### International Journal of Pharmacy and Pharmaceutical Sciences

ISSN- 0975-1491

Vol 7, Issue 6, 2015

**Original Article** 

# ISOLATION AND IDENTIFICATION OF POTENTIAL CYTOTOXIC COMPOUND FROM KEMBANG BULAN [TITHONIA DIVERSIFOLIA (HEMSLEY) A GRAY] LEAVES

# MAE SRI HARTATI WAHYUNINGSIH<sup>1</sup>, MAHARDIKA AGUS WIJAYANTI<sup>2</sup>, ARIEF BUDIYANTO<sup>3</sup>, MUHAMMAD HANAFI<sup>4</sup>

<sup>1</sup>Department of Pharmacology and Therapy, <sup>2</sup>Department of Parasitology, <sup>3</sup>Department of Dermatology, Faculty of Medicine, Universitas Gadjah Mada, Indonesia, <sup>4</sup>Research Centre for Chemistry, Indonesian Institute of Sciences (LIPI), Kawasan PUSPIPTEK Serpong, Banten 13510, Indonesia Email: maeshw98@yahoo.com

### Received: 24 Feb 2015 Revised and Accepted: 28 Apr 2015

#### ABSTRACT

**Objective:** The leaves of Kembang Bulan [*Tithonia diversifolia* (Hemsley) A. Gray] are used traditionally to treat various deseases in Indonesia. Initial study showed that chloroform (CHCl<sub>3</sub>) extract of the leaves inhibited the growth of cancer cells *in vitro* on HeLa cells (half maximal inhibitory concentration/IC<sub>50</sub>, 16,61µg/ml), and the cytotoxic compounds appeared present in the petroleum ether (PE) insoluble fraction (IC<sub>50</sub>, 3,078µg/ml) of the CHCl<sub>3</sub> extract. Objective of the present study was to screen cytotoxic activity of its further fractions and isolates.

**Methods:** The PE insoluble fraction was fractionated by vacuum column chromatography to give 5 combined fractions (I-V). Fraction III, containing 3 isolates (A, B, and C) on thin-layer chromatrography (TLC) displays highest cytotoxic activity that is then subjected into preparative TLC.

**Results:** According to cytotoxic bioassay, B isolate was the active one ( $IC_{50}$ , 47.074±4.79 ug/ml), and further purification of B isolate results in 3 isolates (B1, B2 and B3). B2 isolate was tested on several human cancer cell lines, and shows the most cytotoxic *in vitro* on HeLa ( $IC_{50}$ = 9.776±0.77µg/ml) and WiDR cell lines ( $IC_{50}$ , 0.585±0,08 ug/ml).

**Conclusion:** B2 isolate was a major cytotoxic compound and identified as Tagitinin C, based on its spectroscopic data and comparison with the previous reported, data.

Keywords: T. diversifolia, Fractionation, Cytotoxic, WiDR, Tagitinin C.

#### INTRODUCTION

The leaves of Kembang bulan [Tithonia diversifolia (Hemsley) A. Gray] are traditionally used by Indonesia to cure stomachache, diarhea, liver diseases, wounds and to reduce the blood sugar [1-3]. Antimalarial activity of this plant has been reported and also antimicrobial activity of this extract and a germacranolidetype sesquiterpene lactone has been reported from Tithonia diversifolia leaf extract [4,5]. On the other study [6] showed the cytotoxic effect in vitro on adenocarcinoma colon cell line (HCT-116) of MeOH extract of this species. Whereas [7] revealed that this species extract has antiproliferative effect on human colon cancer (Col-2) and can induce human celluler differentiation on human promyelositic leukemia (HL-60) in vitro. Furthermore, initial study has been done by [8] on this species, CHCl<sub>3</sub> extract of the leaves displays cytotoxic effect (IC  $_{50}, 16.61\ \mu\text{g/ml})$  on HeLa cells better than that on methanol extract (IC<sub>50</sub>, 1006.99 µg/ml). This result suggests that cytotoxic compounds are present in the CHCl3 extract. Fractionation of the dried CHCl<sub>3</sub> extract with petroleum ether (PE) is able to separate the active compounds in the PE insoluble fraction (IC<sub>50</sub>, 3.078µg/ml) rather than in the PE soluble fraction (IC  $_{50},\ 325.331\ \mu g/ml).$ Therefore, further study is emphasized on the PE insoluble fraction and here we are reporting isolation of its cytotoxic compound and its effect on several human cancer cell lines.

