

## QUALITATIVE AND QUANTITATIVE PHYTOCHEMICAL ANALYSIS AND DPPH RADICAL SCAVENGING ACTIVITY OF DIFFERENT EXTRACTS OF FLOWER OF *BAUHINIA ACUMINATA* LINN

SREEJA SANJEEV, REGI RAPHAEL K.

Department of Botany, St. Mary's College, Thrissur  
Email: puthumanasandeep@gmail.com

Received: 20 Aug 2017, Revised and Accepted: 13 Oct 2017

### ABSTRACT

**Objective:** *Bauhinia acuminata* L (Fabaceae) is a species of flowering shrub native to tropical southeastern Asia. The flower of *B. acuminata* are used against various skin diseases, worms, tumours and diabetes. The Indian vaidyas recommended a remedy to treat severe biliousness.

The qualitative and quantitative estimation of phytochemical constituents in various extracts and the antioxidant potential of the flower of *Bauhinia acuminata* were evaluated.

**Methods:** All the analysis was done based on standard protocols.

**Results:** The phytochemical screening reveals the presence of various primary and secondary metabolites like sugar, carbohydrate, amino acid, fat, quinone, steroids, phenol, saponin, alkaloids, and acid content. In quantitative analysis, the important secondary metabolites such as alkaloids and phenolic compounds were estimated in all the extracts.

The ethanol and chloroform extract produced significant antioxidant properties in a dose-dependent manner. DPPH free radical scavenging assay of ethanol and chloroform extracts of flower exhibited IC<sub>50</sub> values of 24.44±1.201µg/ml and 196.68±0.456µg/ml respectively. At 1000 µg/ml concentration both extracts shows maximum radical scavenging activity (98.97% and 85.67 %).

**Conclusion:** The findings of this study indicate that this plant is medicinal with the prominent antioxidant property.

**Keywords:** *B. acuminata*, Photochemicals, Antioxidant, DPPH radical scavenging

© 2017 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open-access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)  
DOI: <http://dx.doi.org/10.22159/ijcpr.2017v9i6.23423>

### INTRODUCTION

Nature has provided a complete storehouse of remedies to cure all ailments of humankind [1]. Traditional medicine use is a common practice in developed and developing countries at the primary healthcare level [2]. Herbal medicines are prepared from various plant parts like leaves, stem, roots, barks and seeds, which usually contain many bioactive compounds and used primarily for treating mild or chronic ailments. Due to the increasing demand in the field of herbal medicines, it has become necessary and pertinent to probe into the area of systematic knowledge about herbal drugs. There is a need for the application of this knowledge in authentication, detailed study and practical utilization of crude drugs [3].

*Bauhinia acuminata* L, belongs to the family Fabaceae is a species of flowering shrub native to tropical southeastern Asia. In Malaysia and Indonesia the plant is used in the treatment of common cold and cough [4]. While in India the leaves and bark of this plant are used for treating asthma [5]. Moreover, the leaf of *B. acuminata* is used to treat bladder stone, venereal diseases, leprosy, asthma and digestive diseases. Different part of this plant such as bark, leaves, stem, flowers and roots have been used in traditional medicine [6]. The plant was also used as febrifuge, vermifuge, anticonvulsant and against chicken pox, guinea worm and black quarter [7-8]. Though different parts of this plant were reported to possess good medicinal properties [9], there is no published study particularly on the phytochemical and antioxidant activity of *B. acuminata* flower. Hence, the focus of this study was to investigate qualitative and quantitative phytochemical analysis and antioxidant activities in flower extracts of *B. acuminata*.

### MATERIALS AND METHODS

#### Collection and identification of plant materials

The plant *B. acuminata* were collected from Edapal, Malappuram District of Kerala, India. Taxonomic identification with Flora of Presidency of Madras by JS Gamble [10].

