

Original Article

IN VITRO ANTIOXIDANT CAPACITIES AND ANTIDIABETIC PROPERTIES OF PLUCHEA LEAVES AND GREEN TEA MIXTURES AT VARIOUS PROPORTIONS

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ABSTRACT

Objective: This study was done to determine the effect of pluchea leaves and green tea mixtures at various proportions to antioxidant and antidiabetic properties.

Methods: The research used a single factor randomized block design. Research factors were pluchea and green tea mixture proportions, including 100:0; 75:25; 50:50; 25:75; and 0:100% (w/w), respectively. Each of factors was repeated five times. The parameters observed in this study were phytochemical compounds, total phenols, total flavonoids, free radical DPPH scavenging activity, iron ion reducing power, alpha amylase and alpha glycosidase inhibition activities. The data were statistically $\alpha = 5\%$, if the data analysis showed a significant analyzed using analysis of variance (ANOVA) at effect then it was continued with duncan's multiple range test (DMRT).

Results: The phytochemical compounds containing in pluchea leaves and green tea mixtures were alkaloids, flavonoids, phenolics, saponins, tannins, triterpenoids, sterols, and cardiac glycosides. The qualitative assay of phytochemical compounds from aqueous extract had a different pattern with total phenolic content (TPC) and total flavonoid content (TFC), especially at 75:25; 50:50 and 25:75 % (w/w) from pluchea leaves and green tea mixture proportions. The 50:50% (w/w) proportions of samples had the highest TPC and TFC, it was proposed that there was an interaction between bioactive compounds of pluchea leaves and green tea. TPC and TFC were correlated with free radical DPPH scavenging activity, iron reducing power and alpha glycosidase inhibition activity, but the alpha amylase inhibition activity had the different graph pattern with the TPC and TFC.

Conclusion: The antioxidant capacity of pluchea leaves and green tea mixtures at various proportions had the graph pattern similar to the alpha glycosidase inhibitory activity and different with the alpha amylase inhibitory activity.

Keywords: Pluchea leaves, Green tea, Antioxidant, Anti diabetic

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INTRODUCTION

Pluchea leaves (*Pluchea indica* Less) and green tea (*Camellia sinensis* (L) O. Kuntze) are two commodities that are potentially as antioxidant and anti-diabetic sources. These are related to phytochemical contents of these plants.

Pluchea indica Less are a herb plant from *Asteraceae* family that are potentially as antioxidant, especially DPPH free radical scavenging activity, iron reducing power, reactive oxygen species scavenging activity, and β -carotene linoleic acid system inhibition activity [1-3], anti-warmed over flavor [4], anti-inflammatory [5], and anti-diabetic [6]. Pluchea leaves contain the phytochemical compounds, such as flavonoids, sterols, saponins, tannins, alkaloids, and cardiac glycosides [1, 2, 6, 7].

Green tea contains a major active group of tea leaves' components (up to 30% of the dry weight of the water-extractable material) [8-10]. They are comprised on epigallocatechin gallate (23.4–112.4 lg/ml) as the major component, followed by epigallocatechin (18.4–78.9 lg/ml), epicatechin gallate (5.6–29.6 lg/ml), epicatechin (4.6–14.5 lg/ml) and catechin (3.2–8.2 lg/ml) [11]. The catechins effectively scavenge oxy-radicals involved in the pathogenesis of many chronic diseases, including cardiovascular diseases, cancer, neurodegenerative diseases, and ageing [12], antioxidant and prebiotic activities [13], anticancer [8], antidiabetic [14], anti-inflammatory and analgesic effect [15].

Pluchea leaves and green tea usually are used as a health beverage. Many people generally utilize this commodity as herbal tea that is packed in tea bag packaging. The potency of each commodity as antioxidant and anti-diabetic has been determined [4,5,13,14]. Therefore it is necessary to know the potency of pluchea leaves and

green tea mixtures as antioxidant and antidiabetic activities. The pluchea leaves and green tea mixtures can allow the interaction among bioactive compounds that can give synergism or antagonism effects of antioxidant and antidiabetic activities. Until now this interaction is not fully understood. Therefore it is necessary to investigate the effect of a mixture of them at various proportions to antioxidant and antidiabetic activities. This study was done to determine the effect of pluchea leaves and green tea mixtures at various proportions to antioxidant and antidiabetic properties.

MATERIALS AND METHODS

Chemicals and reagents

Pluchea leaves were harvested and collected from a pluchea garden in Mangrove areas, Wonorejo, Rungkut, Surabaya, East Java, a city in the middle of Indonesia in Java Island. The plant was authenticated in the Herbarium of Biology and Food Industry Microbiology Laboratory at the department of Food Technology, Agricultural Technology Faculty, the Widya Mandala Catholic University of Surabaya with voucher specimen no FTP-UKWMS-0001 for future reference.

