

Original Article

RP-HPLC METHOD DEVELOPMENT AND VALIDATION FOR THE SIMULTANEOUS ESTIMATION OF CLINDAMYCIN PHOSPHATE AND CLOTRIMAZOLE IN PHARMACEUTICAL DOSAGE FORMS

M. SUDHAKAR, K. VIJAYASRI, SRIDHAR SIDDIRAJU¹, MATURI NIRUPAMA^{2,*}

Department of Pharmaceutical Analysis and Quality Assurance²; Department of Pharmaceutical Chemistry¹, Malla Reddy College of Pharmacy, Hyderabad, Telangana, India.
Email: maturi.nirupama@gmail.com

Received: 16 Aug 2014 Revised and Accepted: 15 Sep 2014

ABSTRACT

Objective: The aim of this work was to develop and validate a simple Reverse Phase-High Performance Liquid Chromatography method for the simultaneous estimation of Clindamycin and Clotrimazole in pharmaceutical dosage forms.

Methods: The mobile phase consists of phosphate buffer and Acetonitrile in the ratio of (48:52) with gradient programming, Hypersil BDS (250×4.6 mm, 5μ) column used as stationary phase with a flow rate of 1 ml/min, injection volume 10 μl and the run time was 10 min. Detection wavelength was at 220 nm by using Photo Diode Array detector.

Results: The retention times of Clindamycin and Clotrimazole were found to be 2.2 min and 5.7 min respectively. The method was validated according to ICH guidelines. Validation parameters like accuracy, precision, linearity, range, limit of detection, limit of quantification and robustness all were within the limits. The linearity responses of Clindamycin and Clotrimazole were found to be in the concentration ranges of 25-150 μg/ml and 50-300 μg/ml. The percentage recovery for both drugs was found in the range of 99-100%. The LOD & LOQ values for were found to be 1.29 μg/ml and 3.93 μg/ml and Clotrimazole were found to be 1.31 μg/ml and 3.96 μg/ml, respectively.

Conclusion: The results obtained are accurate and within the limits. Hence this method can be applicable for the estimation of Clindamycin and Clotrimazole in pharmaceutical dosage forms.

Keywords: Clindamycin and Clotrimazole, RP-HPLC, Validation.

INTRODUCTION

Clindamycin is an antibacterial, broad-spectrum antibiotic derived from lincomycin. Clindamycin is used for the topical and systemic treatment. And it is effective as an anti-aerobic and anti-protozoal. May be useful in respiratory tract infections, and also to treat gynecological infection. Chemically Clindamycin is methyl 7-chloro-6,7,8-trideoxy-6-(1-methyl-trans-4-propyl-L-2-pyrrolidine carboxamido) 1-thio-L-threo-α-D-galacto-octopyranoside 2-(dihydrogen phosphate)[3] and the structure shown in fig. 1.

Clotrimazole is an imidazole derivative with a broad spectrum of antimycotic activity. It is used for the local treatment of oropharyngeal candidiasis and vaginal yeast infections, and also used in fungal infections of the skin such as ringworm, athlete's foot, and jock itch. Chemically Clotrimazole is 1-[(2-chlorophenyl) diphenylmethyl]-1H-imidazole[4] and the structure shown in figure-2.

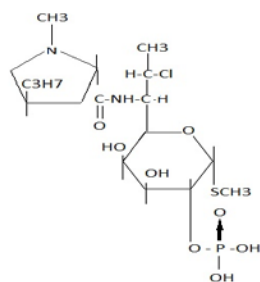


Fig. 1: Structure of Clindamycin phosphate

The literature survey reveals that the few HPLC methods are developed and validated for the estimation of Clindamycin and Clotrimazole combination with other drugs. The reported methods available for the estimation of Clindamycin individually are new

validated bio-analytical high performance liquid chromatography method[5], spectrophotometric method[6], HPLC-UV[7,8], derivative spectrophotometric method[9], RP-HPLC[10-16], liquid chromatography/electro spray ionization mass spectrometry[17], capillary electrophoresis with an end-column electro chemiluminescence[18], gas liquid chromatography[19] and ultra high performance liquid chromatography-electro spray ionization tandem mass spectrometry [20].

