

## EUDRAGIT COATED ALGINATE BEADS BEARING OXALIPLATIN LOADED LIPOSOMES: FORMULATION, OPTIMIZATION AND *IN VITRO* CHARACTERIZATION

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### ABSTRACT

**Objective:** The present investigation aimed to develop and characterize Eudragit S-100 coated alginate beads bearing oxaliplatin loaded liposomes for colon-specific drug delivery.

**Methods:** Liposomes were formulated by the thin-film hydration method. The process and formulation variables were optimized by Box-Behnken design (BBD) with the help of Design-Expert® Software. Three independent variables taken were HSPC: Chol molar ratio ( $X_1$ ), hydration time ( $X_2$ ), and sonication time ( $X_3$ ). The response variables selected were entrapment efficiency of oxaliplatin, polydispersity index, and vesicle size.

**Results:** The liposomes possessed an average vesicle size of  $110.1 \pm 2.8$  nm, PDI  $0.096 \pm 0.3$ , zeta potential of  $-6.70 \pm 1.4$  mV, and entrapment efficiency of 27.65%. The beads were characterized for their size, *in vitro* drug release, and swelling index. The degree of swelling of the beads was found to be 2.3 fold higher at pH 7.4 than at pH 1.2. The *in vitro* drug release depicted a sustained drug release in 48 h.

**Conclusion:** The outcomes of the study proposed that the developed system can be effectively used for site-specific drug delivery to the colon via the oral route.

**Keywords:** Oxaliplatin loaded liposomes, Formulation, Optimization, *In vitro* characterization

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### INTRODUCTION

Colorectal cancer (CRC) originates from the inner wall of the colon epithelium. It ranks fourth amongst the various malignant tumors. Despite the advancements in its diagnostic methods, it is fatal. Usually, it is marked by the development of colorectal polyps, which are abnormal growths in the intestinal lining. This is followed by its invasion in the muscular tissues and lymph nodes, subsequently to other body organs [1]. Although the treatment of colorectal cancer has greatly improved over time, still it remains one of the leading causes of cancer-related deaths. Oxaliplatin (OHP)-based chemotherapy is the first-line treatment for colorectal cancer [2]. OHP is a third-generation platinum chemotherapeutic drug that can form Pt-DNA adducts with DNA chains to produce inter-chain cross-linking and intra-chain cross-linking, resulting in DNA damage [3, 4]. It can also inhibit the synthesis of DNA and RNA, and trigger systemic immune reactions, leading to apoptosis [5].

Liposomes are biocompatible, biodegradable, and non toxic. Moreover, liposomes can entrap both hydrophobic and hydrophilic molecules and can release the entrapped drug, therefore it could be a useful drug carrier system [6]. But, they can degrade during their passage via the GIT. This can be prohibited by entrapping them in eudragit coated alginate beads that are degraded by the microflora present in the large intestine. Because of the enteric coating, the beads will remain protected in the GIT, and reach the colon [7]. This approach uses a combination of the pH-sensitive property of enteric polymer (i.e. eudragit S-100) and the biodegradability of alginates for colon-specific targeting. On entering the colon, alginate is degraded by the polysaccharidases and eudragit coating gets dissolved in the ileocaecal region of the small intestine in the colon, releasing the liposomes [8, 9].

The developed liposomes were optimized using Box Behnken Design (BBD). BBD is response surface designs based on three-level incomplete factorial designs. They are particularly designed since they need only 3 levels, coded as -1, 0, and +1. BBD is available for 3–10 factors. They are formed by combining two-level factorial designs with incomplete block designs. This generates designs with suitable statistical attributes, but most significantly, with only a fraction of

the experiments necessitated for a three-level factorial. Since there are only three levels, the quadratic model is appropriate. A comparative analysis of BBD and other response surface designs (central composite, Doehlert matrix, and three-level full factorial design) has portrayed that the BBD and Doehlert matrix are somewhat more efficient than the central composite design but much more efficient than the three-level full factorial designs where the efficiency of one experimental design is termed as the number of coefficients in the estimated model divided by the number of experiments performed [10].

One more advantage of the BBD is that it does not have combinations for which all factors are simultaneously at their highest or lowest levels. Therefore, these designs help to evade experiments conducted under extreme conditions, for which undesirable outcomes may be obtained.

The primary aim of this research work was to optimize the various process and formulation variables to prepare colon-specific oxaliplatin-loaded liposomes for colon cancer delivery.