Dried green tea was purchased by PT. Rolas Nusantara Mandiri Surabaya, East Java, Indonesia. Methanol, ferric (II) chloride, potassium ferricyanide, sodium carbonate, ethanol, petroleum ether, copper sulphate, Folin ciocalteus's phenol, aluminium chloride, sodium nitrite, sodium hydroxide, amyl alcohol, chloroform, ammonium hydroxide, sulphuric acid, mercury chloride, potassium iodide, iodine, magnesium, acetic acid, chloride acid, ether, chloroacetic acid, sodium dihydrogen phosphate, disodium hydrogen phosphate, sodium acetate, and starch were purchased from Merck Company (Darmstadt, Germany). Potassium sodium

tartrate tetrahydrate, 1,1-diphenyl-2-picrylhydrazyl free radical (DPPH), alpha amylase enzyme and yeast alpha glycosidase enzyme, gallic acid, (+)-catechin and p-nitrophenil- α -D-glucopyranoside were purchased from Sigma-Aldrich Company Ltd. (Gillingham, UK). Commercial mineral water with pH 7.65 was purchased from PT Erindo Mandiri (Surabaya, East Java, Indonesia) and aqua dest was bought from PT. Megah Sejahtera Scientific (Surabaya, East Java, Indonesia).

Sampling

Pluchea indica less leaves were harvested from 1-6 age level [16]. The leaves were dried at room temperature and ground to a fine powder (45 mesh). The moisture content was around 15.06-16.72 % db. And then dried leaves powder was mixed before extraction.

Extraction

Pluchea leaves powder was mixed with green tea powder at 100:0; 75:25; 50:50; 25:75; and 0:100 % (w/w) proportions, respectively. And then samples were extracted by Deetae *et al.* [16] method with slight modification. *Pluchea* leaves were sorted from 1-6 segment level, and then they were washed and dried at ambient temperature for 7 d. After dried *pluchea* leaves were done crushing to get 45 mesh particle size. *Pluchea* leaves powder was homogenised before they were mixed by the similar particle size of green tea powder. The mixed samples (2g) in tea bag were macerated with 100 ml commercial mineral water at 95 °C temperature for 5 minutes with constant stirring. The residue of samples in tea bag was separated from the filtrate. And then the filtrate was analyzed further. Each analysis of water extract was always fresh condition.

Phytochemical composition

Phytochemical compounds content in samples were detected by Harborne method [17]. The compounds identified included alkaloids, flavonoids, phenolics, triterpenoids, sterols, saponins, tannins, and cardiac glycosides (fehling test).

Total phenol content analysis

Total phenol content (TPC) in samples was analyzed with folin ciocalteu's phenol reagent. The redox reaction between antioxidant compounds having an aromatic ring and phosphomolybdate compound in folin ciocalteu's phenol reagent is the principle of this analysis. The formation of the blue colour solution was determined by UV-Vis spectrophotometer (Shimadzu spectrophotometer UV-Vis 1700) at λ 760 nm, the result was stated as mg gallic acid equivalent/l samples (mg GAE/l samples) [19].

Total flavonoid content analysis

Total flavonoid content (TFC) was analyzed based on Al-Temimi and Choudary [19] method. This principle analysis is based on a stable acid complex compound formation of reaction between $AlCl_3$ and Oxo group at C_4 ring and a hydroxyl group at C_3 or C_5 ring of flavones and flavonol. This reaction produced solution coloured yellow and NaOH solution addition resulted in quinoid compound colored red that measured by UV-Vis spectrophotometer (Shimadzu spectrophotometer UV-Vis 1700) at λ 510 nm, the result was stated by mg catechin equivalent/l samples (mg CE/l samples).

DPPH free radical scavenging activity analysis

DPPH free radical scavenging activity analysis was measured by Brand William *et al.* [20]. The principal analysis is electron or hydrogen donating of antioxidant compounds to DPPH free radical colored purple to form DPPH-H non-radical colored yellow that was measured by UV-Vis spectrophotometer (Shimadzu spectrophotometer UV-Vis 1700) at 517 nm with a gallic acid compound as standard.

Iron reducing power analysis

Iron reducing power analysis was determined by Chanda and Dave [21] method that was used to measure the antioxidant capacity to reduce Fe^{3+} to Fe^{2+} . The complex compound from ferri ferrocyanide coloured Berlin blue was determined by UV-Vis spectrophotometer (Shimadzu spectrophotometer UV-Vis 1700) at λ 700 nm with a gallic acid compound as standard [20].

α -Amylase inhibition assay

The α -amylase inhibition was determined by an assay described from Bernfeld [22] with slight modification. The 500 μ L of water extract was incubated with 500 μ L of a α -amylase solution (enzyme solution (12.5 units/ml) was obtained by dissolving 0.1 g of α -amylase in 100 ml of 0.2 M sodium acetate buffer pH 5 at ambient temperature (32 °C) for about 10 min. After incubation, 500 μ L of 1 % starch solution (dissolving 1 g of amyllum starch in 100 ml of distilled water with boiling and stirring for 15 min) was added and incubated at ambient temperature (32 °C) for about 10 min. To the above, 2 ml of NaOH 1 M was added to stop the reaction and was incubated in a water bath (37 °C) for 10 min. After 10 min, reaction mixture color changed to orange-red and was removed from the water bath and cooled to ambient temperature. Absorbance was measured at 540 nm in Shimadzu spectrophotometer UV-Vis 1700. Extracts of *pluchea* leaves and green tea mixtures at different proportions (100:0; 75:25; 50:50; 25:75; and 0:100 mg/ml, respectively) were performed in five repetitions. Individual blank was performed by replacing enzyme with buffer. Control was performed by replacing extract with solvent. The inhibition percentage of α -amylase was assessed by the following formula:

$$\text{Inhibition (\%)} = \frac{(Ac(0) - Ac(t)) - (As - Ab)}{Ac(0) - Ac(t)} \times 100\% \dots \dots \dots (\text{Equation 1})$$

Where, Ac (0) that absorbance of 100% enzyme activity (only solvent with enzyme), Ac (t) that absorbance of 0% enzyme activity (only solvent without enzyme), As that absorbance of test sample with an enzyme, Ab that absorbance of test sample without enzyme.

