at the  $2\theta$  angle were  $11.29^\circ$ ,  $43.9^\circ$  and  $64.29^\circ$ . The diffused peaks at higher angles confirm the semi-crystalline nature of

the polymer [39]. However, the characteristic crystalline peak at the  $2\theta$  angle of  $20.3^\circ$  became lower intensity peak, and strongest peaks of  $17.2^\circ$ ,  $19.8^\circ$  disappeared in the pattern of lyophilized powder as in fig. 18 producing another peak at the  $2\theta$  angle of  $5.69^\circ$  and  $20.6^\circ$ . This study suggested that there was no significant difference in characteristic peaks position of lafutidine in the lyophilized powder. Accordingly, all types of energy input during preparation and the attendance of stabilizer did not change the crystallinity of lyophilized nanosuspension powder. A just difference detected between the coarse powder, and lafutidine nanosuspension was in the peak intensity and presented of high-intensity peaks, probably attributed to particle size reduction, stabilizer at the surface [40].

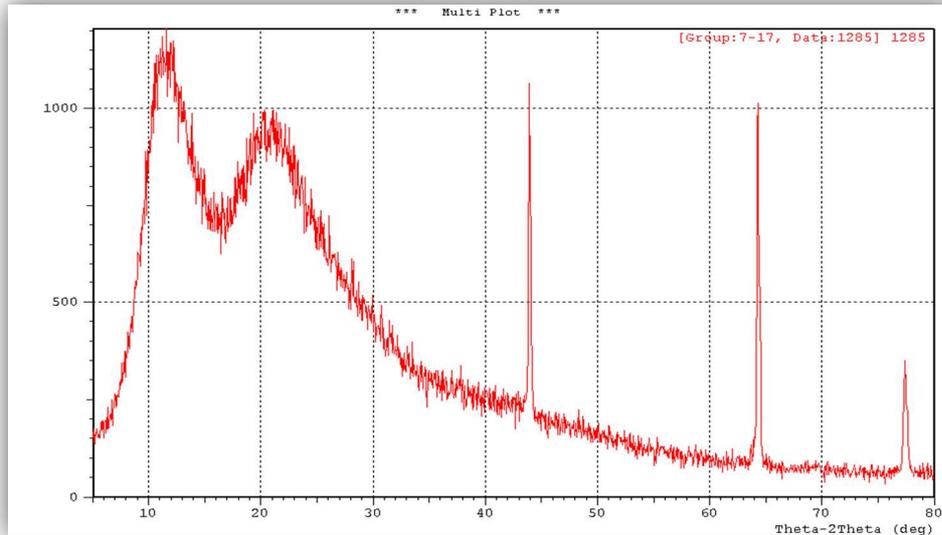


Fig. 17: Powder x-ray diffraction analysis of the PVP K-90

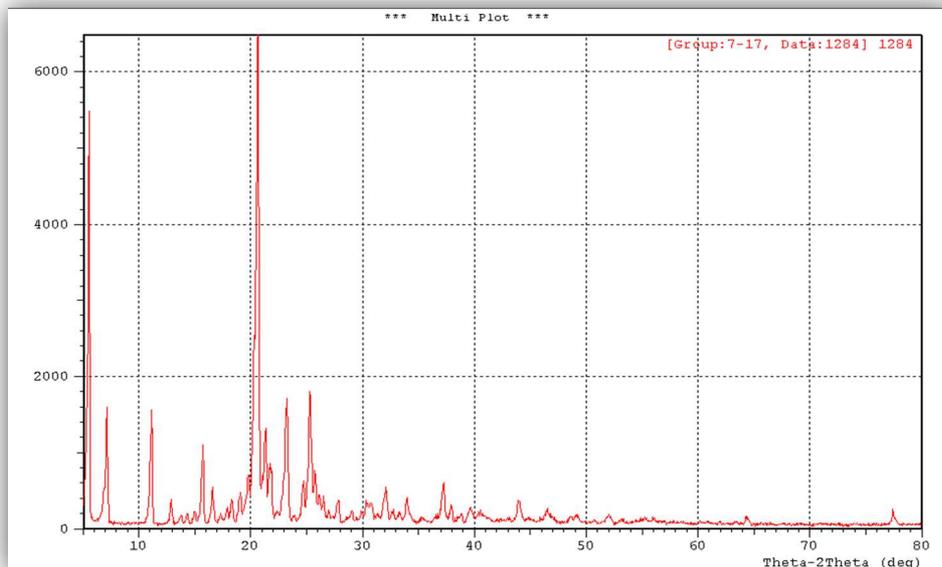


Fig. 18: Powder X-ray diffraction analysis of the selected formula F7

### CONCLUSION

It may be concluded from the results of this study that nanosuspensions of particle water-insoluble drug lafutidine can be prepared using solvent anti-solvent precipitation method and using PVP K90, tween 80, and tween 20 as stabilizers.

The process parameters, such as stabilizer type, the combination of the surfactant with the polymer, stabilizer concentration, stirring speed and combination of other stabilizer were investigated and optimized to produce the smallest drug nanoparticles. The dissolution rate of the nanosuspension significantly enhances as compare with the pure drug.

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**AUTHORS CONTRIBUTIONS**

All the author have contributed equally

**CONFLICT OF INTERESTS**

Declared none

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