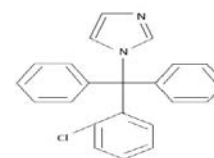


Fig. 2: Structure of Clotrimazole

Clotrimazole has been determined in different pharmaceutical preparations by stability-indicating high performance liquid chromatography method[21], high performance thin layer chromatography[22] and spectrophotometric method[23,24], simultaneous estimation of Clotrimazole combination with other drugs[25-27]. Since there is no reported method on simultaneous estimation of Clindamycin and Clotrimazole in combined tablet dosage forms. The main objective of this study was to develop and validate the assay method of Clindamycin and Clotrimazole in tablet dosage forms.

MATERIALS AND METHODS

Materials

All standard purified materials were used for this study. The solvents which are used in the preparation of solutions should be HPLC grade and obtained from Merck Specialties Private Limited,

Mumbai. Active pharmaceutical ingredients Clindamycin Phosphate and Clotrimazole were supplied by Spectrum Pharma Research Solutions, Hyderabad. The formulation was purchased in a local market.

Instrumentation

The High performance liquid chromatography system consists of water's 2695 with 2996 module Photo Diode Array detectors equipped with a quaternary solvent delivery pump, automatic sample injector and column thermostat. The system was controlled by Empower 2 software and it is used for the analysis.

Chromatographic conditions

The chromatographic separation was performed on Hypersil BDS (250×4.6 mm, 5 μm). The column temperature was maintained at 30°C. The mobile phase consisting of buffer and acetonitrile in the ratio of 48:52 with a flow rate of 1.0 ml/min. The detection wavelength was set at 220 nm and had given acceptable retention time and good resolution in between Clindamycin and Clotrimazole. The run time was taken as 10 min.

Standard solution preparation

Accurately weighed and transferred 10mg of Clindamycin and 20mg of Clotrimazole working standards into a 10 ml clean dry volumetric flask, add 7 ml of diluents, Sonicated for 5 minutes and make up to the final volume with diluents. 1 ml from the above two stock solutions was taken into a 10 ml volumetric flask and made up to 10 ml. The standard solution consists of 100 μg/ml of Clindamycin and 200 μg/ml of Clotrimazole, respectively.

Sample preparation

25 tablets were weighed and calculate the average weight of each tablet than the weight equivalent to 25 tablets was transferred into a 100 ml volumetric flask, 70 ml of diluent added and sonicated for 25 min, further the volume made up with diluents and filtered. From the filtered solution 0.2 ml was pipette out into a 10 ml volumetric flask and made up to 10 ml with diluents.

Preparation of buffer: 0.01N (KH₂PO₄)

Accurately weighed 1.36 gm of Potassium dihydrogen Ortho phosphate in a 1000 ml of volumetric flask add about 900 ml of milli-Q water added and degas to sonicate and finally make up the volume with water and pH adjusted to 3 with dil. OPA.

Method validation

The optimized chromatographic method was validated according to the ICH guidelines for the validation of parameters like linearity, precision, accuracy, limit of detection (LOD), limit of quantification (LOQ) and robustness. Validation of different types of analytical procedures, as defined in the ICH Q2A guideline "Text on validation of analytical procedure."

System suitability parameters

System suitability parameters are validated by injecting five times of 100 μg/ml and 200 μg/ml concentrations of standard solutions of Clindamycin phosphate and Clotrimazole in to the system. The parameters like resolution factor, theoretical plate and tailing factor were calculated.

Linearity

The range of linearity was evaluated by injecting the standard solution of Clindamycin and Clotrimazole in five different concentrations into the HPLC system. The linearity concentrations of Clindamycin and Clotrimazole were prepared in the ranges of 25-150 μg/ml and 50-300 μg/ml. Plot a graph in between the peak area versus concentration (on X-axis and Y-axis). The linear regression coefficient, correlation coefficient was calculated. The correlation coefficient (r^2) should be 0.999.

Precision

Repeatability

Repeatability of Clindamycin and Clotrimazole was validated by injecting the six times of standard solution and sample solutions in

to the system. The areas of all the injections were taken and their corresponding values for mean, standard deviation and %RSD are calculated.