α -Glucosidase inhibition assay

The inhibition of yeast α -glucosidase was determined by using the method described by Da Silva Pinto *et al.* [23] with slight modification. Briefly, 0.0019 mg yeast α -glucosidase (0.0833 unit/ml) was dissolved in 100 ml of 0.2 M phosphate buffer (pH 7). The substrate solution, 2 mmol p-nitrophenyl- α -D-glucopyranoside, was prepared in the same buffer. The water extract of *pluchea* leaves and green tea at various proportions (50 μ L), 50 μ L of enzyme solution, 50 μ L of substrate solution, and 100 μ L buffer phosphate solution (pH 7) were mixed well and incubated at 37 °C for 15 min. After incubation, the solution was added by 1000 μ L of 0.2 M Na_2CO_3 solution to stop the reaction. Absorbance was measured at 410 nm in Shimadzu spectrophotometer UV-Vis 1700. Extracts of *pluchea* leaves and green tea mixtures at different proportions (100:0; 75:25; 50:50; 25:75; 0:100 mg/ml) were performed in five repetitions. Individual blank was performed by replacing enzyme with buffer. Control was performed by replacing extract with solvent. The inhibition percentage of α -amylase was assessed by the following formula:

$$\text{Inhibition (\%)} = \frac{(Ac(0) - Ac(t)) - (As - Ab)}{Ac(0) - Ac(t)} \times 100\% \dots \dots \dots (\text{Equation 2})$$

Where, Ac (0) that absorbance of 100% enzyme activity (only solvent with enzyme), Ac (t) that absorbance of 0% enzyme activity (only solvent without enzyme), As that absorbance of the test sample with an enzyme, Ab that absorbance of test sample without enzyme.

RESULTS AND DISCUSSION

Phytochemical analysis

Phytochemicals are primary and secondary natural constituents occurring in the medical plants, leaves, vegetables and roots that have defence mechanism and protect from various diseases [25]. The qualitative analysis of the water extract from *pluchea* leaves and green tea mixtures at various concentrations confirmed the presence of alkaloids, flavonoids, phenols, saponins, tannins, triterpenoids, and cardiac glycosides (table 1). Data showed that *pluchea* leaves were very rich in alkaloids, flavonoids, phenolics, saponins, tannins, triterpenoids, and cardiac glycosides compared with green tea. The increasing of green tea proportion in samples decreased the phytochemical quality. Keerthana *et al.* [25] said that the phytochemical constituents can be used to treat diabetes if they have good antioxidant and antidiabetic properties. Dastjerdi *et al.*

[26] and Nanumala *et al.* [27] said that the capacity to inhibit alpha amylase and alpha glycosidase enzymes are a result of the existence of various phytochemicals like flavonoids, tannins, saponins, anthraquinones, sterols, phlobatannins, terpenoids and polyphenols.

Total phenol content

Total phenol content (TPC) in the water extract from pluchea leaves and green tea mixtures at various proportions was showed in fig. 1. Generally, the TPC of pluchea leaves water extract was significantly ($p < 0.05$) higher compared to the TPC of green tea water extract. This condition was similar to observation of the phytochemical assay, however, there were different phenomena that were observed of samples at 75:25, 50:50, and 25:75 proportions, respectively. These differences may be indicated by the differences in solved phenolic compounds and interaction between phenolic compounds from pluchea leaves and green tea mixtures. These phenomena determined structure of phenolic compounds, such as free or bound phenolics. Many studies have informed that phenolic compounds of fruits and vegetables are

usually in free structure higher than in bound structure [29,30]. There was a free form of phenolics of pluchea leaves and green tea in aqueous media showed that there was hydroxyl group free. The functional group can interact to form bound phenolics or hydrogen bound so that interferes with the reaction among hydroxyl group of phenolic compounds, tungsten and molybdenum ions of Folin Ciocalteu's reagent.

The effectiveness of TPC assay is depended on the number and position of hydrogen-donating hydroxyl groups on the aromatic ring of the phenolic compounds [31].

Green tea comprises phenolic compounds, especially catechin [8-11], however, pluchea leaves contain phenolic acids i.e. chlorogenic acid and caffeic acid [4] and flavonoid compounds i.e. kaempferol, myricetin, and quercetin [32]. Natural properties of phenolic compounds have biological and pharmacological properties, such as antioxidant, anticancer, anti-aging, and anti-inflammatory activities [33,34]. Phenolic compounds have been used as antioxidants and mainly inhibit free radicals and block their chain reactions [32-34].

