ABSTRACT

Objective: The objective of the present article is to identify the most suitable Indian millet inulin for the growth of probiotic Lactobacillus casei and to evaluate the effects of the fortification vectors (probiotics and probiotic-prebiotic combination in immobilized conditions) and immobilization methods on the sustainability of L casei in a fortified Indian sweet (milk cake) preserved under different conditions.

Methods: Inulin was extracted from pearl, finger and great millets. The concentrations of L casei, grown on three millet inulins, were compared in 24 h batch culture. The L casei and probiotic-prebiotic combinations namely L casei-commercial inulin and L casei-pearl millet inulin were immobilized using entrapment, external and internal microencapsulation methods. The Indian milk cake samples were fortified with the immobilized probiotic cells, co-immobilized probiotic-prebiotic combinations. The fortified samples were preserved at different conditions (temperature: 4 °C and-20 °C; Time: 1-4 w). The sustainability of L casei in the preserved samples was determined using spread plate method and the cell concentrations were compared among all fortified samples.

Results: Pearl millet inulin is determined to be the most suitable millet inulin for the growth of L casei. The synergistic combination of L casei-pearl millet inulin, co-immobilized with internal gelation technique is the best fortification vector for the viability of L casei in preserved food samples.

Conclusion: The L casei, co-immobilized with pearl millet inulin through internal gelation technique, can be utilized as an effective fortification vector for the sustainability of probiotic cells in preserved Indian milk cakes and similar food samples.

Keywords: Probiotic, Lactobacillus casei (L casei), Prebiotic, Inulin, Pearl millet, Immobilization, Viability, Indian Milk cakes
understandable that the sustainability of probiotic cells is expected to increase by co-immobilization of prebiotics along with probiotics, no such research studies is available in the literature. To the best of our knowledge, this is the first study addressing the above topics to increase by co-immobilization of prebiotics along with probiotics, understandable that the sustainability of probiotic cells is expected to increase by co-immobilization of prebiotics along with probiotics, no such research studies is available in the literature. To the best of our knowledge, this is the first study addressing the above topics.

**Materials and Methods**

**Microorganisms**

*Lactobacillus casei* (2651 1951 RPK) culture purchased from NCIM, Pune were used.

**Materials**

Food grade inulin, peptone, yeast extract, sodium acetate, dipotassium hydrogen phosphate, tri-ammonium citrate, magnesium sulphate, manganese sulphate, sodium chloride, disodium hydrogen phosphate, potassium dihydrogen phosphate and sodium chloride purchased from Himedia, India, were used. Sodium alginate, acetic acid, tween80, potassium dihydrogen phosphate, disodium hydrogen phosphate, calcium chloride, glycerol, sodium dodecyl sulphate (SDS), beef extract, calcium carbonate, lactose and ethanol purchased from Merck, India, were used.

**Growth medium and other reagents**

**MRS medium**

**Composition**

Beef extract: 10 g/l; yeast extract: 5 g/l; peptone: 10 g/l; sodium acetate: 5 g/l; di-potassium hydrogen phosphate: 2 g/l; tri-ammonium citrate: 2 g/l; magnesium sulphate: 0.05 g/l; manganese sulphate: 0.05 g/l and lactose: 20 g/l; pH 7.

**Phosphate buffer (0.1M)**

**Composition**

Potassium di-hydrogen phosphate: 1.361 g/l; di-sodium hydrogen phosphate: 35 g/l (to adjust the pH at 7).

**Peptone water**

**Composition**

Peptone digest: 10 g/l; sodium chloride: 5 g/l.

**Equipments**

Magnetic stirrer (Remi, India), constant temperature bath (S. C Dey and co., India), digital weighing machine (Sartorius), autoclave (Gurpreet engineering works), hot air oven (G. B. Enterprises, Kolkata, India), B. O. D incubator (G. B. Enterprises, Kolkata, India), laminar airflow (G. B. Enterprises, Kolkata, India), mixer-grinder (Crompton Greaves, Kolkata, India) and high speed homogenizer (Plasto Crafts (Model: Superspin R-V/FM), India), refrigerator (Whirlpool; Corona Deluxe, India), freezer-20 °C (Blue Star, India) were used.

