NEW FLAVONOIDS FROM THE AERIAL PARTS OF POLYGONUM EQUISETIFORME SM (POLYGONACEAE)

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ABSTRACT

Objective: The current study was to deal the isolation and identification of secondary metabolites from Polygonum equisetiforme and evaluation of antioxidant activity of its extract.

Methods: The methanol-water extract (7:3) of the air-dried aerial parts of Polygonum equisetiforme was fractionated and separated to obtain the isolated compounds by different chromatographic techniques. Structures of these compounds were elucidated by UV and 1D⁄2D H⁄C NMR spectroscopy and compared with the literature data. The crude extract was evaluated for in vitro antioxidant activity using the 2,2 diphenyl dipicryl hydrazine (DPPH) method.

Results: Ten secondary metabolites were isolated from Polygonum equisetiforme in this study. Of which three new flavonoids named as 3,5,7,2',5' pentahydroxyflavone 3-O-β-D-glucopyranoside (1), 3,5,7,2',5' pentahydroxyflavone 3-O-β-D-glucopyranoside 8 O-sulphated (2) and quercetin 3′-O-β-D-glucorinde methyl ester 8-sulphated (3) as well as quercetin 3-O-β-D-glucorinde methyl ester (4), quercetin 3′-O-β-D-glucopyranoside (5), quercetin 7-O-β-D-glucopyranoside (6), quercetin (7), myricetin (8), P. methyly gallic acid methyl ester (9) and gallic acid (10). The antioxidant potential of P. equisetiforme extract was evaluated by investigating it's total phenolic and flavonoid content and DPPH radical scavenging activity whereby the extract showed significant antioxidant activity (IC₅₀ = 37.45 μg/ml). The total phenolic and flavonoid content was found to be 130.79±5.502 and 45.8±1.63 μg/ml, respectively.

Conclusion: Polygonum equisetiforme is a promising medicinal plant, and our study tends to support the therapeutic value of this plant as an antioxidant drug.

Keywords: Polygonum equisetiforme, Aerial parts, New flavonoid, Antioxidant activity

INTRODUCTION

Family Polygonaceae comprises about 40 genera and 800 species [1], distributed throughout the world, especially in temperate and warm regions. Polygonum is the largest genus belonging to the family Polygonaceae, which comprises about 300 species. According to Boulos [2] only six species are found in Egypt. P. arena strum, P. avicular, P. bellardii, P. equisetiforme, P. maritimmn and P. plebeium. The secondary metabolites identified in the species of the Polygonum genus are flavonoids [3–6], anthraquinones [7], lignan glycosides [8], polysaccharides [9], phenylpropanoid glycosides [10], sesquiterpene [11-13] and stilbenes [14, 15]. Numerous polygonum species are frequently used in traditional medicine. Some species are used in the treatment of cough, diarrhea, diuretic and to treat urinary inflammation [16]. Turkish ethnomedicine reported the use of P. cognatum for the treatment of urinary inflammation and as diuretic agents [17]. The root extracts of P. multiflorum possess anti-inflammatory [18], antioxidant [19], anti-HIV [20], and liver protective effects [21] as well as some monomeric compounds isolated from its roots. The medicinal properties of Polygonum species are due to its high constituents of bioactive compounds. Drimane sesquiterpenoids, non-sesquiterpenoids and sulphated flavonoids are characteristic compounds of some Polygonum species which reveal various biological activities, such as antiinflammatory, antitumour and less aldose reductase inhibitory [22-23]. Polysaccharide derivatives with radical scavenging and anti-tumour activities have been isolated from many Polygonum species [24, 25]. Flavonoids and chalcones isolated from some species of Polygonum on the other hand exhibit strong antioxidant effects role against oxidative stress damages, such as atherosclerosis or cancer [26-29]. No comprehensive screening studies have yet been published on P. equisetiforme species. The current study was focused on the isolation and identification of flavonoids from the aqueous methanol extract of P. equisetiforme
Mediterranean region, A sample of P. equisetiforme purchased from Merck Co. (Santa Ana, CA, USA). Extraction and isolation of the plant was confirmed at low temperature. Identification of the plant was confirmed at the National Research Centre, Dokki, Cairo, Egypt.

The aerial parts of P. equisetiforme (aerial parts) were collected from Egypt during April 2013 (flowering date). A voucher specimen (P75) is deposited in the Herbarium of the Botany Department, Faculty of Science, Cairo University, Egypt.