### MATERIALS AND METHODS

Oxaliplatin was procured as a gift sample from Khandelwal Laboratories Ltd. (Mumbai, India), Hydrogenated soy phosphatidylcholine (HSPC); Cholesterol (CH) was obtained from Sigma (St. Louis, MO). Eudragit S-100 was obtained as a gift sample from Rohm GmbH and Co KG., Darmstadt, Germany, Methanol (Merck Life Science Pvt. Ltd., Mumbai; India), chloroform (Merck Life Science Pvt. Ltd., Mumbai; India), ultrapure water (Millipore, Bedford, MA) was used throughout the experiment. All other chemicals employed were of the highest grade commercially available.

### Preparation of liposomes

Liposomes were formulated by the thin-film hydration method reported by Bangham *et al.* and Mezei and Gulasekhelam. In brief, HSPC and CH were dissolved in chloroform: methanol (3:1) in a round-bottomed flask. A thin film was formed on the inner sides of the flask by evaporating the solvent system at 45 °C by continuously

rotating the round bottom flask for 40 min at 100 rpm using a rotary evaporator (Buchi type, York Sci Co., Bombay) under reduced pressure. The solvent was completely removed. OHP was dissolved in HEPES buffer (pH 7.4). The film was hydrated with the drug-containing HEPES buffer by vortexing the flask for 30

min to get multilamellar vesicles. The suspension was kept aside for 4-5 h for the complete swelling of the liposomes. It was sonicated for 5 min at  $4\pm 1$  °C. The liposomes were passed through Sephadex G-50 columns to remove the untrapped drug [11, 12].

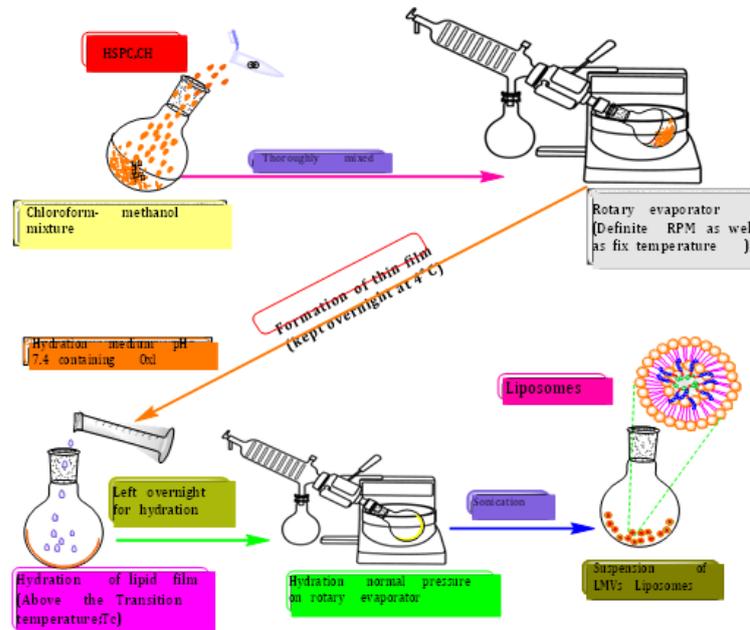


Fig. 1: Schematic representation of formulation of liposomes using the thin-film hydration method

**Optimization**

Design of experiment (DoE) is helpful in the identification of main variables influencing the quality characteristics of interest in the process. A designed experiment is a sequence of tests in which the input variables of a process are altered purposefully so that corresponding changes in the output responses can be observed and identified. It is widely employed in the research and development domains for process optimization [13].

Systematically DoE optimization principle was implemented by response surface methodology (RSM) for analyzing second-order polynomial models, using various feasible experimental runs. With the help of Design-Expert® software (10.0, Stat-Ease Inc., Minneapolis, USA) and 3-factors, 3-levels BBD was utilized for analyzing the effect of independent variables i.e. Molar ratio of HSPC: Chol ( $X_1$ ), Hydration time ( $X_2$ ), and sonication time ( $X_3$ ) to estimate the 3-D response surfaces by scrutinizing the effect of independent variables on selected dependent variables i.e., percent entrapment efficiency OHP ( $Y_1$ ), polydispersity index (PDI) ( $Y_2$ ) and vesicle size ( $Y_3$ ). All independent variables are shown as +1, 0, and -1, which portray high, medium, and low levels, respectively (table 1). All the independent and dependent (response) variables have been outlined with their coded and decoded levels in table 1. In the design matrix 17 runs were produced on Three-level, Three-factors (table 2). Few model parameters such as multiple

correlation coefficient ( $R^2$ ), adjusted multiple correlation coefficient/(adjusted  $R^2$ ), predicted multiple correlation coefficient (predicted  $R^2$ ), coefficient of variation (CV), and predicted residual sum of the square were estimated and compared to explore the generated model. The response surface analysis (RSA) was carried out by 3D response surfaces (fig.) using Design-Expert® software (10.0, Stat-Ease Inc., Minneapolis, USA). 3D response surface plots help to establish desirable responses and operating conditions. They usually display a three-dimensional view that can impart a clear image of the response. These 3D surface plots aid the experimenters to comprehend the nature of the relationship between the factors and the responses [14].