Reproducibility

Reproducibility of the method evaluated by injecting six times of 100mg/ml, 200mg/ml concentrations of standard solutions and sample solution of Clindamycin and Clotrimazole on different days and different analysts or by different instruments. The areas of all injections were taken and their corresponding values for mean, standard deviation and % RSD of reproducibility are calculated.

Accuracy

The accuracy was validated by using a minimum of three different concentrations of standards, Clindamycin and Clotrimazole, 50%, 100% and 150%. The percentage recoveries are analyzed of the obtained amount of Clindamycin and Clotrimazole in pharmaceutical formulations.

Method robustness

The robustness can be determined by varying the following parameters

1) Flow rate: It can be determined by altering the flow rate from 1 ml/min to 1.2 ml/min. The standard solution of Clindamycin and Clotrimazole was prepared and was injected by varying the flow rate along with the optimized method.

2) Column temperature: It can be determined by varying the column temperature ±5%. The standard solution of Clindamycin and Clotrimazole was prepared and was injected by changing organic mobile phase composition along with the optimized method.

3) Mobile phase: It can be determined by changing the organic mobile phase composition by ±10%. The standard solution of Clindamycin and Clotrimazole was prepared and was injected by changing an organic mobile phase composition along with the optimized method.

LOD and LOQ

Limit of detection (LOD) and limit of quantification (LOQ) of Clindamycin and Clotrimazole was calculated from the calibration curve method. The linearity solutions of Clindamycin and Clotrimazole were prepared and injected. LOD and LOQ were calculated by using following equations. $LOD = (3.3\sigma/S)$, $LOQ = (10\sigma/S)$.

Where σ = standard deviation of the response; S = slope of the calibration curve of the analyze.

RESULTS AND DISCUSSION

Method development and optimization

A simple RP-HPLC method for the simultaneous estimation of Clindamycin and Clotrimazole in pharmaceutical dosage forms. In method development the solubility of the active pharmaceutical ingredient was checked in different solvents like methanol and water. The Clindamycin was completely soluble in water and Clotrimazole is methanol soluble. Finally the standards are diluted by methanol and water in the ratio of 80:20.

The different mobile phases like acetonitrile and potassium dihydrogen ortho phosphate buffer were used in different compositions with a flow rate of 1 ml/min but the peak resolution, retention time and tailing factor were not satisfactory. Finally by changing the composition ratio (48:52) of the mobile phase was selected at a flow rate of 1 ml/min.

The chromatographic separation was tested by using different columns like Hybersil, ultima, kromosil (150×4.6 mm; 5 μm) columns maintained at different temperatures like 25,30,35 were used, but the retention time, peak resolution and tailing were not in the desired limits. Actual chromatographic separation was achieved on BDS Hybersil (250×4.6 mm; 5 μm) using mobile phase composition of (48:52) buffer and acetonitrile.

Table 1: System suitability parameters of standard solution

S. No.	Clindamycin				Clotrimazole			
	Rt	Area	USP Plate count	USP tailing	Rt	Area	USP Plate count	USP tailing
1	2.215	751380	2852	1.31	5.638	7168163	4470	1.26
2	2.277	745868	3143	1.27	5.957	7228132	4859	1.25
3	2.286	749940	3099	1.27	6.004	7233293	6242	1.24
4	2.293	758163	3428	1.29	6.004	7233293	6242	1.24
5	2.293	758163	3428	1.29	6.123	7248529	4875	1.22
Mean		752702.8				7222282		
SD		5378.77				31202.8		
%RSD		0.71				0.43		

SD=Standard deviation; RSD= Relative Standard deviation; Rt= Retention Time

System suitability parameters

The system suitability tests were conducted before performing the validation and the parameters were within the acceptance criteria like retention times of Clindamycin and Clotrimazole were 2.2 minutes and 5.7 minutes, respectively. The plate count was >2000, peak tailing was <2 and the %RSD of peak areas of standard were ≤ 2.(Table 1), (fig. 3). Hence the proposed method was successfully applied to routine analysis without any problems.

