ABSTRACT

Objective: To evaluate the antimicrobial activity of methanolic extract from the peel of the fruit of Citrus Limon (Family-Rutaceae) in conjugation with phytochemical analysis.

Methods: The methanolic extract from the peel of the fruit of Citrus Limon (Family-Rutaceae) was separated from fruits, shade dried, powdered and extracted using methanol, analysed for phytochemical constituents using standard methods. The antimicrobial activity of the plant extract was examined against 2 bacterial strains among one is Gram-positive (Staphylococcus aureus) and other is Gram-negative (Escherichia coli) and 1 fungal strain (Candida albicans) using agar well diffusion method.

Results: The present investigation shows the phytochemical analysis, antimicrobial activity of the methanolic extract of the fruits peel Citrus limon. Various phytochemical analyses revealed the presence of alkaloids, saponin, flavonoids, carbohydrates, glycosides and citric acids and tannins. The antimicrobial activity of the methanolic extract of the plant showed significant result against all the test organisms.

Conclusion: The present study concluded that methanolic extract of the peel of Citrus Limon contains the high significance of phytochemicals. The methanolic extract of the plant was found to possess promising antimicrobial activity when compared with the standards.

Keywords: Citrus Limon, Antimicrobial, Zone of Inhibition, Agar disc diffusion method

INTRODUCTION

For a long period of time, there are many naturally occurring materials which are having bioactive substance and show biological activity for the health of human beings and they have a great potential for producing new drugs. In plant chemotherapy, the use of naturally occurring antimicrobial substances is gaining more importance and have higher significant values [1].

According to WHO medicinal plants are used in order to the therapeutic purpose and be used as a pioneering the synthesis, semi-synthetic chemical drugs [2]. About 80 % of the world population use herbal medicine to treat the ailment. From the statistics, it is under in developed countries and higher in less developed countries [3].

Medicinal plants have an important role for the health of individuals and communities. These plants have a great medicinal value that lies various chemical substances which produce physiological action on the human body. Medicinal plants contains many chemical compounds such as alkaloids, flavonoids, glycosides, saponins, resins, oleoresins, sesquiterpene, phenolic compounds, fats and oils [4].

Citrus fruit is an important medicinal plant of the family Rutaceae. It is used mainly for its alkaloids, which are having anticancer activities and the antibacterial potential in crude extracts of different parts (leaves, stem, root, juice, peel and flower) of lemon against various bacterial strains. Citrus fruits have a broad spectrum of biological activity including antibacterial, antifungal, antidiabetic, anticancer and antiviral activities due to alkaloids [5].

The lemon peel extracts is done by different solvents such as ethanol, methanol and acetone which are subjected to antibacterial assay. Methanolic extract shows higher antimicrobial activity against tested microorganisms (E. coli, S. aureus, Candida albicans and Trichophyton rubrum) [6].

The citrus peels are rich in nutrients and contain many phytochemicals, these are β and γ-s-tocopherol, glycosides and volatile oils. Some polyethoxylated, phenolic compound, ascorbic acid, flavones have several important activities, which are very rare in other plants. Citrus peels also used in scurvy, digestion, respiratory disorders, peptic ulcer, eye infections, gums, gout, skin care, piles, urinary disorders, weight loss traditionally. In addition, it also used in disinfect and sterilizing reagent. So the target of this present study is to identify the phytochemical components of Citrus limon and to determine the antimicrobial effects of the dried peel extract on E. coli, S. aureus, Candida albicans and Trichophyton rubrum.

MATERIALS AND METHODS

Collection of plant materials

Citrus limon was collected from Azara market, Guwahati India. The plant specimen was authenticated by Dr. G. C. Sarma, Curator, Department of Botany, Gauhati University, Assam. The voucher specimen has been deposited in the Herbarium of the Department of Botany, Gauhati University with voucher no Acc. No.-18219 dated 28-10-2016 for future reference.

Chemicals and reagents

Methanol, Ethanol, Petroleum Ether(PE), Chloroform, hydrochloric acid, Dragendorff reagent, Mayer’s reagent, Benedict’s reagent, sulphuric acid, lead acetate, Moliš’s reagent, Fehling solution A and B, sodium citrate, copper sulphate, ferric chloride, sodium hydroxide, glacial acetic acid, benzene, chloroform, ammonium, nitric acid, dimethyl sulfoxide (DMSO), potassium nitrate, gelatine, Beef extract, Peptone, Sodium Chloride, Dextrose and agar. All the chemicals and solvents used were of standard analytical grades.

Preparation of extracts of Citrus lemon peels

The Citrus lemon peels were dried under shade and then undergone crushing in an electric blender to form powdered. Then it was defatted by Petroleum ether dried the marc again extracted with chloroform then again dried the marc and finally extracted with methanol using Soxhlet’s extractor. The percent yield of methanolic extract was then calculated out.