**Characterization of optimized liposomes**

The optimized liposomes were characterized for various attributes such as morphology, vesicle size, zeta potential, PDI, *in vitro* assessment.

**Surface morphology**

The morphology of the liposomes was determined by transmission electron microscopy (JEM-200 CX; JEOL, Tokyo, Japan). The samples were placed on a grid by the drop-casting technique. It was dried and observed under a transmission electron microscope [15] (fig. 6B).

Table 1: Factors and their levels as per BBD for optimization of liposomes

Factors (Independent variables)	Levels		
	Low (-1)	Medium (0)	High (+1)
X1 Molar ration of HSPC: Chol	1.5:0.5	2:1	2.5:1.5
X2 Hydration time (Minutes)	60	120	180
X3 Sonication time (Minutes)	6	7	8
Response (Dependent variables)	Constraints		
R1: Entrapment efficiency OHP	Maximum		
R2: Polydispersity index (PDI)	Minimum		
R3: Vesicular size (nm)	In range		

Table 2: Design matrix with compositions of liposomal formulations

Runs	Factor 1	Factor 2	Factor 3	Responses		
	Molar ratio HSPC: Chol	Hydration time	Sonication time	%EE OHP	PDI	Vesicle size
1	-1.00	0.00	-1.00	24.55	0.218	116.6
2	1.00	1.00	0.00	28.00	0.172	115.8
3	-1.00	0.00	1.00	20.99	0.115	108.5
4	-1.00	1.00	0.00	24.44	0.192	111.0
5	0.00	0.00	0.00	28.50	0.099	111.6
6	1.00	-1.00	0.00	26.60	0.092	113.0
7	0.00	0.00	0.00	27.90	0.079	112.8
8	0.00	0.00	0.00	28.72	0.085	112.2
9	0.00	0.00	0.00	29.22	0.084	112.3
10	0.00	-1.00	-1.00	28.21	0.166	119.4
11	0.00	-1.00	1.00	23.60	0.096	112.0
12	0.00	1.00	-1.00	29.40	0.248	121.0
13	1.00	0.00	-1.00	28.59	0.175	120.0
14	-1.00	-1.00	0.00	22.35	0.125	112.9
15	1.00	0.00	1.00	25.80	0.093	111.3
16	0.00	0.00	0.00	28.58	0.089	112.1
17	0.00	1.00	1.00	27.38	0.136	111.1

### Size, zeta potential, and PDI

Vesicle size (z-average), size distribution (polydispersity index), and zeta potential of drug-loaded liposomes were determined by NanoPlus-3 (Version 5.01, Micromeritics Instrument Corporation, Particulate Systems, Norcross, GA, USA) by Photon Correlation Spectroscopy (PCS). Diluted preparation was placed in the cuvette and size was determined. The zeta potential of the liposomes was computed by Helmholtz-Smoluchowski equation from their electrophoretic mobility at 20 V/cm field strength and 50  $\mu$ S/cm conductivity [15].

### Entrapment efficiency

The entrapment efficiency of prepared liposomes was estimated by separating the untrapped drug with the help of Sephadex G-50 minicolumn by centrifugation technique [16]. Sephadex G-50 (1.0 g) was allowed to swell in 0.9% NaCl at 25 °C with shaking, for 4 h, leading to the gel formation. It was stored at 4 °C and filled in minicolumn. Liposomes were added to the prepared column and centrifuged at 2000 rpm for 2 min. The separated liposomes were lysed using 0.1% Triton X-100 and filtered via a 0.2-micron membrane filter. The filtrate was assessed for drug content using HPLC (Shimadzu Corporation, Kyoto, Japan) employing 0.15 mol/l sodium dodecyl sulphate (SDS)/6% (v/v) pentanol mobile phase at a detection wavelength of 325 nm. The experiment was performed in triplicate [17]. The percentage entrapment efficiency was calculated using the following formula:

$$\text{Entrapment efficiency (\%)} = \frac{\text{Total drug (mg)} - \text{free drug (mg)}}{\text{Total drug (mg)}} \times 100$$

### Preparation of liposomes loaded eudragit s-100 coated calcium alginate beads

Liposomes were pelleted by centrifugation, suspended in water, and mixed with alginate solution. This dispersion was passed through a syringe (22 gauge) into the calcium chloride solution (100 mmol/l) with slow stirring for an hour. The beads formed were collected and washed with distilled water, and dried in a vacuum desiccator overnight to attain a constant weight [18]. The enteric coating of the beads was done as per the method given by Huyghebaert *et al.* [19]. The coating solution was prepared by stirring Eudragit-S 100 and 1M ammonia for 1 hour. Then triethyl citrate was added to the solution and stirred for 1 hour. The coating solution was passed by a 0.3 mm sieve before use. The alginate beads were dip-coated and air-dried.