# MATERIALS AND METHODS

The leaves of *Tithonia diversifolia* were collected from Pakem-Yogyakarta Special District of Indonesia in Februari 2009, identified at the Department of Biology Pharmacy, and voucher specimen (no FA/BF/182/Ident/VIII/09) was deposited in Department of Biology Pharmacy, Faculty of Pharmacy, Universitas Gadjah Mada. *Cervical cancer Cell line* (HeLa) was obtained from Prof. Tatsuo Takeya (NAIST-Japan), whereas, *Burkitt's lymphoma* (Raji), myeloma, and SiHa cell lines (LPPT-UGM), Breast cancer (T47D, MCF7 and EVSA-T), colon cancer (WiDR), and melanoma (M19) cell lines were obtained from Prof. Kees Nooter (Erasmus Medical Center, the Netherlands). UV spectrum was recorded on UV spectrometer (Shimadzu UV-365), IR spectrum on Perkin Elmer Spectrum 1000, and NMR spectra were recorded on Bruker HX 500 (LIPI, Jakarta). 3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) Sigma-Cat. Number: M-6494.

#### Cytotoxic bioassay

#### Stock solution preparation

Sample (1.0 mg) was dissolved in DMSO (100,0  $\mu l)$  (E. Merck, Germany) and then RPMI-1640 medium was added to obtain a stock solution (10 mg/ml).

#### In vitro cells culture

Human cancer cells (HeLa, MCF7, EVSA-T, T47D, WIDR, Raji, Myeloma, SiHa, and M19) were maintained in RPMI-1640 medium (Sigma Chemical Co., USA), supplemented with 10% fetal bovine serum (Gibco Invitrogen,USA), 100  $\mu$ g/ml of Streptomycin (Gibco Invitrogen, USA), 100 unit/ml of penicillin (Gibco Invitrogen, USA), and 2 mM of glutamin in tissue culture flask. The cells were incubated in 5% of CO<sub>2</sub> incubator set at 37°C [9].

#### Cytotoxic assay

Several human cancer cell lines were cultured in RPMI 1640 supplemented with 10% heat inactivated fetal bovine serum (FBS). Cultures were maintained in a humidified incubator at 37 °C in an atmosphere of 5% CO2. A hundred  $\mu$ l of media containing 2x10<sup>3</sup>cells was added to 96-well plate and incubated for 2 hours then samples at various concentrations were added. Following 24 hours of incubation, cells were gently washed with 1X PBS, and 100  $\mu$ l of MTT 0.5 mg/ml was added to the well. The cells were incubated for 4 hours at 37 °C and the reaction was stopped by adding 100  $\mu$ l SDS 10%. Each sample was repeated three times. The plates were incubated overnight and read in the microplate reader (Bio-Rad) at 595 nm. Data generated were used to plot a dose-response curve of which the concentration of samples required to kill 50% of cell population (IC<sub>50</sub>) was determined.

Isolation of cytotoxic active compound was done according to Bioassay Guided Isolation method (fig. 1). Each extract, fractions, or compounds obtained were monitored by cytotoxic assay.

# Extraction of *T. diversifolia*'s leaves

Dried-powdered leaves (500 g) were macerated (24 hours, room temperature) with  $CHCl_3$  (1 L). Maceration was done 3 times, each was filtered in vacuo. The combined filtrates was evaporated by rotary evaporator to give sticky  $CHCl_3$  extract (28.5 g) (A).

#### Isolation and identification of cytotoxic compound

The CHCl<sub>3</sub> extract (A) was triturated with petroleum ether (PE) to give PE soluble (A1) and PE insoluble (6,8 g). (A2) fractions. The latter that displayed cytotoxic activity was fractionated by vaccum liquid column (VLC) chromatography (SiO<sub>2</sub>, n-hexane–EtOAc with increasing amount of EtOAc) results in 5 combined fractions (I-V). Cytotoxic active compound was isolated from fraction III (0,91 g) that displayed the best cytotoxic activity among the other fractions by preparative TLC (SiO<sub>2</sub> GF<sub>254</sub> prep. grade, n-heksane: EtOAc 1:1 v/v) to give 2 isolates (**1** and **2**) that were separated. These 2 compounds were tested for their cytotoxic activity. Compound **1**, the cytotoxic active compound was identified according its spectroscopic (UV, IR, <sup>13</sup>C-and <sup>1</sup>H-NMR) data and comparison with data in the literatures.