Plant material
A sample of P. equisetiforme (aerial parts) was collected from Mediterranean region, Egypt during April 2013 (flowering date). The samples were separately air-dried, powdered and kept in tightly-closed amber coloured glass containers and protected from light at low temperature. Identification of the plant was confirmed by Botany Department, Faculty of Science, Cairo University, Egypt [2]. A voucher specimen (P75) is deposited in the Herbarium of the National Research Centre, Dokki, Cairo, Egypt.

Extraction and isolation

The aerial parts of P. equisetiforme (1.75 kg) were crushed and extracted with 70% methanol by soaking at room temperature and the methanol extract was evaporated under reduced pressure and lyophilized (120 gm). A sample (100 gm) of the dry extract was fractionated by chromatography on polyamide 65 column. The column was eluted with water and with water–methanol step gradient. The obtained fractions (500 ml of each fraction) were subjected to paper chromatography using BAW and 15% acetic acid as developing solvents, and the similar fractions were collected together to give six major fractions (I–VI). Separation of fractions II (200 mg) on Sephadex column LH-20 CC using 3:6% EtOH gave two sub-fractions. The first sub-fraction was purified on Sephadex column LH-20 CC using EtOH–H2O (1:1) as solvent system and gave a pure sample of 1 (25 mg), the second sub-fraction was subjected to Sephadex column LH-20 CC using EtOH–H2O (3:7) to give two compounds which were further purified on Sephadex CC using EtOH HPLC as solvent to give pure samples of 2 (18 mg) and 3 (30 mg). Fraction III (180 mg) have been applied on Sephadex column LH-20 CC using butanol-water saturated as solvent system and gave one compound which was further purified on Sephadex LH-20 CC using methanol HPLC to afford the purified sample of 4 (25 mg). Elution of fraction IV (150 mg) by n-BuOH–water saturated; afforded two compounds which were separated by preparative paper chromatography using 15% AcOH as eluent and were further purified on Sephadex LH-20 CC using EtOH–H2O (1:1) to give pure samples of 5 (30 mg) and 6 (28 mg). Consecutive CC on Sephadex LH-20 with n-butanol-water saturated for elution of fraction V (240 mg) give two sub-fractions which were separated by preparative paper chromatography using BAW (4:1:5) as eluent. These two compounds were further purified on Sephadex LH-20 CC using saturated n-butanol-water as eluent to afford pure samples of 7 (30 mg) 8 (26 mg). Fraction VI (70 mg) has been separated on sephadex LH-20 CC using n-butanol-water saturated to give two compounds which were further purified on sephadex LH-20 CC using MeOH: H2O (1:1) to give pure samples of 9 (30 mg) and 10 (25 mg).

3, 5,7,2′,5′ pentahydroxyflavone 3-0-β-D-glucopyranoside (1)

\( R_1 \) values (x100) 0.6 in (BAW), 0.8 in (H2O-ac 15%); UV Spectral Data \( \lambda_{max} \) (nm): MeOH: 253, 263sh, 344, NaOMe: 284, 322sh, 372, NaAc: 272, 336sh, 390, NaNaAc/H2O: 260, 290sh, 375, AlCl3: 260, 415sh, 408, AlCl3/NaOMe: 390 nm; 1H NMR (400 MHz, DMSO-d6) \( \delta \) (ppm): 8.28 (1H, broad singlet, H-6'), 7.35 (1H, dd,
RESULTS AND DISCUSSION

The methanol-water extract (7:3) of the aerial parts of Polygonum esquifolium was subjected to fractionation on a polyamide 6s column gave three new compounds besides seven known compounds (1-10, fig. 1). The isolated compounds undergo conventional chemical and spectroscopic methods of analysis (UV, 1/2D NMR) as well as chromatography to elucidate their chemical structures.