### Physical characterization of liposomes loaded eudragit coated alginate beads

#### Determination of the size of beads

The size of uncoated and coated alginate beads was determined using optical microscopy. The diameters of beads were measured with an

eyepiece micrometre on an optical microscope (40X magnification). Randomly 100 selected beads were measured for each sample [20].



Fig. 2: Liposomes loaded eudragit coated alginate beads

### Drug content

Approximately 0.5 g of alginate beads were ground in a pestle. The powder was dissolved in 100 ml of phosphate buffer (pH 7.4) containing 0.1% Triton X 100 by stirring for 6-7 h. The solution was filtered through a 0.2  $\mu$ m millipore membrane filter [21]. The concentration of the drug was determined by HPLC method as discussed above.

### In vitro drug release profile

*In vitro* drug release from coated beads was estimated according to Souder and Ellenbogen extraction method [22].

The *in vitro* drug release was performed in the simulated gastric fluid pH 1.2 in the 1<sup>st</sup> hour, a mixture of simulated gastric and intestinal fluid pH 4.5 (2-3 h), simulated intestinal fluid pH 7.5 (4-5 h), and simulated colonic fluid pH 6.8 (6-8 h) using a dialysis bag [17]:

An accurately weighed amount of enteric-coated beads were placed in a dialysis bag which was placed in a beaker having 100 ml of simulated fluids at 37 $\pm$ 1 °C with magnetic stirring maintained under sink conditions, and media were changed as per Souder and Ellenbogen's scheme. Samples from the beaker (one ml each) were withdrawn and replaced immediately with the same volume of fresh medium at fixed times. The study was continued for 48 h. These samples were analyzed for drug content using HPLC method as discussed above.

### Swelling study

The swelling behavior of coated and uncoated alginate beads was estimated in buffer solutions of pH 1.2 and 7.4 at 37 $\pm$ 1 °C. The beads

were accurately weighed ( $W_0$ ) and immersed in 100 ml of buffer solution. At definite time intervals, the swollen beads were wiped with tissue paper and weighed ( $W_g$ ) [23]. The percentage swelling degree of the beads was computed by the equation:

$$\% \text{ Swelling} = \frac{W_g - W_0}{W_0} \times 100$$

where,  $W_0$  is the initial weight of the beads and  $W_g$  is the weight of the alginate beads at equilibrium swelling.

## RESULTS

The developed liposomes were investigated for dependent variables like %EE of OHP ( $Y_1$ ), PDI ( $Y_2$ ), and vesicle size ( $Y_3$ ) on the chosen independent variables, HSPC: Chol molar ratio ( $X_1$ ), hydration time ( $X_2$ ), and sonication time ( $X_3$ ) as per BBD. The ten polynomial coefficients ( $\beta_1$ - $\beta_{10}$ ) were suggested during mathematical modelling, with  $\beta_0$  as intercept has been shown in table 3. All the

polynomial coefficients were compliant with the second-order quadratic polynomial model for the assessment of the chances of remarkable interaction(s) between the examined responses as portrayed in the equation.

$$Y = \beta_0 + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3 + \beta_4 X_1 X_2 + \beta_5 X_1 X_3 + \beta_6 X_2 X_3 + \beta_7 X_1 X_2 X_3 + \beta_8 X_1 X_2 (X_1 - X_2) + \beta_9 X_1 X_3 (X_1 - X_3) + \beta_{10} X_2 X_3 (X_2 - X_3)$$

A high significant statistical value ( $p > 0.001$ ) was achieved for the three response variables in the generated model. The  $r^2$  values for all the models were found in the range 0.9875 and 0.9951 which proposed an excellent fit of polynomial equation produced for response data ( $p < 0.0001$  in all the cases). The "lack of fit", ranging between  $p = 0.6371$  and  $p = 0.7335$  for all the produced models were noticed to be insignificant which affirm that the suggested model was appropriate. The closeness of magnitude in the adjusted (Adj) and predicted (Pred)  $r^2$  (ranging from 0.9833-0.9888 and 0.9232-0.9746) also confirm an excellent fit of the data to the generated models.

