

Original Article

ESTROGENIC AND ANTIESTROGENIC ACTIVITIES OF COMMERCIAL DIETARY SUPPLEMENTS CONTAINING HERBAL INGREDIENTS AND ISOFLAVONES

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ABSTRACT

Objective: This study aimed to assess the estrogenic and antiestrogenic activities of five dietary supplements, commercially available for treatment of menopausal symptoms, before and after gastrointestinal digestion by employing a yeast steroid-regulated transcription system.

Methods: Supplements (S) were extracted with either 80% methanol or water. Water extracts were subjected to simulated gastrointestinal (GI) digestion. Estrogenic and antiestrogenic activities were assessed by a steroid-regulated transcription system in *Saccharomyces cerevisiae* expressing the human estrogen receptor alpha.

Results: The highest estrogenic activities were detected in both S1 methanol (2342.5±20.83 MU) and water (1225.6±20.6 MU) extracts (400 estradiol equivalents). Extracts showed antiestrogenic properties by reducing the transcriptional activity induced by estradiol in transgenic yeast. The highest antiestrogenic activity was detected in S2 methanol extract and S3 water extract, which inhibited estradiol activity by 76% and 64%, respectively. After GI digestion, S1, S2 and S3 extracts showed significantly higher estrogenic and antiestrogenic activities as 'serum-available' than 'colon-available' samples and S4 and S5 extracts showed significantly higher activities as 'colon-available' than 'serum-available' samples.

Conclusion: All dietary supplements revealed estrogenic and antiestrogenic activities. The GI digestion demonstrated the availability of phytoestrogens for absorption in the blood stream. Supplements containing soy isoflavones and alfalfa ingredients had the highest estrogenic activities and could be more effective than supplements with complex plant formulation in alleviating menopausal symptoms and treating osteoporosis. The transgenic yeast assays proved to be a powerful tool for assessing the *in vitro* estrogenic and antiestrogenic activities of dietary supplements.

Keywords: Antiestrogenic, Dietary supplement, Estrogenic, Phytoestrogens, Simulated gastrointestinal digestion

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INTRODUCTION

Estrogen hormones are important for the development of sexual characteristics, reproduction, and overall health, especially for women's health, as estrogens are the primary hormones responsible for a majority of functional pathways in reproduction, cardiovascular, neuronal and skeletal systems [1]. Women between the ages of 40-60 are affected by changes in estrogen levels that result in the onset of menopause. Menopausal women suffer from a variety of symptoms, such as hot flashes, night sweats, mood swings, insomnia, vaginal dryness, in addition to long-term complications such as osteoporosis [2]. This period of transition is often eased by undergoing hormone replacement therapy (HRT) or taking specially formulated dietary supplements containing phytoestrogens. In either case, the objective of the treatment is to replenish estrogenic levels to alleviate menopausal symptoms and prevent osteoporosis. In 2002, the Women's Health Initiative (WHI) publicized the increased risk of developing breast cancer due to the hormone replacement therapy [3], which has led to a rise in alternative and complementary therapies for menopause, most of which include botanical supplements [4, 5].

Plants are known to produce estrogen-like compounds or phytoestrogens, which are non-steroidal polyphenolic compounds structurally similar to the mammalian estrogen 17 β -estradiol. Phytoestrogens, which are present in dietary supplements that are widely marketed as natural alternatives to hormone replacement therapy, function similarly to animal estrogens in that they bind to estrogen receptors (ER) inducing transcription of target genes in human cells [6]. The chemical structures and functions of phytoestrogens and their effects in mammals have been studied extensively since their discovery in the 1920s [7, 8]. More recently, the publicized increased risk of developing breast cancer due to the hormone replacement therapy [3, 9], also led to a rise in

development and use of supplements containing phytoestrogens for use in alleviating menopausal symptoms. However, not enough studies on the effectiveness of supplements in replenishing the estrogen levels needed to lessen the menopausal symptoms have been published so far.

