

THE EFFECTS OF MIXTURES OF VARIOUS CONCENTRATIONS OF CARBOXYMETHYL CHITOSAN/AMORPHOUS CALCIUM PHOSPHATE WITH GYPSUM ON DENTIN REMINERALIZATION

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ABSTRACT

Objective: Carboxymethyl chitosan/amorphous calcium phosphate (CMC/ACP) is a non-collagen protein analog with the ability of dentin remineralization, and gypsum is a compound material than can facilitate application of CMC/ACP. CMC/ACP concentration is claimed to increase calcium and phosphate concentration, which indicates dentin remineralization. The aim of this study is to observe the effects of different concentrations of CMC/ACP mixed with gypsum on dentin remineralization.

Methods: Occlusal cavities were created in 18 freshly extracted teeth, and the teeth were divided into three groups. The first group consisted of untreated demineralized dentin, and the second and third groups were treated with mixtures of 2.5% and 5% CMC/ACP and gypsum. All groups were checked and analyzed on the 14th day using scanning electron microscopy/energy-dispersive X-ray spectroscopy and one-way ANOVA ($p<0.05$).

Results: Significant differences ($p=0.001$) of calcium and phosphate concentrations were found between Groups I, II, and III ($p=0.001$), whereas no differences were found between Groups II and III.

Conclusion: Gypsum and CMC/ACP work synergistically to induce dentin remineralization. The highest dentin remineralization was shown by 5% CMC/ACP-gypsum mixture, but the level was not statistically different from that of 2.5% CMC/ACP-gypsum mixture.

Keywords: Carboxymethyl chitosan, Amorphous calcium phosphate, Gypsum, Calcium sulfate, Remineralization.

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INTRODUCTION

Gypsum, or calcium sulfate, is a biocompatible regenerative material that releases calcium ions. It is absorbed easily without inducing an inflammatory response [1,2]. Gypsum is often used as a compound material for medicines [3]. Amorphous calcium phosphate (ACP) is a saturated solution of calcium phosphate particles in amorphous form. As it acts as a bioapatite precursor in the transition phase of biomineratization, stabilizing it requires the use of an analogous material with several carboxyl groups, such as carboxymethyl chitosan (CMC), to bind the calcium ions [4,5]. Chen *et al.* [6] and Annisa [7] used CMC/ACP as the material for intrafibrillar dentin remineralization *in vitro*.

Moreover, Subhi *et al.* mixed gypsum with chitosan and bone morphogenetic protein 2 (BMP2) to create pulp capping materials [8]. They analyzed the physical, mechanical, and cellular properties of dental pulp stem cells (DPSCs) and obtained pH results close to neutral and a setting time and compressive strength that were both better than found with *Dycal*. The cellular analysis of DPSCs showed good viability and increased alkaline phosphatase activity on the 3rd day [8].

Dentin remineralization is formed conventionally as guided tissue remineralization (GTR)/bottom-up/non-classical remineralization. GTR is a biomimetic mineralization process, with natural formation of mineral crystals and non-collagen proteins and no residual apatite. When the non-collagen protein has been damaged by caries, a substitute analog material is required to bring calcium and phosphate particles in amorphous form to the gap zone area [9]. Annisa *et al.* demonstrated that CMC/ACP can improve the GTR process [10]. Varying the CMC/ACP concentration can increase the dentin remineralization, with 2.5% being the optimum

concentration [11]. In contrast, addition of gypsum to the CMC/ACP may facilitate its application on cavities. Therefore, using scanning electron microscopy (SEM)/energy-dispersive X-ray spectroscopy (EDX), this study analyzes the effects of various concentrations of CMC/ACP mixed with gypsum on dentin remineralization.

METHODS

This research was approved by the Dentistry Research Ethics Commission of the Faculty of Dentistry, Universitas Indonesia, with protocol number 051171218. Molar teeth without abnormalities were immersed after extraction in a solution containing phosphate-buffered saline (PBS) at 4°C and stored for 14 days. Three cavity samples were made in each tooth to obtain 18 cavities. Each cavity was inundated with 17% ethylenediaminetetraacetic acid (EDTA) solution (*MD Cleanser™*, Meta Biomed Co. Ltd., Chungbuk, Korea) for 1 week, stored in a shaker incubator (100 rpm) at 37°C, and rinsed with aquabidest for 30 min. In addition, each cavity was inundated with 1 M NaCl solution (pH 7.0) for 8 h at 25°C.