**Table 3: Generated model summary statistics of all responses determined according to BBD**

Coefficient code	Polynomial coefficients for the response variable		
	% EE OHP	Hydration time	Vesicular size
$\beta_0$	+28.58	+0.087	+112.20
$\beta_1$	+2.08	-0.0147	+1.39
$\beta_2$	+1.06	+0.033	+0.20
$\beta_3$	-1.62	-0.045	-4.26
$\beta_4$	-0.17	-1.11	+1.17
$\beta_5$	+0.19	+0.71	-0.15
$\beta_6$	+0.65	-0.015	-0.63
$\beta_7$	-2.70	-0.023	-0.40
$\beta_8$	-0.54	+0.034	+1.37
$\beta_9$	-0.90	+0.039	+2.30

**Table 4: Constraint for numeric optimization and predicted solution**

Factors (Independent variables)	Goal	Levels	
		Low (-1)	High (+1)
$X_1$ Molar ratio of HSPC: Chol	In range	1.5:0.5	2.5:1.5
$X_2$ Hydration time	In range	60	180
$X_3$ Sonication time	In range	6	8
Response (Dependent variables)			
$Y_1$ : Entrapment efficiency OHP	Maximum	20.990	29.400
$Y_2$ : PDI	Minimum	0.079	0.250
$Y_3$ : Vesicular size (nm)	In range	108.500	121.000

## DISCUSSION

The present study was aimed at developing and optimizing oxaliplatin-loaded liposomes and entrapping them in Eudragit S-100 coated beads that could be used for colonic delivery. OHP-loaded liposomes were prepared by thin-film hydration method using Hydrogenated phosphatidylcholine and cholesterol. OHP is hydrophilic thus; it was entrapped in the aqueous core surrounded by a lipid bilayer of HSPC and cholesterol rendering its structural stability. The optimization of various formulation and process variables was done using Quality by design approach, Design-expert® software. The Box-Behnken design was applied for the optimization of formulation and process parameters, namely the molar ratio of HSPC: Chol, Hydration time, and sonication time. It was found that with an increase in the molar ratio of HSPC: Chol, a rise in EE (%) of OHP was noticed up to a high level, followed by a constant value which could be due to saturation of all the aqueous fluid of the aqueous core [24]. Likewise, increasing the hydration time was accompanied by an increase in the EE (%) which could be the result of the high amount of the entrapment of aqueous fluid. However, with an increase in sonication time, there was a decrease in the EE (%) which can be because of the drug leakage due to rupture of liposomes during sonication [25] (fig. 3A-C). Fig. 4 A-C depict the dependence of PDI on various factors like the molar ratio of HSPC: Chol, hydration time, and sonication time. At a low ratio of HSPC: Chol, high PDI was noticed. With an increase in HSPC: Chol ratio and sonication time there was a decrease in PDI. Nevertheless,

with an increase in hydration time, there was an increase in PDI. Fig. 5 A-C show the dependence of the vesicular size of liposomes on the factors like the molar ratio of HSPC: Chol, hydration time, and sonication time. They show that as the molar ratio of HSPC: Chol and hydration time increased, the particle size also increased. This could be due to the increased bilayer molecules and hydration fluid in the liposomes. While an increase in sonication time led to a decrease in vesicular size because of the high shear stress, which resulted in the rupturing liposomes into smaller vesicles [26].

The optimum preparation has a maximum % EE of OHP, minimum PDI, and vesicle size in range  $Y_1 = (20.990 \leq 29.400)$ ,  $Y_2 = (0.079 \leq Y \leq 0.250)$  and  $Y_3 = (108.5 \leq Y \leq 121.0)$

The morphology of liposomes was confirmed by optical microscopy and transmission electron microscopy images (fig. 6A and B). The spherical shape of the vesicles was revealed by transmission electron microscopy. The mean diameter of the liposomes was found to be  $110.1 \pm 2.8$  nm. Low polydispersity index, i.e.  $0.096 \pm 0.3$  showed that liposomes had a narrow size distribution and were homogeneously distributed. Zeta potential is one of the most important attributes to assess the stability of a liposomal suspension, which was found to be  $-6.70 \pm 1.4$  mV. The liposomes were entrapped in sodium alginate beads which were further coated with Eudragit S-100. Tri-ethyl citrate was used as a plasticizer during coating due to its good affinity for eudragit. The mean diameters of the uncoated and coated beads were found to

be  $1.375 \pm 0.106$  mm and  $1.608 \pm 0.113$  mm, respectively. The increase in the size of the coated beads could be attributed to the application of eudragit S-100 coating on the beads. The drug content was in

proportion to that of loaded drug ranging from  $21\% \pm 1.8\%$  to  $28\% \pm 2.7\%$  wt/wt [27].