Phytoestrogens also are found to be inhibitors of estrogen-mediated tumorigenesis [10]. Besides treating/preventing the menopausal symptoms and osteoporosis, botanical supplements containing isoflavone phytoestrogens are known to be associated with decreased postmenopausal breast cancer risk [8]. The above-mentioned study reported that women consuming isoflavone-containing supplements had a reduced risk for breast cancer in Ontario, Canada [11]. Clinical studies are also supported by *in vitro* studies indicating that isoflavones, mainly genistein, exhibited antiestrogenic activities at higher concentrations (>20 μ M) and thus inhibited the proliferation of breast cancer cells [12, 13].

Given the significant interest in the dietary supplements aimed at alleviating menopausal symptoms, the present study was undertaken to assess the estrogenic and antiestrogenic activities of five commercial dietary supplements before and after simulated GI digestion, in an effort to evaluate the bioavailability of phytoestrogens in the supplement formulations. To the best of our knowledge, this is the first study reporting the estrogenic and antiestrogenic activities of dietary supplements before and after GI digestion by employing an estrogen-regulated transcription system.

MATERIALS AND METHODS

Chemicals

Ortho-Nitrophenyl- β -galactoside (ONPG), 17 β -estradiol, sodium phosphate dibasic heptahydrate (Na₂HPO₄·7H₂O), sodium phosphate

monobasic, monohydrate (NaH₂PO₄ · H₂O), potassium chloride (KCl), magnesium sulfate (MgSO₄·7H₂O), D-(+)-glucose, 2-Mercaptoethanol (C₂H₆OS) and 200-proof ethyl alcohol (CH₃CH₂OH) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Yeast nitrogen base (YNB) was from United States Biological (Salem, MA, USA). Cassamino acids were from BD Difco (San Jose, CA, USA). Adenine sulfate (C₅H₅N₅O₅·H₂SO₄) was from Acros Organics (NJ, USA). Dietary supplements were purchased from two suppliers in Denton, Texas, USA.

Preparation of dietary supplement extracts

Dietary supplements were extracted in 80% methanol or water as the daily dosage recommended by the manufacturers (table 1) at room temperature for 2 d and then centrifuged at 3,000-4,000 rpm for 20 min. All supernatants were filtered through Whatman # 54 filter paper, and water extracts were filter sterilized. The filtrates were centrifuged at 10,000 rpm, 4 °C for 15 min and stored at -20 °C for further study.

Table 1: Recommended dose and composition of dietary supplements containing herbal ingredients and isoflavones

Supplement	Recommended dose/day	Content
S1 Alfalfa	3 tablets	Organic Alfalfa (<i>Medicago sativa</i> , <i>Fabaceae</i>) leaf, 1800 mg
S2 Solaray®'s Black Cohosh	4 tablets	Black Cohosh (<i>Cimicifuga racemosa</i> , <i>Ranunculaceae</i>) root, 540 mg
S3 Estro Pause Menopause Support	4 liquid soft-gels	Calcium as calcium carbonate and calcium citrate, 500 mg; Magnesium as magnesium oxide and magnesium citrate, 200 mg; Fish oil, 2158 mg; John's Wort (<i>Hypericum perforatum</i> , <i>Hypericaceae</i>) extract, 300 mg; Soy isoflavones, 100 mg; Black cohosh extract, 80 mg; Red clover (<i>Trifolium pretense</i> , <i>Fabaceae</i>) powder, 40 mg; Asian Ginseng (<i>Panax ginseng</i> , <i>Araliaceae</i>) extract, 31 mg; Chaste Tree (<i>Vitex agnus-castus</i> , <i>Lamiaceae</i>) fruit extract, 20 mg
S4 Advanced Therapeutics Isoflavone Rx-Phytoestrogen®	2 tablets	Soy isoflavone concentrate, 125 mg; Genistin, 25 mg; Daidzin, 19.25 mg; Glycitin, 5.75 mg
S5 Meno Care for Menopausal Comfort®	4 capsules	Herbal blend (800 mg) containing: Asoka tree (<i>Saraca asoca</i> , <i>Fabaceae</i>) bark; Shatavari (<i>Asparagus racemosus</i> , <i>Asparagaceae</i>) root; Chebulic Myrobalan (<i>Terminalia chebula</i> , <i>Combretaceae</i>) fruit rind; Heart-leaf Sida (<i>Sida cordifolia</i> , <i>Malvaceae</i>) root; Licorice (<i>Glycyrrhiza glabra</i> , <i>Fabaceae</i>) root; Gotu Kota (<i>Centella asiatica</i> , <i>Apiaceae</i>) whole plant

