

Original Article

EVALUATION OF HEPATOPROTECTIVE AND ANTIOXIDANT EFFECT OF *COMBRETUM ALBIDUM* G. DON AGAINST CCl₄ INDUCED HEPATOTOXICITY IN RATS

D. RAJALINGAM^{1,2*}, R. VARADHARAJAN^{1,2}, S. PALANI³

¹Dept of Pharmaceutical Chemistry, Kamalakshi Pandurangan College of Pharmacy, Tiruvannamalai, Tamil Nadu, India, ²Research Centre, Manonmanium Sundaranar University, Tirunelveli, Tamil Nadu, India, ³Dept of Biotechnology, Anna BioResearch Foundation, Arunai Engineering College, Tiruvannamalai, Tamil Nadu, India
Email: drlingam2007@gmail.com

Received: 29 May 2016 Revised and Accepted: 22 Jul 2016

ABSTRACT

Objective: The present investigations were undertaken to evaluate the hepatoprotective and antioxidant activity of the ethanolic extract of the whole plant of *Combretum albidum* G Don against CCl₄-induced hepatotoxicity in rats.

Methods: Hepatoprotective effect of ethanolic extract of *Combretum albidum* (EECA) was determined by using carbon tetrachloride (CCl₄) intoxication of rats as experimental models. The extent of liver damage and effect of the plant extract was assessed by various biochemical parameters like alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), total bilirubin (TB) and total protein (TP) in blood serum and concentration of thiobarbituric acid reactive substances (TBARS), reduced glutathione (GSH), superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), glutathione-s-transferase (GST) in liver were determined. Histopathological changes in the liver of different groups were also studied.

Results: The administration of EECA at dose levels of 250 and 500 mg/kg/b.w., orally had decreased the rise of ALT, AST, ALP, TB and TBRAS levels and the effects were comparable to standard drug (Silymarin 25 mg/kg/b. w.) the GSH, SOD, CAT, GPx, GST and TP levels were significantly increased in the animals received EECA. The histopathological studies show decreased necrosis and hepatocellular degeneration when compared to the CCl₄ intoxicated liver.

Conclusion: This study demonstrates that the hepatoprotective and the antioxidant activity of the whole plant of *Combretum albidum* therefore scientifically supports the use of this plant in traditional medicine for treatment of liver disorders.

Keywords: *Combretum albidum*, Hepatoprotective, Antioxidant, Liver regeneration

© 2016 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)
DOI: <http://dx.doi.org/10.22159/ijpps.2016.v8i9.13136>

INTRODUCTION

The liver is the most powerful metabolic organ. The continuous exposure and a variety of toxic environmental agents, certain drugs enhance hepatic injury, identified as a toxicological problem [1]. Most of the toxic chemicals damage the liver cells mainly by inducing lipid peroxidation and other oxidative damages in liver [2]. The choice of treatment for common liver diseases such as cirrhosis, fatty liver, and chronic hepatitis is problematic [3]. In spite of amazing development in modern medicine no effective drugs are available, which activate liver functions and often protect the liver from the damage or helps to reconstruct hepatic cells [4]. In the lack of reliable liver protective drugs in modern medicine, Plants traditionally used in the relief of liver dysfunction might, therefore, provide a useful source of new hepatoprotective compounds for development as pharmaceutical entities or as simple dietary adjuncts to existing therapies.

The *Combretum albidum* G. Don belongs to the family Combretaceae, commonly known as Buffalo calf in English, Vragay and odaikodi in Tamil, is a large woody climbing, deciduous shrub up to 30 m high, found in semi-evergreen and deciduous forests, along the river banks of peninsular, India and Sri Lanka [5]. In the ethnobotanical claims, leaves of the medicinal plant *Combretum albidum* G. Don are used in treating patients with jaundice and bark used for treating various skin diseases [6]. A decoction of the fruit is used for treating dysentery and diarrhea [7]. Its wiry stem, seed oil, root reported to cure eye problems, eczema and malarial fever [8].

The muthuvans tribe in Kerala prefers water extract of the stem bark as a remedy for both normal and severe jaundice [9]. The literature review revealed that the pharmacognostic standardization, physicochemical analysis. Preliminary phytochemical studies and isolation of five triterpenoids, beta-sitosterol, gallic acid, ellagic acid, and antibacterial activity of the plant were reported by several researchers [9-13].