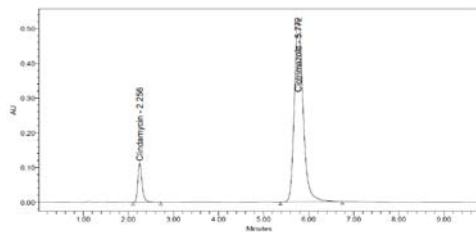


Fig. 3: HPLC chromatograph of standard solution (i. e. Clindamycin and Clotrimazole)

Linearity

The linearity of Clindamycin and Clotrimazole was prepared in the range of 25-150 µg/ml and 50-300µg/ml. These were represented by linear regression equation y (Clindamycin) =7319. x+2881 (r²=0.999), y (Clotrimazole)=34358. x+13610 (r²=0.999). From the calibration curve the regression line for both drugs was linear. (Table 2), (fig. 4,5).

Table 2: Linearity of Clindamycin and Clotrimazole

Clindamycin Clotrimazole			
Concentration (ppm)	Area	Concentration (ppm)	Area
0	0	0	0
25	180013	50	1890194
50	374437	100	3699949
75	557979	150	5280411
100	744271	200	7108478
125	904671	250	8675551
150	1101466	300	10373947

Precision

Injected standard preparation six times in same concentration in to the system. The precision of analytical method expresses closeness of agreement between a series of measurements obtained from multiple sampling of the homogenous under the prescribed

conditions. Reproducibility and Repeatability for Clindamycin and Clotrimazole were shown in table 3. This indicated the method was highly precise.

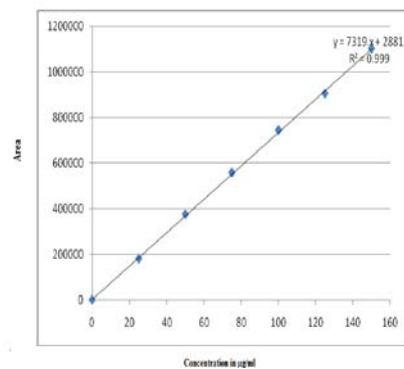


Fig. 4: Linearity graph of Clindamycin

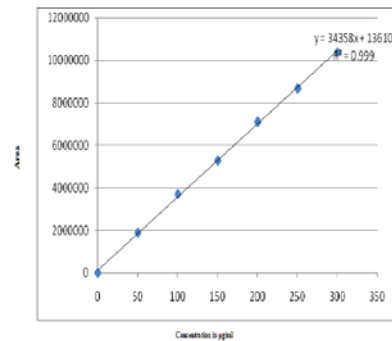


Fig. 5: Linearity graph of Clotrimazole

Accuracy

The percentage recoveries for Clindamycin and Clotrimazole were found to be 99-100% and the %RSD for Clindamycin and Clotrimazole were found to be 0.37 and 0.67. The results of recovery studies were shown in table 4.

Robustness

Robustness data for Clindamycin and Clotrimazole by changing the parameters like flow rate, temperature and mobile phase ratio. It was shown in table 5.

Table 3: Determination of repeatability and intermediate precision for Clindamycin and Clotrimazole

Drug	Repeatability			Intermediate precision		
	Peak area	Std. Dev	%RSD	Peak area	Std. Dev	%RSD
Clindamycin	753768	2130.09	0.28	750846	8073.8	1.1
Clotrimazole	7246639	32443.1	0.44	7174052	75631.9	1.1

Table 4: Accuracy for standard solution

Clindamycin				Clotrimazole			
Standard concentration (µg/ml)	Amount added (µg/ml)	Amount recovered (µg/ml)	%Recovery	Standard concentration (µg/ml)	Amount added (µg/ml)	Amount recovered (µg/ml)	%Recovery
100	50	49.79	100.49	200	100	100.02	98.83
100	50	50.15	100.25	200	100	100.25	99.90
100	50	50.23	100.15	200	100	100.50	100.23
100	100	99.83	99.83	200	200	199.90	100.12
100	100	100.55	100.55	200	200	199.47	98.85
100	100	99.75	99.75	200	200	199.98	99.92
100	150	150.10	100.68	200	250	249.90	100.46
100	150	150.54	100.81	200	250	249.47	100.48
100	150	150.33	100.07	200	250	249.98	100.67
Mean			100.28				99.94
SD			0.371				0.673
% RSD			0.37				0.67

