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**Original Article** 

# PRELIMINARY PHYTOCHEMICAL SCREENING AND ANTIMICROBIAL ACTIVITY OF ETHANOLIC EXTRACT OF STEMS OF *IPOMOEA AQUATICA* FORSK. (FAMILY-CONVOLVULACEAE)

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# ABSTRACT

**Objective:** To evaluate the antimicrobial activity ethanolic extracts of stems of the plant *Ipomoea aquatica* Forsk. (family-Convolvulaceae) along with preliminary phytochemical analysis.

**Methods:** The ethanolic extract of stems of the plant *Ipomoea aquatica* Forsk. (family-Convolvulaceae) were prepared and analyzed for phytochemical constituents using standard methods. The antimicrobial activity of the plant extracts was examined against 2 bacterial strains among one is gram positive and other is gram negative and 2 fungus using agar well diffusion method.

**Results:** The present experiment shows the phytochemical analysis, antimicrobial activity of the ethanolic extract of stems of the plant *Ipomoea aquatic* Forsk. (Family-Convolvulaceae). Various phytochemical analysis revealed the presence of alkaloids, saponin, flavonoids, carbohydrates, glycosides, Steroids, proteins and amino acids and tannins. All the extracts of the plant showed antimicrobial activity against most of the test organisms.

**Conclusion:** The present study concluded that ethanolic extract of stems of the plant *Ipomoea aquatica* Forsk. Contain the high presence of phytochemical. The ethanolic extract of the plant was found to possess promising antimicrobial activity when compared with the standards.

Keywords: Ipomoea aquatic Forsk., Antimicrobial, Zone of Inhibition, Agar disc diffusion method

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# INTRODUCTION

Knowledge of herbs has been handed down from generation to generation for thousands of years. The revival of interest in natural drugs started in last decade mainly because of the widespread belief that green medicine is healthier than synthetic products. In the recent past, there has been a tremendous increase in the use of plant-based health products in developing as well as developed countries resulting in an exponential growth of herbal products globally. According to WHO, about80% of the population in the world relay on the traditional medicine for the treatment of various diseases [1].

In the present era of drug development and discovery of newer drug molecules, many plant products are evaluated on the basis of their traditional uses. In this regard, one of the many plants which are being evaluated for their therapeutic efficacies is *Ipomoeaaquatica* (I. aquatica) (Convolvulaceae) which is a perennial herb found throughout India, Ceylon, Tropical Asia, Africa, and Australia [2].

This plant is grown as an aquatic plant which grows abundantly in marshy areas. Various parts of the *l. aquatica* plant are used medicinally in Southeastern Asia and reported to be useful for the treatment of high blood pressure, as an emetic in the treatment of opium and arsenic poisoning [3]. The dried juice has reported to be a purgative, while the leaves and stems possess cooling action. Moreover, it is also traditionally used in the treatment of nervous and general debility, piles, worm infections, leucoderma, leprosy, jaundice and liver complaints [4].

Very few studies have been done in this plant. This includes the inhibition of prostaglandin synthesis, eye diseases, constipation and hypoglycemic effects [5]. Phytochemical investigations of this plant have revealed the presence of carotenes such as  $\beta$ -carotene, cryptoxanthin, lutein, lutein epoxide, violoxanthin and neoxanthin, flavonoids such as mycertin, quercetin, luteolin and apigenin and some alkaloids [6].

In the Indian system of traditional medicine (Ayurveda) it is recommended to consume I. aquatica to mitigate disorders like jaundice. The effectiveness of these plant products from traditional claims must be proved to help develop novel drugs acting against these disorders. Nevertheless, no literature was found on the hepato-protective action of this plant. Keeping this in view, the present study was aimed to evaluate the hepato-protective activity of I. aquatica on Thioacetamide-induced liver damage in rats [7].

