DEVELOPMENT AND VALIDATION OF CARBAMAZEPINE PLASMA CONCENTRATIONS MEASUREMENT AND ITS APPLICATION ON EPILEPSY PATIENTS

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ABSTRACT

Objective: To develop and validate high-performance liquid chromatography with photodiode array (HPLC-PDA) detector as a method for measuring carbamazepine plasma concentrations in epilepsy patients treated with monotherapy or polytherapy.

Methods: Carbamazepine was extracted from epilepsy patients’ plasma through liquid-liquid extraction, using protein precipitation with chloroform. Analysis was performed using HPLC with Inertsil DS-4 C18 (4.6 x 150 mm), 5 μm particle size column. The optimal condition for separation was established in a mobile phase consisting of acetonitrile:water (50:50) at a flow rate of 1.0 ml/min, detected by PDA detector at 220 nm. Propylparaben was used as the internal standard. The retention time was 3.5 min.

Results: Linearity was obtained over a concentration range of 0.5-16 μg/ml with r = 0.999. The method showed good intra- and inter-day precision and accuracy of more than 90% difference (% diff) and 95% relative standard deviation (RSD). Lower limit of quantification (LOQ) was 0.5 μg/ml and lower limit of detection (LOD) was 0.2 μg/ml with 100% accuracy and more than 90% precision. Recovery test was nearly 100%. Stability of carbamazepine plasma concentration in 3 epilepsy patients was measured on the first and third month of treatment, ranging between 83.5 to 98.7%. When used to compare carbamazepine as a monotherapy versus polytherapy, the method showed good selectivity.

Conclusion: The present HPLC method was valid for measuring carbamazepine plasma concentrations in epilepsy patients treated with monotherapy or polytherapy. This method meets the standard in the EMEA guideline in terms of linearity, precision, and accuracy, also selectivity in epilepsy patients treated with polytherapy.

Keywords: Carbamazepine plasma concentration, HPLC, Validation, Epilepsy patients, Monotherapy or polytherapy

INTRODUCTION

Carbamazepine is a hydrophobic, weakly acidic, non-polar aromatic ester, highly soluble in alcohol (fig. 1). It is a first-generation anti epileptic drug (AED), used as a first-line therapy for focal epilepsy. However, 30% of patients with focal epilepsy do not respond to maximum dosages of carbamazepine, resulting in the need to administer polytherapy with additional AEDs to control seizures, which may lead to drug interactions. Measurements of specific drug concentrations in the plasma of patients on polytherapy may not be reliable due to the selectivity of multidrug measurements.

There are various methods to measure levels of carbamazepine in plasma. One of the simplest and most accurate methods is high-performance liquid chromatography (HPLC) [1]. Previous studies indicated that HPLC was adequate, effective, suitable, simple, rapid, highly sensitive, also had a good intra-day and inter-day precision and an accuracy of >90% [1-6]. Considering many patients also use other AEDs, the physician has to make sure that the measurement of carbamazepine plasma levels is unaffected by the presence of other AED. Only few publications on methods of plasma concentration measurements described the implementation of measurements in patients, especially in polytherapy. The rationale of this study was to develop and validate a simple, accurate, rapid, precise, reproducible HPLC method with photodiode array (PDA) detection for quantification of carbamazepine plasma concentration. In this study, we developed a method to measure carbamazepine plasma concentration in epilepsy patients treated with carbamazepine as a monotherapy or polytherapy.

MATERIALS AND METHODS

Instruments

We used a HPLC system with separation module by Waters 2695, Waters 2998 PDA Detector (Waters, USA), Inertsil column ODS-4 C18 (4.6 x 150 mm), 5 μm pore size (GL Sciences, Japan), a vacuum filter, and Whatman cellulose nitrate and 0.45 μm nylon membrane filter.

Fig. 1: Chemical structure of carbamazepine [1]

Chemical solutions and reagents

Carbamazepine (cat no.94496, Fluka), propylparaben, acetonitrile, methanol, aquabidest, and hydrochloric acid were purchased from E-Merck. Gobazam, phenytoin, phenobarbital, clonazeapam, and folic acid are generic drugs produced by Indonesian drug manufacturers. Other drugs used include Depakote™ containing valproic acid (Abbott), Lamictal™ containing lamotrigine and Keppra™ containing levetiracetam (Glaxo Smith Kline), and Topamax™ containing topiramate (Soho).

Preparation of standard solutions

Five mg of carbamazepine and propylparaben were weighed, separately put in a 0.5 ml measuring flask, then diluted with methanol until the desired concentrations were reached. The concentration of each standard solution was 1 mg/ml and each solution was stored at 4 °C.