Simulated gastrointestinal (GI) digestion

Simulated GI digestion of supplement water extracts was performed by the method of McDougall *et al.* [14] (fig. 1). Each supplement extract (2.5 ml) was added to the 17.5 ml simulated stomach solution (1.6 g pepsin, 1 g NaCl, pH 2). The mixture was incubated at 37 °C and 100 rpm in a shaking water bath for 2 h (Gastric Phase, fig. 1). The intestinal phase of the simulated GI was assembled as follows: the gastric phase solution was placed in a glass beaker along with 4.5 ml of a mixture of 4 mg/ml pancreatin, and 25 mg/ml bile salts and a segment of cellulose dialysis tubing (molecular weight cut off 12 kDa) containing 1M NaHCO₃ to neutralize the titratable acidity was placed in the gastric phase solution and the beaker was incubated at 37 °C for 2 h. At the end of the incubation period, the solution in the beaker constituted the 'OUT SAMPLE,' representing material that remains in the gastrointestinal tract (colon-available) and the solution that diffused into the dialysis tubing constituted the 'IN SAMPLE,' representing the serum available material. All samples were centrifuged at 16,000 rpm, filtered through 0.45 µm nylon syringe filters and stored at -20 °C for further analyses.

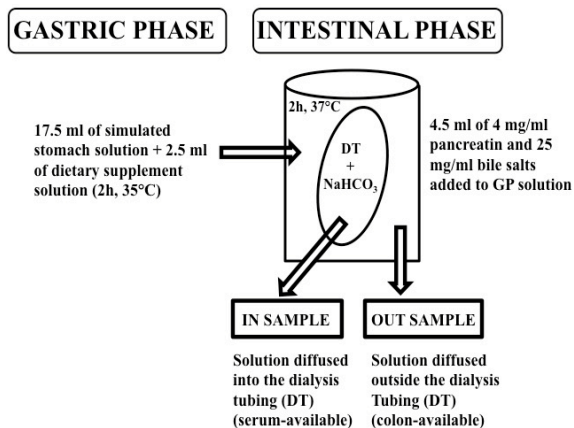


Fig. 1: Schematic representation of simulated GI digestion. GP, gastric phase; DT, dialysis tubing

Estrogenic and antiestrogenic assays

The estrogenic and antiestrogenic assays were performed according to the method of Maier *et al.* [15]. In short, estrogen (E) equivalents in each non-digested and digested dietary supplement extracts were estimated based on a 17β-estradiol standard curve. A steroid-regulated *Saccharomyces cerevisiae* system [BJ3505 (MAT a, pep4::His 3, prbl-D1.6R, his3-D200, lys2-801, trpl-D101 (gal3), ura3-52(gal2), can1)] was used to determine the estrogenic and antiestrogenic activities of extracts. Yeast cells contained the expression plasmid YEPE10 expressing the human ERα and YRPE2, a reporter plasmid expressing the *E. coli* β-galactosidase enzyme (*lacZ*) gene. Yeast cultures were grown in a cassamino acid-glucose medium (CAA medium-20% glucose, 10% yeast nitrogen base, and 5% adenine sulfate) at 230 rpm, 30 °C, overnight in an incubator-shaker. For the estrogenic assays, cultures were inoculated with 100 µg, 200 µg, 300 µg, or 400 µg E equivalents of non-digested or digested dietary supplements extracts. Cultures inoculated with 2 µg estradiol or with 2 µg genistein were used as positive controls and yeast cultures without any treatments were used as negative controls for all experiments. Average estradiol activity was 2,500 MU, and average genistein activity was 900 MU. For antiestrogenic assays, cultures were inoculated with 100 µg, 200 µg, 300 µg, or 400 µg E equivalents of non-digested or digested supplement extracts and 2 µg of estradiol. The antiestrogenic activity of extracts translates in a reduction of the transcriptional activity induced by estradiol in transgenic yeast, as active compounds in extracts compete with E for the ligand-binding site of the ER. All cultures were allowed to grow in the incubator-shaker for six hours. After six hours, the cells were disrupted with glass beads, the protein concentrations in supernatants were estimated, estrogenic and antiestrogenic assays were performed as previously described [15]. Estrogenic activity was calculated according to the formula:

$$\text{MU} = \{O. D. \text{ at } 420 \text{ nm}\} / \{\text{Protein concentration (g)} \times \text{Time (min)}\} \times 1000 \text{ and expressed in Miller Units (MU).}$$