Medicinal plants have been tested for various kinds of activities such as antimicrobial, hypoglycemic, antihelmintic, hepatoprotective, antioxidant, analgesic, antipyretic activities. Antipyretic, antileishmanial and insecticidal activities. Medicinal plants such as Ocinumgratissimum and Eugenia uniflora are rich in volatile oils. It is also reported that they contain 75% thymol which has an antimicrobial effect against Staphylococcus sp., Escherichia coli and Shigella sp. They are also used in the treatments like diarrhea human ear infection [8].

Herbal medicines also offer therapeutics for age-related disorders like memory loss, osteoporosis, immune disorders, etc. for which no modern medicine is available. The chemical constituents present in them are a part of the physiological functions of living flora and hence they are believed to have better compatibility with the human body [9].

The therapeutic plants continually collaborate with the quickly changing and possibly harming outside ecological variables. In spite of the fact that living beings without versatility, plants have advanced expand elective safeguard methodologies, which include a huge assortment of compound metabolites as devices to conquer stretch conditions. The capacity of plants to complete combinatorial chemistry by mixing, coordinating and developing the quality items required for secondary metabolite biosynthetic pathways, makes a boundless pool of chemicals, which people have abused to their advantage. Consequently, the utilization of plants by people in both conventional and present day therapeutic frameworks, in this manner, to a great extent utilize this standard [10].

*Ipomoea aquatica* Forsk, commonly called water spinach, belongs to the family Convolvulaceae is a perennial herb and tender, trailing or floating perennial aquatic plant found on moist soil along the margins of fresh water. It is also commonly used as a green leafy vegetable in rural India Studies with Ipomoea aquatica Forsk revealed that the inhibition of prostaglandin synthesis, effects on liver disease and constipation [11].

So the point of this present study is to recognize the phytochemical of *Ipomoea aquatica* Forsk. and to decide the antimicrobial effects of the stem extract on *E. coli, S. aureus, Candida albicans* and *Trichophyton rubrum.* 

### MATERIALS AND METHODS

#### **Collection of plant materials**

*Ipomoea aquatica* Forsk. was collected from Azara of Kamrup district of Assam in October 2016 and the plant was thoroughly washed with water; roots and leaves were discarded and the stems were dried in shade for 2 w and finally dried in a thermostatic oven at a considerably low temperature not exceeding 30 °C for 24 h. The plant was authenticated by Prof. Dr. G. C. Sharma, Department of Botany, Gauhati University. A voucher specimen (Acc. No.18210, Dated: 28.10.2016) was kept in Department of Botany, Gauhati University for future reference.

#### **Chemicals and reagents**

Ethanol (ET), Petroleum Ether(PE),dimethyl sulfoxide (DMSO), hydrochloric acid, Dragondorff reagent, Mayer's reagent, Wagner's reagent, Benedict's reagent, sulphuric acid, lead acetate, Molisch's reagent, Fehling solution A and B, sodium citrate, copper sulphate, ferric chloride, sodium hydroxide, glacial acetic acid, benzene, chloroform, ammonia, nitric acid, dimethyl sulfoxide (DMSO), potassium nitrite, gelatine, Beef extract, Peptone and agar. All the chemicals and solvents used were of standard analytical grades.

#### Preparation of plant extracts

*I. aquatica* was carried out using ethanol (99.5%) by maceration method at room temperature. 100 gm of the ground sample was taken in the round bottom flask. To that above flask, 500 ml of ethanol was poured and was kept aside under cold maceration technique for about 7 d with intermittent shaking. The liquid extract was then distilled off and the remaining solvent was allowed to evaporate in a water bath at a constant temperature. Then it was poured in a tarred Petri dish was dried for several days. The concentrated extract was then weighed and the percentage yield was calculated and finally stored in airtight container at 4 °C. Thus total ethanolic extract is obtained which was used for investigation. The percent yield of ethanolic extract was then calculated out.

# Preparation of extract/drug stock solution

The stock solution of Ipomoea aquatica stem extract was prepared on each occasion by careful weighing and dissolving in a suitable volume of Dimethylsulphoxide (DMSO) to get a concentration of 100 mg/ml. A tablet of ciprofloxacin was dissolved in an appropriate volume of water to get 5 mg/ml of stock solution.