Table 1: The phytochemical content of pluchea leaves and green tea mixture at various proportions

Phytochemical constituents	Pluchea leaves and green tea mixtures (% w/w)				
	100:0	75:25	50:50	25:75	0:100
Alkaloids	++++	+++	+++	++	+
Flavonoids	++++	+++	++	+	+
Phenolics	++++	+++	+++	++	+
Saponins	++++	+++	+++	++	+
Tannins	++++	+++	+++	++	+
Triterpenoids	++++	+++	+++	++	+
Sterols	-	-	-	-	-
Cardiac Glycosides	++++	+++	++	+	+

Note: += indicates presense of phytochemicals and -= indicates absence of phytochemicals, +++++ shows high concentration, += shows low concentration

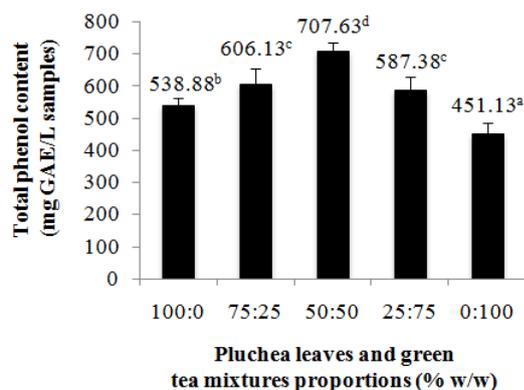


Fig. 1: Total phenolic content of aqueous extract of pluchea leaves and green tea mixtures at various proportions (n=5, Values expressed as mean±SD. Data were analyzed by one way analysis of variance (ANOVA) followed by duncan's multiple range test (DMRT))

Total flavonoid content

Total flavonoid content (TFC) was determined by aluminum chloride method that was based on the formation of a complex between the aluminum ion, Al (III), and the carbonyl and hydroxyl groups of flavones and flavonols that produce a yellow color [38]. In this study, the flavonoid compounds measured are flavones and flavonols groups, the TFC was ranged from 95.73 to 413.73 mg of quercetin equivalent L-1 of samples (fig. 2). The results showed that pluchea leaves contained higher TFC than green tea leaves. The pluchea leaves and green tea mixtures at 50:50 % (w/w) proportions had the highest TFC compared with the other proportions. The TFC of samples had a similar pattern to the TPC, because the flavonoid compounds are the major component of phenolic compounds [33]. The existence of a functional group from the carbonyl and hydroxyl groups of flavones and flavonols in pluchea leaves and green tea in samples can cause synergism effect so that the formation of

a complex with aluminium ion increased in samples at 75:25; 50:50; 25:75 proportions, respectively. The flavonoid compounds can act as free radical scavenging, inhibition of hydrolytic and oxidative enzymes and anti-inflammatory action [39, 40].

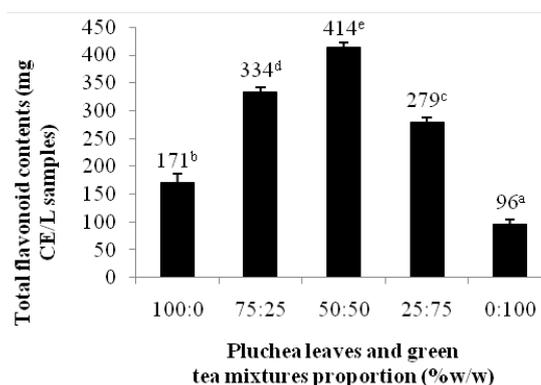


Fig. 2: Total flavonoid content of aqueous extract of pluchea leaves and green tea mixtures at various proportions (n=5, Values expressed as mean±SD. Data were analyzed by one way analysis of variance (ANOVA) followed by duncan's multiple range test (DMRT))

Andarwulan *et al.* [31] informed that the flavonols contained in pluchea leaves are kaempferol, myricetin, and quercetin. Peres *et al.* [11] said that flavanols are the major flavonoid compounds in green tea so that they aren't detected by aluminium chloride method.

DPPH free radical scavenging activity

The method is based on the reduction of the methanolic-DPPH solution because of the presence of antioxidant substances having hydrogen donating groups (RH) such as phenolics and flavonoids

compounds due to the formation of non radical DPPH-H form [12, 41]. The total antioxidant activities (TAA) determined by DPPH of water extract of pluchea leaves and green tea mixtures at various proportions were expressed as mg of quercetin equivalent L of samples (fig. 3). Total antioxidant activity (DPPH) was highest in 50:50 proportions, followed by 75:25 proportions, followed by 25:75 proportions, followed by 100:0 proportions, and then 0:100 proportions of pluchea leaves and green tea mixtures. There was trended that hydroxyl groups among phenolic compounds in pluchea leaves and green tea were antagonism effect so that the TAA of them was lower than the TAA sum of pluchea leaves and green tea. The graph pattern of this TAA was similar to the graph pattern of the TPC. The TAA of pluchea leaves was higher than it of green tea. The differences of TAA among all proportions of pluchea leaves and green tea mixtures were statistically significant. There were significant differences between total antioxidant activities of all proportions of pluchea leaves and green tea mixtures depending on TPC of an extract of samples. Widyawati *et al.* [2] said that the different antiradical activity of the extract was depended on the polarity of solvent and solute. These determined the intrinsic compounds to be extracted. The different structure of phytochemical constituents determines scavenging activity. Rohman *et al.* [40] informed that the phytochemicals responsible for the scavenging activity in this species are phenolic and flavonoid constituents. Har and Ismail [41] also explained that phenolic acid and flavonoids are known to be contributing to the antioxidant activity. Chlopicka *et al.* [42] said that among polyphenols the greatest antioxidant efficacies in pseudocereals breads are showed for quercetin, tannic acid, caffeic acid and gallic acid, while catechin and resveratrol have the lowest ones.