**Analytical instrument**

Optical microscope (Optika, Italy) was used.

**Extraction of inulin from Indian millets**

The prebiotic native sources, namely, pearl (bajra), finger (ragi) and great (jowar) millets were washed properly with distilled water. The same protocol followed for extraction of inulin from wheat, oat, daliah etc. has been used [15].

**Sterilization procedure**

MRS medium, 2 % sodium alginate solution and home-made sweet were autoclaved at 121 °C for 15 min.

**Pre-adaptation of Lactobacillus casei to different carbohydrate sources**

The probiotic bacteria, namely, *Lactobacillus casei* was pre-adapted to different carbohydrate sources, namely, lactose, commercial inulin and three millet inulins. For lactose, sterile modified de-Man Rogosa Sharpe (MMRS) medium was prepared by using 20 g/l lactose as the carbohydrate source. Four other stock cultures were also prepared using MMRS media in which the food grade commercial inulin and three millet inulins at a concentration of 0.3225 g/l were separately used as the carbohydrate source. Each medium was inoculated with 1 % (v/v) of *L. casei* and was incubated for 24h at 37 °C. The concentration of 0.3225 g/l was chosen for each inulin sample because the maximum growth of *L. casei* on commercial inulin had been observed by the present researchers at this level [15]. Since the purity of each inulin extract is dependent on the natural source, different quantity of inulin extracts was required to prepare the inulin solution of equal strength.

**Determination of the effectiveness of millet inulins for the growth of L. casei**

Batch studies were conducted in three 250 ml conical flasks using 100 ml modified MRS medium containing lactose and different millet inulins. The concentration of inulin in each conical flask was 0.3225 g/l and 20 g/l respectively. The pearl millet inulin, finger millet inulin and great millet inulin were respectively used in the first, second and third flasks. Each flask was seeded with 10 % (v/v) stock culture of *L. casei*. The flasks were kept in an incubator for 24 h at 37 °C. The cell concentration in each flask was measured by using spectrophotometric method determining the optical density of the growth medium at 600 nm. The cell concentrations of growth medium in three flasks at 24 h were compared to identify the most effective millet inulin for the growth of *L. casei*.

**Characteristics of millet extracts**

FTIR analysis of the extracts of three millet sample has been carried out to determine the presence of inulin qualitatively. The quantity of inulin in each extract has been determined spectrophotometrically following the protocols described by Samanta Koruri et al [16].

**Co-encapsulation and entrapment of Lactobacillus casei with and without inulin**

The protocols described by Chan et al., 2011 and Yoo et al., 1996 have been principally followed [17, 18] for the entrapment and co-entrainment. *L. casei* cultures, pre-adapted to lactose and commercial inulin were respectively spiked up to 20 g/l lactose and 0.3225 g/l commercial inulin and were individually mixed with 2% sterile sodium alginate solution in the volumetric ratio of 1:4. Each mixture was then added dropwise into a 1% CaCl₂ solution at room temperature and stirred continuously. The beads containing entrapment *L. casei* and co-entrapped *L. casei* and inulin were allowed to harden and were washed with saline water and stored at 4 °C.

**Co-encapsulation and encapsulation of L. casei with and without inulin through external gelation**

For the co-encapsulation and encapsulation of *L. casei* with and without inulin, via external gelation method, the protocol suggested by Song et al., 2013 have been principally followed [19]. *L. casei* cultures, pre-adapted to lactose and commercial inulin were spiked with the respective carbohydrates up to the concentration levels as used for pre-adaptation, i.e., 20 g/l and 0.3225g/l for lactose and inulin respectively. Each culture was mixed with sodium alginate solution in the volumetric ratio of 1:4. 1 ml of the mixture was stirred with 5 ml of vegetable oil. Tween 80 (emulsifier) and SDS were added to the resulting solution to maintain their concentrations at 0.2 % and 0.25 % respectively. This mixture was then stirred at 200 rpm for 30 min. CaCl₂ was added quickly but gently down the side of a beaker, in which the mixture was taken. The formed microbeads were kept undisturbed for 30 min. They were then filtered with muslin cloth. Finally, the microspheres were washed with 0.9% saline water containing 0.05% glycerol. The washed microbeads were stored at 4 °C.
Co-encapsulation and encapsulation of *L. casei* with and without inulin through internal gelation