Compound 1 was obtained as a yellow amorphous powder which showed chromatographic properties (dark purple spot on paper chromatogram under UV light). It gave lemon yellow color with Nature staff reagent [33] characteristic of flavonoids bearing a free hydroxyl at 5-position. UV spectral properties of 1, in methanol and after the addition of diagnostic reagents detected the presence of free 7-OH group [34]. H NMR spectrum of 1 (DMSO-d6, room temperature) displayed aromatic signals at δ ppm (d, J = 8.8 Hz) 7.35 (d, J = 8.4, 1.9 Hz) and 6.84 (d, J = 8.4 Hz) describable to H-6, H-4 and H-3 of 2', 5'-dihydroxy B-ring [35]. The AM-spin coupling system of two different proton signals at 6.20 and 6.40 ppm with Jvalue 1.5 Hz, assignable to H-6 and H-8, respectively concluded the 7, 3 dihydroxy A-ring. Also, H NMR displayed one signal, a hexose anomeric proton resonance at δ 5.20 (d, J = 6.8 Hz, H-1') specifying the presence of sugar with β-configuration. 1C experiments showed methylene group, 10 methine and 10 quaternary carbons. The 1C NMR spectrum of compound 1 showed one signal at δ 177.59 assigned to the carbonyl carbon. Heteronuclear 1H-13C correlation experiments (HMOC, HMBC) led to full assignments of the 1H and 13C NMR chemical shifts of compound 1. HSQC proton resonances are in good agreement with those of each corresponding carbon. However, the unequivocal assignment could be confirmed by HMOC and HMBC which proved that the signals at 156.50, 146.77, 160.94, 157.60, 148.45 and 144.81 are assignable to C-2, C-7, C-5, C-9, C-2' and C-5' respectively. In the HMBC experiments, the correlation was observed between δ H 5.20 (H-1') with δ C 134.07 (C-1). Investigation of the electron-difference chromatogram showed the migration of 2 but with degradation, indicating the presence of a sensitive sulphate group. 1H NMR data of 2 (DMSO-d6, room temperature) displayed aromatic signals at δ ppm 8.37 (broad singlet) 7.42 (dd, J = 8.4, 1.9 Hz) and 6.83 (d, J = 8.4 Hz) describable to H-6, H-4 and H-3 of 2', 5'-dihydroxy B-ring. The A-ring has been concluded based on the spin coupling system of one singlet upfield proton at δ 5.93 ppm assignable to H-6 together with the absence of meta coupling suggested that the position 8 is substituted. Also, H NMR displayed one signal, a hexose anomeric proton resonance at δ 5.17 (d, J = 6.8 Hz, H-1') indicating the presence of sugar with β-configuration.

Compound 2 was obtained as a yellowish powder which showed a dark purple spot on PC under UV light. UV spectral data showed the intrinsic chromatographic behaviour of flavonoid 3-O-glycoside with/without substitution in the A-ring. Compound 2 gave a positive sulphate test (a white ppt. with BaCl2) [36]. Investigation of the electrophoresis chromatogram showed the migration of 2 but with degradation, indicating the presence of a sensitive sulphate group. 1H NMR data of 2 (DMSO-d6, room temperature) displayed aromatic signals at δ ppm 8.37 (broad singlet) 7.42 (dd, J = 8.4, 1.9 Hz) and 6.83 (d, J = 8.4 Hz) describable to H-6, H-4 and H-3 of 2', 5'-dihydroxy B-ring. The A-ring has been concluded based on the spin coupling system of one singlet upfield proton at δ 5.93 ppm assignable to H-6 together with the absence of meta coupling suggested that the position 8 is substituted. Also, H NMR displayed one signal, a hexose anomeric proton resonance at δ 5.17 (d, J = 6.2 Hz, H-1') indicating the presence of sugar with β-configuration.

Compound 3 was obtained as a yellowish powder which showed a dark purple spot on PC under UV light. UV spectral data showed the intrinsic chromatographic behaviour of flavonoid 3-O-glycoside with/without substitution in the A-ring.
about 4 ppm proving a sulphate substitution at C-8 [37]. Heteronuclear 1H-13C correlation experiments (HMQC, HMBC) exhibited confirmation of the 1H and 13C NMR chemical shifts of compound 2. Also, the HSQC proton resonances are in good agreement with those of each corresponding carbon. In the HMBC spectrum (fig. 2) the proton signal at δ H 5.17 (H-1) displayed a long-range correlation with the carbon appeared at δc 133.60 (C-3) suggesting a glucosyl moiety is attached to C-3. The proton signal at δ N 8.37 (H-6) showed long-range correlation with δc 120.38 (C-1) and δc 151.79 (C-2) indicated that the flavones with unusual B-ring. On the basis of these results, compound 2 was elucidated to be 3,5,7,2',5' pentahydroxyflavone 3-O-P-D-glucopyranoside 8-C-sulphated. It is the first time for the isolation of this compound from any plant.