Antiestrogenic activity was expressed as percentage using the formula:

$$\{(\text{Activity of supplement+estradiol}) / (\text{Activity of estradiol})\} \times 100$$

Statistical analysis

Means and standard deviations (SD) of three experiments were calculated. One-way ANOVA was performed, and significance of differences among means was determined by Tukey's test (*P ≤ 0.05).

RESULTS AND DISCUSSION

Both non-digested methanol and water supplement extracts showed increased estrogenic and antiestrogenic activities with increasing concentrations (table 2). The highest estrogenic activities were detected in S1 methanol (2342.6±20.8 MU) and water (1225.6±20.6 MU) extracts (400E equivalents). The estrogenic activities of other methanol extracts of non-digested dietary supplements ranked as follows: S3 and S4>S2 and S5 and of water extracts: S3>S2>S4>S5 (table 2).

Supplement extracts showed antiestrogenic properties by reducing the transcriptional activity induced by estradiol in transgenic yeast. The highest antiestrogenic activities were detected in S2 methanol extract and in S1 and S3 water extracts, which inhibited estradiol activity by 76% and 60% and 64%, respectively (table 2).

The antiestrogenic activities of other methanol extracts at 400E equivalents ranked as follows: S3 and S4>S1 and S5 and of water extracts: S5>S2 and S4 (table 2).

Table 2: Estrogenic and antiestrogenic activities of non-digested dietary supplement extracts

Estrogenic activities (MU)				
80% Methanol Extracts	100E Equiv	200E Equiv	300E Equiv	400E Equiv
S1	188.9±8 ^{(a)(*)}	1101.8±20.6	1453.3±3.4	2342.6±20.8
S2	13.3±3.3	86.1±9.5 ^(b)	221.1±16 ^(b)	273.3±1.1 ^(b)
S3	214.4±15 ^(c)	353.9±27.2	944.2±30	1653.6±26 ^(c)
S4	402±3.1	567±20	720±17.2	1620.8±11 ^(c)
S5	152.2±2.2 ^(a)	175.6±13.3 ^(b)	239.4±4 ^(b)	361.7±7.3 ^(b)
Water Extracts				
S1	282±9 ^{(a)(*)}	308.3±3.9 ^(a)	311.7±2 ^(*)	1225.6±20.6
S2	353.3±6.7 ^(*)	376.7±8.9 ^(b)	638.1±19 ^(b)	829.5±20
S3	273.9±17 ^(ac)	310±2.2 ^(*)	594.5±23 ^(b)	960.8±12
S4	241±0.3 ^(ac)	366.1±10.6 ^(b)	401.2±3 ^(*)	443.9±6.1
S5	157.2±1.7	208.6±0.8	213.3±6 ^(*)	281.1±6.7
Antiestrogenic activities (%)				
80% Methanol Extracts	100E Equiv	200E Equiv	300E Equiv	400E Equiv
S1	14.5±1.4 ^(*)	30.5±2.3 ^{(a)(*)}	42.2±2.2 ^(a)	52.7±1.9 ^{(a)(*)}
S2	4.8±1.4 ^(b)	35.5±4.6 ^(ab)	49.1±2 ^(b)	76.4±3.3 ^(*)
S3	26.5±0.2 ^(*)	35.2±0.3 ^(abc)	39.3±1 ^(abc)	62.7±4.9 ^(c)
S4	8.3±0.6 ^(b)	18.3±0.8 ^(*)	40±0.3 ^(abcd)	62.1±0.4 ^(c)
S5	4.1±0.6 ^(b)	28.6±1 ^{(abc)(*)}	44±0.2 ^(abcd)	54.8±3 ^{(a)(*)}
Water Extracts				
S1	9.4±3.5 ^{(a)(*)}	27.1±1.8 ^(a)	28.9±4 ^{(a)(*)}	60.2±9.2 ^{(a)(*)}
S2	18.2±1 ^{(b)(*)}	29.5±2 ^{(a)(*)}	32±8 ^{(ab)(*)}	39.9±7 ^{(b)(*)}
S3	4.6±0.1 ^(abc)	20±0.3 ^{(ac)(*)}	32±0.3 ^(abc)	64.2±1 ^{(a)(*)}
S4	21±1 ^{(bcd)(*)}	20.9±3.3 ^(acd)	30±1 ^{(abcd)(*)}	37.4±4 ^{(b)(*)}
S5	14.3±1 ^(abcd)	20.3±1.1 ^(acd)	30±5.2 ^(abcd)	46.9±1