To the best of our knowledge, there is no scientific report of the hepatoprotective and antioxidant effect of *Combretum albidum*. The present investigations are mainly emphasized on exploration and exploitation of the hepatoprotective and antioxidant activity of ethanolic extract of the whole plant of *Combretum albidum* against CCl₄-induced hepatotoxicity in rats.

MATERIALS AND METHODS

Plant material

The whole fresh plant of *Combretum albidum* G. Don was collected from Thirunelveli district of Tamilnadu, India in the month of February. The plant was identified and authenticated by Dr. V. Chelladurai, Research officer, Botany C. C. R. A. S. Govt. of India, (Retired). The voucher specimen (KPCP3/2014), was deposited in our pharmaceutical chemistry laboratory for future reference. The whole plant was dried under shade, made into a coarse powder with a mechanical grinder, passed through 40 mesh sieves and stored in closed containers for further use.

Extraction procedure

The dried, coarsely powdered *Combretum albidum* whole plant (500g) was extracted with ethanol [90%v/v] in soxhlet apparatus for 24 h. Then the solvent was completely recovered on the ethanol extract of *Combretum albidum* (EECA) under reduced pressure by a rotary vacuum evaporator. The concentrated extract was dried on a water bath and preserved in a vacuum desiccator.

Animals

Studies were carried out using Wister albino male rats (180-200g), obtained from Indian Veterinary Preventive Medicine (IVPM), Ranipet, Tamilnadu, India. The animals were grouped and housed in polyacrylic cages (38 x 23 x10 cm) with not more than six animals per cage and

maintained under standard laboratory conditions (temperature $25\pm 2^\circ\text{C}$) with dark and light cycle (12/12 h). The animals were fed with standard pellet diet supplied by the Poultry Research Station, Nandhanam, India. and fresh water *ad libitum*. All the animals were acclimatized to laboratory condition for a week before the commencement of the experiment. All animal studies were performed in accordance to guidelines of CPCSEA and Institutional Animal Ethical Committee (IAEC) of Kamalakshi Pandurangan college of Pharmacy, Tiruvannamalai (Tamilnadu). CPCSEA registration number was 745/03/ac/CPCSEA, and all the procedures were followed as per rules and regulation.

Drugs and chemicals

Silymarin was purchased from Micro labs, Tamilnadu. India. ALT, AST, ALP, Bilirubin and Total Protein kits were procured from Span Diagnostics, Surat, India. Thiobarbituric acid (TBA), nitro blue tetrazolium chloride (NBT), Phenazine methosulphate was purchased from Central Drug House, New Delhi, India and 5,5'-dithiobis-2-nitrobenzoic acid (DTNB), reduced glutathione (GSH), carbon tetrachloride purchased from SICCO Research Laboratory, Mumbai, India. All other chemicals and solvent were of analytical grade and commercially available.

Acute toxicity studies

An acute oral toxicity study was performed as per OECD-423 guidelines (acute toxic class method), Albino rats (n=6) of single-sex were selected for the acute toxicity study. Which received a single oral dose of 2000 mg/kg body weight of ethanolic extract of *Combretum albidum*. The dose was administered to overnight fasted rats, and the food was withheld for a further 3-4 h after administration of the drug and observed for signs of toxicity for a period of 14 d [14].

Experimental design

CCL₄-induced hepatotoxicity study

After acclimatization, the rats were divided into 6 groups of 6 rats each.

Group I: Served as Normal control which received liquid paraffin 2 ml/kg body weight (b. w), intraperitoneal (I. P).

Group II to V: Were administered with CCl₄ in liquid paraffin (1:2) in the dose 1 ml/kg body weight I. P, once in every 72 h for 16 d (1, 4, 7, 10, 13, 16 d).

Group III and IV: Were administered ethanolic extract of *combretum albidum* (EECA) at the dose of 250 mg/kg and 500 mg/kg body weight orally once in every 24 h for 16 d respectively.

Group-V: Was administered with reference drug Silymarin at the dose of 25 mg/kg body weight orally once in every 24 h for 16 d [15].

