

Original Article

ANTIOXIDANT AND ANTI-INFLAMMATORY POTENTIAL OF QUCERTIN

MUTHUKALA B, SIVAKUMARI K*, ASHOK K

Department of Advanced Zoology and Biotechnology, Presidency College, Chennai 600005

Email: dr.sivakumari@rediffmail.com

Received: 02 May 2015, Revised and Accepted: 05 Jun 2015

ABSTRACT

Objective: In the present study the antioxidant potential and anti-inflammatory effect of quercetin compound was assessed.

Methods: The antioxidant potential of the drug was determined by ORAC assay and DPPH assay. Likewise for anti-inflammatory studies, RBC's were collected from healthy volunteers and the hemo protective activity of the drug was carried out at various concentrations.

Results: The results showed that quercetin has a positive effect on both the parameters. The probable reasons governing the facts are discussed in the light of previous literature.

Conclusion: The results show that quercetin seems to be a good replacement for chemical therapeutic drugs as it has antioxidative and anti-inflammatory properties.

Keywords: Quercetin, Antioxidant, Anti-inflammatory, Hemoprotective.

INTRODUCTION

Flavonoids, a large group of natural polyphenolic compounds, are powerful antioxidants found in various fruits, vegetables, tea, red wine, and medicinal herbs. Flavonoids can scavenge free radicals and other oxidizing intermediates because of their phenolic hydroxyl groups and thus contribute to the counteraction of body against a great variety of diseases [1].

Quercetin is a unique bioflavonoid that has been extensively studied by researchers over the past 30 years. Bioflavonoids were first discovered by Nobel Prize laureate Albert Szent Gyorgyi in the year 1930. Flavonoids belong to a group of natural substances with variable phenolic structure and are found in the fruits, vegetables, grains, bark roots, stem, flowers, tea and wine [2]. These natural products were known for their beneficial effects on health long before flavonoids were isolated as the effective compounds. More than 4000 varieties of flavonoids have been identified, many of which are responsible for their attractive colors of flowers, fruits and leaves [3].

In view of this, the present study has been taken to determine the antioxidant potential and anti-inflammatory effects of quercetin, to elucidate it as a potential drug for various ailments.

MATERIALS AND METHODS

Quercetin compound was purchased from Sigma Aldrich, USA and used for the present study.

In vitro antioxidant activity

ORAC assay

The antioxidant potential of quercetin was determined by ORAC assay following the methods of Huang *et al.* (2002, 2005) [4, 5].

In vitro Anti-inflammatory activity

Membrane stabilization assay

Quercetin was subjected to human red blood cell (HRBC) membrane stabilization method to study the anti-inflammatory activity according to the method of Gandhidasan *et al.* (1991) [6].

RESULTS

In vitro antioxidant activity

Table-1 presents the data on antioxidant potential of quercetin when tested with ORAC assay. The data reveals that rutin has antioxidant potential, as the value of Net Relative Fluorescence Unit (NRFU)

increased with an increase in the concentration of the drug. The NRFU values are Maximum at 100 μ M (75.77+0.047) and decreases as the concentration is increased further.

Similarly when DPPH assay of quercetin was carried out, among all the concentrations of rutin, 12.5 μ M showed the maximum % DPPH inhibition (Table-2).

The results indicate that quercetin has antioxidant potential and can be used as a drug for combating various ailments.

Table 1: ORAC Assay for Quercetin in different concentrations

| S. No. | Concentration of quercetin (μ M) | Net relative fluorescence unit |
|--------|---------------------------------------|--------------------------------|
| 1 | 12.5 | 49.7+0.808 |
| 2 | 25 | 69.44+0.074 |
| 3 | 50 | 73.32+0.038 |
| 4 | 100 | 75.77+0.047 |
| 5 | 200 | 69.7+0.027 |

Values are mean+SE of six individual observations.

Table 2: DPPH Assay of Quercetin

| S. No. | Concentration of Quercetin (μ M) | % DPPH inhibition |
|--------|---------------------------------------|-------------------|
| 1 | 12.5 | 0.294+0.003 |
| 2 | 25 | 0.148+0.047 |
| 3 | 50 | 0.079+0.041 |
| 4 | 100 | 0.077+0.160 |
| 5 | 200 | 0.070+0.050 |

Values are mean+SE of six individual observations.

Table 3: Haemoprotective activity of Quercetin

| S. No. | Concentration of Quercetin (μ M) | % Protection |
|--------|---------------------------------------|--------------|
| 1 | 62.5 | 97.348+0.370 |
| 2 | 125 | 62.219+0.907 |
| 3 | 250 | 97.028+0.140 |
| 4 | 500 | 96.896+0.252 |
| 5 | 1000 | 96.967+0.121 |

Values are mean+SE of six individual observations.

Haemoprotection of quercetin

When quercetin was tested for its hemoprotective activity, the per cent hemoprotection was directly proportional to concentration; the values being 97.348±0.370 at 62.5 µM and 96.967±0.121 at 1000 µM (table 3). The results thus reveal us that quercetin has hemoprotective effect.