#### Preparation of extracts

Flower of the plant was shade dried for several days. The dried plant material was ground to a coarse powder and 50 gm of the powdered plant material was soaked in solvents of increasing polarity starting petroleum ether, chloroform, ethyl acetate, ethanol and distilled water (1:5) for 72 h [11]. The solvent was then removed by rotary evaporation. Each residue was weighed and the yield percentage [12] was determined. The dried extract was stored in the refrigerator for further studies.

#### Qualitative phytochemical analysis

The phytochemical screening of the plant extract in various solvents was performed by using standard protocol given by Harbone [13].

#### Quantitative phytochemical analysis

##### Determination of total phenolic content

Total phenolic content analysis was performed by the method of Malick *et al.* [14]. Dissolve the test extract in the concentration of 1 mg/1 ml and make up this test solution up to 3 ml with distilled water. 0.5 ml Folin-Ciocalteu reagent and 2 ml of 20% Na<sub>2</sub>CO<sub>3</sub> were respectively added. The samples were thereafter incubated in boiling water bath exactly for one minute. The absorbance was measured by using spectrophotometer at 650 nm against reagent as blank.

A standard calibration plot was generated at 650 nm using known concentrations of catechol. The concentrations of phenols in the test samples were calculated from the calibration plot and expressed as catechol equivalent of phenol/g of sample. Each extract was assayed in triplicates.

##### Determination of total alkaloid content [15].

The plant extract (1 mg/1 ml) was dissolved in 2N HCl and then filtered. 1 ml of this solution was transferred to a separating test

tube, and then 5 ml of Bromocresol green solution along with 5 ml of phosphate buffer with neutral PH were added. The mixture was shaken and the complex formed was extracted with chloroform by vigorous shaking. The extract was collected in a 10 ml volumetric flask and diluted with chloroform. Now, the absorbance of the complex in chloroform was measured at 470 nm against the blank prepared as above but without atropine. The whole experiment was conducted in triplicates.

A standard calibration plot was generated at 470 nm using known concentrations of atropine. The concentrations of alkaloids in the test samples were calculated from the calibration plot and expressed as atropine equivalent of alkaloid/g of sample.

#### Antioxidant property screening

##### DPPH radical scavenging assay

The free radical scavenging activity of the plant extracts assessed based on the radical scavenging effect of the stable 1, 1-diphenyl-2-picrylhydrazyl (DPPH), by a method given by Braca *et al.*, [16]. The diluted test solutions (10 µg/ml-1000 µg/ml concentration) and 6.34 µM solution of DPPH were prepared in methanol, and 100µl test, along with 100µl DPPH solution and 800µl of methanol was taken in a test tube and mixed well. These solution mixtures were kept in dark for 20 min and optical density was measured at 517 nm using Cecil-Elect Spectrophotometer. Methanol (900µl) with DPPH solution (6.34µM, 100µl) taken as control and methanol as blank. The optical density recorded and percentage of inhibition calculated using the formula given below [17].

Percentage (%) inhibition of DPPH activity =  $A-B/A \times 100$

Where A = optical density of the control and B = optical density of the sample.

Sample concentration providing 50% inhibition (IC<sub>50</sub>) was calculated from the graph plotting inhibition percentage against extract concentration.

##### Statistical analysis

All the analysis were performed in triplicate and the results were statistically analyzed and expressed as mean (n=3)±standard deviation.

#### RESULTS AND DISCUSSION

##### Yield of extract

Comparatively, flower ethanol extract exhibited higher extraction yield. The extraction ability of different solvents for recovering extractable components from flower followed the order: ethanol (2.00g)>chloroform (1.26g)>petroleum ether (1.13g)>ethyl acetate (1.10g)>distilled water (1.02g). The variation in yield may be due to the polarity of the solvents used in the extraction process.

##### Preliminary phytochemical screening

The phytochemical analysis is of paramount importance in identifying a new source of therapeutically and industrially valuable compounds having medicinal plants have been chemically investigated [18]. The preliminary Phytochemical screening showed the presence of primary and secondary metabolites like sugar, carbohydrate, amino acid, fat, quinone, steroids, phenol, saponin, alkaloids, and acid content (table 1).