Estimation of biochemical parameters

The biochemical parameters were determined after 24 h fasting of the last dose. Blood was obtained from all animals by puncturing retro-orbital plexus. The blood samples were allowed to clot for 45 min at room temperature. Serum was separated by centrifugation at 2500 RPM at 30 °C for 15 min and used for the estimation of various biochemical parameters namely alanine transaminase (ALT), aspartate transaminase (AST), alkaline phosphatase (ALP) [16], total bilirubin (TB) [17] and total protein content (TP) [18]. The dissected liver was washed with 0.9% saline and homogenated (5%) in ice-cold phosphate buffer, and then centrifuged at 1000 RPM for 10 min followed by centrifugation of the supernatant at 12000 RPM for 15 min to get the mitochondrial fractions. These fractions were used for the estimations of thiobarbituric acid reactive substances (TBARS) [19]. Reduced glutathione (GSH) [20], superoxide dismutase (SOD) [21], catalase (CAT) [22], glutathione peroxidase (GPx) [23], glutathione-s-transferase (GST) [24].

Statistical analysis

The results are expressed as mean \pm SD of six animals from each group. One-way ANOVA followed by Dunnet multiple comparison tests have used to analyze the data by Graph pad prism. $P < 0.05$ was considered statistically significant.

Histopathological study

After the collection of blood samples, the rats were killed and their livers were excised, rinsed in ice-cold normal saline and processed separately for histological observation. Initially, the materials were

fixed at 10% buffered neutral formalin solution for 48 h and then with a bovine solution for 6 h. Paraffin sections were taken at 5 mm thickness processed in alcohol-xylene series and was stained with alum hematoxylin and eosin [25]. The sections were examined under photomicroscope for histopathological changes, necrosis, steatosis and fatty changes of hepatic cells.

RESULTS

Acute toxicity

It was observed that the administration of single oral dose 2000 mg/kg/body weight of ethanolic extract of *Combretum albidum* to a rat, didn't induce drug-related toxicity and mortality in the animals, and it was safe up to the dose of 2000 mg/kg/body weight.

Biochemical parameters

The effect of the ethanolic extract of *Combretum albidum* of CCl₄ induced hepatotoxicity and oxidative stress in rats were represented in the fig. 1-5. The animals administrated only with CCl₄ resulted in a significant increase ($P < 0.001$) in serum ALT, AST, ALP and TB levels as shown in the fig. 1-2. However, the total serum protein [TP] level was decreased when compared to a normal control group, indicating hepatocellular damage. The toxic effects of CCl₄ were controlled in the animals treated with ethanolic extract of *Combretum albidum* at the doses of 250 and 500 mg/kg b.w., produced significant ($P < 0.01$ and $P < 0.001$) dose-dependent decreases in serum marker enzyme ALT, AST, ALP (fig. 1) and TB (fig. 2) respectively as well as increases total protein level (fig. 2) as compared with normal control by the way of restoration of the level of the liver function similar to that of the reference drug silymarin (25 mg/kg. b. w).

The activity of lipid peroxidation (LPO) level was significantly ($P < 0.001$) increased (fig. 3) and CAT, GPx, GSH (fig. 4), GST and SOD activity were significantly ($P < 0.001$) decreased (fig. 5) in the serum level of rats treated with CCl₄ when compared with that of the normal control that received only liquid paraffin. Treatment of rats with ethanolic extract of *Combretum albidum* at the dose of 250 and 500 mg/kg b.w., significantly ($P < 0.01$, $P < 0.001$) decreased the elevated lipid peroxidation levels and the decreased levels of CAT, GPx, GSH, GST and SOD were restored to the normal levels in a dose-dependent manner when compared with standard drug (Silymarine) treated group.

Histopathology

Histological observation of liver tissue of the normal control group animal showed (fig. 6A) hepatic cells with well-preserved cytoplasm, nucleus, nucleolus, and central vein. In rats treated with CCl₄ (fig. 6B), histological observation showed fatty degeneration, damage of parenchymal cells, steatosis and hydropic degeneration of liver tissue. The prominent damage in the central lobular region appeared in the liver. The animals treated with the ethanolic extract of *Combretum albidum* (250 and 500 mg/kg) showed an improvement in the pathological changes, reduced the fatty degeneration and inflammation at dose-dependent manner (fig. 6C-E)

