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Original Article

INSIGHTS INTO THIORIDAZINE FOR ITS ANTI-TUBERCULAR ACTIVITY FROM MOLECULAR DOCKING STUDIES

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ABSTRACT

Objective: Thioridazine (TZ) is a drug that has been used for over 35 years as a psychoactive drug, is now potentially utilized in combination with certain anti-TB drugs to cure MDR, XDR and TDR TB. The current study explores the plausible reasons for its anti-tubercular activity through molecular docking procedure.

Methods: Molecular docking were performed by using the molecular modeling software Glide® from the suite of Schrödinger Inc., Molecular docking studies were performed to study the binding affinity of Thioridazine on the active sites of various *Mycobacterium tuberculosis* enzymes in an effort to increase the understanding of the action of Thioridazine (TZ) as an antitubercular agent. Seventeen enzymes from different mechanisms were docked and the resulting glide scores (G-Scores) were tabulated.

Results: The enzyme CmaA2 - Cyclopropane mycolic acid synthase (PDB id: 1KPI) scored lowest binding energy which means the greater stability of the Thioridazine's ability to bind to the receptor. MmaA2 (1TPY), InhA (2NSD) and PknG (2PZI) enzymes of *Mycobacterium tuberculosis* gave the best G-scores.

Conclusion: The docking study results revealed that Thioridazine may act by more than one possible mechanism to exert anti-tubercular activity against MDR (Multi Drug Resistant), XDR (Extensively Drug Resistant) and TDR (Totally Drug Resistant) -TB.

Keywords: Thioridazine (TZ), Efflux pumps, Molecular docking, Enzymes, Binding energy, MDR, XDR.

INTRODUCTION

The World Health Organization (WHO) declared TB a global public health emergency in 1993[1]. Tuberculosis (TB) still continues to be a major global health problem. Globally in 2012, an estimated 450, 000 people developed MDR-TB and there were an estimated 170, 000 deaths from MDR-T band also there was reported cases of XDR and TDR-TB. On an average, an estimated 9.6% of MDR-TB cases have converted to XDR-TB [2]. Udwadia ZF et al the clinical physicians of Hinduja Hospital, Mumbai cited the use of Thioridazine (TZ) as salvage therapy in 4 Indian patients with XDR-TB (near total drug resistance to current therapy) with advanced disease. They found the drug to be done well tolerated, even in the malnourished and ill patient population. It also led to clinical improvement in 3 of the 4 patients [3]. Thioridazine, a neuroleptic drug, which is milder and less toxic than chlorpromazine (CPZ), kills intracellular M. tuberculosis isolates that are resistant to two or more antibiotics. Thioridazine shows anti tuberculous effects in-vitro and in-vivo mouse models [4]. Thioridazine is effective when used in combination with antibiotics to which the initial Mycobacterium tuberculosis was resistant. Because Thioridazine is cheap, it should be taken into account in therapy of XDR and TDR-TB patients in economically disadvantaged countries [5, 6]. At the Hinduja Hospital (Mumbai, India), Ethical committee approval was obtained to proceed with a trial involving inclusion of Thioridazine in the treatment regimen on a 'compassionate' basis. Leonard Amaral et al [7] have now made pleas for larger collaborative studies with Thioridazine in patients with XDR-TB who have exhausted all available drug options. Van Ingen J [8] suggests that Thioridazine itself may be patented for its "new use". New drugs for treatment of mycobacterial disease, most notable multidrug- and extensively drug-resistant tuberculosis, are urgently needed; phenothiazines and their targets should be exploited for this use. The current study aims to explore the possible mechanism of action of Thioridazine through molecular docking studies. Though Thioridazine was found to act through efflux pump mechanism, still it was not well explored [9]. Present study tries to throw light on this by molecular docking studies.

MATERIALS AND METHODS

In this work Glide (grid based ligand docking with energy) program [10, 11] was used for molecular docking provided by Schrödinger Inc.,

Protein and ligand preparation

3D Crystal structures of all the enzymes reported in this work were downloaded from the Protein Data Bank (PDB) and the *in-silico* models were subjected to the Protein Preparation Wizard workflow implemented in the Schrödinger package. They were preprocessed by the addition of hydrogen, assigning the bond order, identifying overlaps, creating zero order bonds to metals and creating disulfide bonds. The co-factors, unwanted water molecules and chains were deleted. Then the energy minimization was done followed by the grid generation. Ligand was prepared by a series of steps that includes conversions, generation of variations on the structure, optimization of the structures.