# **Phytochemical screening**

The Ethanolic extract of *Ipomoea aquatica* was subjected to preliminary phytochemical screening for their presence or absence of active constituents utilizing a standard method of analysis.

#### Test for alkaloids

Extracts were dissolved individually in dilute Hydrochloric acid and filtered. The filtrate was carefully tested with various alkaloidal reagents as follows:

(a) *Dragondroff's Test:* To the filtrates, few drops of dilute Dragondroff's reagent (Potassium bismuth iodide solution) was added. An orange-brown precipitate indicates the presence of alkaloids.

(b) *Wagner's Test:* To the filtrates few drops of Wagner's reagent (Iodine-potassium iodide solution) was added. Reddish brown precipitate indicates the presence of alkaloids.

(c) *Mayer's Test:* To the filtrates few drops of Mayer's Reagent (Potassium mercuric iodide solution) was added. Cream precipitate indicates presence of alkaloids.

#### **Test for saponins**

*Foam Test:* 0.5 gm of extract was shaken with 2 ml of water. If foam produced persists for ten minutes it indicates the presence of saponins.

# Test for flavonoids

The ethanol extract 5 ml was added to a concentrated sulphuric acid (H2SO4) (1 ml) and 0.5g of Mg. A pink or red coloration that disappear on standing 3 min. indicates the presence of flavonoids.

# Test for tannins

*Gelatin Test:* To the extract, 1% gelatin solution containing sodium chloride was added. Formation of white precipitate indicates the presence of tannins

#### Test for carbohydrates

Extract was dissolved individually in 5 ml distilled water and filtered. The filtrates were used to test for the presence of carbohydrates.

a) *Molisch's Test:* Filtrates were treated with 2 drops of alcoholic  $\alpha$ -naphthol solution in a test tube. Formation of the violet ring at the junction indicates the presence of Carbohydrates.

b) *Benedict's Test:* Filtrates were treated with Benedict's reagent and heated gently. Orange red precipitate indicates the presence of reducing sugars.

c) *Fehling's Test:* Filtrates were hydrolysed with dil. HCl, neutralized with alkali and heated with Fehling's A and B solutions. Formation of red precipitate indicates presence of reducing sugars.

#### Test for glycoside

2 ml of concentrated H2SO4 was added carefully and shaken gently. A reddish brown colour indicated the presence of steroidal ring i. eglycone portion of the glycoside.

# Test for phytosterols

*Salkowski's Test:* Extract was treated with chloroform and filtered. The filtrates were treated with few drops of Conc. Sulphuric acid, shaken and allowed to stand. Appearance of golden yellow colour indicates the presence of triterpene.

#### Test for proteins and amino acids

*Xanthoproteic Test:* The Extract was treated with few drops of conc. Nitric acid. Formation of yellow colour indicates the presence of proteins.

# Fluorescence analysis

Table 2 represents the results of the fluorescence characteristics of *lpomoea aquatica* Forsk. Powdered drug in day light, short UV light ( $\lambda$ max 254 nm) and long UV light ( $\lambda$ max 365 nm). The identification and comparison of the colors was done using the standard colour index chart.

#### **Microbial strains**

Three bacterial strains [*Staphyllococcusaureus* (ATCC 25923), *Escherichia coli* (ATCC 25922) and two fungi [*Candida albicans* (ATCC 10231), and *Trichophytonrubrum* (ATCC 28188)] were used. Stains were obtained from Girijananda Chowdhury Institute of Pharmaceutical Science, Azara, Guwahati.

# Sterilization of materials

The petri dishes and pipettes packed into metal canisters were appropriately sterilized in the hot air oven at 170  $^\circ C$  for 1 h at each

occasion. A solution of the extract and culture media were autoclaved at 121  $^{\circ}\mathrm{C}$  for 15 min.

# Culture media

The media employed for the study was solid agar media.