Preparation of extract/drug stock solution

The stock solution of Citrus limon peel extract was prepared on each occasion by careful weighing and dissolving in a suitable volume of Dimethylsulphoxide (DMSO) to get a concentration of 100 mg/mL A
tablet of ciprofloxacin was dissolved in an appropriate volume of water to get 5 mg/ml of stock solution.

**Phytochemical screening**

Phytochemical screening were done using standard method. All the experiment has been repeated in triplicate for final confirmation of the result.

1. **Test for saponins:** To 1 ml of aqueous extract was added to few volume of distilled water in a test tube. The solution was shaken vigorously and observed for a stable persistent froth for 20 min.

2. **Test for alkaloids:** Two methods were used to test for alkaloids. First, evaporate 20 ml of ethanol extract, the dry residue dissolved in 5 ml of HCl (2N) and filtered. A few drops of Mayer’s reagent and Wagner was added, the presence of precipitate indicates the alkaloids.

Second, 3 to 15 ml of the aqueous extract was added 2 ml of NH\textsubscript{3} reagent and Wagner was added, the presence of precipitate indicates the alkaloids.

3. **Test for sterols and steroids:** Extracts were treated with chloroform and filtered. The filtrates were treated with few drops of Conc. Sulphuric acid, shaken and allowed to stand. Appearance of golden yellow colour indicates the presence of triterpene.

4. **Test for the phenolic compounds:** Flavonoids: The ethanol extract 5 ml was added to a concentrated sulphuric acid (H\textsubscript{2}SO\textsubscript{4}) (1 ml) and 0.5g of Mg. A pink or red coloration that disappears on standing 3 min. indicates the presence of flavonoids.

5. **Tannins:** Two methods were used to test for tannins. First, about 1 ml of the ethanol extract was added in 2 ml of water in a test tube. 2 to 3 drops of diluted ferric chloride (Fecl\textsubscript{3}) solution was added and observed for green to blue-green or blue-black coloration. Second, 2 ml of the aqueous extract was added to 2 ml of water, 1 to 2 drops of diluted ferric chloride (Fecl\textsubscript{3}) solution were added. A dark green or blue-green coloration indicates the presence of tannins.

6. **Test for Glycoside:** 2 ml of concentrated H\textsubscript{2}SO\textsubscript{4} was added carefully and shaken gently. A reddish brown colour indicated the presence of steroidal ring i.e. glycone portion of the glycoside [13, 14].

**Culture media**

The media employed for the study was solid agar media.

**Microbial strains**

Three bacterial strains [Staphylococcus aureus (ATCC 25923), Escherichia coli (ATCC 25922) and two fungi] Candida albicans (ATCC 10231), and Trichophyton rubrum (ATCC 28186) were used. Stains were obtained from Girijananda Chowdhury Institute of Pharmaceutical Science, Azara, Guwahati.

**Sterilization of materials**

The petri dishes and pipettes packed into metal canisters were appropriately sterilized in the hot air oven at 170 °C for 1 h at each occasion. A solution of the extract and culture media were autoclaved at 121 °C for 15 min.

**Antibacterial activity**

The antimicrobial activity of the different extracts of the plant was assayed by agar-well diffusion method as described in NCCLS, 1993 [15]. Petri plates containing 20 ml nutrient agar medium was seeded with bacterial strains. Wells of approximately 10 mm was bored using a well cutter. Plant extracts were prepared in DMSO (stock: 1 mg/ml DMSO). The plant extracts of 25, 50, and 100 µl concentrations were added. Ciprofloxacin (20 µl) and DMSO (100 µl) were used as positive and negative controls respectively.

The plates were then incubated at 37 °C for 24 h. The antagonists present in the plant extract are allowed to diffuse out into the medium and interact with the test organisms in the freshly seeded plate. The diameter of the zone of inhibitions was measured in millimetres after 24 h.

**Antifungal activity**

The dextrose agar plates were prepared and inoculated with a fungal culture. Wells of approximately 10 mm was bored using a well cutter and samples of different concentration were added. The zone of inhibition was measured in millimetres after overnight incubation and compared with that of standard antifungal (Fluconazole) (10 µl) which was used as positive control and DMSO (10%) as the negative control [14].

**RESULTS**

The present systematic examination shows the phytochemical analysis, antimicrobial activity of the methanolic extract of the peel of the fruit of *Citrus limon*. The yield % of the extraction of methanol was 18.05%. It was dark brown solid powder in appearance.

**Phytochemical analysis**

The phytochemical analysis with the methanolic extracts of the dried fruit of *Citrus limon* showed the presence of alkaloids, saponin, sterols, Steroids, terpenoids, protein and amino acid, tannins, carbohydrate are given in (table 1).

**Antimicrobial activity**

Generally, Plant extracts are rich in antimicrobial compounds. The in vitro antimicrobial study of the methanolic extracts of fruits peel of *Citrus limon* under different concentration with the standard are mentioned in (table 2). The methanolic extracts of dried fruits peel of the plant *Citrus limon* showed antimicrobial activity against most of the test organisms.