Results represent means±SD of three independent experiments (each experiment had two replicates; n=6). In each column, mean values with no superscript letters are significantly different from each other at $P \leq 0.01$; mean values with * are significantly different from each other at $P \leq 0.05$; mean values with same superscript letters are not significantly different (Tukey's test).

The high estrogenic activity of S1 dietary supplement extracts (table 2) was induced by the phytoestrogens in its active ingredient, alfalfa (table 1). This plant is known to contain coumestrol, apigenin, luteolin, quercetin and other chemicals that exhibit estrogenic activity [16, 17]. A study employing Italian women showed that use of alfalfa extracts for three months completely alleviated hot flashes and night sweating in 20 out of 30 women [18]. The Health, Eating, Activity, and Lifestyle (HEAL) study showed that 31 alfalfa users had a substantially lower risk of hot flashes demonstrating the effect of alfalfa in alleviating the menopausal symptoms [19].

The S2 dietary supplement contains black cohosh root as an active ingredient (table 1). Although S2 methanol extract showed low estrogenic activity, the water extract had a significantly higher estrogenic activity (table 2). The root and rhizome of black cohosh have been researched for over 30 y for the relief of menopause-related symptoms and have been shown to be effective for the treatment of menopausal symptoms such as hot flushes, profuse sweating, sleeping disorders and nervous irritability [20]. Thus, The North American Menopause Society recommended black cohosh for the treatment of women with menopausal symptoms [21]. A 2011-2012 randomized, double-blind, placebo-controlled clinical trial conducted on 84 early post-menopausal participants with Greene climacteric scale (GCS) scores of 15 to 42 showed that black cohosh extract reduced the total and subscale scores (vasomotor, psychiatric, physical, and sexual symptoms) during the 4-8 w of treatment [22-25]. Some studies found that chemicals in black cohosh extract do not bind to ERs in ligand binding assays with recombinant estrogen receptors [26]. However, we think that the estrogenic and antiestrogenic activities exhibited by S3 in the transgenic yeast system are induced by triterpene glycosides,

especially cimicifugoside phytoestrogens specific to black cohosh [27, 28]. It was suggested, based on preclinical findings that chemicals in black cohosh extracts with dopaminergic or serotonergic activity rather than estrogenic activity ameliorate menopausal symptoms. Thus, black cohosh is considered a safe alternative to hormone replacement therapy even in patients with a history of hormone-responsive neoplasias since it induces apoptosis in human mammary tumor cells and does not have proliferative effects on crucial estrogen-responsive tissues [29-31].

The estrogenic and antiestrogenic activities of S3 and S4 are most likely due to the soy isoflavones in their formulations. S3 also contains plant materials known for their phytoestrogens, such as red clover and black cohosh, and S4 contains isoflavone glycosides besides a soy isoflavone concentrate of unknown composition (table 1). Red clover has been known to contain phytoestrogens, mostly isoflavones and some coumestans, which could act as either estrogen agonists or antagonists, depending on the phytoestrogen and its serum concentration [32]. The isoflavones formononetin, biochanin A, genistein and daidzein are present in red clover plants as glycosides and malonates [32]. The largest randomized study conducted by Tice *et al.* with 252 postmenopausal women reporting ≥ 35 hot flashes per week to Promensil (red clover isoflavones; Novogen Ltd.), Rimostil (red clover isoflavones; Novogen Ltd.), or placebo for 12 w showed no beneficial effects of red clover products over placebo in relieving hot flash [33]. Similarly, other studies reported by Barber *et al.* [34], and Knight *et al.* [35] showed no effects of red clover in alleviating the menopausal symptoms as compared to placebo. In contrast to the above studies, other studies reported a statistically significant reduction in hot flash frequency among women taking Promensil compared to placebo. Van de Weijer