DISCUSSION

According to Meena *et al.* (2008) [7], more than 2000 flavonoids have been reported among woody and non-woody plants [8]. Biosynthesis, isolation techniques and preparative chromatography [9], TLC, UV and IR spectral studies have provided new dimensions to the chemistry of flavonoids to such an extent that their presence have become important taxonomically [10]. Presence of flavonoids has been reported from many plant species like *Lycium barbarum* [11]; *Passiflora plamer* [12]; *Cassia angustifolia* [13]; *Jatropha curcas* L.[14].

Quercetin has been reported from many plant species like *Cicer arietinum* Linn. [15] and *Acacia catechu* [16]. As mentioned earlier, since quercetin has anti-inflammatory, antioxidant and anticancer properties, isolation and extraction of this compound *in vivo* (leaf, stem, fruit, root) and *in vitro* callus from *Citrullus colocynthis* and its enhancement by addition of elicitors in culture, can be exploited further for largescale production of this medicinally important compound [7].

The antioxidant activity of quercetin is well known as it possesses a suitable structure for free radical scavenging and ion chelation. However, the results of the TBARS assay indicate that unlike galangenin, both analogues do not show significant antioxidant activity. This may be due to the presence of an additional 5-OH group in galangenin generating a structure for effectively scavenging the free radical. Absence of 5-OH group in Q-Cl and Q-OCH₃ leads to a drop in the antioxidant activity. Quercetin is also known to chelate iron, which is responsible for the production of free radicals. The chelation involves 3', 4'-hydroxy groups thus highlighting the importance of the catechol-like moiety [17].

CONCLUSION

The results show that quercetin seems to be a good replacement for chemical therapeutic drugs as it has antioxidative and anti-inflammatory properties.

ACKNOWLEDGMENT

The authors are thankful to V Clin Bio Labs (P) Ltd, Sri Ramachandra University, Porur, Chennai for the technical support.

CONFLICT OF INTERESTS

Declared None.

REFERENCES

1. Roeder E. Medicinal plants in Europe containing pyrrolizidine alkaloids. *Pharmazie* 1995;50:83-98.
2. Middleton EJ. Effect of plant flavonoids on immune and inflammatory cell functions. *Adv Exp Med Biol* 1998;439:175-82.
3. De Groot H, Rauen U. Tissue injury by reactive oxygen species and the protective effects of flavonoids. *Fundam Clin Pharmacol* 1998;12:249-55.
4. Huang D, Ou B, Hampsch-Woodill M, Flanagan J, Prior R. High-throughput assay of oxygen radical absorbance capacity (ORAC) using a multichannel liquid handling system coupled with a microplate fluorescence reader in 96-well format. *J Agric Food Chem* 2002;50:4437-44.
5. Huang D, Ou B, Prior RL. The chemistry behind antioxidant capacity assays. *J Agric Food Chem* 2005;53:1841-56.
6. Gandhidasan R, Thamarachelvan A, Baburaj S. Anti-inflammatory action of *Lannae coromandelica* by HRBC membrane stabilization. *Fitoterapia* 1999;62:81-3.
7. Meena MC, Patini V. Isolation and identification of flavonoid "Quercetin" from *Citrullus colocynthis* (Linn.) Schrad. *Asian J Exp Sci* 2008;22(1):137-42.
8. Harborne JB. Plant phenolics. In: secondary plant products (Eds) Bell EA, Charlewood BV, Springer Verlag Berlin; 1981. p. 320.
9. Casteel HW, Wender SM. Identification of flavonoid compounds, R_f values and colour tests. *Anal Chem* 1953;25:508.
10. Smith EB. In: Prospective in phytochemistry (Eds) Harborne JB, Swain T. Academic Press; London: 1969.
11. Harsh ML, Nag TN, Jain S. Arid zone plants of Rajasthan a source of antimicrobials. *Comp Physiol Ecol* 1983;8:129-31.
12. Ulubelen A, Mabry JJ, Dellamonicas G, Chopin J. Flavonoids of *Passiflora plamer*. *J Nat Prod* 1984;47:384-5.
13. Goswami A, Reddi A. Antimicrobial activity of flavonoids of medicinally important plant *Cassia angustifolia* *in vivo* and *in vitro*. *J Phytol Res* 2004;17:179-81.
14. Saxena S, Sharma R, Rajore S, Batra A. Isolation and identification of flavonoid "Vitexin" from *Jatropha curcas* L. *J Plant Sci Res* 2005;21:116-7.
15. Joshi RS. Biosynthesis of primary and secondary products from *in vivo* and *in vitro* tissue cultures of some medicinal plants. Ph. D. Thesis, University of Rajasthan, Jaipur, India; 1985.
16. Jain R, Patni V, Arora DK. Isolation and identification of flavonoid "quercetin" from *Acacia catechu* (L. f.) Willd-A Katha yielding plant. *J Phytol Res* 2007;20:43-5.
17. Ferrali C, Signorini B, Caciotti. Protection against oxidative damage of erythrocyte membrane by the flavonoid quercetin and its relation to iron chelating activity. *FEBS Lett* 1997;416:123-9.