Fig. 1: Flow chart of bioassay guided for cytotoxic compound isolation from the leaves of *T. diversifolia* 

### **RESULTS AND DISCUSSION**

The leaves of *Tithonia diversifolia* (Hemsley), A. Gray., localy known as Kembang bulan (Fam. Asteraceae) have been used extensively by Indonesian to reduce blood sugar, but most of the published data shows that *T. diversifolia* contains potential cytotoxic compounds that are potential to be developed for future anticancer drug. Our previous study on this species indicates that those cytotoxic compounds are present in the non polar fraction [8], that can be localized by trituring with PE, appeared in the PE insoluble fraction rather than that of PE soluble fraction (IC<sub>50</sub>, 3.078µg/ml v/s 325.331 µg/ml). Upon fractionation of this active fraction in the VLC, 5 combined extracts are obtained (fr. I–V). Fraction III, containing 3 isolates (A, B, C) on TLC displays highest cytotoxic activity that are then subjected into preparative TLC.

The cytotoxic activity is best performed by B isolate with an average of  $IC_{50}$  is 47.047±4.79 µg/ml followed by C isolate ( $IC_{50}$ , 136.579±7.47 µg/ml) and A isolate ( $IC_{50}$ , 146.886±8.29 µg/ml) (fig. 2).



Fig. 2: Cytotoxic (+/-SD) of  $IC_{50}$  values of A, B, and C isolates on HeLa cells

The cytotoxic effect difference is understandable as their TLC profile (at similar TLC system) is quite different, then their activity values are predicted to be different. According to TLC figure, B isolate is suspected to be a mixture of compounds, so further purification was done by preparative TLC to give 3 compounds (B1, B2 and B3). B1 isolate is unrecovered due to its limited amount, whereas B2 isolate and B3 is recovered as the major compounds.

Based on the cytotoxic assay, B2 isolate is the most active compound among others ( $IC_{50}$ ,  $9.776\pm0.98\mu g/ml$ ) even though shows lower cytotoxic effect than that of Doxorubicine (positive control) ( $IC_{50}$ ,  $1.046\pm0.18 \mu g/ml$ ) (fig. 3).



Fig. 3: Cytotoxic (+/-SD) of  $IC_{50}$  values of B1, B2, B3 isolates compared to that of Doxorubicine as positive control on HeLa cells *in vitro* 

The B2 isolate displays IC50 higher than that of clinically used Doxorubicine on HeLa cell lines; however it may be different when it is tested on the different human cancer cell lines. Therefore, further cancer cells selectivity were applied. The result shows that B2 isolate turns out to be selective for its cytotoxicity effect on other human cancer cell lines that is shown by IC 50 value on HeLa cells line that is much higher than that of the other tested human cancer cell lines. Cytotoxic property of B2 isolate was further evaluated on several human cancer cell lines (WIDR, Myeloma, Raji, MCF7, T47D, M19, and EVSA-T) in vitro. In this study, sensitivity of B2 isolate on certain kind of human cancer is identified. Among those human cancer cell lines tested, B2 isolate is most sensitive on WiDR that is shown by the  $IC_{50}$  value on this cell cancer with average of  $IC_{50}$  is 0.585±0.08µg/ml, and the second sensitive cancer cells lines is M19 (IC  $_{50}$ , 0.996±0.39µg/ml) and the rests (myeloma, Raji, MCF7, T47D and AVSA-T) are on the range of 1.7-4.6 µg/ml although these values are still appreciable below IC<sub>50</sub> of HeLa cell lines (fig.4).

Other studies using the same cell lines is T47D and MCF-7 on 1.3.11 fungi isolated from the fruit of "Tanaman Buah Makassar" shows the results of IC50 31 ug/ml against MCF-7 cells, and not in T47D cells. It is indicated that the IC<sub>50</sub> value of B2 isolates still below then IC<sub>50</sub> of 1.3.11 fungi isolated from the fruit of "Tanaman Buah Makassar" [10].