**Table 1: Phytochemical screening of flower of *B. acuminata* in different solvents**

Primary and secondary metabolites	Test/Reagent	Petroleum ether	chloroform	Ethyl acetate	Ethanol	Distilled water
Carbohydrate	Molisch's reagent	+	+	+	+	+
Starch	Iodine test	-	-	-	-	-
Sugar	Benedict's test	-	+	+	+	+
Ketose	Seliwanoff's test	-	-	-	-	-
Proteins	Biuret test	-	-	-	-	-
Amino acid	Ninhydrin test	+	-	-	-	-
Fat	Filter paper test	+	+	+	+	+
Quinone	H <sub>2</sub> SO <sub>4</sub> test	-	-	-	-	+
Cardiac glycoside	Kellar-killani test	-	-	-	-	-
Steroids	Salkowski test	-	-	+	-	-
Flavonoids	Fluorescent test	-	-	-	-	-
Phenols	Folin test	+	+	+	+	+
Saponins	Foam test	-	-	-	-	+
Alkaloids	Dragendroff's reagent	+	+	+	+	+
Tannin	FeCl <sub>3</sub> test	-	-	-	-	-
Terpenoids	Salkowski test	-	-	-	-	-
Acid	NaHCO <sub>3</sub> test	+	-	+	+	+

+indicates the presence of metabolite,-indicate the absence of metabolite

#### Quantitative estimation of phenol and alkaloid compounds

Secondary metabolites analysis is necessary for extraction, purification, separation, crystallization, identification of various phytochemicals. Calibration curve of catechol and atropine is given in fig. 1 and fig. 2. Phenol and alkaloid are present in all the samples extracted by five different solvents. The total phenol content was highest with (1.26±0.01) catechol/g in ethanol extract followed by (1.17±0.02) catechol/g of ethyl acetate extract. Phenols are antiseptic and reduce inflammation when taken internally. These bioactive agents have an irritant effect when applied to the skin. Above of all, phenols have a high affinity to chelate metals and scavenge the free radicals in cells [19]. Polyphenols act as antioxidants, which protect cells and body chemicals against damage, caused by free radicals and reactive atoms that contribute to tissue damage in the body. It has been reported that these compounds deactivate the substances that promote the growth of tumours [20]. Consumption of diets rich in plant polyphenols offers

protection against the development of cancer, cardiovascular diseases, diabetes, osteoporosis and neurodegenerative diseases.

Alkaloids protect against chronic diseases [21] and earlier recorded that bitter leaf contains an alkaloid that is capable of reducing headaches associated with hypertension. Alkaloids are a diverse group of secondary metabolites found to have antimicrobial activity by inhibiting DNA topoisomerase [22]. Alkaloid constituents in flower showed that the chloroform extract had higher amounts (0.72±0.03) atropine/g of the extract followed by (0.66±0.02) atropine/g of the ethyl acetate extract. The results are tabulated in table 2.

#### Antioxidant property of flower

Antioxidant compounds may function as free radical scavengers, initiator of the complexes of pro-oxidant metals, reducing agents and quenchers of singlet oxygen formation [23]. Therefore, the importance of the search for natural antioxidants has increased in the recent years so many researchers focused the same [24].