The protocol suggested by Song et al., 2013 and Sultana et al., 2000 was principally followed with some modifications for the encapsulation of cells through internal gelation method [19, 20]. *L. casei* cultures pre-adapted to lactose, commercial inulin and the millet inulin, most effective for the growth of the probiotic microorganism, were at first spiked with the respective carbohydrate sources as described in the case of entrapment and external gelation i.e. up to 20 g/l and 0.3225 g/l for lactose and inulin samples respectively. Each culture was then individually mixed with 2% sodium alginate solution in the volumetric ratio of 1:4. CaCO₃ powder and Tween-80 were added to each mixture up to a concentration level of 0.8 g/l and 0.5% (v/v). This mixture was then combined with vegetable oil in the volumetric ratio of 1:5 and stirred at 300 rpm for 30 min. The temperature of the mixture was maintained at 25 °C by circulating water from a constant temperature bath and acetic acid was next added dropwise to the solution to release the Ca²⁺ from the insoluble CaCO₃ powder. Microcapsules were formed and were filtered with a muslin cloth and washed with 1% (v/v) aqueous Tween-80 solution and distilled water. The microcapsules were then stored at 4 °C.

Microscopic characterization of beads and microcapsules

The size analysis of the beads and microcapsules were done microscopically.

**Preparation of Indian cottage cheese for milk cakes**

Indian cottage cheese (chhena) was prepared from 1L milk. Firstly the milk was allowed to boil, followed by addition of lemon juice into it stirring continuously. When the milk curdled well, cottage cheese was strained using a muslin cloth and washed with water. After removing the excess water, 200 g chhena was obtained. The chhena was added to a pan and was mixed well with sugar and cardamom powder on a low flame until a mixture of thick consistency was obtained. The mixture was then cooled and kneaded properly and was autoclaved at 120 °C.

**Preparation of milk cakes with and without fortification with probiotic or probiotic–prebiotic microcapsules**

The sterile 200 g chhena, as described above, was divided into eight parts—first part was mixed with probiotic Ca-alginate beads spiked by lactose; second part was mixed with Ca-alginate beads spiked by commercial inulin; third part was mixed with probiotic microcapsules (external gelation) spiked by lactose; fourth part was mixed with probiotic microcapsules (external gelation) spiked by commercial inulin; fifth part was mixed with probiotic microcapsules (internal gelation) spiked by lactose; sixth part was mixed with probiotic microcapsules (internal gelation) spiked by commercial inulin; seventh part was mixed with probiotic microcapsules spiked by millet inulin, most effective for the growth of the probiotic microorganism and the eighth part was kept unfortified.

From each part, one milk cake weighing 25 g was prepared using sterile molds. All operations for the preparation of milk cake were conducted in a UV-sterilized hood. During the fortification, 10% (w/w) beads or microcapsules were used. The milk cakes are shown in fig. 1. All milk cakes were wrapped with sterile aluminium foils (fig. 1) before preservation.

**Viability assay of the probiotic cells in food products**

The fortified Indian milk cakes were stored at 4 °C and 20 °C over a period of 4 w. Unfortified samples were used as controls. Viability assay was performed every one-week interval as per the following protocol: in a UV-sterilized hood, 5 ml of phosphate buffer was first mixed with 1 g of food samples, and then 1 ml from this mixture was serially diluted with peptone water by 10 fold for six times. 0.1 ml of the diluted mixture was plated on MRS agar using spread plate technique. The agar plates were incubated under anaerobic condition for 24 h at 37 °C and the colonies on each plate were counted. According to this technique the concentration of cells per millilitre of the original phosphate buffer solution of the food sample, 10 times the count obtained from the spread plate method. Since 1g of food sample was originally present in 5 ml phosphate buffer, the concentration of cells in the food sample is 5×10⁷ times that obtained from spread plate method.

All experiments were carried out thrice, and the averages of three replicate experimental results have been reported.

**RESULTS AND DISCUSSION**

**Characteristics of millet extracts**

The presence of functional groups of inulin in the three millet samples was clearly indicated through the FTIR results (not shown). The % (w/w) of inulin in great, pearl and finger are provided in table 1. From the analysis of the table 1, it is clear that the inulin content of pearl millet extract is the highest among all millet extracts. The results could not be compared with the literature data due to unavailability.