Fig. 4: The cytotoxic (+/-SD) of IC<sub>50</sub> values of B2 isolate on several human cancer cells

The B2 isolate appeared as white amorphous powder, showing maximum UV<sub>MeOH</sub> absorption at  $\lambda$  241 nm, indicating the present of non-conjugated of unsaturated carbonyl. IR spectra (KBr, cm<sup>-1</sup>) of B2 isolate displays characteristic–OH band (3456),-CH<sub>2</sub> (2924), 2 ester–C=O absorption bands (1766 and 1735) and 1 unsaturated–C=O group (1658). The present of ester groups was further shown by a characteristic absorption band of–C-O-C-(1512, 1458 and 1373) [11, 12] (fig. 5).



Fig. 5: IR spectrum (KBr, cm<sup>-1</sup>) of B2 isolate

The <sup>13</sup>C-NMR (CDCl<sub>3</sub>-TMS, 125 MHz) spectra of B2 isolate and table 1 shows the presence of 19 carbons, and there are some characteristic carbonyl (–C=O) as ketone at  $\delta$  196.8 ppm (C-3), ester at  $\delta$  176.2 (C-1') and lactone at  $\delta$  169.8 (C-12). It is a clue present in the <sup>13</sup>C-NMR spectra, 2 ketone ester–C=O signals present in the spectra but the absence of alcoholic type is not recognized in the spectra data therefore the present of a lactone functional group and methine from the main structure skeleton are possible. A pre assumption then can be drawn that B2 isolate is a sesquiterpene having an ester and a lactone functional groups, specifically to be an ester of a sesquiterpen lactone

The <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 500 MHz) spectra of B2 isolate (fig. 6) determines the absolute structure of compound B2. There are some olefinic signals in the region of  $\delta$  6.93 (H-1,d) and 6.23 (H-2,d) are not characteristic conjugated double bounds but trans olefinic with each have / value 16,8 Hz; that is paralel to that information data of UV spectra. Instead, a terminal in the spectra  $\delta$ , 5.80 (H-13a, d, J= 1.6Hz) and 6.36 (H-13b, J=d, 1.6Hz) that is a bit downfield due to an electron withdrawing group (-C=O) causing deshielded signals of protons in the =CH<sub>2</sub>. There spesific signals of methyl protons at  $\delta$ 1.04 (H-3', d, J 6.9 Hz) and 1.05 (H-4', d, J, 6,9 Hz) and methine proton at  $\delta$ , 2,44 (H-2',m, J=6,9Hz) indicated that the presence of isopropyl functional group. In the spectra, <sup>1</sup>H-NMR also indicated the presence of two methyl singlets at d 1.53 (H-14) and 1.95 (H-15) ppm. Two carbinil proton (-CH-O-) on δ, 5,39 (broad) dan 5,34 ppm can be identified as first order splitting pattern with nighboring protons. In addition, a broad signal  $\delta$ , 2,45 ppm is identified as an-OH group (table 1).



Fig. 6: 1H-NMR (CDCl3-TMS, 500 MHz) spectra of B2 isolate

Table 1: Chemical shift of B2 isolate based on the data <sup>1</sup> H-NMR (CDCl<sub>3</sub>, 500 MHz) and <sup>13</sup> C-NMR (CDCl<sub>3</sub>-TMS, 125 MHz)

C-H number	[13]C-NMR (δ, ppm)	<sup>1</sup> H-NMR (δ, ppm), J (Hz)
1	129,8	6.93 (d. 16.8)
2	160.1	6.23 (d.16.8)
3	196.8	-
4	139.1	-
5	137.2	5.84 (d. 9.3)
6	76.1	5.39 (d.8.4)
7	47.2	3.54 (br. s)
8	74.0	5.34 (m)
9	48.6	1.98 (dd 10.14); 2.46(m)
10	72.0	-
11	136.1	-
12	169.8	-
13	124.7	5.80 (d 1.6); 6.36 (d 1.6)
14	29.2	1.53 (s)
15	19.8	1.95 (d 1.2)
1'	176.2	
2'	34.0	2.44 (m 6.9)
3'	18.6	1.04 (d 6,9)
4'	18.8	1.05 (d 6,9)
10-0H		2.45 (br. s)

Based on the spectra data (UV, IR and NMR) and comparison with reported literature data [13], B2 isolate is identified as Tagitinin C (fig. 6). There are 3  $\alpha$ - $\beta$  unsaturated-C=O groups in the Tagitinin C skeleton. It is believed that nucleophilic attack occurs at the empty electron carbon (Michael additon).