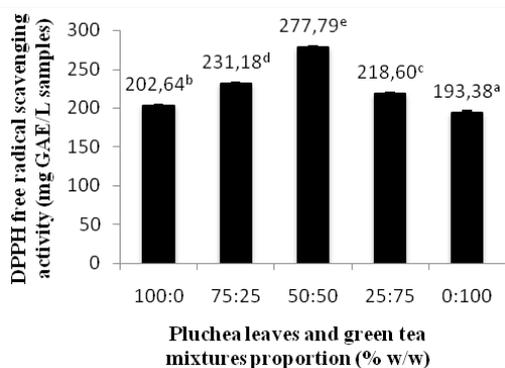


Fig. 3: DPPH free radical scavenging activity of aqueous extract of pluchea leaves and green tea mixtures at various proportions (n=5, Values expressed as mean±SD. Data were analyzed by one way analysis of variance (ANOVA) followed by duncan's multiple range test (DMRT))

Widyawati *et al.* [2] showed that aqueous extract of pluchea leaves is potential to scavenge free radical. Ruskin *et al.* [43] added that there is a positive correlation between phenolic content and free radical scavenging activity. The high phenolic content of *Chukrasia tabularis* (17.2 mg GAE/g) shows the linear correlation between phenolic content and antioxidant activity. The general assessment of the analytical results for the plant extracts definitely shows the individual specificity of each sample and a rich diverse spectrum of phenolic compounds differing from flavonoid group.

Iron ion reducing power

The total antioxidant activities (TAA) determined by reducing power (RP) of aqueous extract of pluchea leaves and green tea mixtures at various proportions were expressed as mg of gallic acid equivalent per L of samples (fig. 4). Among samples evaluated, the 50:50 proportions of pluchea leaves and green tea mixtures had the highest reducing power, this graph pattern of the RP was similar to the graph pattern of DPPH free radical scavenging activity. There was trended that the RP had the same graph pattern as it of TFC.

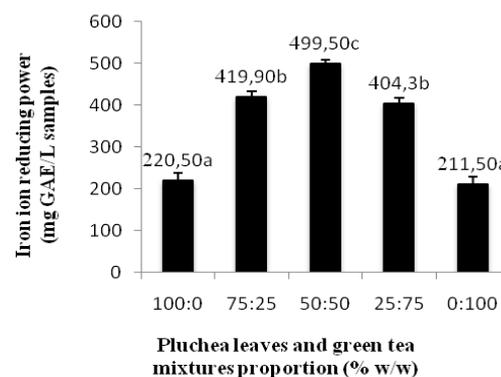


Fig. 4: Iron ion reducing power of aqueous extract of pluchea leaves and green tea mixtures at various proportions (n=5, Values expressed as mean±SD. Data were analyzed by one-way analysis of variance (ANOVA) followed by duncan's multiple range test (DMRT))

This estimated that the hydroxyl group of flavonoid compounds between pluchea leaves and green tea mixtures was interacted to give the synergism effect of RP. The differences of TAA among all proportions of pluchea leaves and green tea mixtures were statistically significant. There were significant differences between TAA of all proportions of pluchea leaves and green tea mixtures depending on TFC of an extract of samples. Rohman *et al.* [40] said that TAA of extracts and fractions of red fruit (*Pandanus conoideus* Lam) determines the total phenolics content and total flavonoid content. He also suggested that phenolics compounds more likely contribute to its reducing activity (RP) than flavonoid compounds. On the contrary, the opposite results were found in this research that the RP was more likely contributed by flavonoid compounds than it of phenolic compounds. Gholivand and Piryaei [44] studies that the electron donation capacity reflects the reducing power of bioactive compounds which is related to antioxidant activity.

Antioxidants can be explained as reducers, and inactivation of oxidants by reducers can be described as redox reaction in which one reaction species is reduced at the expense of the oxidation of the other. Fe³⁺-reduction is often used as an indicator of electron donating activity, which is an important mechanism of the phenolic antioxidant action.

α-Amylase inhibitory activity

The ability of aqueous extract from pluchea leaves and green tea mixtures at various proportions in inhibiting the α-amylase is presented in fig. 5.

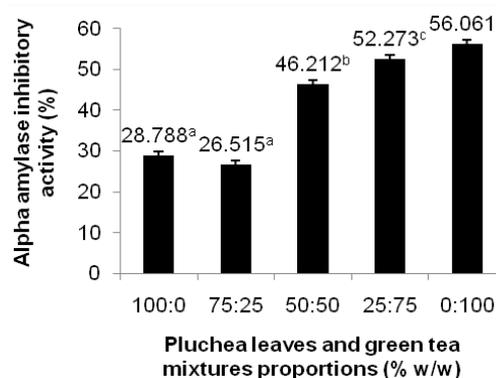


Fig. 5: α-Amylase inhibitory activity of aqueous extract of pluchea leaves and green tea mixtures at various proportions (n=5, Values expressed as mean±SD. Data were analyzed by one way analysis of variance (ANOVA) followed by duncan's multiple range test (DMRT))