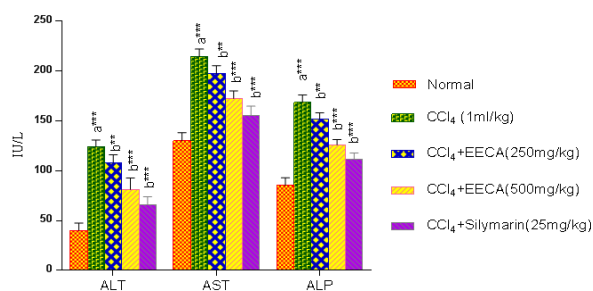


Fig. 1: Effect of ethanolic extract of *Combretum albidum* on serum activity levels of ALT (IU/L), AST (IU/L) and ALP (IU/L) in CCl₄ induced hepatotoxicity in rats

Values are expressed mean \pm SD for six rats in each group. a As compared with control, b as compared with CCl₄, ***represents $P < 0.001$, **represents $P < 0.01$

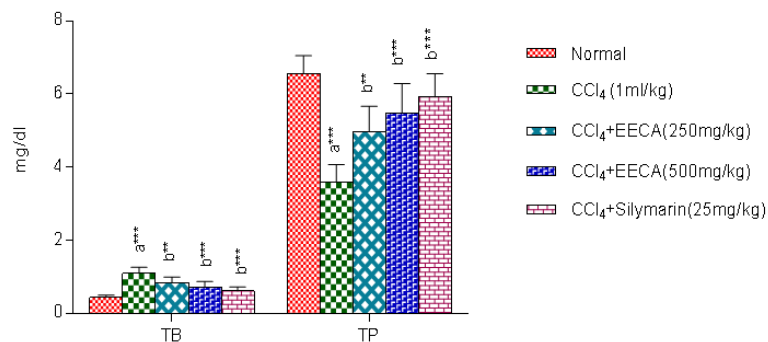


Fig. 2: Effect of ethanolic extract of *Combretum albidum* on serum levels of total bilirubin (mg/dl) and total protein in CCl₄ induced hepatotoxicity in rats

Values are expressed mean±SD for six rats in each group. a As compared with control, b As compared with CCl₄, ***represents P<0.001, **represents P<0.01.

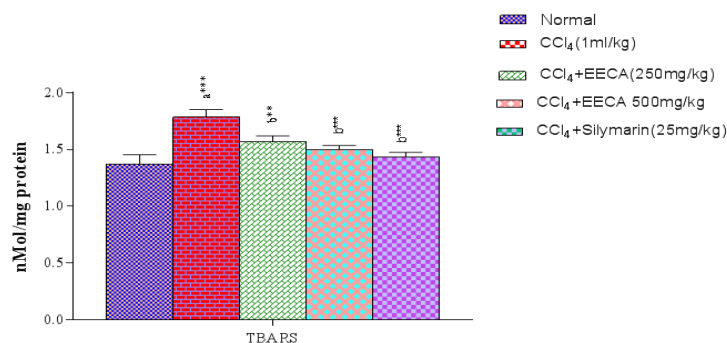


Fig. 3: Effect of ethanolic extract of *Combretum albidum* on serum levels of TBARS (nMol/mg protein) [Lipid peroxidation (LPO)] level of hepatic tissue in CCl₄ induced hepatotoxicity and oxidative stress in rats.

Values are expressed mean±SD for six rats in each group. a As compared with control, b As compared with CCl₄, ***represents P<0.001, **represents P<0.01

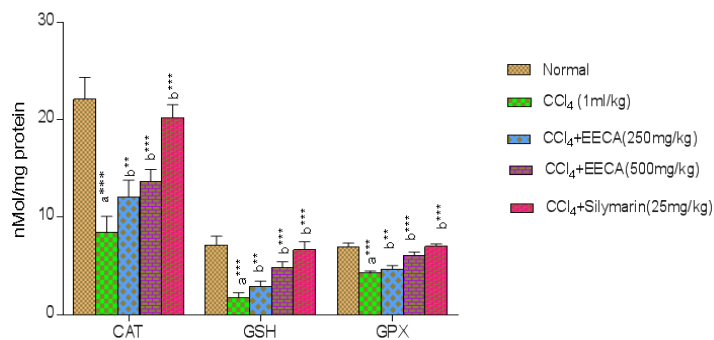


Fig. 4: Effect of ethanolic extract of *Combretum albidum* on hepatic levels of CAT (U/mg protein), GSH (U/mg protein) and GPx (micrograms of glutathione utilized/min/mg protein) in CCl₄ induced hepatotoxicity and oxidative stress in rats.