We made a mixture of CMC/ACP and gypsum by heating the gypsum for 3 h at 110°C in an electric oven (*Universal Oven Memmert Life 600*, Schwabach, Germany) to form a hemihydrate. The CMC/ACP was made as follows: 2.5 g of CMC (*University of North Sumatra Research Center Laboratory*, Medan, Indonesia) were mixed into 40 ml of water. The mixture was stirred at 1000 rpm until the CMC powder dissolved and a gel was formed. Next, 0.498 g of K₂HPO₄ was added to the gel, and the mixture was stirred at 500 rpm. Then, a mixture of 0.555 g of CaCl₂ in 10 ml deionized water was dropped slowly into the CMC gel, and the resultant mixture was stirred for 5 min until a CMC/ACP gel was formed. The solution was then frozen for 2 h at -80°C and lyophilized

by freeze-drying for 6 h. This was followed by the production of a concentrated solution of 2.5% and 5% CMC/ACP ready to be mixed with 1 g of gypsum.

The 18 cavities were divided into three groups: Group I was demineralized dentin, Group II was application of 2.5% CMC/ACP-gypsum mixture, and Group III was application of 5% CMC/ACP-gypsum mixture. The cavities were filled with light-curing temporary restoration, immersed in PBS solution, and stored in a shaker incubator for 14 days at 37°C.

All teeth were cut to the cavity base and fixed using a multilevel dehydration method. The samples were also soaked with 50%, 70%, 80%, and 90% ethanol for 20 min and 96% ethanol for 2 h. The surfaces were coated with gold (Au), and the morphology of the dentin surfaces was analyzed using a SEM connected with EDX (TESCAN VEGA3, Brno-Kohoutovice, Czech Republic) to measure the content of calcium and phosphate ions. The data were analyzed using Integrated Calibration and Application Tool software (*Oxford Instruments*). EDX analysis of the cavity base was performed twice for each sample.

RESULTS AND DISCUSSION

Studies of CMC/ACP in gel-based preparation have proven that GTR can remineralize dentin within 14 days [6,7]. However, it is difficult for clinicians to apply the required additional material. Therefore, this study selected gypsum, due to its biocompatible and regenerative characteristics, as a CMC/ACP compound material to obtain applicable preparations.

Gypsum is a mineral that consists of calcium sulfate dihydrate ($\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$) and shows a good biological response. It is absorbed perfectly *in vivo*, with no inflammatory response and promotes the release of growth factors (BMP-2, BMP-7, transforming growth factor-beta, and platelet-derived growth factor), angiogenesis, and migration of fibroblasts, which improve the process of bone healing [1]. Moreover, as reported by Lewis *et al.*, the addition of thick polymers (i.e., hyaluronan and CMC) improves the handling of gypsum and its mechanical properties [3]. The addition of CMC will enhance flexural and compressive strength. According to Reynolds *et al.*, adding polymers to gypsum increases osteogenesis and calcification, also affects bone regeneration [3].

The addition of gypsum to the CMC/ACP gel will form a paste with a smooth texture, making it easy to put on cavity. Subhi *et al.* showed that a mixture of gypsum, CMC/ACP, and BMP-2 as a pulp-capping material increased alkaline phosphatase activity on the 3rd day [8]. Erliyana *et al.* also analyzed that adding gypsum to 5% CMC/ACP concentration could increase the remineralization of dentin, but the results were not significant [11]. Increasing CMC concentration in the ACP mixture improved its remineralization ability, and the concentration of 2.5% CMC showed significant results [12].

In this study, the demineralized dentin was used as a sample which imitated clinical condition. Approximately 17% EDTA was chosen as the demineralization material for its chelation ability against calcium ions, using crosslinking of intact collagen [13].

EDX (EDS and EDAX) is used to analyze either chemical elements or the characteristics of specimens. Each element has a unique atomic structure that allows its identification by X-rays.

Statistical analysis was conducted using Statistical Package for the Social Sciences 22 software. The phosphate and calcium concentrations were analyzed using one-way ANOVA and the between-group comparisons were made using the *post hoc* Tamhane test with a significance level of $p < 0.05$. The highest average value of calcium and phosphate (Table 1) was observed with 5% CMC/ACP-gypsum mixture (Group III), the next highest value was observed with 2.5% CMC/ACP-gypsum mixture (Group II), and the lowest value was observed with the demineralized

dentin (Group III). Both calcium and phosphate show significant increases between groups.