and Barentsen showed that consumption of Promensil (80 mg/day isoflavones) for 12 w significantly reduced hot flashes in 30 randomized women reporting ≥ 5 hot flashes per day [36]. Another 12-week randomized, controlled trial conducted on 72 postmenopausal women showed the effectiveness of dried leaves of red clover in reducing the severity of menopausal symptoms [37]. The systematic review and meta-analysis by Ghazanfarpour *et al.* [38] concluded that consumption of red clover is effective in decreasing hot flashes, especially in women with severe hot flashes (≥ 5 per day).

S3 and S5 contain ingredients from several different plant species used in folk medicine to alleviate menopausal and other symptoms. Most of these ingredients possess estrogenic and/or antiestrogenic properties in different assay systems [39-46]. Although S5 is a pure herbal blend compared to S3, which contains mineral components and fish oil besides plant material, its extracts had the lowest estrogenic and antiestrogenic activities in the recombinant yeast system (table 2) indicating that some of the ingredients are very weak phytoestrogens and/or non-estrogenic ingredients interfered with the activity of phytoestrogens as reported before [47].

Except for S2 extract, all other non-digested dietary supplements showed higher estrogenic activities as methanol extracts than water extracts in the transgenic yeast system, whereas except for S2, S4 and S5 extracts (at 100E equivalents), all other supplement extracts displayed significantly higher antiestrogenic activities as methanol extracts than water extracts (table 2). It seems that methanol is a better solvent in extracting the active ingredients of the dietary supplements under study than water. In general, methanol is more efficient in the extraction of lower molecular weight phytochemicals such as flavonoids, proanthocyanidins, and tannins as compared to water [48]. Therefore, methanol formulations (tinctures) of dietary supplements may be more potent in delivering phytoestrogens for alleviating menopausal symptoms.

After GI digestion of dietary supplements, the 'IN SAMPLE' represents the serum-availability, and the 'OUT SAMPLE' represents the colon-availability of phytoestrogens. It is known that bioavailability and biological activities of supplement phytochemicals are affected by digestion and absorption in the blood stream. During the digestion process, bioactive compounds may undergo structural and chemical changes, which may result in variations in their biological activities [49]. Phytoestrogens exist in the form of inactive glycosides in plant

tissues. A glycoside is a molecule containing a sugar bound to an aglycone, a non-sugar moiety and the active form of phytoestrogens. The isoflavone aglycones genistein and daidzein are present in soy primarily as the *p*-D glycosides genistin and daidzin, respectively [50]. The glycosidic forms of isoflavones are hydrolyzed by intestinal bacteria and enzymes and further metabolized through glucuronidation within the liver [50]. Daidzein is metabolized in the liver to equol or *o*-desmethylangolensin (*O*-DMA) and genistein to pethyl phenol [51], three potent phytoestrogens derivatives that can be detected in human serum and urine [37]. In our study, S4 contains glycosidic forms of isoflavones, namely genistin, daidzin, and glycerin. The GI digestion of these glycosides resulted in aglycones that induced significantly higher estrogenic activities at 400E equivalents (760.8 \pm 13 MU as 'IN SAMPLE' and 1402.2 \pm 1 MU as 'OUT SAMPLE', table 3) than the non-digested extract (443.9 \pm 6.1 MU, table 2), which contains only phytoestrogen glycosides.