Ligand docking

After the generation of the grid, prepared ligand was docked to identify the interaction between the ligand and the active site of the protein. The interactions were hydrophobic, hydrophilic and Van der Waal's interactions. The strength of ligand interaction differs from enzyme to enzyme based upon affinity. During the docking procedure, conformation of the ligand was retained and extra precision (xp) mode was selected.

Visualization of the docking results

Once the ligand was docked against all the enzymes of interest, they were visualized for interactions, G-score and some other parameters with the use of Glide XP visualiser.

RESULTS AND DISCUSSION

Comparative docking analysis of Thioridazine with various pathophysiological enzymes responsible for tuberculosis is a feasible method to study the Thioridazine potential as antimycobacterial agent. Though, Thioridazine is reported for its efficacy towards tuberculosis in an unusual mechanism, the docking results showed it may act on other mechanisms possibly. The binding efficiency of the drug Thioridazine with 17 pathophysiological enzymes is shown in table 1 and the G-Score has ranged between -3.1 to -9.5. Hydrophobic interaction of Thioridazine when positioned at the target is shown in fig. 1. The non-bonding interactions/hydrophobic region include the key amino acid residues like tyrosine (TYR), isoleucine (ILE), leucine (LEU) and phenylalanine (PHE) in top scored three enzymes against Thioridazine. From the virtual screening results, it is seen that Thioridazine docked well with the enzymes Cyclopropane Mycolic Acid Synthase (lipid metabolism), Methoxy Mycolic Acid Synthase (lipid metabolism), NADH-Dependent Enoyl ACP (Acyl Carrier Protein) Reductase (lipid metabolism), and Protein Kinase-G (Regulatory Protein) with the G-Score of -9.5 Kcal/mol, -9.4 Kcal/mol, -8.4 Kcal/mol and -8.2 Kcal/mol respectively. A new indication for an old antidepressant molecule Thioridazine, against *Mycobacterium tuberculosis* through an unusual mechanism has prompted the study.



Fig. 1: Ligand Interaction diagram



Table 1. Mole	cular docking	results of	Thioridazine	from (lide
Table 1: Mole	culai uocking	lesuits of	I moi iuazine	nome	mue

S. No.	Name of the enzyme (PDB ID)	G-Score(Kcal/mol)	Functional category
1.	CmaA- 2 Cyclopropane Mycolic Acid Synthase[12] (1KPI)	-9.5	Lipid Metabolism
2.	MmaA2 - Methoxy Mycolic Acid Synthase 2[13](1TPY)	-9.4	Lipid Metabolism
3.	InhA - NADH-Dependent Enoyl ACP (Acyl Carrier Protein) Reductase[14] (2NSD)	-8.4	Lipid Metabolism
4.	PknG - Protein Kinase[15, 16] (2PZI)	-8.2	Regulatory Protein
5.	LdtB – L, D-Transpeptidase-2[17] (3VAE)	-6.4	Cell wall and Cell Processes
6.	GlnA1 – Glutamine Synthetase[18] (4ACF)	-6.0	Intermediary Metabolism
			and Respiration
7.	PcaA - Mycolic Acid (Cyclopropane) Synthase[19] (1L1E)	-5.4	Lipid Metabolism
8.	Mmr – Multi-Drug Resistant Protein[20] (2IQ4)	-4.9	Cell wall and Cell Processes
9.	AdoK – Adenosine Kinase[21] (2PKK)	-4.5	Intermediary Metabolism and
			Respiration
10.	Glf - UDP-Galactopyranose Mutase[22] (1V0J))	-4.1	Cell wall and Cell Processes
11.	FabH - (3-Oxo-acyl)-(Acyl Carrier Protein) ACP Synthase[23] (1HZP)	-3.7	Lipid Metabolism
12.	MbtK - Lysine-N(Epsilon)-acyl transferase[24] (1YK3)	-2.2	Lipid Metabolism
13.	FabD - Malonyl CoA- Acyl Carrier Protein transacylase [25](2QC3)	-3.8	Lipid Metabolism
14.	ThyX – Thymidylate Synthase[26] (3GWC)	-2.0	Intermediary Metabolism and
			Respiration
15.	TmK – Thymidylate Kinase [27](1G3U)	-3.8	Intermediary Metabolism and
			Respiration
16.	DprE1 - Decaprenylphosphoryl-B-D-Ribose 2'-Epimerase[28] (4FD0)	-3.8	Lipid Metabolism
17.	EmbC- Arabinosyl-Indolyl Acetyl Inositol Synthase[29] (3PTY)	-3.1	Cell wall and Cell Processes