Values are expressed mean±SD for six rats in each group. a As compared with control, b As compared with CCl₄, ***represents P<0.001, **represents P<0.01

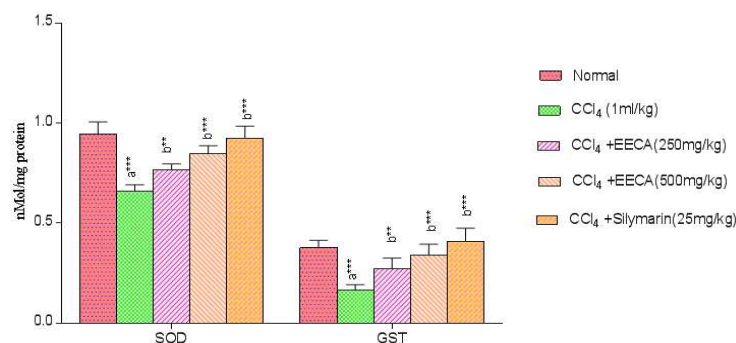


Fig. 5: Effect of ethanolic extract of *Combretum albidum* on hepatic levels of SOD (units of activity nMol/mg protein) and GST (Units nMol/mg protein) in CCl₄ induced hepatotoxicity and oxidative stress in rats.

Values are expressed mean±SD for six rats in each group. a As compared with control, b As compared with CCl₄, ***represents P<0.001, **represents P<0.01

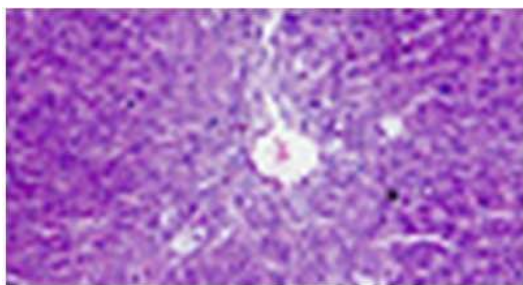


Fig. 6A): Histology of normal hepatic tissue

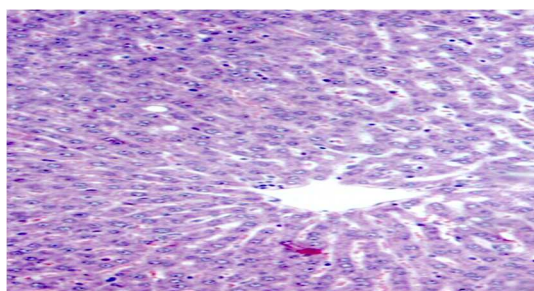


Fig. 6B): CCl₄ induced damage in hepatic tissue

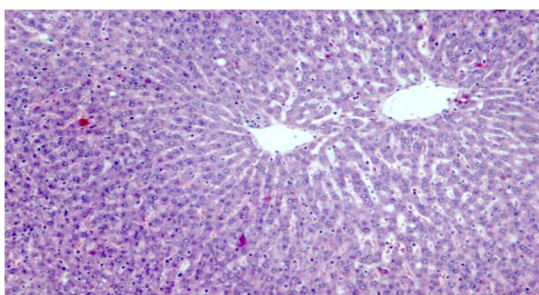


Fig. 6C): Effect of EECA (250 mg/kg) dose On CCl₄ induced hepatic damage

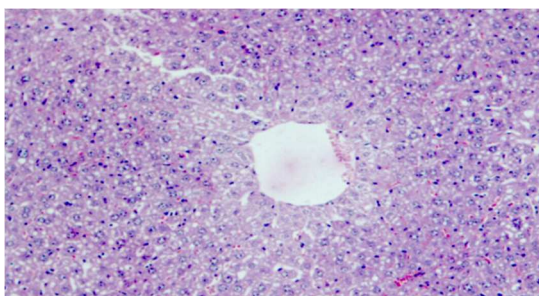


Fig. 6D): Effect of EECA (500 mg/kg) dose On CCl₄ induced hepatic damage

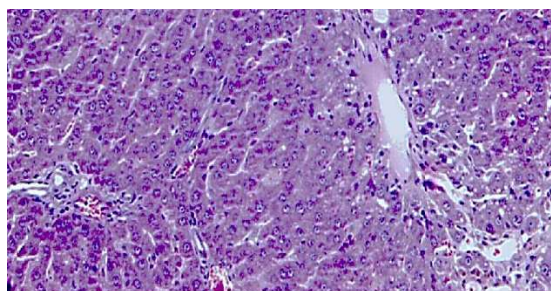


Fig. 6E): Effect of Silymarin On CCl₄ induced hepatic damage

DISCUSSION

Prophylactic action in liver damage induced by carbon tetrachloride has widely been used as an indicator of the liver protective activity of drugs in general [26]. Since the changes associated with CCl₄-induced liver damage are similar to that of acute viral hepatitis [27]. Investigation of chronic administration of CCl₄ induced liver damage in animals was chosen as an experimental model.