**Table 1:** Phytochemical screening of methanolic extract of dried fruits peel of *Citrus limon* (family-Rutaceae)

<table>
<thead>
<tr>
<th>S. No.</th>
<th>Phytochemical test</th>
<th>Reagent used (test performed)</th>
<th>Observation</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Alkaloids test</td>
<td>Mayer’s test</td>
<td>Frothing observed</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>Saponin test</td>
<td>Wagner’s test, Dragendroff’s test</td>
<td>Turbidity obtained</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>Flavonoid test</td>
<td>Foam test</td>
<td>Golden yellow colour observed</td>
<td>+</td>
</tr>
<tr>
<td>4</td>
<td>Carbohydrate test</td>
<td>Lead acetate test</td>
<td>Yellow colour observed</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>Glycoside test</td>
<td>Modified Borntrager’s</td>
<td>Brownish black ppt not observed</td>
<td>+</td>
</tr>
<tr>
<td>6</td>
<td>Steroid test</td>
<td>Sandwiski’s test</td>
<td>Ring not formed</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>Tannins test</td>
<td>Tannins test</td>
<td>Gelatin test</td>
<td>+</td>
</tr>
<tr>
<td>8</td>
<td>Proteins and amino acids</td>
<td>Xanthoproteic test</td>
<td>Reddish black not seen</td>
<td>+</td>
</tr>
</tbody>
</table>

+ sign indicates the presence and - sign indicates absence.
Table 2: Antimicrobial activity methanolic extract of dried fruits peel of *Citrus limon* (family-Rutaceae) using disc diffusion assay

<table>
<thead>
<tr>
<th>Name of the compounds and their concentration</th>
<th>Antibacterial activity diameter of inhibition zone (mm)</th>
<th>Antifungal activity diameter of inhibition zone (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Standard Ciprofloxacin (20 µl)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Standard Fluconazole (20 µl)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Plant extract (25 µl)</td>
<td>13.5</td>
<td>10.3</td>
</tr>
<tr>
<td>Plant extract (50 µl)</td>
<td>18.35</td>
<td>16.5</td>
</tr>
<tr>
<td>Plant extract (100 µl)*</td>
<td><strong>20.6</strong></td>
<td><strong>22.2</strong></td>
</tr>
<tr>
<td><em>significant activity</em></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Fig. 1: Zone of inhibition vs concentration

**DISCUSSION**

Antimicrobial activity of dried fruits peel of *Citrus limon* has been evaluated. Phyto-constituents present in plants namely alkaloids, saponin, sterols, terpenoids are having an exciting set of circumstances that makes it possible to do something for more extensive of modern therapies against a wide range of microorganisms. The present study was done against the variety of Gram-positive, Gram-negative bacteria, and fungal strains were selected for screening antimicrobial impact of the extracts to see the antimicrobial spectrum. Results of this study shows that the methanolic extracts of the peel of the fruit of *Citrus limon* were highly successful in producing the desired result against most of the Gram-positive bacteria, Gram-negative bacteria, and fungal strains in agar well diffusion method.

Phytochemical constituents of plants such as tannins, alkaloids, flavonoids, phenolic compounds and several other aromatic compounds are secondary metabolites which can be used in achieving a defence mechanism against plundering by many micro-organisms.

The practical exhibition and explanation of antimicrobial activity against both Gram-positive and Gram-negative bacteria and on various fungal strains may be an indication of something presence of broad spectrum antibiotic compounds in the extracts which is shown in fig. 3. The 100µg/ml concentration of dried fruits peel of *Citrus limon* have an influencing antimicrobial activity.

**CONCLUSION**

Medicinal Plant in plant kingdom are the important source of drug and plays a great role for the health of individual and communities. Many plants have been tested for the presence of compounds with therapeutic activity. Therefore, it is absolutely necessary to evaluate the antimicrobial activity of fruits peel of *Citrus limon*.

From the study, the antibacterial activity of the peel of the dried fruit of *Citrus limon* was judged by using disk diffusion method. The microorganisms that have been selected for study were Gram-positive, *S. aureus* and Gram-negative *E. coli* and fungus like *C. albicans*, and *T. rubrum*. Study of this micro-organism was done as they have the capacity of rapidly developed antibiotic resistance as antibiotic use increases.

After doing the study of a methanolic extract of *Citrus limon* peel showed the presence of various phytochemicals of which include alkaloids, saponin, sterols, Steroids, and terpenoids.

At the end of the conclusion we know about the significant antimicrobial activity of the plant extracts when estimated with standards. So, after lots test, decision and thoughts it was found that the minimum inhibitory concentration of the plant extracts was less than 100µg/ml and therefore the plant was considered to have a very good antimicrobial activity.

However, further studies are definitely required to put light on the unknown biological activities of *Citrus lemon* peel and its effect on living organisms, tissues or cells against various diseases.

**CONFLICT OF INTERESTS**

Declare none

**REFERENCES**


**How to cite this article**