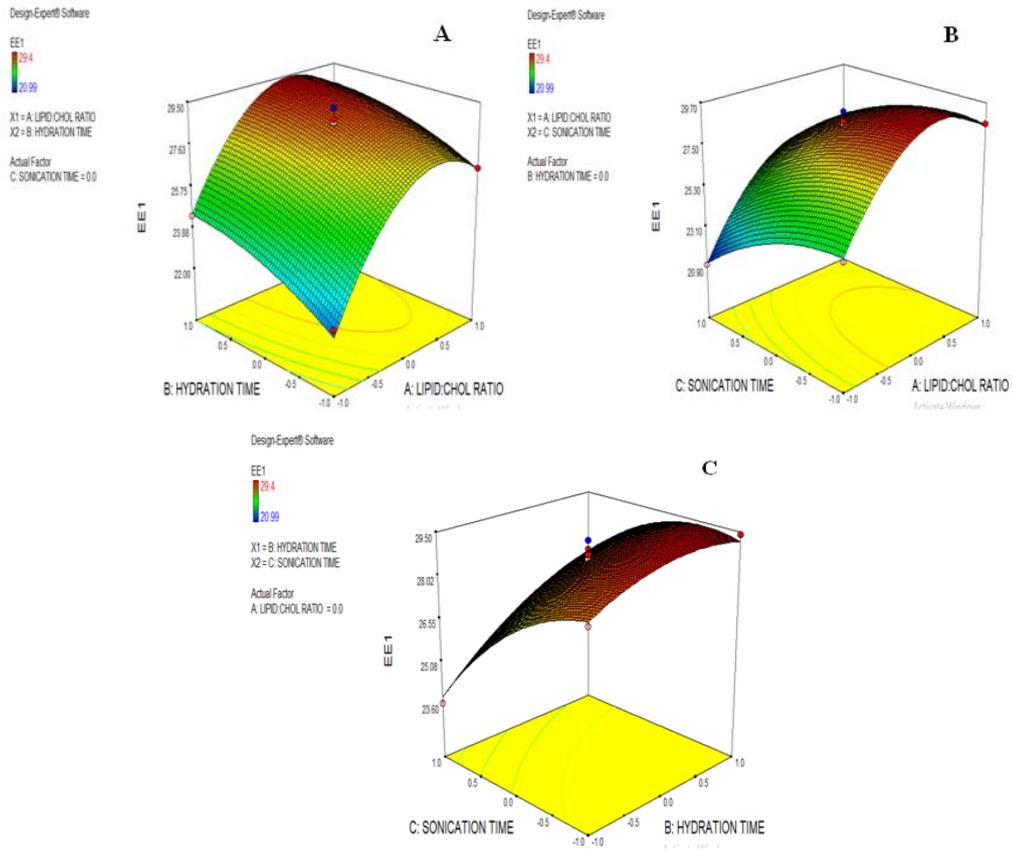
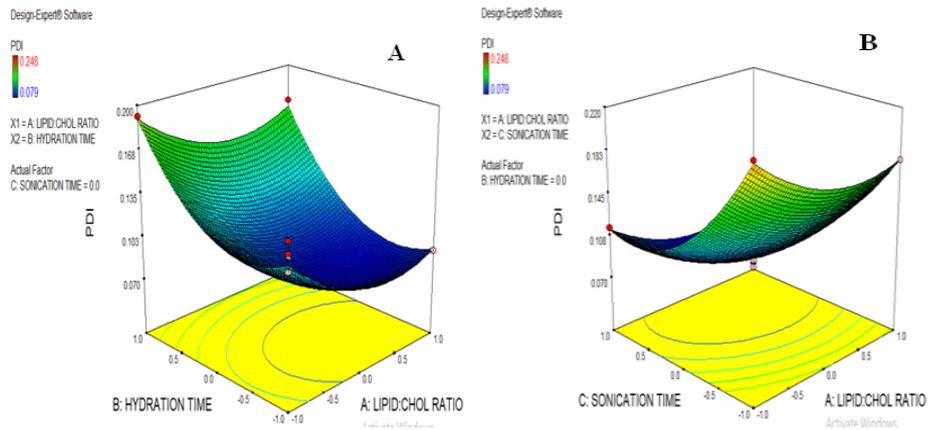