It is well documented that carbon tetrachloride is biotransformed under the action of cytochrome *P*-450 system in the microsomal compartment of liver to trichloromethyl or peroxy trichloromethyl free radical. These free radicals bind covalently to the macromolecules and induce peroxidative degradation of the membrane lipids of endoplasmic reticulum rich in polyunsaturated fatty acids. This leads to the formation of lipid peroxides followed by pathological changes such as triacylglycerol accumulation, polyribosomal disaggregating, and depression of protein synthesis, cell membrane breakdown and even death [28, 29].

In general, the extent of liver damage is assessed by histopathological evaluation and serum levels of ALT, AST, ALP, TB and TP release in circulation [30, 31]. When the liver cell plasma membrane is damaged, a variety of enzymes normally located in the cytosol are released into the blood stream. Their estimation in the serum is a useful quantitative marker of the extent and type of hepatocellular damage [32].

In the present study, it was observed that administration of CCl₄ elevates the levels of serum marker enzymes ALT, AST, ALP and total serum bilirubin as well as decreases total serum protein level significantly. Ethanolic extract of *Combretum albidum* and reference drug silymarin-treated groups exhibited lower serum levels of ALT, AST, ALP and total bilirubin as well as increases total protein as compared to CCl₄ treated groups. The stabilization of serum ALT, AST, ALP, and total bilirubin and the restoration of total protein levels by ethanolic extract of *Combretum albidum* is a clear indication of the improvement of the functional status of the liver cells.

Hepatoprotective activity correlated with antioxidant activity since it is free radical mediated damage [33]. An elevated level of malondialdehyde (MDA) reflects an enhanced lipid peroxidation leading to tissue damage and failure of antioxidant defense mechanisms to prevent the formation of excessive free radicals [34]. Treatment with ethanolic extract of *Combretum albidum* significantly reversed these changes. Hence it may be possible that the mechanism of hepatoprotection by ethanolic extract of *Combretum albidum* is due to its antioxidant effect.

The enzymatic antioxidant defense systems are the natural protector against lipid peroxidation. SOD, CAT and GPx enzymes are important scavengers of superoxide ion and hydrogen peroxide. These enzymes prevent generation of hydroxyl radical and protect the cellular constituents from oxidative damage [35]. In the present study, it was observed that the ethanolic extract of *Combretum albidum* significantly increased the hepatic SOD activity in CCl₄ induced liver damage in rats. This shows that the ethanolic extract of *Combretum albidum* can reduce reactive free radicals that might lessen oxidative damage to the tissues and improve the activities of the hepatic antioxidant enzyme.

Earlier studies regarding mechanism of CCl₄ induced hepatotoxicity have shown that GSH plays a key role in detoxifying the reactive, toxic metabolites of CCl₄ and that liver necrosis begins when the GSH stores are marked in a depleted state [36, 37]. Administration of the ethanolic extract of *Combretum albidum* increased the content of GSH significantly as compared to CCl₄ treated groups.

Catalase (CAT) is an enzymatic antioxidant widely distributed in all animal tissues and the highest activity, is found in the red cells and in the liver. CAT decomposes hydrogen peroxide and protects the tissue from highly reactive hydroxyl radicals [38]. Therefore the reduction in the activity of these enzymes may result in a number of deleterious effects due to the accumulation of superoxide radicals and hydrogen peroxide. Administration of the ethanolic extract of *Combretum albidum* increased the activities of CAT in CCl₄ induced liver damage in rats to prevent the accumulation of excessive free radicals and protected the liver from CCl₄ intoxication.

These findings can be further corroborated by histopathological studies. The histopathological examination clearly reveals that the hepatic cells, central vein, and portal triad are almost normal in the liver section of rats treated with an ethanolic extract of *Combretum albidum* in contrast to the liver section of rats which received CCl₄ only. Thus *Combretum albidum* can be considered to be an effective hepatoprotective as it ameliorates almost to normalcy the damage caused by CCl₄ to hepatic function.