CONCLUSION

Thioridazine as an antidepressant is reported to act by inhibiting the efflux pump mechanism. Our docking studies indicate that it might be acting by inhibition of several enzymes notably Cyclopropane Mycolic Acid Synthase (lipid metabolism), Methoxy Mycolic Acid Synthase (lipid metabolism), NADH-Dependent Enoyl ACP (Acyl Carrier Protein) Reductase (lipid metabolism), and Protein Kinase-G (Regulatory Protein) with the G-Score of -9.5 Kcal/mol, -9.4 Kcal/mol, -8.4 Kcal/mol and -8.2 Kcal/mol respectively.

Therefore the Anti-TB activity of Thioridazine is not because of Efflux pump mechanism but may be by more than one mechanism possibly. Our computational studies also conclude that the novel approach on Thioridazine derivatives may lead to newer anti-tubercular agents.

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CONFLICT OF INTERESTS

We declare that there is no conflict of interest with any organization regarding the materials discussed in this work.

REFERENCES

- 1. Global tuberculosis report; World Health Organization (WHO), Geneva, Switzerland, Retrieved; 2014. p. 8-10.
- 2. World Health Organization, Global tuberculosis report, Geneva, Switzerland, Retrieved; 2014. p. 8-10.
- Udwadia ZF, Sen T, Pinto LM. Safety and efficacy of Thioridazine as salvage therapy in Indian patients with XDR-TB. Rec Pat Anti-Infect Drug Dis 2011;6(2):88-91.
- Diane Ordway, Miguel Viveiros, Clara Leandro, Rosa'rio Bettencourt, Josefina Almeida, Marta Martins, *et al.* Clinical Concentrations of Thioridazine Kill Intracellular Multidrug-Resistant *Mycobacterium tuberculosis.* J Antimicrob Chemother 2003;47(3):917–22.
- Amaral L, Molnar J. Why and how Thioridazine in combination with antibiotics to which the infective strain is resistant will cure totally drug-resistant tuberculosis. Exp Rev Anti Infect Ther 2012;10:869-73.
- 6. Leonard A, Jette EK, Miguel V, Jorge A. Activity of phenothiazines against antibiotic resistant *Mycobacterium tuberculosis*: a review supporting further studies that may elucidate the potential use of Thioridazine as antituberculosis therapy. J Antimicrob Chemother 2001;47:505-11.
- Leonard A, Martin J, Stephen H, Udwadia ZF, Soolingen DV. Thioridazine cures extensively drug-resistant tuberculosis (XDR-TB) and the need for global trials is now. Int J Antimicrob Agents 2010;35:524–6.
- Van Ingen J. The broad-spectrum antimycobacterial activities of phenothiazines, *in vitro*: somewhere in all of this there may be patentable potentials. Rec Pat Anti infect Drug Dis 2011;6:104–9.
- 9. Dutta NK, Mazumdar K, Dastidar SG, Karakousis PC, Amaral L. New patentable use of an old neuroleptic compound Thioridazine to combat tuberculosis: a gene regulation perspective. Rec Pat Anti infect Drug Dis 2011;6(2):128-38.
- 10. Parvathy NG, Manju P, Mukesh M, Leena T. Design, Synthesis and Molecular docking studies of Benzothiazole derivatives as Anti microbial agents. IJPPS 2013;5(2):101-6.
- 11. Friesner RA, Banks JL, Murphy RB, Halgren TA, Klicic JJ, Mainz DT, *et al.* Glide: A new approach for rapid, accurate docking and scoring. J Med Chem 2004;47(7):1739-49.
- Daniel B, Zhen L, James CS, Michael SG. Mycolic acid cyclopropanation is essential for viability, Drug Resistance, and Cell Wall Integrity of *Mycobacterium tuberculosis*. Chem Biol 2009;16:499–509.
- 13. Michael SG. The mmaA2 Gene of *Mycobacterium tuberculosis* Encodes the Distal Cyclopropane Synthase of the α -Mycolic Acid. J Biol Chem 2003;278(10):7844–9.
- 14. Xin H, Akram A, Paul R, Montellano O. Inhibition of the *Mycobacterium tuberculosis* enoyl acyl carrier protein reductase InhA by arylamides. Bioorg Med Chem 2007;15(21):6649–58.
- 15. Nicole S, Srinivas H, Gabriele K, Philipp M, Rajesh J, Fritz W, *et al.* Structural basis for the specific inhibition of protein kinase

G, a virulence factor of *Mycobacterium tuberculosis*. PNAS 2007;104(29):12151-6.