Optimum wavelength analysis

Carbamazepine standard solution was diluted in methanol to make a
solution with a concentration of 10.0 μg/ml. Wavelength curve of the carbamazepine standard solution was measured with PDA to obtain optimum wavelength analysis.

Sample preparation

Mobile phase composition

Water, acetonitrile and methanol were filtered with a vacuum pump and Whatman 0.45 μm filter and then degassed for 5 min. A 30 μl solution consisting of 1 mg/ml carbamazepine and propylparaben was injected to the HPLC instrument with a mobile phase containing water:acetonitrile in a 50:50 ratio. The flow rate was 1 ml/min.

Carbamazepine plasma sample preparation

Working solution consisted of 250 μl blank plasma, 50 μl carbamazepine standard in different concentrations, 50 μl propylparaben (100 μg/ml) and 50 μl HCl 10% was added with 3 ml of chloroform after a 15-s vortex. After another 1-min vortex, the working solution was centrifuged in 4000 rpm for 10 min. Then the supernatant was removed to glass tubes to be evaporated with nitrogen in 40 °C for 20 min. Dry extract was reconstituted with mobile phase solution and 30 μl of this solution was injected to the HPLC.

Methods of validation

The validation method was done according to The 2011 Guideline on Bioanalytical Method Validation by the European Medicines Agency. Typical validation characteristics to be considered according to the guidelines include calibration curve, the lower limit of quantification, within-and between-run accuracy and precision, carry-over, dilution integrity, selectivity and stability [7].

Application for epilepsy patients

This study was part of a clinical research approved by the Ethics Committee of Faculty of Medicine Universitas Indonesia with ethical approval number 895UN2. F1/ETIK/2014. A number of epilepsy patients undergoing monotherapy or polytherapy were included in this study as a part of method development and validation.

RESULTS

The HPLC method

Despite the many options available to measure carbamazepine plasma levels, HPLC was considered the most appropriate. Separation was performed using a 150 mm column. Optimum wavelength for analysis detected by PDA was λ=220 nm. The composition of 50:50 water: acetonitrile was chosen as the mobile phase. Retention time was 3.5 min at a flow rate of 1 ml/min for a 30 μl-injected volume. Total run time for each assay was approximately 9 min (fig. 2).

Range, linearity and calibration curve

The desired range was selected based on the need to detect a specific range of carbamazepine concentration for clinical use. The therapeutic range used in clinical practice based on an international epilepsy guideline is 4-12 μg/ml [8]. To date, there is no available data about therapeutic concentration range of carbamazepine in Indonesian subjects. Based on clinical experience, some patients may respond to doses lower than the recommended posology. In contrast, some patients may show a partial response to a certain therapeutic dose, but failed to achieve complete response despite being given a higher dose; thus the range of 0.5 to 16.0 μg/ml was decided for this study.

A minimum of 5 concentrations was recommended by the international conference of harmonisation (ICH) guideline to establish linearity [9]. In this study, linearity was evaluated across the range of 0.5, 1.0, 2.0, 4.0, 8.0, 16.0 μg/ml and low, medium and high concentration at 1.5, 3.0 and 12.0 μg/ml, respectively, along with a blank sample without internal standard, and a zero sample with internal standard.

Extraction was done following the sample preparation procedure. Plot signal of peak area (y) produced by the HPLC method at each concentration (x) was calculated using linear regression. The correlation coefficient of calibration curve was $y = 0.2254x + 0.0321$ (r = 0.999) (fig. 3).
Lower limit of quantification (LOQ) and detection (LOD)

Lower limit of quantification (LOQ) was determined at 0.5 μg/ml, the minimum level at which carbamazepine can be reliably detected. Lower limit of detection (LOD) was determined at 0.2 μg/ml. The accuracy of LOQ was 100.31% and precision was 92.5%. Accuracy of LOD was 101.94% and precision was 96.56%.

**Precision**

The ICH recommends assessing precision using a minimum of 9 assessments covering specific ranges, for instance, assessing 3 different concentrations, each with 3 replicates, for repeatability. Intermediate precision was expressed in different days. Assessment of intra-day precision in this study was done on 5 replicates of samples with low, medium, and high concentration.

The intra-day precision was more than 90% difference (% diff) and 95% relative standard deviation (RSD). Inter-day precision was assessed using 6 replicates in 9 d, also resulting in more than 90% diff and 95% RSD (table 1).