**Table 1: Content of inulin in millet extracts**

<table>
<thead>
<tr>
<th>Millet type</th>
<th>Inulin in extract % (w/w)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Great</td>
<td>30.147±0.127000</td>
</tr>
<tr>
<td>Pearl</td>
<td>47.27±0.155885</td>
</tr>
<tr>
<td>Finger</td>
<td>32.003±0.076376</td>
</tr>
</tbody>
</table>

*Values are mean (n=3)±standard deviation (Standard deviation for sample population of three replicate observations corresponding to each mean result was calculated)

**Effect of naturally extracted inulin on the growth of *Lactobacillus casei***

The concentrations of *L. casei* obtained using the three different millet inulins, following the procedure described in the experimental section, were compared in fig. 2. From the analysis of the fig., it is clear that the pearl millet inulin is the most effective among all millet inulins with respect to the growth of *L. casei*. The effectiveness of pearl millet inulin was followed by that of great millet inulin and finger millet inulin. Better assimilation of the pearl millet inulin by *L. casei* in comparison to inulins derived from other millet sources was the underlying fact. This might be due to the difference in the degree of polymerization of inulin derived from different millet sources. This could be elucidated by the determination of the degree of polymerization defined by a number of monomer units (fructose) per molecule of prebiotic polymer, i.e., inulin derived from great, pearl and finger millets.

**Characterization of entrapped beads and microcapsules**

Table 2 shows the results of microscopic size analysis of beads and microcapsules obtained by entrapment and external/internal emulsification techniques. It is clear that the size of beads is much larger than those of microcapsules. On the other hand, the
microcapsules obtained through internal gelation method are much smaller than those obtained through external gelation. The results are in agreement with those reported by previous researchers [17, 20, 21].

Table 2: Microscopic size analysis of beads and microcapsules*

<table>
<thead>
<tr>
<th>Microencapsulation processes</th>
<th>Bead/microcapsule diameter</th>
</tr>
</thead>
<tbody>
<tr>
<td>Entrapment method</td>
<td>2.98333 mm (±0.028868 mm)</td>
</tr>
<tr>
<td>Emulsification/external gelation</td>
<td>298.333 µm (±2.886751 µm)</td>
</tr>
<tr>
<td>Emulsification/internal gelation</td>
<td>23.9333 µm (±0.11547 µm)</td>
</tr>
</tbody>
</table>

*Values are mean (n=3)±standard deviation

Viability of the probiotic cells in fortified food products after preservation

In fig. 4a and 4b the photographs of the spread plates obtained for unfortified food samples and that fortified with encapsulated L. casei are provided. It was clearly indicated that while the control was fully contaminated, no contamination was present in the case of the fortified one. This establishes the anti-microbial activity of the immobilized probiotic culture against contaminating microorganisms and the elongation of shelf life. Due to unavailability of literature data, the results could not be compared.
In table 3, the concentrations of probiotic cells in the fortified Indian milk cakes have been provided.

<table>
<thead>
<tr>
<th>Weeks</th>
<th>Microcapsules (External gelation) (x10⁷)/ml</th>
<th>Beads (Entrapment) (x10⁷)/ml</th>
<th>Microcapsules (Internal gelation) (x10⁷)/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>PRO</td>
<td>PRO+CI</td>
<td>PRO</td>
<td>PRO+CI</td>
</tr>
<tr>
<td>A</td>
<td>B</td>
<td>A</td>
<td>B</td>
</tr>
<tr>
<td>1</td>
<td>2.2±0.5</td>
<td>0.08±0.00</td>
<td>13.6±2.5</td>
</tr>
<tr>
<td>2</td>
<td>12.7±3</td>
<td>9±0.4</td>
<td>10.6±2.7</td>
</tr>
<tr>
<td>3</td>
<td>6.7±1</td>
<td>3±0.5</td>
<td>6.6±1.2</td>
</tr>
<tr>
<td>4</td>
<td>3.3±1</td>
<td>0.577±0.00</td>
<td>14.6±6.1</td>
</tr>
</tbody>
</table>

For table 3, PRO: fortified with only probiotic; PRO+CI: fortified with probiotics and commercial inulin; PRO+PMI: fortified with probiotics and pearl millet inulin; A: preservation at 4 °C; B: -20 °C. *Values are mean (n=3)±standard deviation.