Fig. 3: (A–C) shows the 3-D response surfaces for Entrapment efficiency (EE) of OHP



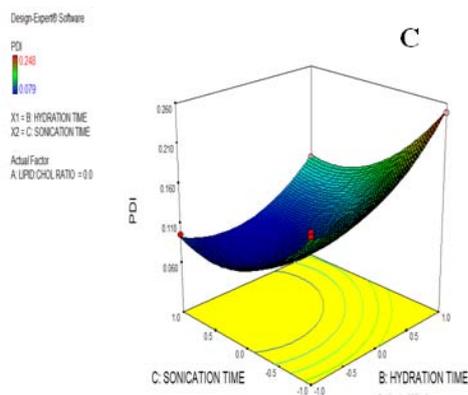


Fig. 4: A-C 3D surface response curves representing PDI

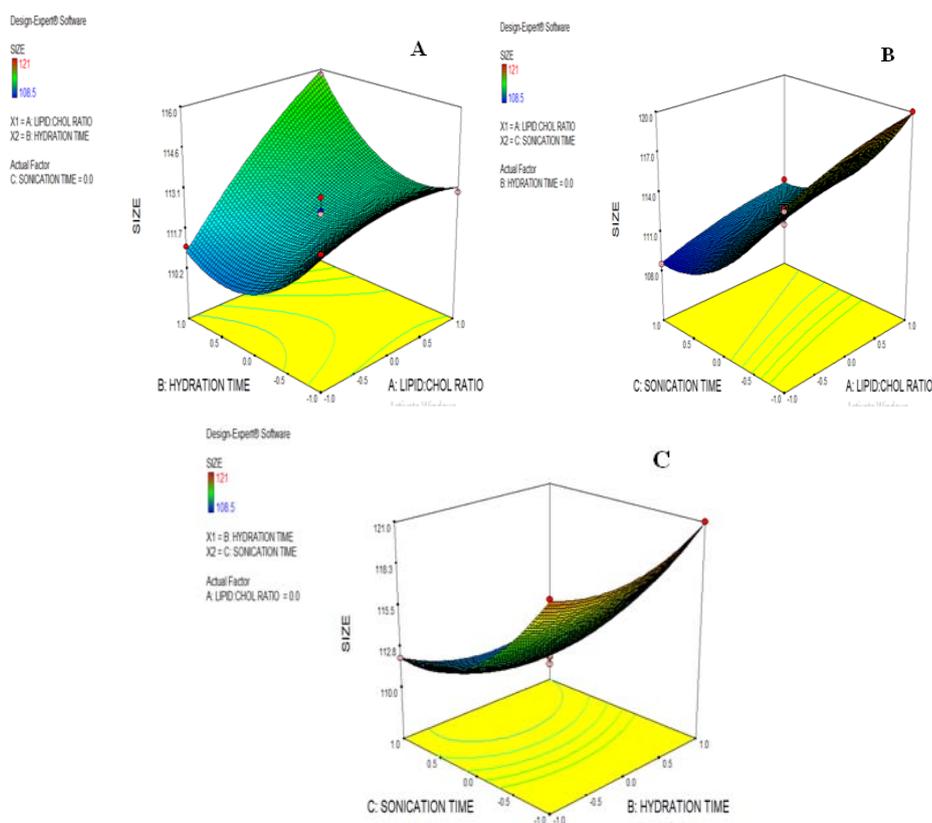


Fig. 5: A-C 3D surface response curves representing the vesicle size,

The cumulative percentage of drug release from Eudragit coated beads is shown in fig. 7. It was found that there was no drug release up to 6 h; it started releasing the drug after the completion of 6.5 h when these beads came in contact with simulated intestinal fluid pH 6.8. There was no drug release in the upper parts of GIT i.e. stomach and intestine from Eudragit S-100 coated beads since they were safeguarded in the upper GIT due to the Eudragit-S-100 coating which undergoes dissolution only at pH>6.8. The coating of the beads starts dissolving after reaching the distal part of the small intestine and the polysaccharidase enzyme present in the colon digests the uncoated alginate beads leading to the release of the entrapped liposomes. At the end of 48 h, 71.6% of oxaliplatin was released from the liposomes. The results attained are concordant with those obtained by Chaurasia *et al.* 2007. They prepared guar gum microspheres bearing methotrexate and reported that maximum drug release was

observed after 7 h, which is due to the digestion of guar gum by the colonic microflora [28].