The α -amylase inhibition activity in this study was ranged from 26.15 \pm 1.52 to 56.06 \pm 3.91%. The 0:100 proportions had the highest inhibitory α -amylase activity compared to the other proportions. The inhibition activity of the green tea extract showed the significantly higher ($p < 0.05$) than it of pluchea leaves extract. In this study, there was no significant correlation found between the TAA (DPPH radical and RP assay) with the α -amylase inhibition activity of pluchea leaves and green tea mixtures at various proportions. This showed that the graph pattern of the TAA and the α -amylase inhibition activity wasn't similar. The same study also is done by Lim and Loh [45]. However, McCue et al. [46] suggested that the antioxidant activity of phenolic from clonal oregano extracts is affected the five sets of disulphide bridges located on the outer surface of α -amylase. The reduction of these cysteine residues causes inhibition by modifying in the structure of the enzyme. Thus, the α -amylase inhibition activity of the pluchea leaves and green tea mixtures at various proportions may perhaps not due to the TAA but it also involves other mechanisms that worth to be disclosed. This argument was supported by Lim and Loh [45]. Akah et al. [47] informed that the phytochemical compounds, such as terpenoids, saponins, flavonoids, glycosides and carbohydrate have similarly been implicated in the anti-diabetic activities of the plant. Therefore the alpha amylase inhibition activity of aqueous extract from pluchea leaves and green tea mixtures at various proportions may be contributed by phytochemical compounds (table 1).

α -Glucosidase inhibitory activity

The α -glucosidase inhibitory activity of the extracts is summarised in fig. 6. Data showed that the 50:50 proportions of pluchea leaves and green tea mixtures had the highest inhibitory α -amylase activity compared to the other proportions. The green tea extract had the higher alpha glucosidase inhibitory activity than it of pluchea leaves extract. However, the TPC and TFC of green tea extract were lower than them of pluchea leaves extract. This phenomenon indicated that the alpha glucosidase inhibitory activity was no be caused by phenolic compounds activity but it was contributed the other phytochemicals. Lim and Loh [45] reported that the α -glucosidase inhibition activity of bound phenolic extract has higher than free soluble phenolic extract because the bound phenolic mostly exists in a β -glycosides form that prefers aqueous phase than the free phenolic which is in the form of aglycones, a non-sugar group. Thus, direct enzyme-inhibitor interaction is predicted to be higher in bound phenolic extracts in the α -glucosidase assay. McCue et al. [48] reported that the mechanisms of inhibition for α -glucosidase may different from α -amylase. There is none disulphide bridges especially not on the surface of the molecule (possible site for interaction with antioxidants) on the structure of Baker's yeast α -glucosidase. This indicated that the α -glucosidase inhibition activity of phenolic extracts doesn't involve the interaction of the antioxidant capacity with the disulphide bridges. Therefore, the inhibition is caused by the phenolic extracts through another mechanism. There was a correlation among the TPC, TFC and alpha glucosidase inhibitory activity. Oboh et al. [49] said that the higher inhibitory activities also are contributed by the presence of some non-phenolic phytochemicals that probably act as enzyme inhibitors, exhibiting an additive or synergism effect with the phenolics present in the sample.

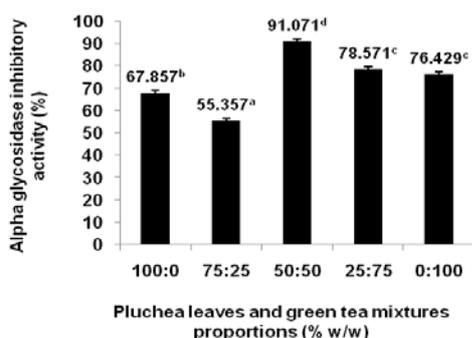


Fig. 6: α -Glucosidase inhibitory activity of aqueous extract of pluchea leaves and green tea mixtures at various proportions (n=5, Values expressed as mean \pm SD. Data were analyzed by one-way analysis of variance (ANOVA) followed by duncan's multiple range test (DMRT))

CONCLUSION

The antioxidant activity and alpha glycosidase inhibitory activity of aqueous extract in pluchea leaves and green tea mixtures at various proportions were correlated with TPC and TFC, but the alpha amylase inhibitory activity wasn't. There was contributed by the other phytochemicals to the alpha amylase inhibitory activity. The inhibition mechanism of alpha amylase and alpha glycosidase was different.

AUTHORS CONTRIBUTION

The first and second authors have participated in the work including participation in the concept, design, analysis, writing, and revision of the manuscript.