Table 5: Robustness: Flow rate

Parameters		Clindamycin			Clotrimazole		
		Mean Area	Std. Deviation	%RSD	Mean Area	Std. Deviation	%RSD
Flow rate	1 ml/min	868401	19025.2	1.9	8499057	17994.8	1.8
	1.2 ml/min	638079	9864.3	0.1	6322925	9967.9	0.2
Temperature	25°C	285762	4211.0	1.5	6831608	174864.2	1.6
	35°C	703537	266104	1.8	6745830	194296.5	1.2
Mobile phase	-10%	645494	2780.3	0.4	6334026	5732.0	0.1
	+10%	861098	13188.1	1.5	8465523	125860.6	1.5

Table 6: Limit of quantification and Limit of detection

S. No.	Parameters	Clindamycin	Clotrimazole
1	LOD	1.29	1.31
2	LOQ	3.93	3.96

Table 7: Assay of Clindamycin and Clotrimazole

Brand Name	Label Claim	Amount found	% Assay
CANDID-CL	Clindamycin(100mg)	100.04± 0.28	100.04
(glenmark)	Clotrimazole(200mg)	200.26± 0.44	100.13

Limit of detection (LOD) and limit of quantification (LOQ)

The values of LOD and LOQ were calculated by using the slope and y-intercept. The LOD and LOQ values for Clindamycin were found to be 1.29 µg/ml and 3.93µg/ml and Clotrimazole were found to be 1.31 µg/ml and 3.96µg/ml, respectively (Table 6).

Assay

The content of Clindamycin and Clotrimazole in the pharmaceutical dosage forms by using the developed method. The percentage purity of Clindamycin and Clotrimazole were found to be 100.04 % and 100.13% and %RSD values for both Clindamycin and Clotrimazole were within limit of ≤2. (Table 7).

CONCLUSION

The proposed analytical technique of RP-HPLC is simple, accurate and precise method for the simultaneous estimation of Clindamycin and Clotrimazole in pharmaceutical dosage forms has been developed. The method was validated as per ICH guidelines. Statistical analysis proves that method is repeatable, sensitive for the analysis of Clindamycin and Clotrimazole in pharmaceutical dosage forms.

ACKNOWLEDGEMENT

The authors express their attitude to Spectrum Pharma research solutions, Hyderabad. For providing best samples of pure Clindamycin and Clotrimazole.

CONFLICT OF INTERESTS

Declared None

REFERENCES

1. Gurdeep R. Chatwal, Instrumental method of Chemical Analysis, 5th edition, page no: 2.44, Himalaya Publishing House; 2006.
2. Qualitative analysis of drugs in pharmaceutical formulations. PD. Sethi. 3rd edition. pg no; 2008. p. 51-60.
3. <http://www.drugbank.ca/drugs/DB01190>.
4. <http://www.drugbank.ca/drugs/DB00257>.
5. Kareem M. younes, Ehab F Elkady. New validated bio-analytical hplc method for the clindamycin in human plasma. Int J Anal Bio-Anal Chem 2013;3(4):151-9.
6. KS Nataraj, GNV Surya, Narasimha Raju, B Anusha. UV spectrophotometric method development of clindamycin phosphate in dosage forms. Int J Pharm Bio Sci 2013;3(4):164-7.
7. Martina mifsud. A simple HPLC-UV method for the determination of clindamycin in human plasma. J Chem Pharm Res 2014;6(1):696-704.
8. Batzias GC, Delis GA, Koutsoviti-Papadopoulou M. A new HPLC/UV method for the determination of clindamycin in dog blood serum. Pharm Biomed Anal 2004;35(3):545-54.
9. Maliheh Barazandeh Tehrani, Melika Namadchian, Sedigheh Fadaye Vatan, Effat Souri. Derivative spectrophotometric method for the simultaneous estimation of clindamycin phosphate and tretinoin in pharmaceutical dosage forms. DARU J Pharm Sci 2013;1:21-9.
10. Roy. Residue determination of clindamycin phosphate and tretinoin on the surface of manufacturing equipment by RP-HPLC. J Pharm Res 2012;7(5):3665.
11. Ye YR, Bektic E, Buchta R, Houlden R, Hunt B. Simultaneous determination of tretinoin and clindamycin phosphate and