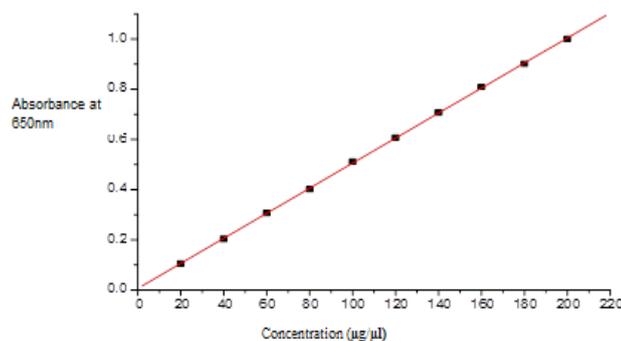


Fig. 1: Catechol calibration curve

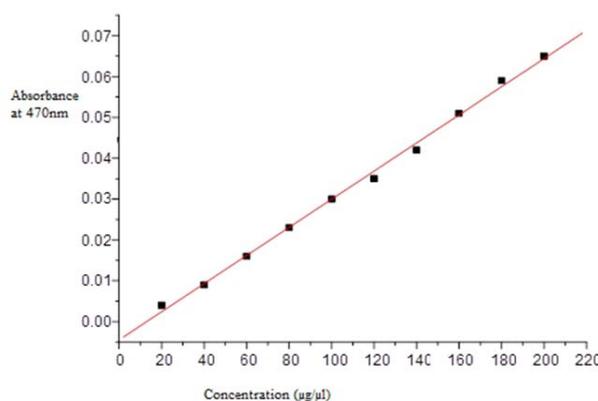


Fig. 2: Atropine calibration curve

Table 2: Quantitative estimation of total phenol and alkaloid content of different extracts

Extracts	Phenol (Catechol/gm)	Alkaloid (Atropine/gm)
Petroleum ether	0.13±0.03	0.35±0.01
Chloroform	1.05±0.01	0.72±0.03
Ethyl acetate	1.17±0.02	0.66±0.02
Ethanol	1.26±0.01	0.41±0.03
Distilled water	0.92±0.01	0.37±0.02

Values are means of three independent determinations±standard deviations (SD).

#### DPPH radical scavenging assay

Among the five different extracts, ethanol and chloroform extracts showed higher phenol and alkaloid contents respectively. Therefore the free radical scavenging potential of these two extracts tested by DPPH assay is given in the table 3. Reduction of the DPPH radicals was observed by a decrease in absorbance where a change in the color to yellow denotes quenching of the free radicals by the plant extracts. The analysis of the radical scavenging activity of the extracts increases with increase in concentration. Ethanol extract of the flower was found

to have the most potent antioxidant property with IC<sub>50</sub> value of 24.44±1.201µg/ml than chloroform extract with 196.68±0.456 µg/ml. The high activity of ethanol extract of the flower is generally attributed to the presence of alkaloids and phenols, as the majority of active antioxidant compounds are observed in these classes of phytochemical compounds. The secondary metabolites phenol and alkaloids which carry a major importance to increase an antioxidant potential. Information obtained from these studies can be used as markers in the identification and standardization of this plant as an herbal remedy and also towards monograph development of the plant.

Table 3: DPPH scavenging activities of extracts of flower (values represent mean±SD, n=3)

S. No.	Concentration (µg/ml)	Percentage of inhibition	
		Ethanol	Chloroform
1	10	45.20±0.644	15.52±0.898
2	15	48.32±0.597	17.24±0.941
3	25	55.42±1.325	25.59±0.716
4	50	63.57±0.580	29.70±1.290
5	75	76.68±1.154	31.56±1.087
6	100	87.13±1.038	38.37±1.402
7	250	95.20±0.689	59.17±0.691
8	500	96.78±0.395	74.21±0.907
9	750	97.79±0.358	81.35±1.026
10	1000	98.97±0.740	85.67±0.544
	IC 50	24.44±1.201	196.68±0.456

**CONCLUSION**

Phytochemical studies portray the presence of several biologically active secondary metabolites such as phenol and alkaloids in the bark of *B. acuminata* for the first time. The antioxidant efficacy of ethyl acetate extract is very high with IC<sub>50</sub> value of 42.62±0.859 µg/ml, indicates that this plant can have great scope for isolation and identification of important antioxidant molecules which can be formulated to make antioxidant dosage forms. Significant correlations were found between the antioxidant capacities and phenolic contents indicating that phenolic compounds are the major contributors to antioxidant capacity. On top of that, these natural antioxidants can have potential advantages among various diseases with oxidative stress. So, further study is necessary to get maximum benefit from this plant.