Fig. 7: Structure of Tagitinin C

Nitrogen atom that is dominant in the protein and receptor as the binding site, possesses a pair of free electrons that is capable to react

according to michael addition reaction, causes abnormalities in the cancer cells and kill the cells. Unfortunately, normal cells also intoxicated by this type of compounds, then this type of compounds should be the point of attention in developing cancer drugs.

#### CONCLUSION

In conclusion, this study demonstrates that B2 isolate is a major cytotoxic compound from the leaves of *T. diversifolia* and is identified as Tagitinin C on the basis of spectroscopic data and comparison with literature data. Tagitinin C is the most sensitive on colon cancer (WiDR,  $IC_{50}$ = 0,585±0.08 ug/ml).

#### ACKNOWLEDGEMENT

The Authors are grateful to staff of Research Center for Chemistry, Indonesian Institute of Science, for <sup>1</sup>H NMR and <sup>13</sup>C NMR measurements and Hibah Risbin Iptekdok 2010 for providing the grant for the work.

# **CONFLICT OF INTERESTS**

# Declared None

#### REFERENCES

- 1. Moronkola DO, Ogunwade IA, Walker TM, Setzer WN, Oyewole IO. Identification of the main volatile compounds in the leaf and flower of Tithonia diversifolia (Hemsl) Gray. J Nat Med 2006;61:63-6.
- 2. Tona L, Kambu K, Mesia K, Cimanga K, Apers S, De Brynne T, *et al.* Biological screening of traditional preparation from some medicinal plants used as antidiarrhoeal in Kinshsa Kongo. Phytomedicine 1999;6(1):59-66.
- 3. Miura T, Furuta K, Yasuda A, Iwamoto N, Kato M, Ishihara E, *et al*. Antidiabetic effect of nitobegiku in KK-Ay diabetic mice. Am J Chin Med 2002;30(1):81–6.
- 4. Madureira MC, Martins AP, Gomes M, Paiva J, Cunha AP, Rosario V. Antimalarial activity of medicinal plants used in

traditional medicine in S. Tome and Principe islands. J Ethnopharmacol 2002;81:23–9.

- Obafemi CA, Sulaimon TO, Akinpelu DA, Olugbade TA. Antimicrobial activity of extract and a germacranolidetype sesquiterpene lactone from Tithonia diversifolia leaf extract. Afr J Biotechnol 2006;5(12):1254-8.
- Goffin E, Ziemons E, De Mol P, de Madureira Mdo C, Martins AP, da Cunha AP, et al. In vitro antiplasmodial activity of Tithonia diversifolia and identification of its main active constituent: Tagitinin C. Planta Med 2002;68(6):543-5.
- Gu JQ, Gills JJ, Park EJ, Mata-Greenwood E, Hawthorne ME, Axelrod F, *et al.* Sesquiterpenoids from Tithonia diversifolia with potential care chemo preventive activity. J Nat Prod 2002;65(4):532-6.
- Wahyuningsih MSH, Syarif RA, Suharmi S, Murini T, Saputra F, Adiguno Suryo W. Selectivity of purified extract from the leaves of tithonia diversifolia (Hemsley) A. gray against hela cells. Trad Med J 2013;18(1):22-8.
- 9. Boersma AWM, Nooter K, Oostrum RG, Stoter G. Quantification of apoptotic cells with Fluorescein Isothiocyanate-labeled Annexin V in chinese hamster ovary cell cultures treated with cisplatin. Cytometry 1996;24:123-30.
- Kumala S, Septiana EP, Meiyanto E. Cytotoxic effect of secondary metabolites produced by endopytic fungi 1.3.11, 1.1.6 and 1.2.6 isolated from the fruit of "Tanaman Buah Makassar" (Brucea Janvanica (L.) Merr) on *in vitro* T47D and MCF7 Intact Cells and Identification of The Fungus 1.3.11 by Ribosomal DNA Sequence Analysis. IJPPS 2010;2(2):80-4.
- 11. Silverstein RM, Bassler GČ, Morrill TC. Spectrometric identification of organic compounds, fifth ed. John Wiley, Sons, Inc, New York; 1991.
- 12. Dyer JR. Applications of absorption spectroscopy of organic compounds. Prentice Hall, Inc, Englewood Cliffs, New Jersey; 1965.
- 13. Ragasa CY, Tepora MM, Rideout JA. Terpenoid from Tithonia diversifolia. J Res Sci Comput Eng 2009;4(1):1-7.