It is well established that the phytoconstituent such as Flavonoids, triterpenoids and tannins are well known for their hepatoprotective activities [39, 40]. The literature review revealed that preliminary phytochemical analysis of heartwood of *Combretum albidum* showed the presence of the higher percentage of tannins, flavonoids, triterpenes, saponins, and glycosides, and five triterpenoids namely betulin, betulonic acid, oleanolic acid, arjunolic acid, ellagic acid and another constituent beta-sitosterol, gallic acid were isolated and reported [12,13]. The hepatoprotective activity of *Combretum albidum* G Don, may be attributed due to the presence of these constituents. This study supports the traditional claims and the plant CA could be added in traditional preparations for the various liver diseases.

CONCLUSION

In conclusion, the present study demonstrated that the Ethanolic extract of *Combretum albidum* possesses dose-dependent strong antioxidant activities and significant protective effect against chronic hepatotoxicity induced by CCl₄. The histopathological studies also substantiate the activity of the EECA. The results suggested that the possible mechanism of this activity may be due to free radical scavenging and antioxidant activity. Therefore, the study scientifically supports the use of this plant in traditional medicine for treatment of liver disorders.

CONFLICT OF INTERESTS

Declared none

REFERENCES

- Zimmerman HJ. Hepatotoxicity. Dis Mon 1993;39:675-87.
- Ross, Wilson. Anatomy and physiology in health and illness. Ninth Edition. Published by Elsevier Ltd; 2005. p. 307-10, 333.
- Scote luper ND. A review of plants used in the treatment: of liver disease Part-1. Alternative Med Rev 1998;16:410-7.
- Chattopadhyay RR. Possible mechanism of hepatoprotective activity of *Azadirachta indica* the whole plant extract, Part II. J Ethnopharmacol 2003;89:217-9.
- Sajeev KK, Sasidharan N. Ethnobotanical observations on tribals of the chinnar wildlife sanctuary. Ancient Sci Life 1997;16:284-92.
- Ganesan S, Ponnuchamy M, Kesavan L, Selvaraj A. Floristic composition and practices on the selected sacred groves of the palla patty village (reserved forest), Tamil Nadu. Indian J Traditional Knowledge 2009;8:154-62.
- Karuppusamy S. Medicinal plants used by paliyan tribes of sirumalai hills of southern India. Indian J Nat Prod Resour 2007;6:436-42.
- Kadavul K, Dixit AK. Ethnomedicinal studies of woody species of kalrayan and shervarayan hills, Eastern Ghats Tamilnadu. Indian J Traditional Knowledge 2009;8:592-7.
- Sreedhar S, Kumar UP, Shree ABR. Pharmacognostic standardization of heartwood of *Combretum albidum* G don an important ethnomedicinal liana. Int J Pharmacogn Phytochem Res 2013;5:106-12.
- Bokhad MN, Rothe SP. Preliminary phytochemical investigation of *Combretum albidum* G. Don. An ignored medicinally important liana. J Exp Sci 2012;3:1-4.
- Sailaja T, Rao ML, Savithamma N. Qualitative and quantitative analysis of phytochemicals of *Combretum albidum* G Don from rare medicinal plant taxon. J Phytochem Photon 2013;114:220-5.
- Kumar UP, Sreedhar S, Purushothaman E. Secondary metabolite from the heartwood of *Combretum albidum* G Don. Int J Pharmacogn Phytochem Res 2015;7:319-24.
- Sreedhar S, Nitha B, Shree ABR. Antimicrobial activity of stem bark of *Combretum albidum* G. Don. A traditional medicinal liana. Int J Pharm Sci Res 2013;4:3184-8.
- OECD. Acute oral toxicity. Acute oral toxic class method guideline 423 adopted 23.03.1996. In: Eleventh Addendum to the OECD, guidelines for the testing of chemicals organization for economical co-operation and development, Paris; 2000.
- Shukla Mukherjee, Sur A, Maiti BR. Hepatoprotective effect of *Swertia chirata* on rat. Indian J Exp Biol 1997;35:384-8.
- King J. The hydrolases-acid and alkaline phosphatase, practical clinical enzymology. Van D. ed. London: Nostrand company Lt; 1965. p. 191-208.