**Table 1: Precision intra-and inter-day of carbamazepine plasma**

<table>
<thead>
<tr>
<th>Concentration (μg/ml)</th>
<th>Intra-day precision</th>
<th>Inter-day precision</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>% diff</td>
<td>RSD(^a) (%)</td>
</tr>
<tr>
<td>1.5</td>
<td>98.1</td>
<td>3.40</td>
</tr>
<tr>
<td>3</td>
<td>103.6</td>
<td>3.57</td>
</tr>
<tr>
<td>12</td>
<td>104.6</td>
<td>4.13</td>
</tr>
</tbody>
</table>

\(^a\)RSD = relative standard deviation

**Accuracy**

Accuracy was determined using a minimum of 9 assessments for precision. Accuracy is reported as percent recovery. The US food and drugs administration (FDA) criteria suggested a mean percent recovery of 100±2% at each concentration and within 80-120% of target concentration [8]. In this study, the mean of recovery at 1.5, 3 and 12 μg/ml was 97%, 91%, and 94%, respectively (table 2).

**Table 2: Accuracy and mean of recovery**

<table>
<thead>
<tr>
<th>Concentration (μg/ml)</th>
<th>Recovery (%)</th>
<th>% diff</th>
<th>% RSD(^a)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.5</td>
<td>97</td>
<td>10.88</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>91</td>
<td>8.11</td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>94</td>
<td>3.92</td>
<td></td>
</tr>
</tbody>
</table>

\(^a\)RSD = relative standard deviation

**Carry-over**

Carry-over was assessed through a 5-time consecutive injection of blank samples followed by highly concentrated calibration standard. The carry-over was less than 20% of the lower limit of quantification and 5% for the internal standard.

**Dilution integrity**

Dilution integrity was demonstrated by spiking the matrix with an analyte with a concentration above the LOQ, then the sample was diluted with the blank matrix [7]. Every dilution factor had 5 determinants. The accuracy and precision of the dilutions were within the 15% range.

**Selectivity**

Identification of carbamazepine was determined by evaluating the presence of peak area as a positive result, and no-presence as negative. Impurities were evaluated by adding pure substances of other antiepileptic drugs such as clobazam 10 ppm, phenytoin 10 ppm, phenobarbital 10 ppm, clonazepam 10 ppm, folinic acid 10 ppm, valproic acid 10 ppm, lamotrigine 10 ppm, levetiracetam 10 ppm, and topiramate 10 ppm. Peak area of carbamazepine was unaffected by the presence of other antiepileptic drugs (fig. 4).

**Stability**

Stability was determined at the first and third month of the study. The sample was stored at -20 °C for 90 d. Across all sample concentrations, % diff was above 80% except for 0.5 μg/ml at the first month showed 21% diff. Carbamazepine plasma concentrations of three drug-resistant epilepsy patients were also determined at the first and third month. All samples showed acceptable standard deviation and RSD (table 3).

**Table 3: Stability of carbamazepine plasma concentration of drug-resistant epilepsy patients**

<table>
<thead>
<tr>
<th>Sample</th>
<th>Concentration (μg/ml)</th>
<th>First month</th>
<th>Third month</th>
<th>% diff</th>
<th>% RSD(^a)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>9.24</td>
<td>10.88</td>
<td>98.8</td>
<td>11.53</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>6.73</td>
<td>6.64</td>
<td>99.04</td>
<td>0.95</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>15.50</td>
<td>12.94</td>
<td>98.2</td>
<td>12.73</td>
<td></td>
</tr>
</tbody>
</table>

\(^a\)RSD = relative standard deviation
Application in epilepsy patients

Ten patients undergoing monotherapy or polytherapy with two or three drugs were selected (table 4). Carbamazepine plasma concentrations were unaffected by the presence of other drugs (fig. 5) and showed consistent results in each dosage and measurement.