# Antibacterial activity

The antimicrobial activity of the different extracts of the plant was assayed by agar-well diffusion method as described in NCCLS, 1993 [12]. Petri plates containing 20 ml nutrient agar medium was seeded with bacterial strains. Wells of approximately 10 mm was bored using a well cutter. Plant extracts were prepared in DMSO (stock: 1 mg/ml DMSO). The plant extracts of 25, 50, and 100  $\mu$ l concentrations were added. Ciprofloxacin (20  $\mu$ l) and DMSO (100  $\mu$ l) were used as positive and negative controls respectively. The plates were then incubated at 37 °C for 24 h. The antimicrobials present in the plant extract are allowed to diffuse out into the medium and interact with the test organisms in the freshly seeded plate. The diameter of the zone of inhibitions was measured in millimeters after 24 h.

# Antifungal activity

The potato dextrose agar plates were prepared and inoculated with a fungal culture. Wells of approximately 10 mm was bored using a well

cutter and samples of different concentration were added. The zone of inhibition was measured in millimeters after overnight incubation and compared with that of standard antifungal (Fluconazole) (10  $\mu$ l) which was used as positive control and DMSO (10%) as the negative control [13].

# RESULTS

The present investigation shows the phytochemical analysis, antimicrobial activity of the ethanolic extract of the plant *Ipomoea aquatic* Forsk. The yield % of the extraction of ethanol was %. It was a dark green solid powder (ET) in appearance.

# Phytochemical analysis

Phytochemical investigations with the ethanolic plant extract have demonstrated the presence of alkaloids, saponin, flavonoids, glycosides, Steroids, carbohydrate, proteins and amino acids and tannins (table 1).

# Antimicrobial activity

The Plant extract is generally rich in antimicrobial compounds. The *in vitro* antimicrobial activity of the ethanolic extract of *Ipomoea aquatic* Forsk. Under different concentration with the standard are appeared at table 3. The ethanolic extract of stems of the plant showed antimicrobial activity against a large portion of the test organisms.

# Table 1: Phytochemical screening of Ethanolic (ET) extract of stems of *Ipomoea aquatica* Forsk

S. No.	Phytochemicaltest	Reagentsused (testperformed)	Observation	Result
1	Alkaloidstest	Mayer's test	Creamy white precipitate	+
		Wagner's test	Reddish brown colour yellow precipitate	+
		Dragendoroff's test	Orange brown precipitate	+
2	Saponin test	Foam test	Froth formation	_
3	Flavonoidstest	LeadAcetate test	Yellow precipitate	+
4	Carbohydrate test	Molisch'stest	Formation of violetring	+
		Benedict's test	Formation of orange redprecipitate	+
		Fehling's test	Formation of redprecipitate	+
5	Glycosides test	Modified Borntrager's test	Formation of pink colour	_
5	Steroidtest	Salkowski's test	Formation of greencolour	+
7	ProteinsandAmino Acids	Xanthoproteic test	Formation of yellowcolour	+
8	Tanninstest	Gelatin test	Formation of greencolour	+

Note: +sign indicatespresenceand-sign indicatesabsence.

# Table 2: Fluorescence analysis power drug of Ipomoea aquatica Forsk. Stems

S. No.	Powder+reagent	Visible light	Short UV light (254 nm)	Long UV light (365 nm)
1	Powder as such	Green	Green	Dark Green
2	Powder+1N NaOH in methanol	Light yellow	Leaf green	Dark green
3	Powder+Conc. HNO3	Light yellow	Light brown	Reddish brown
4	Powder+Conc. HCl	Gold	Emerald	Blue
5	Powder+Conc. H2SO4	Dark green	Black	Black
6	Powder+KOH (5%)	Greenish yellow	Dark green	Blue
7	Powder+FeCl3 (5%)	Yellowish Brown	Brown	Dark green
8	Powder+Picric acid	Yellowish green	Green	Dark green