The average values of calcium and phosphate increased in Groups II and III and also in the mixtures of gypsum and CMC/ACP. This suggests that gypsum and CMC/ACP work synergistically to induce remineralization of dentin. From Group I (demineralized dentin stored for 14 days in a shaker incubator) to Group II (demineralized dentin with 2.5% CMC/ACP-gypsum mixture), there was a significant increase in both calcium and phosphate because of the addition of gypsum, which also contains calcium.

In Table 2, there are significant differences in the values between Group I and Group II ($p = 0.019$) and between Group I and Group III ($p=0.001$). However, there are no significant differences in values between Group II and Group III ($p=0.394$).

Moreover, Table 3 shows significant differences in the values for phosphate groups between Group I and Group II ($p=0.05$) and between Group I and Group III ($p=0.01$), whereas there are no significant differences between Group II and Group III ($p=0.921$).

Table 1: Mean value, standard deviation, and significance of calcium (%) and phosphate (%) in each group

Group	n	Mean (SD)	p
Calcium			
Group I	6	22.38 (1.67)	0.001*
Group II	6	34.81 (1.73)	
Group III	6	37.04 (2.18)	
Phosphate			
Group I	6	11.57 (0.45)	0.001*
Group II	6	17.29 (0.89)	
Group III	6	17.68 (0.32)	

*One-way ANOVA test $p < 0.05$

Table 2: Mean value of differences between calcium groups

Group	Group II	Group III
Group I	0.019*	0.001*
Group II		0.394

Post hoc Tamhane test $p < 0.05$ ()

Table 3: Mean value of differences between phosphate groups

Group	Group II	Group III
Group I	0.005*	0.001*
Group II		0.921

Post hoc Tamhane test $p < 0.05$ ()

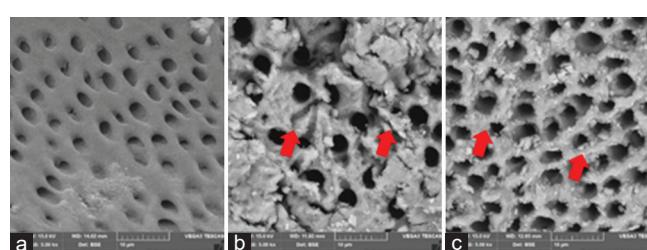


Fig. 1: Scanning electron microscopy results of each group with $\times 5000$. (a) Demineralized dentin, (b) demineralized dentin with 2.5% carboxymethyl chitosan (CMC)/amorphous calcium phosphate (ACP)-gypsum mixture, (c) demineralized dentin with 5% CMC/ACP-gypsum mixture. The remineralization denoted by the red arrows shows the irregular shape of the dentinal tubule walls

In analyzing the significance values between the groups, both calcium and phosphate ions showed a mixture of gypsum, with a significant increase in CMC/ACP amount and phosphate ions. Calcium and phosphate ions can form a hydroxyapatite precursor that allows the ions to enter a narrow yet complex area in the dentin structure. However, the formation of hydroxyapatite is not always preceded by an apatite precursor. ACP is one of the hydroxyapatite precursors [14]. Moreover, the concentration between 2.5% and 5% despite the differences in values was not statistically significant (Tables 2 and 3). However, to verify this, further research using transmission electron microscope observation is required.

Fig. 1 shows different SEM images of the surface morphology among untreated demineralized dentin (a), demineralized dentin after application of 2.5% CMC/ACP-gypsum mixture (b), and demineralized dentin after application of 5% CMC/ACP-gypsum mixture (c). Untreated demineralized dentin shows the thickened dentin and intratubular walls, while demineralized dentin after application of 2.5% CMC/ACP-gypsum mixture shows the mineral deposits on dentinal tubule walls; and the one after application of 5% CMC/ACP-gypsum mixture shows denser mineral deposits with more irregular edges of dentin tubules.

CONCLUSION

About 2.5% and 5% CMC/ACP and gypsum mixtures tended to enhance the dentin remineralization process to the same extent, characterized by an increase in the concentration of calcium and phosphate ions and morphological changes on the dentin surface. Although 5% CMC/ACP-gypsum mixture showed higher dentin remineralization, the difference was not significant.

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CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

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