The concentrations of viable _L. casei_ cells in milk cake samples fortified by either beads or microcapsules containing either only probiotics or mixture of probiotics and probiotic commercial inulin derived from chicory and pearl millet, were experimentally determined for 1, 2, 3 and 4 weeks. As per expectation, the prebiotic inulin was effective in prolonging the preservation period. This might be due to the formation of ice crystals on prolonged preservation causing the destruction of probiotic cells. When the effect of immobilization method on the viability of cells was analysed, it was revealed that whatever might be the fortification vector and the preservation time and condition, the best viability of cells was shown by the microcapsules produced through internal gelation method. This was justified by the fact that the immobilized probiotic cells in the food matrices were protected from external stresses and the biodegradation by the food enzymes.

The concentrations of _L. casei_ of the probiotic microorganism, in fortified Indian milk cake intake even after 4 w of preservation. The viability of probiotic cells in milk cakes fortified by microcapsules produced through internal gelation containing combinations of _L. casei_ and either commercial inulin or pearl millet inulin was indicated by the fig. 5.

**Fig. 5:** Comparison of effect of microcapsules obtained by internal gelation method for both the combination probiotic-commercial inulin and probiotic-pearl millet inulin at 4 °C and-20 °C (Results are average of those obtained from three replicate observations. Standard deviation varies from 0 to 0.57735)

It was clearly revealed from the close analysis of the fig. that the effectiveness of pearl millet inulin in the enhancement of sustainability of probiotic cells was marginally better than that of commercial inulin. This might be due to the difference in the degree of polymerization of inulin derived from chicory and pearl millet inulin. However, more experiments have to be conducted to reveal the exact fact. Therefore, the substitution of commercial inulin by this nonconventional variety of inulin should be attempted in commercial scale.

**CONCLUSION**

Under this study effects of different immobilization methods—Ca-alginate entrapment; external and internal microencapsulation, the fortification vectors, namely the probiotic cells, combination of probiotic cells and commercial inulin/pearl millet inulin, preservation temperature, 4 °C and-20 °C and period of 1-4 w on the sustainability of the probiotic microorganism, _L. casei_ in fortified Indian milk cake has been investigated systematically. The pearl millet inulin has been selected based on its best performance among three millet inulins (great, pearl and finger millet) with reference to the growth of the probiotic cells, namely, _L. casei_ in free (not immobilized) culture. To the best of our knowledge, this is the first investigation of this type. This study indicates that microencapsulation techniques including internal gelation, external gelation and direct calcium-alginate entrapment methods can significantly support the survival of the probiotic microorganisms in the fortified food product, namely Indian milk cake over a preservation period of up to 4 w at both 4 °C and-20 °C.
However, on the basis of the viability of probiotic cells, the internal gelation technique is found to be superior to the other two immobilization techniques. The results show that the contribution of different immobilization methods in increasing the viability of probiotic bacteria in preserved and fortified milk cake follows the order: internal gelation-external gelation-Ca-entrapped. Co-immobilized probiotics with prebiotics, namely, commercial inulin/pearl millet inulin markedly increase the viability of probiotic cells over that obtained through fortification by immobilized probiotics. Thus, the use of both commercial food grade inulin and naturally derived pearl millet inulin has positive effects on the survival of the microencapsulated probiotic cells in the food products during storage. Results of the study suggest that pearl millet inulin offer marginally more potential as prebiotic than the commercial food grade inulin. Overall, the fortification results in elongation of the shelf life of the preserved due to the sustenance of growth of probiotic microorganisms which ultimately inhibit the growth of pathogenic microorganisms in the food products into which they are added. It is expected that the outcome of the present research study can be utilized for the application of pearl millet and the conventional chicory inulin (commercial) for the production of fortified food products using synergistic probiotic-prebiotic combinations co-immobilized through internal gelation technique.

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CONFLICT OF INTERESTS

Declared none

REFERENCES


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