# Table 3: Antimicrobial activity Ethanolic (ET) extract of stems of Ipomoea aquatica Forsk. using disc diffusion assay

Name of the compounds and their concentration	Antibacterial activity diameter of inhibition zone (mm)		Antifungal activity diameter of inhibition zone (mm)	
	Staphylococcus aureus	Escherichia coli	C. albicans	Trichophyton rubrum
Standard Ciprofloxacin (20 µl)	44	42		
Standard Fluconazole (20 µl)			31.30	30.10
Plant extract (25 $\mu$ l)	13.2	11.3	11.90	10.10
Plant extract $(50 \ \mu l)$	17.53	14.92	16.70	12.55
Plant extract $(100 \ \mu l)^*$	22.64	18.20	20.04	15.12

\*significant activity

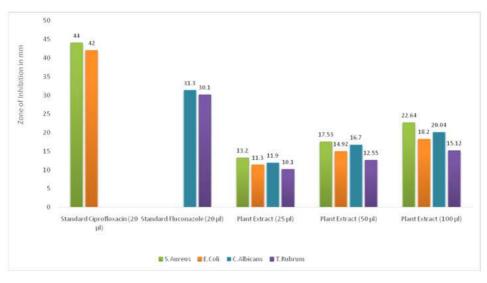


Fig. 1: Zone of inhibition vs concentration

### DISCUSSION

The antimicrobial activity of *Ipomoea aquatic* Forsk. Has been evaluated. Phyto-constituents exhibit in plants to be specific alkaloids, flavonoids, tannins, and triterpenoids are delivering an energizing open door for expansion of modern therapies against extensive variety of microorganisms. The present study was done against the variety of Gram-positive, Gram-negative bacteria, and fungal strains were selected for screening antimicrobial impact of the extracts to see the antimicrobial spectrum. The consequences of this study shows that the ethanolic extracts of the plant *Ipomoea aquatic* Forsk. Were highly successful in producing the desired result against most of the Gram-positive bacteria, Gram-negative bacteria, and fungal strains in agar well diffusion method.

Phytochemical constituents of plants such as tannins, alkaloids, flavonoids, phenolic compounds and several other aromatic compounds are secondary metabolites of plant that fill in as a barrier component against predation by numerous microorganisms.

The exhibition and explanation of antimicrobial activity against both Gram-positive and Gram-negative bacteria and on various fungal strains may be an indication of something presence of broad spectrum antibiotic compounds in the extracts which is shown in fig. 1. The  $100\mu$ g/ml concentration of extract having the most affecting antimicrobial activity.

#### CONCLUSION

The results obtained from this work showed that plant extracts of *Ipomoea aquatica* Forsk. Exhibit antimicrobial effects. In the concluding part, the future scope of Ipomoea species has been emphasized with a view to establish their biological activities and mode of action. Therefore, it is absolutely necessary to evaluate the antimicrobial activity of *Ipomoea aquatica* Forsk.

During the study, the antibacterial activity of the stems of *Ipomoea aquatica* Forsk. was judged by using disk diffusion method. The microorganisms that have been selected for study were Gram positive, *S. aureus* and Gram-negative *E. coli* and fungus like *C. albicans*, and *T. rubrum*. Study of this micro-organism were done as they are important pathogens and also they have the capacity of rapidly developed antibiotic resistance as antibiotic use increases.

At the end the study concluded that ethanolic extract of *Ipomoea aquatica* Forsk. showed the presence of various phytochemicals of which include alkaloids, saponin, carbohydrate, Steroids, and terpeniods.

The present study makes us know about the significant antimicrobial activity of the plant extracts when estimated with standards. So, after lots test, decision and thoughts it was found that the minimum inhibitory concentration of the plant extracts was less than  $100\mu g/ml$  and therefore the plant was considered to have a very good antimicrobial activity.

Further studies are needed to determine the chemical identity of the bioactive compounds responsible for the observed antimicrobial activity and unknown biological activities of *Ipomoea aquatic* Forsk. and its effect on living organisms, tissues or cells against various diseases.

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# **CONFLICT OF INTERESTS**

Declare none

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