All digested supplement extracts induced increasing estrogenic and antiestrogenic activities with increasing concentrations. The S2 'IN SAMPLE' (400E equivalents) showed the highest estrogenic activity (972.2 \pm 7.2 MU), whereas the highest antiestrogenic activity was displayed by S1 and S3 'IN SAMPLES' (400E equivalents) (table 3). S1 extract inhibited estradiol activity by 94%, whereas S3 extract completely inhibited the estradiol activity (table 3). Similarly, S4 'OUT SAMPLE' (400E equivalents) showed the highest estrogenic activity (1402.2 \pm 1 MU), whereas the highest antiestrogenic activity was detected in S5 'OUT SAMPLE' (table 3), which inhibited estradiol activity by 90%. The estrogenic activities of the other 'IN SAMPLE' extracts of dietary supplements ranked as follows: S3 and S4>S1>S5, and for 'OUT SAMPLES' the ranking was: S5>S2 and S3>S1 (table 3). The antiestrogenic activities of other 'IN SAMPLE' extracts of dietary supplements ranked as follows: S4 and S5>S2 and for 'OUT SAMPLES' the ranking was: S5>S3 and S4>S1>S2 (table 3). All extracts (400E equivalents) showed significantly higher estrogenic activities as 'IN SAMPLES' compared to the corresponding 'OUT SAMPLES,' except for S4 and S5 extracts. The much higher estrogenic activity in S4 and S5 'OUT SAMPLES' than 'IN SAMPLES' could be explained by the fact that both gastric digests of these supplements had high concentrations of aglycones and a good portion of them did not diffuse in the dialysis tubing during the 2-h intestinal digestion phase. For the same reason, except for S1 and S3 extracts, the other extracts showed significantly higher antiestrogenic activities as 'OUT SAMPLES' than 'IN SAMPLES'.

Table 3: Estrogenic and antiestrogenic activities of simulated GI digestion extracts of dietary supplements

Estrogenic activities (MU)				
In samples	100E Equiv	200E Equiv	300E Equiv	400E Equiv
S1	125.6 \pm 9.7	333.3 \pm 3 ^(a)	422.9 \pm 5 ^(a)	610.8 \pm 28
S2	403.3 \pm 15 ^(*)	405.1 \pm 10.6	437.8 \pm 25 ^(ab)	972.2 \pm 7.2
S3	470 \pm 23 ^(*)	564 \pm 9 ^(*)	620.8 \pm 4.3 ^{(a)(*)}	712.2 \pm 8.8 ^(c)
S4	564.8 \pm 30.9	614.4 \pm 9 ^(*)	660.8 \pm 5.4	760.8 \pm 13 ^(c)
S5	239.7 \pm 13.8	342.2 \pm 23 ^(a)	362.2 \pm 25 ^{(ab)(*)}	466.2 \pm 19.2
Out samples				
S1	107.4 \pm 1.4	130.7 \pm 4.1	192.2 \pm 2	215.6 \pm 5
S2	169.3 \pm 2.3	214.4 \pm 2.4	365.6 \pm 10	370 \pm 5 ^(b)
S3	411.5 \pm 4.3	444.1 \pm 2.1	812.2 \pm 3 ^(c)	369.6 \pm 1 ^(b)
S4	329.6 \pm 3.2	414.1 \pm 3	808.5 \pm 2 ^(c)	1402.2 \pm 1
S5	364.1 \pm 4	542 \pm 1	1092.2 \pm 2	1242.6 \pm 3
Antiestrogenic activities (%)				
In samples	100E equiv	200E equiv	300E equiv	400E equiv
S1	73.9 \pm 0.3	76.5 \pm 1 ^{(a)(*)}	85.6 \pm 1	93.9 \pm 0.3 ^(a)
S2	20 \pm 0.4 ^(b)	26.7 \pm 2 ^(b)	26.9 \pm 0.7	30.2 \pm 0.2
S3	28.1 \pm 0.1 ^(bc)	58.6 \pm 1 ^(ac)	58.2 \pm 0.3 ^(c)	100 \pm 0.1 ^(a)
S4	21.5 \pm 0.5 ^(bcd)	34.5 \pm 0.1 ^{(b)(*)}	69.3 \pm 7.2	71.6 \pm 4.2 ^(d)
S5	12.2 \pm 0.2 ^(bcd)	51 \pm 0.6 ^{(c)(*)}	58.3 \pm 0.02 ^(c)	63.3 \pm 0.2 ^(d)
Out samples				
S1	29.6 \pm 0.2	51.2 \pm 0.1	64.3 \pm 0.1	72.2 \pm 0.2
S2	32 \pm 1	34.4 \pm 0.1	50 \pm 0.03	60.6 \pm 0.2
S3	61.7 \pm 0.8	69.6 \pm 0.4	74.5 \pm 0.2	80.3 \pm 0.1 ^(c)
S4	26.6 \pm 0.1	42.3 \pm 0.2	60.4 \pm 0.2	81 \pm 0.1 ^(c)
S5	19.6 \pm 0.3	38.9 \pm 0.1	75.7 \pm 0.1	90.2 \pm 0.5