Table 4: Carbamazepine plasma concentration in epilepsy patients

<table>
<thead>
<tr>
<th>No.</th>
<th>Carbamazepine dosages</th>
<th>Other drugs</th>
<th>Carbamazepine plasma concentrations</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>400 mg</td>
<td>None</td>
<td>7.62 μg/ml</td>
</tr>
<tr>
<td>2.</td>
<td>400 mg</td>
<td>Folic acid</td>
<td>6.14 μg/ml</td>
</tr>
<tr>
<td>3.</td>
<td>400 mg</td>
<td>Levetiracetam</td>
<td>9.35 μg/ml</td>
</tr>
<tr>
<td>4.</td>
<td>400 mg</td>
<td>Clonazepam</td>
<td>9.5 μg/ml</td>
</tr>
<tr>
<td>5.</td>
<td>400 mg</td>
<td>Lamotrigine</td>
<td>13.19 μg/ml</td>
</tr>
<tr>
<td>6.</td>
<td>600 mg</td>
<td>Valproic acid</td>
<td>7.95 μg/ml</td>
</tr>
<tr>
<td>7.</td>
<td>600 mg</td>
<td>Phenytoin</td>
<td>8.62 μg/ml</td>
</tr>
<tr>
<td>8.</td>
<td>800 mg</td>
<td>Clobazam</td>
<td>10.08 μg/ml</td>
</tr>
<tr>
<td>9.</td>
<td>1100 mg</td>
<td>Valproic acid and phenobarbital</td>
<td>9.46 μg/ml</td>
</tr>
<tr>
<td>10.</td>
<td>1500 mg</td>
<td>Topiramat and clobazam</td>
<td>14.23 μg/ml</td>
</tr>
</tbody>
</table>

Fig. 5: Chromatogram of carbamazepine (CBZ) and propylparaben (PP) as internal standard, in epilepsy patients; (A): monotherapy; (B): polytherapy with clobazam; (C): Polytherapy with valproic acid and Phenobarbital

DISCUSSION

Therapeutic drug monitoring (TDM) is important in treating patients with chronic diseases. In epilepsy, patients are treated for at least 3 years after being seizure-free. In many cases, polytherapy is needed to control seizure. The safest way to determine whether a patient needs polytherapy is to first evaluate the plasma levels of the initial medication. If the plasma concentration of the initially administered medication is in the therapeutic range but seizures are not yet controlled, polytherapy may be needed. In polytherapy patients, the plasma concentration of the drug of interest should be unaffected by the presence of other drugs. Only a few publications on various methods of plasma concentration measurements described the implementation of measurements in patients, especially in polytherapy. In pharmaceutical laboratory settings, recent studies that quantify one drug reported a simple, rapid, precise and economical spectrophotometric method using HPLC with UV-Visible [10-11]. Another study quantifies more than one drug, used PDA as drug detector [12]. Meanwhile, a study using patient’s plasma reported a selective, accurate and precise HPLC method with PDA [13]. In this study, we used PDA detector to prove the selectivity of the method.

The HPLC technique was validated using the Guidelines on Bioanalytical Method Validation by the EMEA, for the purpose of measuring carbamazepine plasma concentrations in epilepsy patients. Carbamazepine is a hydrophobic and nonpolar compound, hence acetonitrile was chosen as a solvent for the mobile phase, in which the ratio of water: acetonitrile was equal. C18 column was chosen to be the stationary phase with small porous particles. On the surface of the column’s porous particles, there is a chemically-bonded phase that separates the sample’s compounds. Separation of carbamazepine went well and was detected by the PDA detector, after only 3.5 min following the injection. Other study using solid-phase extraction method followed by reverse phase HPLC report the same mobile phase in different ratios. Using the same column and PDA detector, carbamazepine was detected 1 min longer than this study [13]. Another study used different mobile phase for carbamazepine detection in morphine addicted patients; reported the appearance of interfering components [14]. In this study, PDA detector that able to identify multiple components by a comparison of the spectrum proved the method was highly selective in quantify carbamazepine of epilepsy patients’ plasma. In monotherapy and polytherapy patients, there was no interference on peak area of carbamazepine.

All parameters of validation were tested and showed good results and were in compliance with the international criteria (EMEA). The linearity of the range chosen had a good correlation coefficient of concentration and peak area ratio. Range linearity in this study could measure the concentration of carbamazepine in most of the collected samples. Precision and accuracy of the test is important, including the LOQ and LOD, in order to have a reliable measurement to base clinical judgments on. Selectivity of this method was satisfactory. Compared to another study; anti epileptic drugs used as polytherapy in this study varied in dosage and types [13].

Moreover, carbamazepine in the patients’ plasma was adequately stable. In summary, this method is simple, accurate, and satisfactorily reliable for application in epilepsy patients, especially those undergoing polytherapy.

CONCLUSION

The usage of HPLC for measuring carbamazepine plasma concentrations in epilepsy patients treated with monotherapy or polytherapy was valid. This method meets the EMEA guideline standard in terms of linearity, precision, and accuracy, also selectivity in epilepsy patients treated with polytherapy.

AUTHOR CONTRIBUTION

Astri Budikayanti as the first author had done almost all of the steps in this study. Chiswyta Chaliana had a contribution in preparation and laboratory procedures. Melva Louisa was the laboratory supervisor, and Rianto Setiabudy was the mentor of the study.
CONFLICTS OF INTERESTS

All authors have none to declare.

REFERENCES


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