Swelling is an essential characteristic that determines the release behavior of a drug molecule. The swelling property of the beads was determined in buffer solutions with pH 1.2 and 7.4 (fig. 8). It is noticed that the swelling property of beads is highly affected by the pH of the medium. The degree of swelling of the beads was found to be higher at pH 7.4 than at pH 1.2. It could also be due to the ion exchange reaction between Na<sup>+</sup> ions of the phosphate buffer and Ca<sup>2+</sup> ions linked to carboxylic acid groups of alginate. Monovalent ions substitute bivalent ions, break the "egg-box" structure and increase the distance between the polymer chains, thereby enhancing the fluid absorption and swelling of the beads. The outcomes were according to the study reported by Seeli *et al.* 2016 in which they developed guar gum succinate sodium alginate beads that depicted pH-dependent swelling property [29].

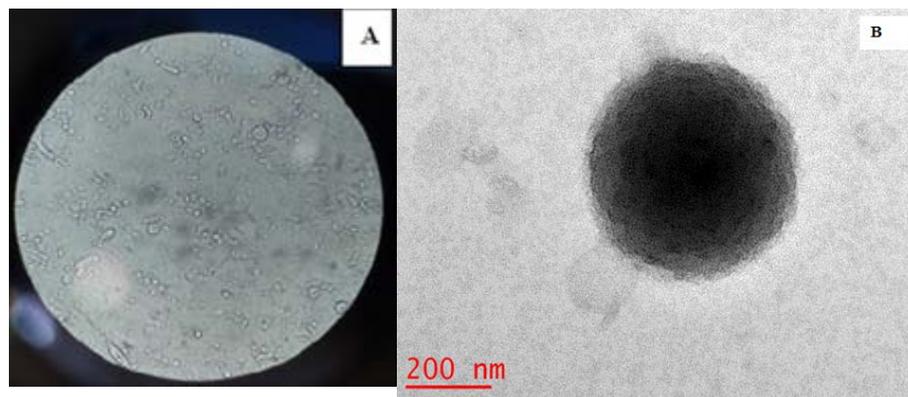


Fig. 6: Images of liposomes (A) Phase-contrast photomicrographs (40x) (B) TEM image

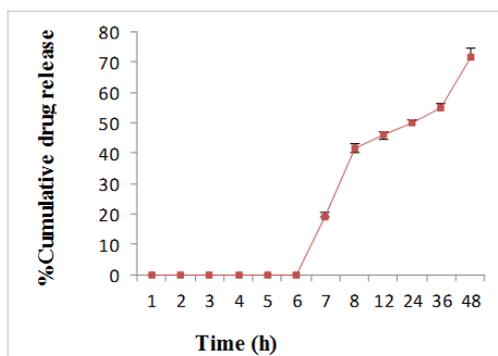


Fig. 7: *In vitro* drug release profile of eudragit coated beads bearing liposomes in various simulated fluids. Simulated gastric fluid (pH 1.2); simulated intestinal fluid (pH 4.5), simulated colon fluid (pH 6.8). Results are presented as mean $\pm$ SD, n = 3

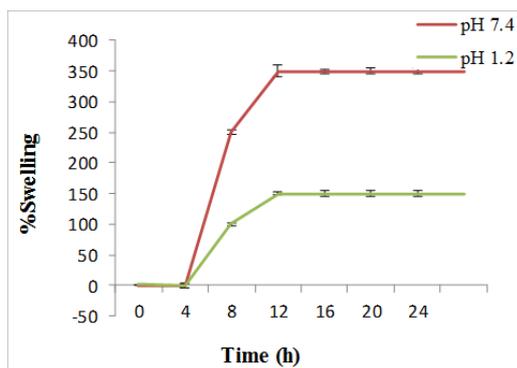


Fig. 8: Determination of % swelling of alginate beads at pH 1.2 and pH 7.4. Results are presented as mean $\pm$ SD, n = 3

## CONCLUSION

In the current study, oxaliplatin-loaded liposomes were formulated using the thin-film hydration method followed by successful optimization using BBD. They were entrapped in alginate beads and dip-coated with Eudragit S-100. The oral delivery of the developed eudragit S-100 coated beads encapsulating oxaliplatin-loaded liposomes can proffer high protection from premature drug release in simulated upper parts of GIT. Whereas, the beads delivered most of the drug in the colon, an environment, abundant in bacterial enzymes that digest the alginate and permit the drug release in the colon. Therefore, the developed system has immense potential in the localized delivery of chemotherapeutic drugs in the colon.

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## AUTHORS CONTRIBUTIONS

Tiwari A: Conducted all the experiments, collected and analyzed the data. Wrote the paper

Jain SK: Designed and supervised the experiments.

## CONFLICT OF INTERESTS

Authors have no conflict of interest.

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