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CONFLICTS OF INTERESTS

Authors declare no conflicts of interest

REFERENCES

- Widyawati PS, Wijaya CH, Hardjosworo PS, Sajuthi D. Antioxidant activity of pluchea (*Pluchea indica* Less) leaves methanolic extract and its fractions. *Agritech* 2012;32:249-57.
- Widyawati PS, Budianta TDW, Kusuma FA, Wijaya EL. Difference of solvent polarity to phytochemical content and antioxidant activity of *Pluchea Indica* less leaves extracts. *Int J Pharm Pharm Res* 2014a;6:850-5.
- Widyawati PS, Budianta TDW, Kusuma FA, Wijaya EL, Yaunatan DI, Wongso RS. Potency of beluntas (*Pluchea indica* Less) leaves to extract as antioxidant and anti warmed over flavor (WOF) of duck meat. *International Congress Challenges of Biotechnological Research in Food and Health*. Surakarta; 2014b.
- Widyawati PS, Budianta TDW, Utomo AR, Harianto I. The physicochemical and antioxidant properties of *Pluchea Indica* Less drink in tea *bag packaging*. *Int J Food Nut Sci* 2016;5:113-20.
- Srisook K, Buapool D, Boonbai R, Simmasat P, Charoensuk Y, Srisook E. Antioxidant and anti-inflammatory activities of hot water extract from *Pluchea indica* Less herbal tea. *J Med Plants Res* 2012;6:4077-81.
- Widyawati PS, Budianta TDW, Gunawan DI, Wongso RS. Evaluation antidiabetic activity of various leaf extracts of *Pluchea indica* Less. *Int J Pharm Pharm Res* 2015;7:597-603.
- Widyawati PS, Wijaya CH, Hardjosworo PS, Sajuthi D. Evaluation of antioxidant activity from pluchea leaves extract (*Pluchea indica* Less) based on leaf segment level. *Rekapangan J* 2011;5:1-14.
- Carvalho M, Jerónimo C, Valentão P, Andrade PB, Silva BM. Green tea: a promising anticancer agent for renal cell carcinoma. *Food Chem* 2010;122:49-54.
- Moderno PM, Carvalho M, Silva BM. Recent patents on *Camellia sinensis*: Source of health promoting compounds. *Recent Patents Food Nut Agric* 2009;1:182-92.
- Yang CS, Lambert JD, Hou Z, Ju J, Lu G, Hao X. Molecular targets for the cancer preventive activity of tea polyphenols. *Mol Carcinog* 2006;45:431-5.
- Peres RG, Tonin FG, Tavares MFM, Rodriguez-Amaya DB. Determination of catechins in green tea infusions by reduced flow micellar electrokinetic chromatography. *Food Chem* 2011;127:651-5.
- Kang KW, Oh SJ, Ryu SY, Song GY, Kim BH, Kang JS, et al. Evaluation of the total oxy-radical scavenging capacity of catechins isolated from green tea. *Food Chem* 2010;121:1089-94.
- Molan AL, Flanagan J, Wei W, Moughan PJ. Selenium-containing green tea has higher antioxidant and prebiotic activities than regular green tea. *Food Chem* 2009;114:829-35.
- Miura T, Koike T, Ishida T. Antidiabetic activity of green tea (*Thea Sinensis L.*) in genetically type 2 diabetic mice. *J Health Sci* 2005;51:708-10.
- Mota MADL, Landim JSP, Targino TSS, Da Silva S FR, Da Silva SL, Pereira MRP. Evaluation of the anti-inflammatory and analgesic effect of green tea (*Camellia sinensis*) in mice. *Acta Cir Bras* 2015;30:242-6.