Results represent means \pm SD of three independent experiments (each experiment had two replicates; n=6). In each column, mean values with no superscript letters are significantly different from each other at $P \leq 0.01$; mean values with * are significantly different from each other at $P \leq 0.05$; mean values with same superscript letters are not significantly different (Tukey's test).

Although the simulated GI digestion of supplement extracts has its own limits, it showed that phytoestrogens in the supplement extracts were available for intestinal absorption and therefore validated the potency of the supplements under study. When the digested supplement extracts were compared to those of non-digested samples, S2, S4 and S5 'IN SAMPLES' showed higher estrogenic activities (972.2±7.2 MU; 760.8±13; and 466.2±4, respectively) (table 3) compared to the corresponding non-digested water extracts (829.5±20 MU, 443.9±6.1 MU; 281.1±6.7 MU, respectively) (table 2). Similarly, all 'IN SAMPLES' except for S2 showed higher antiestrogenic activities (table 3) as compared to the corresponding non-digested extracts at 400E equivalents (table 2). Although the *in vitro* GI digestion does not completely illustrate the *in vivo* digestion process, it represents a simple, affordable, and reproducible method for evaluating the availability of bioactive in the bloodstream [52]. The increased estrogenic and antiestrogenic activities of digested supplement extracts in this study show that the GI digestion increased the availability of phytoestrogens in those extracts.

Chemicals with estrogenic activities found in or isolated from botanical supplements interact with estrogen receptors in human tissues inducing transactivation of certain genes [53]. The endogenous estrogen hormone, 17β-estradiol, is known to be involved in the development of mammary glands and uterus, maintenance of pregnancy and bone density, protection from cardiovascular diseases, and alleviation of menopausal symptoms [54-56]. The high estrogenic and antiestrogenic activities of the five dietary supplements as simulated GI digests in this study show that they may be beneficial in the treatment or prevention of the estrogen-deficient conditions such as menopausal symptoms, osteoporosis. Snelten *et al.* [57] reviewed the effects of popular botanical supplements on the key steps of estrogen metabolism and reported that botanical supplements containing black cohosh, dong gui, ginger, hops, licorice and red clover modulated key estrogen metabolizing enzymes such as aromatase, catechol-O-methyltransferase (COMT), NAD (P) H-quinone oxidoreductase 1 (NQO1), and reactive oxygen species (ROS) and therefore could protect against estrogen carcinogenesis. In our study, S1 and S3 extracts showed higher antiestrogenic activity in 'IN SAMPLES' and S2, S4 and S5 showed the highest antiestrogenic activity in 'OUT SAMPLES' (table 3), consistent with results of other studies on the antiestrogenic and anti-proliferative effects of plant ingredients and isoflavones in recombinant estrogen receptor assays and cancer cell cultures [6, 10-13, 30, 31, 39, 44, 46, 58-60].

CONCLUSION

The dietary supplements investigated in this study revealed estrogenic and antiestrogenic activities in the estrogen-responsive transcriptional system in yeast, confirming the presence of phytoestrogens in the supplement formulations. Moreover, the simulated GI digestion demonstrated the availability of phytoestrogens for absorption in the blood stream. Among the supplements studied, those containing soy isoflavones and alfalfa ingredients had the highest estrogenic activities and could be more effective than supplements with a complex plant formulation in alleviating menopausal symptoms and treating osteoporosis. More studies on a larger number of dietary supplements are necessary to determine if the antiestrogenic activities of candidate phytoestrogens in the supplements are effective in preventing and treating estrogen-induced cancers. The GI digestion coupled with the transgenic yeast assays proved to be a convenient and powerful tool for assessing the *in vitro* estrogenic and antiestrogenic activities of dietary supplements and could be easily adapted to routine quality control procedures for supplements aimed at treating menopausal symptoms.

CONFLICT OF INTERESTS

Authors have no conflict of interests

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