**CONFLICT OF INTERESTS**

Declared none

**REFERENCES**

- Kokate CB, Purohit AP, Gokhale SB. Pharmacognosy. 18<sup>th</sup> edition. Pune India: Nirali Publication; 2002.
- Essawi T, Srouf M. Screening of some Palestinian medicinal plants for antibacterial activity. J Ethnopharmacol 2000;70:343-9.
- Kirikar KR, Basu BD. Indian medicinal plants. Vol. 1. Dehar Dun, India: International books Distributers; 1980.
- Timothy Johnson. CRC ethnobotany desk reference, CRC Press: LLC Boca Taton; 1999. p. 110.
- Khare CP. Indian medicinal plants: an illustrated dictionary. Springer-Verlag Berlin; 2007. p. 85.
- Reyad-Ul-Ferdous M, Azam MG, Hossain MD. Phytochemical screening, in vitro membrane stabilizing activities and thrombolytic activities of *Lophopetalum javanicum*. Int J Pharm Sci Res 2014;5:350-0.
- Fordin DG. History and concept of big plant genera. Taxon 2004;53:753-76.
- Singh S, Singh SK, Yadav AA. Review on cassia speices; pharmacological, traditional and medicinal aspects in various countries. Am J Phytomed Clin Ther 2013;1:291-312.
- Mohammad FK, Rabeya IS, Ridwan BR, Mohammad AR. Evaluation of cytotoxicity of *beta vulgaris* and membrane stabilizing activities. Biomed Pharmacol J 2014;17:99-101.
- Gamble JS. Flora of presidency of Madras. Aldard and son publishing company Ltd; 2004.
- Taleb-Contini SH, Salvador MJ, Balanco JMF, Albuquerque S, De Oliveria DCR. Antiprotozoal effect of crude extracts and flavonoids isolated from *Chromolaena hirsute* (Asteraceae). Phyto Res 2004;18:250-4.
- Patil UH, Gaikwad DK. Phytochemical evaluation and bactericidal potential of *Terminalia arjuna* stem bark. Int J Pharm Sci Res 2010;2:614-9.
- Harbone JB. Phytochemical methods, a guide to modern techniques of plant analysis. India: Springer pvt Hd; 1998.
- Malick EP, Singh MB. Plant enzymology and Hittoenzymology, Kalyani Publishers, New Delhi; 1980.
- Fadhil S, Monsef H. Spectrophotometric determination of total alkaloids in *Peganumharmala* L. using Bromocresol green. Res J Phytochem 2007;1:79-82.
- Braca A, Sortino C, Politi M. Anti-oxidant activity of flavonoids from *Licanialicaniaeflora*. J Ethnopharmacol 2002;79:379-81.
- Bors W, Saran M, Elstner EF. Screening for plant anti-oxidants. In: Linskens HF, Jackson JF. eds. Modern Methods of Plant Analysis-Plant Toxin Analysis-New Series, Berlin: Springer; 1992;13:277-95.
- Ambasta SP, Ramachandran K, Kashyapa K, Chand R. Useful plants of India. Publication and information directorate. Council of Scientific and Industrial Research, New Delhi; 1986. p. 4337.
- Michalak A. Phenolic compounds and their antioxidant activity in plant growing under heavy metal stress. Polish J Environ Studies 2006;15:523-30.
- Rahaman Onike. Phytochemical Screening Tests and Medicinal Values of Plants Active Properties; 2010.
- Ayitey Smith E, Addae Mensah I. Phytochemical, nutritional and medical properties of some leafy vegetables consumed by Edo people of Nigeria. W Afr J Pharmacol Drug Res 1977;4:7-8.
- Bonjean K, De Pauw-Gillet MC, Defresne MP, Colson P, Houssier C, Dassonneville L, et al. The DNA intercalating alkaloid cryptolepine interferes with topoisomerase II and inhibits primarily DNA synthesis in B16 melanoma cells. J Ethnopharmacol 1998;69:241-6.
- Andlauer W, Furst P. Antioxidative power of phytochemicals with special reference to cereals. Cereal Foods World 1998;43:356-9.
- Jayaprakasha GK, Selvi T, Sakariah KK. Antibacterial and antioxidant activities of grape (*Vitisvinifera*) seed extract. Food Res Int 2003;36:117-22.