16. Deetae P, Parichanon. P, Trakunleewatthana P, Chanseetis C, Lertsiri S. Antioxidant and anti-glycation properties of thai herbal teas in comparison with conventional teas. *Food Chem* 2012;133:953-9.
17. Harborne JB. *Phytochemical Method*. Padmawinata K, Soediro I. Eds. Bandung: Institut Teknologi Bandung; 1996.
18. Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods in Enzymol* 1999;99:152-78.
19. Al-Temimi A, Choudhary R. Determination of antioxidant activity in different kinds of plants *in vivo* and *in vitro* by using diverse technical methods. *J Nut Food Sci* 2013;3:1-9.
20. Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. *Lebensm Wiss Technol* 1995;28:25-30.
21. Chanda S, Dave R. *In vivo* models for antioxidant activity evaluation and some medicinal plants possessing antioxidant properties: an overview. *Afr J Microbiol Res* 2009;3:981-96.
22. Bernfeld P. Amylases, alpha and beta. In: Colowick SP, Kaplan NO. Eds. *Methods in Enzymology*. Vol. 1. New York: Academic Press; 1955. p. 149-58.
23. Da Silva Pinto M, Kwon YI, Apostolidis E, Lajolo FM, Genovese MI, Shetty K. Functionality of bioactive compounds in Brazilian strawberry (*Fragaria x ananassa* Duch.) cultivars: evaluation of hyperglycemia and hypertension potential using *in vitro* models. *J Agric Food Chem* 2008;56:4386-92.
24. Wadood A, Ghufuran M, Jamal SB, Naeem M, Khan A, Ghaffar R, et al. Phytochemical analysis of medicinal plants occurring in the local area of mardan. *Biochem Anal Biochem* 2013;2:2-4.
25. Keerthana G, Kalaivani MK, Sumathy A. In vitro alpha amylase inhibitory and antioxidant activities of ethanolic leaf extract of *Croton Bonplandianum*. *Asian J Pharm Clin Res* 2013;6 (Suppl 4): 32-6.
26. Dastjerdi ZM, Namjooyan F, Azemi ME. Alpha amylase inhibition activity of some plants extract of teucrium species. *Eur J Biol Sci* 2015;7:26-31.
27. Nanumala SK, Tulasi P, Sujitha E. *In vitro* antidiabetic activity of seed extracts of *Cassia auriculata* and *Cassia angustifolia*. *Eur J Exp Biol* 2015;5:12-7.
28. Hung PV, Duy TL. Effects of drying methods on bioactive compounds of vegetables and correlation between bioactive compounds and their antioxidants. *Int Food Res J* 2012;19:327-32.
29. Adefegha SA, Oboh G. *In vitro* inhibition activity of polyphenol-rich extracts from *Syzygium aromaticum* (L.) Merr. and perry (Clove) buds against carbohydrate hydrolyzing enzymes linked to type 2 diabetes and Fe²⁺-induced lipid peroxidation in rat pancreas. *Asian Pac J Trop Biomed* 2012;2:774-81.
30. Priečina L, Karlina D. Total polyphenol, flavonoid content and antiradical activity of celery, dill, parsley, onion and garlic dried in convention and microwave-vacuum dryers. 2nd International Conference on Nutrition and Food Sciences IPCBEE. Vol. 53. Singapore: IACSIT Press; 2013.
31. Andarwulan N, Batari R, Sandrasari DA, Bolling B, Wijaya H. Flavonoid content and antioxidant activity of vegetables from Indonesia. *Food Chem* 2010;121:1231-5.
32. Calado JCP, Albertão PA, Aparecida de Oliveira E, Letra MHS, Sawaya ACHF, Marcucci MC. Flavonoid contents and antioxidant activity in fruit, vegetables and other types of food. *Agric Sci* 2015;6:426-35.
33. Stanković MS. Total phenolic content, flavonoid concentration and antioxidant activity of *Marrubium Peregrinum* L. extracts. *Kragujev J Sci* 2011;33:63-72.
34. Stanojević L, Stanković M, Nikolić V, Nikolić L, Ristić D, Čanadanovic-Brunet J, et al. Antioxidant activity and total phenolic and flavonoid contents of *Hieracium pilosella* L. extracts. *Sensors* 2009;9:5702-14.
35. Da Silva MCA, Paiva SR. Antioxidant activity and flavonoid content of *Clusia fluminensis* Planch. and Triana. *An Acad Bras Cienc* 2012;84:609-16.
36. Bag GC, Devi PG, Bhaigiyabati TH. Assessment of total flavonoid content and antioxidant activity of methanolic rhizome extract of three *Hedychium* species of manipur valley. *Int J Pharm Sci Rev Res* 2015;30:154-9.
37. Pontis JA, Da Costa LAMA, Da Silva SJR, Flach A. Color, phenolic and flavonoid content, and antioxidant activity of honey from Roraima, Brazil. *Food Sci Tech Campinas* 2014;34:69-73.
38. Ghasemia K, Ghasemia Y, Ebrahimzadeh MA. Antioxidant activity, phenol and flavonoid contents of 13 citrus species peels and tissues. *Pak J Pharm Sci* 2009;22:277-81.
39. Alaa S, Abdellah M, Hamadi L. Phytochemical screening, contribution to the study of the anti fungal effect of flavonoids from different parts of *Ziziphus lotus* of south west Algeria. *Asian J Pharm Clin Res* 2017;10:13-6.
40. Rohman A, Riyanto S, Yuniarti N, Saputra WR, Utami R, Mulatsih W. Antioxidant activity, total phenolic, and total flavonoid of extracts and fractions of red fruit (*Pandanus conoideus* Lam). *Int Food Res J* 2010;17:97-106.
41. Har LW, Ismail IS. Antioxidant activity, total phenolics, and total flavonoids of *Syzygium polyanthum* (Wight) Walp leaves. *Int J Med Arom Plants* 2012;2:219-28.
42. Chlopicka J, Pasko P, Gorinstein S, Jedryas A, Zagrodzki P. Total phenolic and total flavonoid content, antioxidant activity and sensory evaluation of pseudocereal breads. *LWT Food Sci Technol* 2012;46:548-55.
43. Ruskin RS, Vasanthakumari B, Citarasu T. *In vitro* activity of various leaf extracts of *Canthium coromandelicum* (Burm F.) Alston. *Asian J Pharm Clin Res* 2017;10:214-8.
44. Gholivand MB, Piryaei M. The antioxidant activity, total phenolics and total flavonoids content of *Bryonia dioica* Jacq. *Biologija* 2012;58:99-105.
45. Lim SM, Loh SP. *In vitro* antioxidant capacities and antidiabetic properties of phenolic extracts from selected citrus peels. *Int Food Res J* 2016;23:211-9.
46. McCue P, Vatter D, Shetty K. Inhibitory effect of clonal oregano extracts against porcine pancreatic amylase *in vitro*. *Asian Phram J Clin Nut* 2004;13:401-8.
47. Akah PA, Uzodinma SU, Okolo CE. Antidiabetic activity of aqueous and methanol extract and fractions of *Gongronema latifolium* (Asclepidaceae) leaves in alloxan diabetic rats. *J Appl Pharm Sci* 2011;1:99-102.
48. McCue P, Kwon YI, Shetty K. Anti-diabetic and antihypertensive potential of sprouted and solid-state bioprocessed soybean. *Asian Pharm J Clin Nut* 2005;14:145-52.
49. Oboh G, Akinyemi AJ, Ademiluyi AO, Adefegha SA. Inhibitory effects of aqueous extract of two varieties of ginger on some key enzymes linked to type-2 diabetes *in vitro*. *J Food Nut Res* 2010;49:14-20.

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