- their degradation products in topical formulations by RP-HPLC. *J Sep Sci* 2004;27(1-2):71-7.
12. Brown LW. High-pressure liquid chromatographic assays for Clindamycin, Clindamycin phosphate. and Clindamycin palmitate. *J Pharm Sci* 1978;67(9):1254-7.
 13. Zhou W, Zhang Y, He J. Determination of content and entrapment efficiency of clindamycin phosphate Liposome. *Sheng Wu Yi Xue Gong Cheng Xue Za Zhi* 2009;26(3):566-8.
 14. Nil Froushzadeh MA, Siadat AH, Baradaran EH, Moradi S. Clindamycin lotion alone versus combination lotion of clindamycin phosphate plus tretinoin versus combination lotion of clindamycin phosphate plus salicylic acid in the topical treatment of mild to moderate acne vulgaris: a randomized control trial. *Indian J Dermatol Venereol Leprol* 2009;75(3):279-8.
 15. Platzer DJ, White BA. Development and validation of a gradient HPLC method for the determination of Clindamycin and related compounds in a novel tablet formulation. *J Pharm Biomed Anal* 2006;41(1):84-8.
 16. Geoffrey K Wu, Abhay Gupta, Mansoor A Khan, Patrick J Faustino. Development and application of a validated HPLC method for the determination of clindamycin palmitate hydrochloride in marketed drug products: an optimization of the current USP methodology for assay. *J Anal Sci Methods Instrum* 2013;3:202-11.
 17. Wang SM. Separation and characterization of clindamycin phosphate and impurities by liquid chromatography electro spray ionization mass spectrometry. *Rapid Commun Mass Spectrom* 2009;23(6):899-906.
 18. Nianjun Yang. Determination of clindamycin by capillary electrophoresis with chemiluminescence of tris (2, 2-bipyridine) ruthenium (II). *J Talanta* 2008;75(3):817-23.
 19. Brown LW. High-pressure liquid chromatographic assays for Clindamycin, Clindamycin phosphate and clindamycin palmitate. *J Pharm Sci* 1978;67(9):1254-7.
 20. Yan L. Simultaneous determination of macrolide and lincosamide antibiotics in animal feeds by ultra-performance liquid chromatography-electro spray ionization tandem mass spectrometry. *Se Pu* 2010;28(11):1038-42.
 21. Jagadish PC Prashanth K, Muddukrishna Krishnamurthy. Development and validation of stability indicating HPLC method for clotrimazole formulation. *Int J Pharm Sci* 2014;6(1):121-9.
 22. Parul Parmar, Ankitha Mehta. Development and validation of HPTLC Method for the estimation of clotrimazole in drug and tablet formulation. *J Pharm Sci* 2009;71(4):451-4.
 23. Amit V Patel, Koamal R Dudhasia, Chhaganbhai N Patel. Development and validation of UV Spectrophotometric method for simultaneous estimation of clotrimazole and beclomethasone dipropionate in their combined dosage form. *Inventi Impact Pharm Anal Qual Assur* 2004;54(1-2):1-13.
 24. Arshad MD, Bharath SA, Venugopal Darak, K Kalyan Chakravathy. Development and validation of simultaneous spectrophotometric estimation of clotrimazole and tinidazole in tablet dosage forms. *IJPT'S J Anal Chem* 2011;2(1):3-17.
 25. Varaprasad Bobbarala, K HussainReddy, Somasekhar P. Determination of dipropionate, clotrimazole, chloramphenicol and lidocaine in pharmaceutical formulations using a novel RP-HPLC method. *Int J Pharm Bio Sci* 2011;2(3):453-62.
 26. Manassra A, Khamis M, el-Dakiky M, Abdel-Qader Z, Al-Rimawi F. Simultaneous HPLC analysis of betamethasone and clotrimazole in cream formulations. *Pharm Anal Acta* 2010;1(2):2153-435.
 27. R Rajameena, K Rama, C Muthulakshmi. Method development and validation for estimation of clindamycin phosphate and Clotrimazole in pharmaceutical dosage forms. *Int Res J Pharm* 2013;4(7):141-6.