- Rajesh PK, Rohini K, Srikumar PS. Anti-tuberculosis drugs against *Mycobacterium tuberculosis* PKnB and mutants' 133d/ d76a-A comparative docking study. IJPPS 2014;6(1):662-4.
- 17. Erdemli SB, Gupta R, Bishai WR, Bianchat MA. Targeting the cell wall of *Mycobacterium tuberculosis*: Structure and mechanism of L, D-transpeptidase 2. Struc 2012;20(12):2103-15.
- Nordqvist A, Nilsson MT, Lagerlund O, Muthas D, Gising J, Yahiaoui S, et al. Synthesis, Biological evaluation and x-ray crystallographic studies of imidazo (1, 2-A) pyridine-based Mycobacterium Tuberculosis Glutamine Synthetase Inhibitors. Med Chem Comm 2012;3:620.
- Huang CC, Smith CV, Glickman MS, Jacobs W Jr, Sacchettini JC. Crystal structures of Mycolic acid Cyclopropane synthases from Mycobacterium tuberculosis. J Biol Chem 2002;277(13):11559-69.
- Liliana R, Cristina V, Rebeca B, Miguel V, José AA. Role of the mmr efflux pump in drug resistance in *Mycobacterium tuberculosis.* J Antimicrob Chemother 2013;57(2):751–7.
- William BP, Mary CL, Vincent E. Identification and characterization of a unique adenosine kinase from *Mycobacterium tuberculosis*. J Bact 2003;183(22):6548-55.
- 22. Pan F, Jackson M, Ma Y, McNeil MR. Cell wall core galactofuran synthesis is essential for growth of mycobacteria. J Bact 2001;183(13):3991-8.
- 23. Musayev F, Sachdeva S, Scarsdale JN, Reynolds KA, Wright HT. Crystal structure of a substrate complex of *Mycobacterium tuberculosis*-ketoacyl-acyl carrier protein synthase III (FabH) with lauroyl-coenzyme A. J Mol Biol 2005;346(5):1313-21.
- Qiao C, Gupte A, Bosho HI, Wilson DJ, Bennett EM, Somu RV, et al. O-[(N-Acyl) sulfamoyl] adenosines as antitubercular agents that inhibit mbta: an adenylation enzyme required for siderophore biosynthesis of the mycobactins. J Med Chem 2007;50(24):6080-94.
- 25. Kremer L, Nampoothiri KM, Lesjean S, Dover LG, Graham S, Betts JC, et al. Biochemical characterization of acyl carrier protein (AcpM) and Malonyl-CoA: AcpM Transacylase (mtFabD), Two Major Components of Mycobacterium tuberculosis fatty acid synthase II. J Biol Chem 2001;276:27967-74.
- Sampathkumar P, Turley S, Ulmer JE, Rhie HG, Sibley CH, Hol WG. Structure of *Mycobacterium tuberculosis* flavin dependant Thymidylate Synthase (MtbThyX) at 2.0 A⁰resolution. J Mol Biol 2003;352(5):1091-104.
- Sierra LD, Munier LH, Gliies AM, Barzu O, Delarue M. X-ray Crystal Structure of TMP Kinase from *Mycobacterium tuberculosis* complexed with TMP at 1.95A^oresolution. J Mol Biol 2001;311(1):87-100.
- Vadim M, Giulia M, Katarina M, Ute M. Benzothiazinones Kill *Mycobacterium tuberculosis* by blocking arabinan synthesis. Sci 2009;324(5928):801–4.
- 29. Belanger AE, Besra GS, Ford ME, Mikusova K, Belisle JT, Brennan PJ, *et al.* The embA, B genes of Mycobacterium avium encode an arabinosyl transferase involved in cell wall arabinan biosynthesis that is the target for the antimycobacterial drug ethambutol. Proc Natl Acad Sci 1996;93(21):11919-24.