Compound 3 was isolated as yellow amorphous powder and showed dark purple fluorescent spot turned to yellow and orange fluorescence on PC with ammonia vapours and Naturstoff spray reagents respectively [33]. The UV spectral of compound 3 in MeOH were similar to those of 3-substituted quercetin except for a slight bathochromic shift of band II (≈12 nm) on the addition of NaOAc indicating to a free OH on C-4. Also, the bathochromic shift in band II (≈20 nm) accompanied with the increase in the intensity of band I on the addition of NaOMe indicated a free OH on C-4. Also, the bathochromic shift in band II (≈20 nm) was also proved from the bathochromic shifts in the band I (δ +12 nm) on the addition of NaOAc and δ(17 + nm) in band II on the addition of H2B[4, 38]. Compound 3 gave a positive sulphate test (a white ppt. with BaCl2) [36]. Investigation of the electrophoresis chromatogram showed the migration of 3 but with degradation, indicating the presence of a sensitive sulphate group. The 1H NMR spectrum of 3 (CD3OD, room temperature) showed aromatic signals at δ ppm 7.74 (d, J = 2 Hz), 7.72 (dd, J = 8.6, 1.8 Hz), and 6.86 (dd, J = 8.6 Hz) describable to H-2', H-6' and H-5' of 3, 4'-dihydroxy B-ring of flavonoids [39]. Moreover, the H-6 proton appeared as a singlet (δ 6.34) and absent of meta-coupling, suggesting the presence of the 8'-C-substituted. 13C NMR spectrum showed twenty-two signals seven in the aliphatic region for the sugar moiety and methyl ester group and the remaining for the quercetin unit. The DEPT spectrum of compound 3 revealed for the presence of one methyl group at δ 5.147 and four methine aromatic carbons. The presence of β-conformation of glucuronic acid methyl ester in compound 3 was confirmed by the anomeric carbon resonance at δ 103.06 and up the field of C-6" at δ 169.27. The downfield signals (at δ 177.64 and 169.27) were assigned to the two carbonyl carbons. The signals at δ 103.06 (C-1''), 73.89 (C-2''), 75.68 (C-3''), 71.34 (C-4''), 75.89 (C-5''), and 169.22 (C-6'') were achieved long-range correlation with δ 133.60 (C-3) and δ 151.79 (C-2) indicating the presence of a glucosyl moiety. The presence of the-C-sulphated group substitution at position 8 in A-ring was also proved from the HMBC correlation from H-1 to δ 133.60 that suggested the β-glucuronic acid methyl ester was connected to C-3 of the quercetin moiety. From the above data, compound 3 is deduced to be Quercetin 3-O-P-D-glucuronic 6''-methyl ester 8-C-sulphated. This compound was isolated for the first time from natural source.

The phenolic and flavonoid contents were 130.79±5.502 GAE/G extract and 45.81±1.63 mg RE/G extract, respectively.

The antioxidant activity assay

The imbalance between oxidizing agents and the antioxidant defend system may lead to damage of the macro-molecules [40] which consequently play a critical role in the pathogenesis of various diseases [41]. The plants have flavonoids and polyphenolic constituents that have remarkable antioxidant activity [42]. The hydromethanol extract of the aerial parts of P. equeiforme exhibited a good anti-oxidant ability especially because of their phenolic compounds. It showed the significant scavenging activity of the crude DPPH radical by IC50 = 3.74G AE/G. The antioxidant potential of the plant proved the correlation between the phenolic and the flavonoid content and its antioxidant activity; indicating that these phenolic and flavonoid metabolites may be useful therapeutic agents.

CONCLUSION

The plants have flavonoids and polyphenolic constituents that have remarkable antioxidant activity. In this study, three new flavonoids named as, pentahydroxyflavone 3-O-P-D-glucopyranoside, 3, 5, 7, 2', 5' pentahydroxyflavone 3-O-P-D-glucopyranoside 8-C-sulphated and Quercetin 3-O-P-D-glucorinde 6''-methyl ester 8-C-sulphated isolated from the aqueous methanol extract ofPolygonum equeiforme Harv. The extract of the arial parts of P. equeiforme exhibited a good anti-oxidant ability especially because of their phenolic compounds, including phenolic acids, flavonoids and their derivatives. Thus, further investigations are required in order to make optimal use of this plant.

CONFLICT OF INTERESTS

Declared none

REFERENCES


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