- Malloy HJ, Evelyn KA. The determination of bilirubin with the photoelectric colorimeter. J Biol Chem 1937;119:481.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the folin-phenol reagent. J Biol Chem 1951;193:265-75.
- Okhawa H, Ohigni N, Yagi K. Assay of lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem 1979;95:351-9.
- Sedlak J, Lindsay RH. Estimation of total protein bound and non-protein sulfhydryl groups in tissue with Ellman's reagent. Anal Biochem 1968;25:293-8.
- Misra HP, Fridovich. The role of superoxide anion in the autooxidation of epinephrine and a simple assay of superoxide dismutase. J Biol Chem 1972;247:3170-84.
- Tukahara S, Hamilton BH, Nell J, Ogura Y, Nishmura E. Hypocatalasemia new genetic carrier state. J Clin Invest 1960;29:610-6.
- Rotruck JT, Pope LA, Ganther HE, Swanson AB. Selenium biochemical role as a component of glutathione peroxidase. Science 1973;179:588-93.
- Habig WH, Pabst MS, Jekpoly WB. Gluthathione transferase: a first enzymatic step in mercapturic acid formation. J Boil Chem 1974;249:7130-6.
- Valeer JD. Liver tissue examination. J Hepatol 2003;39:543-9.
- Clauson GA. Mechanism of carbon tetrachloride hepatotoxicity. Pathol Immunopathol Res 1989;8:104-12.
- Rubinstein D. Epinephrine release and liver glycogen levels after carbon tetrachloride administration. Am J Physiol 1962;203:1033-7.
- Recknagel RO, Ghosal AK. Quantitative estimation of peroxidative degeneration of liver microsomal and mitochondrial lipids after carbon tetrachloride poisoning. Exp Mol Pathol 1966;5:413-26.
- Noguchi T, Fong KL, Lai EK. Specificity of phenobarbital-induced cytochrome P450 for metabolism of carbon tetrachloride to the trichloromethyl radical. Biochem Pharmacol 1982;31:615-24.
- Plaa G, Charbonneau M. Detection and evaluation of the chemically induced liver injury. In: Hayes AW. Principals and methods of toxicology. Raven Press: New York; 1994. p. 841-6.
- Portmann B, Talbot IC, Day DW, Davidson AR. Histopathological changes in the liver following a paracetamol over dose; correlation with clinical and biochemical parameter. J Pathol 1975;117:169-80.
- Mitra SK, Venkataranganna MV, Sundaram R, Gopumadhavan S. Protective effect of HD-03, a herbal formulation, against various hepatotoxic agents in rats. J Ethnopharmacol 1998;63:181-6.
- Murthy KNC, Jayaprakash GK, Singh RP. Studies on antioxidant activity of pomegranate (*Punica granatum*) peel extracts using *in vivo* models. J Agric Food Chem 2002;50:4791.
- Souza MF, Rao VSN, Silveira ER. Inhibition of lipid peroxidation by ternatin, a tetramethoxyflavone from *Egletes viscosa* L. Phytomedicine 1997;4:25.
- Scott MD, Lubin BH, Zuo L, Kuypers FA. Erythrocyte defense against hydrogenperoxide: preeminent importance of catalase. J Lab Clin Med 1991;118:7-16.
- Reckengel RO, Glende EA, Britton RS. Free radical damage and lipid peroxidation. In: Histotoxicology. Meeks RG, Harrison SD, Bull RJ. Eds. CRC Press: Florida; 1991. p. 401-36.
- Williams A, Burk RF. Carbon tetrachloride hepatotoxicity: an example of free radical-mediated injury. Semin Liver Dis 1990;10:279-84.
- Chance B, Green Stein DS, Roughton RJW. The mechanism of catalase action I-steady state analysis. Arch Biochem Biophys 1952;37:301-39.

39. Manjunatha BK, Vidya SM. Hepatoprotective activity of *Vitex trifolia* against carbon tetrachloride induced hepatic damage. *Indian J Pharm Sci* 2008;70:241-5.
40. Das S, Sarma G. Study of the hepatoprotective activity of the ethanolic extract of the pulp of *Eugenia Jambolana* (Jamun) in albino rats. *Exp Res* 2009;3:1466-74.

How to cite this article

- D Rajalingam, R Varadharajan, S Palani. Evaluation of hepatoprotective and antioxidant effect of *Combretum albidum* G. Don against CCL₄ induced hepatotoxicity in rats. *Int J Pharm Pharm Sci* 2016;8(9):218-223.